BRIEF REPORT



Rate of Lung Function Decline in People with Cystic Fibrosis Having a Residual Function Gene Mutation

Gregory S. Sawicki · Michael W. Konstan · Edward F. McKone · Richard B. Moss · Barry Lubarsky · Ellison Suthoff · Stefanie J. Millar · David J. Pasta · Nicole Mayer-Hamblett · Christopher H. Goss · Wayne J. Morgan · Margaret E. Duncan · Yoojung Yang

Received: September 12, 2022 / Accepted: October 7, 2022 / Published online: November 1, 2022 \odot The Author(s) 2022

ABSTRACT

Introduction: Cystic fibrosis (CF) is an autosomal recessive disease caused by mutations in the CF transmembrane conductance regulator (*CFTR*) gene. Approximately 5% of people with CF have residual function (RF) *CFTR* mutations that result in partially retained CFTR activity.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s41030-022-00202-y.

G. S. Sawicki (⊠) Boston Children's Hospital, Harvard Medical School, 300 Longwood Avenue, 4th Floor, Boston, MA 02115, USA e-mail: gregory.sawicki@childrens.harvard.edu

M. W. Konstan Case Western Reserve University School of Medicine, Cleveland, OH, USA

M. W. Konstan Rainbow Babies and Children's Hospital, Cleveland, OH, USA

E. F. McKone St Vincent's University Hospital, Dublin, Ireland

R. B. Moss Stanford University, Palo Alto, CA, USA

B. Lubarsky \cdot E. Suthoff \cdot M. E. Duncan \cdot Y. Yang Vertex Pharmaceuticals Incorporated, Boston, MA, USA

Published literature on disease trajectory among those with RF mutations is limited. In this retrospective study, we characterized lung function decline across different age groups in CFTR modulator-untreated people with CF heterozygous for *F508del* and an RF mutation (*F*/RF). *Methods*: Rate of decline in percent predicted

forced expiratory volume in 1 s (ppFEV₁) was analyzed using data from the US CF Foundation Patient Registry (2006–2014) in *F*/RF (all), *F*/RF (excluding *R117H*), and *F508del* homozygous (*F/F*) cohorts. Annual rates of ppFEV₁ decline were estimated over 2-year periods based on

S. J. Millar · D. J. Pasta Formerly of ICON Clinical Research, North Wales, PA, USA

N. Mayer-Hamblett Seattle Children's Hospital, Seattle, WA, USA

N. Mayer-Hamblett · C. H. Goss University of Washington, Seattle, WA, USA

W. J. Morgan University of Arizona, Tucson, AZ, USA calendar year. Subgroup analyses by age [6–12 (children), 13–17 (adolescents), 18–24 (young adults), and \geq 25 years (adults)] were performed.

Results: The estimated annualized rate of $ppFEV_1$ decline was -0.70 percentage points per year (95% CI -1.09, -0.30) in the *F*/RF (all) cohort (N = 1242) versus -1.91 percentage points per year (95% CI -2.01, -1.80) in the *F/F* cohort (N = 11,916) [difference, 1.29 percentage points per year (95% CI 0.88, 1.70); P < 0.001]. In the F/RF (all) cohort, all age groups demonstrated lung function decline ranging from -0.30 to -1.38. In the F/RF (excluding R117H) cohort, the rate of decline was -1.05 percentage points per year (95% CI -1.51, -0.60) [difference versus F/F cohort, 0.95 percentage points per year (95% CI 0.48, 1.41; *P* < 0.001); not statistically significant in children and young adults].

Conclusion: Progressive lung function decline was observed in people with F/RF genotypes across all assessed age groups, reinforcing the importance of early intervention and clinical monitoring to preserve lung function in all people with CF.

PLAIN LANGUAGE SUMMARY

In people with cystic fibrosis, lung function typically decreases over time and is linked to the severity of the disease. How fast lung function decreases (referred to as the rate of lung function decline) in cystic fibrosis depends on the specific mutations (changes) in the CFTR gene (which causes the disease). Lung function decline has been well studied in some mutation groups, but not many previous studies have looked at lung function decline in people with one copy of the F508del-CFTR mutation (which is the most common CFTR mutation and results in little to no functional CFTR protein) and another CFTR mutation called a residual function mutation (referred to as people with F/RFgenotypes). We used data from the US Cystic Fibrosis Foundation Patient Registry (which collects information on the health of people in the USA who have cystic fibrosis), to look at the rate of lung function decline in people with F/RF genotypes. We found that people with cystic fibrosis who have F/RF genotypes experience lung function loss over time. We also found that this lung function loss occurred in people of all ages with F/RF genotypes. This finding supports the importance of early treatment to help prevent lung function loss in all people with cystic fibrosis, including people with F/RF genotypes.

Keywords: Cystic fibrosis; *F508del*; Lung function; Lung function decline; *R117H*; Residual function

Key Summary Points

Although approximately 5% of people with cystic fibrosis (CF) have residual function (RF) *CFTR* mutations, published data on disease trajectory in this population is limited

This retrospective study characterized lung function decline in CFTR modulatoruntreated patients who were heterozygous for *F508del-CFTR* and an RF mutation (*F*/RF genotypes) compared with patients who were homozygous for *F508del-CFTR* (*F*/F genotype)

Lung function decline was observed in all age groups of patients with *F*/RF genotypes

Findings reinforce the importance of early intervention to preserve lung function in all people with CF, including those with *F*/RF genotypes

INTRODUCTION

Cystic fibrosis (CF) is a progressive, life-shortening, multisystem, autosomal recessive disease affecting > 80,000 people worldwide [1]. CF is caused by mutations in the CF transmembrane conductance regulator (*CFTR*) gene that result in reduced quantity and/or function of CFTR protein [1, 2]. Loss of chloride transport activity can lead to various clinical manifestations, including mucus accumulation in the airways, pulmonary exacerbations, reduced lung function, exocrine pancreatic insufficiency, intestinal malabsorption, reproductive dysfunction, and elevated sweat chloride concentration [1]. Disease severity and rate of progression vary in people with CF and may be determined in part by the extent of chloride transport loss associated with each mutation [3–6]. Lung function and its rate of decline are important clinical outcomes in CF [5, 7].

Approximately 5% of people with CF have residual function (RF) CFTR mutations that exhibit residual CFTR-mediated ion transport due to partially retained CFTR activity [6, 8]. Those with RF mutations are most frequently heterozygous with an F508del-CFTR mutation (F/RF genotypes) [9], although RF mutations may be found in combination with other mutations. F/RF genotypes can result in a range of molecular defects, including reduced or variable synthesis of CFTR channels, moderate defects in CFTR processing and trafficking, impaired channel gating, and altered channel conductance [10–12]. These defects result in a reduced quantity of normal CFTR channels at the cell surface or a normal quantity of CFTR channels at the cell surface that exhibit reduced chloride ion transport ability [8, 13–15]. Compared with people with CF who are homozygous for F508del-CFTR (F/F genotype), those with *F*/RF genotypes—including those with the most common RF mutation, R117H-may have slower disease progression and are more likely to be pancreatic sufficient and have lower sweat chloride concentrations, indicative of partially maintained CFTR activity [10, 16, 17]. Nevertheless, people with CF who have F/RF genotypes have been shown to develop progressive lung disease with age and die prematurely [6].

As published literature on the disease trajectory in people with CF with RF mutations is limited, we sought to characterize lung function decline in a CFTR modulator-untreated cohort of people with CF with F/RF genotypes compared with that in people with the F/F

genotype. Here, we report the findings from a retrospective study of longitudinal registry data evaluating the rate of percent predicted forced expiratory volume in 1 s (ppFEV₁) decline in people with CF with *F*/RF genotypes.

METHODS

Analysis Population

Data from participants in the US Cystic Fibrosis Foundation Patient Registry (CFFPR) [18] from 2006 to 2014 were used to assess lung function in people with CF with *F*/RF and *F*/F genotypes, none of whom were receiving CFTR modulator therapy during this study period. RF mutations were identified based on clinical or in vitro evidence of residual ion transport. RF mutations are listed in the Supplementary Material (Table S1). Individuals were excluded if they had presumed CF-related metabolic syndrome, in which an R117H mutation was detected by newborn screening and sweat chloride level was < 60 mmol/L or no sweat test result was recorded. Clinic encounters recorded in the 2006–2014 period were censored at death, lung transplant, ivacaftor treatment initiation, or last encounter, whichever occurred first. The study included individuals who had a qualifying genotype, were aged 6–45 years, and had ≥ 3 $ppFEV_1$ measurements spanning > 0.5 years in a randomly chosen two-calendar year period beginning at the first ppFEV₁ measurement in the calendar year.

Analysis was performed in three subpopulations: (1) *F/F*; (2) *F/*RF (all), comprising participants with *F508del* on one allele and any RF mutation on the second allele; and (3) *F/*RF (excluding *R117H*), comprising participants with *F508del* on one allele and any RF mutation except *R117H* on the second allele. In each subpopulation, the following age groups were analyzed: 6–12 (children), 13–17 (adolescents), 18–24 (young adults), and \geq 25 years (adults). Inclusion age in the study was between 6 and 45 years to ensure balanced proportions of participants with *F/*RF and *F/F* genotypes in each age group.

Data Analysis

Initially, the 2006–2014 period was divided into overlapping two-calendar year periods (i.e., 2006–2007, 2007–2008 ... 2013–2014). We identified analyzable periods in which individuals had \geq 3 ppFEV₁ measurements spanning \geq 0.5 years in a randomly chosen 2-year period beginning at the first ppFEV₁ measurement in the calendar year. Age was defined as age at the first encounter (i.e., baseline) in each of these randomly chosen 2-year periods.

Demographic and baseline clinical characteristics were descriptively summarized for each subpopulation. The F/F subpopulation was compared with the other two subpopulations using the *t*-test for continuous measurements and chi-square test for discrete measurements.

The intercept and slope (rate of change) of ppFEV₁, based on 2012 Global Lung Initiative equations [19], were estimated over 2-year periods using all available measurements. Two repeated-measures models were developed: one comparing F/F with F/RF (all) and one comparing F/F with F/RF (excluding R117H). The dependent variable was ppFEV₁, with mutation group, age group, calendar year, and time (years since baseline) as fixed effects. All interaction terms, except those with both age group and calendar year, were included. Compound symmetry was specified as the covariance structure to account for correlated data within each individual. The intercept and slope estimates were calculated overall and by each age group from these models. The proportion of participants in each age group was used to calculate the overall values.

Sensitivity analyses using Wang–Hankinson equations [20, 21] for $ppFEV_1$ were also performed.

All statistical tests were two sided. *P*-values ≤ 0.05 were considered statistically significant. The analysis was performed using SAS software version 9.4 (SAS Institute Inc., Cary, NC, USA).

Compliance with Ethical Guidelines

This study was conducted in accordance with the current Good Clinical Practice Guidelines as described by the International Council for Harmonisation, the Declaration of Helsinki, and all applicable local regulations. People with CF provided informed consent to participate in the US Cystic Fibrosis Foundation Patient Registry and to have their data included in the registry. The study did not involve primary data collection and used only de-identified data; thus, institutional review board approval was not required.

RESULTS

Participants and Baseline Characteristics

A total of 1242 participants with *F*/RF genotypes and 11,916 with the F/F genotype were included in the analysis. The most common RF mutawere tions R117H [*n* = 353 (28.4%)]. $3849 + 10 \text{KBC} \rightarrow T [n = 228 (18.4\%)], \text{ and}$ $2789 + 5G \rightarrow A [n = 206 (16.6\%)]$ (Supplementary Material, Table S1). The mean (SD) baseline ppFEV₁ was 80.4 (24.8), 78.6 (25.3), and 73.4 (26.5) percentage points (Table 1), respectively, in the F/RF (all), F/RF (excluding R117H), and F/F cohorts. Staphylococcus aureus, Pseudomonas aeruginosa, Haemophilus influenzae, Aspergillus, Stenotrophomonas maltophilia, and Burkholderia cepacia were detected in all genotype groups at baseline (Table 1). The F/RF (all) cohort had significantly lower rates of *P. aeruginosa* (P < 0.001), Aspergillus (P < 0.001), B. cepacia (P < 0.001), S. maltophilia (P < 0.01), and S. au*reus* (P < 0.05) than the *F*/*F* cohort, whereas rates were more similar between the F/RF (excluding *R117H*) and *F/F* cohorts (Table 1).

Lung Function

The *F*/RF (all) cohort exhibited progressive lung function loss, with an estimated annualized ppFEV₁ rate of decline of -0.70 percentage points per year (95% CI -1.09, -0.30). When

Characteristic	F/ F (n = 11,916)	F/RF (all) ($n = 1242$)	P-value ^a	<i>F</i> /RF (excluding <i>R117H</i>) (<i>n</i> = 889)	<i>P-</i> value ^b	
Age, years, mean (SD)	18.0 (9.6)	23.0 (12.1)	< 0.001	23.6 (11.8)	< 0.001	
Female, n (%)	5698 (47.8)	645 (51.9)	0.006	473 (53.2)	0.002	
Height-for-age <i>z</i> -score, mean (SD)	-0.49 (1.03)	0.04 (1.03)	< 0.001	-0.01 (1.02)	< 0.001	
Weight-for-age <i>z</i> -score, mean (SD)	-0.48 (1.11)	0.36 (1.14)	< 0.001	0.29 (1.15)	< 0.001	
BMI-for-age <i>z</i> -score, mean (SD)	-0.29 (1.08)	0.36 (1.09)	< 0.001	0.29 (1.10)	< 0.001	
ppFEV ₁ at baseline visit, %, mean (SD)	73.4 (26.5)	80.4 (24.8)	< 0.001	78.6 (25.3)	< 0.001	
Patients with positive microbio	blogy $n \ (\%)^{c}$					
Staphylococcus aureus	5386 (45.2)	524 (42.2)	0.042	402 (45.2)	0.99	
Pseudomonas aeruginosa	4131 (34.7)	277 (22.3)	< 0.001	234 (26.3)	< 0.001	
Haemophilus influenzae	662 (5.6)	69 (5.6)	1.00	49 (5.5)	0.96	
Aspergillus species	688 (5.8)	42 (3.4)	< 0.001	32 (3.6)	0.007	
Stenotrophomonas maltophilia	677 (5.7)	48 (3.9)	0.008	38 (4.3)	0.078	
Burkholderia cepacia	237 (2.0)	7 (0.6)	< 0.001	7 (0.8)	0.012	

Table 1 Demographic and Clinical Characteristics, by Mutation

BMI body mass index, *F F508del, ppFEV*₁ percent predicted forced expiratory volume in 1 s, *RF* residual function ${}^{a}F/F$ versus *F*/RF (all)

^b*F/F* versus *F*/RF (excluding *R117H*)

^cAt first visit in the 2-year period

participants with an R117H mutation were excluded, the rate of decline was -1.05 percentage points per year (95% CI -1.51, -0.60). The rate of decline in the F/F cohort was -1.91percentage points per year (95%) CI -2.01, -1.80). The difference in the annual rate of decline between the F/RF (all) cohort and the F/F cohort was 1.29 percentage points per year (95% CI 0.88, 1.70; *P* < 0.001; Fig. 1), and the difference between the F/RF (excluding R117H) cohort and the F/F cohort was 0.95 percentage points per vear (95% CI 0.48, 1.41; P < 0.001).

The *F*/RF genotype cohorts [*F*/RF (all) and *F*/RF (excluding *R117H*)] demonstrated lung function decline in all age groups (Fig. 2). The rates of decline in children with *F*/RF (all) and *F*/RF (excluding *R117H*) genotypes were -0.30 (95% CI -0.93, 0.34) and -0.80 (95% CI

-1.58, -0.02) percentage points per year, respectively (Fig. 2). These were similar to the rates of lung function decline in adolescents [F/RF (all), -0.30 (95% CI -1.05, 0.45) percentage points per year; F/RF (excluding *R117H*), -0.57 (95% CI -1.41, 0.28) percentage points per year] (Fig. 2). In young adults and adults, the rates of lung function decline were greater in magnitude, with rates of -1.38(95% CI -2.14, -0.61) and -0.94 (95% CI -1.45, -0.44) percentage points per year in the cohort and -1.85 F/RF (all) (95%) CI -2.69, -1.01) and -1.06(95% CI -1.64, -0.48) percentage points per year in the *F*/RF (excluding *R117H*) cohort, respectively. Although the rates of decline were slower for all age groups in the F/RF (all) cohort than in the F/F cohort, rates between the F/RF (excluding



Fig. 1 Estimated slope of $ppFEV_1$, by mutation. F F508del, $ppFEV_1$ percent predicted forced expiratory volume in 1 s, RF residual function. Annual rates of

R117H) versus *F/F* cohorts were not statistically significantly different in children [difference, 0.52 percentage points per year (95% CI -0.27, 1.31)] or young adults [difference, 0.67 percentage points per year (95% CI -0.19, 1.53)] (Fig. 2). The estimated intercepts and rates of decline in the overall population and by age group were similar in sensitivity analyses using the Wang–Hankinson equations (Table 2).

DISCUSSION

We sought to better understand the rate of lung function decline in people with CF who have an RF CFTR mutation. We used data from 2006 to 2014 to ensure an adequate sample size for CFTR modulator-untreated cohorts. Our data showed a decline in $ppFEV_1$ in those with F/RFgenotypes across all age groups, although the rate of decline varied by age. Overall, the rate of ppFEV₁ decline observed in participants with F/RF genotypes was slower than that in participants with F/F genotypes, consistent with the findings reported by Leung et al. [22]. When participants with an R117H mutation were excluded from the analyses, rates of lung function decline in children and young adults were not statistically significantly different compared with rates in the same age groups in the F/Fcohort. This finding indicates that, despite having better preserved lung function compared with the F/F cohort at baseline, the F/RF(excluding R117H) cohort also experienced a ppFEV₁ decline were based on 2012 Global Lung Initiative equations [19]. Error bars represent SE; values in parentheses are the 95% CI of the slope. *P < 0.001 versus F/F

progressive decline in lung function. The findings of this study demonstrate the progressive nature of CF in people with the *F*/RF genotype, highlighting the importance of beginning treatment to preserve lung function as early as possible. Since the analysis, several CFTR modulators have become available for this patient population, including ivacaftor (Kalydeco), tezacaftor/ivacaftor and ivacaftor (Symdeko), and, most recently, elexacaftor/tezacaftor/ivacaftor and ivacaftor (Trikafta) [23–25].

This retrospective study had several limitations. The severity of disease in individuals with F/RF genotypes was highly variable and influenced by several factors, including environment and the presence of modifier genes [10]. It is possible that in the current era of widespread newborn CF screening, children with F/RF genotypes with mild symptoms are being diagnosed with CF more frequently, while the adult F/RF population in this study may have been diagnosed due to being symptomatic (instead of via newborn screening) and therefore may exhibit greater lung function decline than younger F/RF populations. Phenotypic variability in people with CF who have an R117H mutation has also been shown to be influenced by polymorphisms in the polythymidine tract of intron 8 of the CFTR gene [26]; this could not be examined in these analyses due to lack of data availability in the registry.

Characteristic	$F/F^{\rm a}$ ($n = 11,916$)	F/RF (all) ($n = 1242$)	F/RF (excluding R117H) ($n = 889$)
Intercept (95% CI)			
Overall	74.88 (74.49, 75.28)	86.25 (84.92, 87.58)	85.49 (83.89, 87.10)
6–12 years of age	91.88 (91.21, 92.55)	99.85 (97.34, 102.36)	100.52 (97.32, 103.71)
13–17 years of age	81.18 (80.26, 82.10)	92.02 (88.76, 95.28)	90.12 (86.32, 93.92)
18–24 years of age	64.86 (64.02, 65.70)	80.98 (77.97, 83.99)	79.78 (76.39, 83.17)
\geq 25 years of age	56.02 (55.21, 56.83)	68.34 (66.48, 70.21)	66.58 (64.40, 68.76)
Slope (95% CI)			
Overall	-1.93 (-2.03, -1.82)	-0.74 (-1.14, -0.34)	-1.09 (-1.56, -0.63)
6–12 years of age	-0.75 (-0.90, -0.59)	0.13 (-0.52, 0.78)	-0.43 (-1.23, 0.36)
13–17 years of age	-3.56 (-3.75, -3.37)	-1.26 (-2.03, -0.49)	-1.43 (-2.30, -0.57)
18–24 years of age	-2.51 (-2.70, -2.33)	-1.39 (-2.17, -0.61)	-1.84 (-2.70, -0.98)
\geq 25 years of age	-1.88 (-2.07, -1.69)	-1.01 (-1.52, -0.49)	-1.11 (-1.71, -0.52)

Table 2 Estimated intercept and annualized slope for ppFEV1 using Wang-Hankinson equations (sensitivity analysis)

*F F508del, ppFEV*₁ percent predicted forced expiratory volume in 1 s, *RF* residual function ^aIntercepts and slopes were calculated by weighting each age group category pooled across the *F/F* and *F/*RF (all) cohorts



Fig. 2 Estimated slope of $ppFEV_1$, by mutation and age group. *F F508del*, *ppFEV*₁ percent predicted forced expiratory volume in 1 s, *RF* residual function. Annual rates of $ppFEV_1$ decline were based on 2012 Global Lung

Initiative equations [19]. Error bars represent SE; values in parentheses are the 95% CI of the slope. *P < 0.05 versus F/F; $^{\dagger}P < 0.001$ versus F/F

CONCLUSION

These results demonstrate substantial disease burden characterized by progressive lung function decline in people with CF across all age groups evaluated, regardless of genotype. The decline across all age groups reinforces the importance of early treatment intervention and clinical monitoring to preserve lung function in people with CF, including people with F/RFgenotypes.

ACKNOWLEDGEMENTS

Funding. This study was funded by Vertex Pharmaceuticals Incorporated (Boston, MA, USA), which participated in the design, statistical analysis, and interpretation of the data; provided editorial and writing assistance; and funded the journal's rapid service fee.

Medical Writing, Editorial, and Other Assistance. Editorial coordination and support were provided by Nathan Blow, PhD. NB is an employee of Vertex Pharmaceuticals Incorporated and may own stock or stock options in that company. Medical writing and editorial support were provided by Liz Phipps, PhD, CMPP, and Jackie Highland, PhD, of ArticulateScience, LLC, and was funded by Vertex Pharmaceuticals Incorporated. This work resulted from a scientific advisory group formed by Vertex Pharmaceuticals Incorporated and the US Cystic Fibrosis Foundation. All authors had full access to the study data, and the corresponding author had the final responsibility for the decision to submit for publication.

Author Contributions. All named authors meet the International Committee of Medical Journal Editors criteria for authorship for this article, take responsibility for the integrity of the work as a whole, and have given their approval for this version to be published. Individual author contributions are as follows: **GSS**: conceptualization, writing – original draft, writing – review and editing, and supervision.

Pulm Ther (2022) 8:385-395

MWK: conceptualization, methodology, investigation, writing - original draft, writing review and editing, and visualization. EFM: conceptualization, writing - review and editing, and project administration. RBM: investigation, writing - review and editing, and supervision. BL: conceptualization, methodology, writing review and editing, and visualization. ES: conceptualization, methodology, validation, writing - review and editing, visualization, supervision, and funding acquisition. SJM: methodology, software, formal analysis, and writing - review and editing. DJP: methodology, software, formal analysis, and writing review and editing. NM-H: conceptualization, methodology, and writing – review and editing. CHG: conceptualization, formal analysis, and review - writing and editing. WJM: conceptualization and writing - review and editing. MED: conceptualization, resources, data curation, writing - original draft, writing - review and editing, and supervision. YY: conceptualization, methodology, validation, writing original draft, writing - review and editing, visualization, supervision, project administration, and funding acquisition.

Prior Presentation. This manuscript is based on work that was previously presented at the American Thoracic Society International Conference (Washington, DC; May 19–24, 2017).

Disclosures. All authors received nonfinancial assistance (assistance with manuscript preparation) from ArticulateScience, LLC, which received funding from Vertex Pharmaceuticals. Additional disclosure are as follows: GSS reports grants to his institution, travel support, and participation on an advisory board for Vertex Pharmaceuticals. MWK reports grants to his institution for clinical trial participation, honoraria for advisory board meetings, fees for consulting and speaker bureau, and travel reimbursement/support from Vertex Pharmaceuticals; grants to his institution for clinical trial participation, personal fees, and travel reimbursement/support from the US Cystic Fibrosis Foundation; grants to his institution for clinical trial participation, travel reimbursement/support, and consulting fees

from Laurent Pharmaceuticals; grants and consulting fees from AzurRx; personal fees (honoraria for advisory board meeting) from Insmed, Ionis Pharmaceuticals, Nabriva Therapeutics, and Santhera; consulting and speaker bureau fees and travel reimbursement/support from Chiesi; consulting fees and travel support/reimbursement from Merck; consulting fees from Cystetic Medicines, Mylan, and Paranta Biosciences; and grants to his institution from the National Institutes of Health. EFM reports research grants, consulting fees (steering committee), and payment/honoraria (educational materials) from Vertex Pharmaceuticals; consulting fees (advisory board) from Janssen Pharmaceuticals, Viatris Pharma, and Enterprise Pharma; and payment/honoraria (lecture) from Pfizer. RBM reports chapter authorship royalties from UpToDate; consulting fees from 4D Molecular Therapeutics, Aridis Pharmaceuticals, Nob Hill Therapeutics, and Zambon; and stock or stock options in Pfizer, Regeneron Pharmaceuticals, and Sanofi. BL is a former employee of Vertex Pharmaceuticals. ES, MED, and YY are employees of Vertex Pharmaceuticals and may own stock or stock options in that company. SJM is a former employee and consultant of ICON plc (was an employee/consultant at the time of the study) and may own stock or stock options with that company. SJM is currently an employee of Unlearn.AI. SJM reports payments from Vertex Pharmaceuticals to ICON; grants or contracts from various pharmaceutical, biotech, and device companies to ICON; support (for attending meetings and/or travel) from ICON; and payments from various pharmaceutical, biotech, and device companies to ICON for support for attending meetings and/or travel. DJP is a former employee and consultant of ICON plc (was an employee/consultant at the time of the study), which received funding from Vertex Pharmaceuticals for this study and which receives grants or contracts from various pharmaceutical, biotechnology, and device companies for providing clinical research services; is a former employee and 50% owner of DMA Corporation, which received grants or contracts from various pharmaceutical, biotechnology, and device companies; reports consulting fees and payments for advisory board participation to themself and/or DMA Corporation from the US Cystic Fibrosis Foundation; reports payments or honoraria to themself and/or DMA Corporation from various pharmaceutical, biotech, and device companies; reports support for attending meetings and/or travel from ICON (while an employee); and reports payments from various pharmaceutical, biotech, and device companies to ICON for support for attending meetings and/or travel. DJP may own stock or stock options with ICON plc and DMA Corporation. DJP is currently retired from DMA Corporation. NM-H reports honorarium to herself for participation on the US Cystic Fibrosis Foundation Registry Committee and grants to her institution from the US Cystic Fibrosis Foundation; payments to herself for advisory board participation and grants to her institution from the National Institutes of Health; and consulting fees from and advisory board participation for Vertex Pharmaceuticals. CHG reports funding from the US Cystic Fibrosis Foundation (for conducting exacerbation studies, studies of gallium to treat infections, and clinical trial coordination in CF), the National Institutes of Health (to support clinical research in CF and clinical trials in other disease entities and to support multicenter clinical trials), and the US Food and Drug Administration (for studies of gallium to treat infections in CF); consulting fees (serving on an advisory board) from Enterprise Therapeutics; honoraria (to serve as chair of a grant review committee) from Gilead Sciences; payment or honoraria (to serve as a data safety monitoring board chair for a trial supported by Novartis and the European Commission) from Novartis; payment or honoraria (to serve as a US lead in a phase 2 trial of a novel therapy for CF) from Boehringer Ingelheim; honoraria and travel expenses (for talk at UK LEAD conference) from Vertex Pharmaceuticals; support to travel to grant review sessions from Gilead Sciences; served as a data safety monitoring board chair (for a trial supported by Novartis and the European Commission) for Novartis; served as an advisor for Aer Therapeutics; served on an advisory board for Enterprise Therapeutics; and owns stock or stock options (for serving as an advisor) with Aer Therapeutics. WJM reports grants from the

National Institutes of Health National Heart, Lung, and Blood Institute (funding for Tucson Children's Respiratory Study [Co-I], ORBEX – asthma prevention study [site PI, executive committee), the National Institutes of Health National Institute of Allergy and Infectious Disease (consultant to PARK – asthma prevention study), and the US Cystic Fibrosis Foundation (participation as chair of the CFF data safety monitoring board [does not directly oversee the Vertex studies]); consulting fees (CFF data safety monitoring board and CFF Comparative Effectiveness Research/Registry Committee) from the US Cystic Fibrosis Foun-

dation; honorarium (speaker at biennial Pediatric Pulmonary Board Preparation Course) from the American College of Chest Physicians; and funding for Healthcare.

Compliance with Ethics Guidelines. This study was conducted in accordance with the current Good Clinical Practice Guidelines as described by the International Council for Harmonisation, the Declaration of Helsinki, and all applicable local regulations. People with CF provided informed consent to participate in the US Cystic Fibrosis Foundation Patient Registry and to have their data included in the registry. The study did not involve primary data collection and used only de-identified data; thus, institutional review board approval was not required.

Data Availability. The data sets generated during and/or analyzed during the current study are available from the US Cystic Fibrosis Foundation Patient Registry, https://www.cff. org/researchers/patient-registry-data-requests. The US Cystic Fibrosis Foundation Patient Registry collects and manages its own data and maintains processes for researchers to request summarized data.

Open Access. This article is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License, which permits any non-commercial use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide

a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/bync/4.0/.

Pulm Ther (2022) 8:385-395

REFERENCES

- 1. Ratjen F, Bell SC, Rowe SM, Goss CH, Quittner AL, Bush A. Cystic fibrosis. Nat Rev Dis Primers. 2015;1: 15010.
- 2. Bell SC, Mall MA, Gutierrez H, Macek M, Madge S, Davies JC, et al. The future of cystic fibrosis care: a global perspective. Lancet Respir Med. 2020;8(1): 65–124.
- 3. Dugueperoux I, De Braekeleer M. The CFTR 3849+10kbC->T and 2789+5G->A alleles are associated with a mild CF phenotype. Eur Respir J. 2005;25(3):468–73.
- 4. McKone EF, Goss CH, Aitken ML. CFTR genotype as a predictor of prognosis in cystic fibrosis. Chest. 2006;130(5):1441–7.
- Wagener JS, Millar SJ, Mayer-Hamblett N, Sawicki GS, McKone EF, Goss CH, et al. Lung function decline is delayed but not decreased in patients with cystic fibrosis and the R117H gene mutation. J Cyst Fibros. 2018;17(4):503–10.
- Rowe SM, Daines C, Ringshausen FC, Kerem E, Wilson J, Tullis E, et al. Tezacaftor-ivacaftor in residual-function heterozygotes with cystic fibrosis. N Engl J Med. 2017;377(21):2024–35.
- 7. Cystic Fibrosis Foundation. US cystic fibrosis foundation 2020 annual data report. Bethesda: US Cystic Fibrosis Foundation; 2021.
- Sheppard DN, Rich DP, Ostedgaard LS, Gregory RJ, Smith AE, Welsh MJ. Mutations in CFTR associated with mild-disease-form Cl⁻ channels with altered pore properties. Nature. 1993;362(6416):160–4.

- US CF Foundation JHU, The Hospital for Sick Children. The clinical and functional translation of CFTR (CFTR2). 2011. https://cftr2.org/.
- Castellani C, Cuppens H, Macek MJ, Cassiman J, Kerem E, Durie P, et al. Consensus on the use and interpretation of cystic fibrosis mutation analysis in clinical practice. J Cyst Fibros. 2008;7(3):179–96.
- 11. Clancy JP. Rapid therapeutic advances in CFTR modulator science. Pediatr Pulmonol. 2018;53(S3): S4-11.
- Van Goor F, Yu H, Burton B, Hoffman BJ. Effect of ivacaftor on CFTR forms with missense mutations associated with defects in protein processing or function. J Cyst Fibros. 2014;13(1):29–36.
- Highsmith WE, Burch LH, Zhou Z, Olsen JC, Boat TE, Spock A, et al. A novel mutation in the cystic fibrosis gene in patients with pulmonary disease but normal sweat chloride concentrations. N Engl J Med. 1994;331(15):974–80.
- Sheppard DN, Ostedgaard LS, Winter MC, Welsh MJ. Mechanism of dysfunction of two nucleotide binding domain mutations in cystic fibrosis transmembrane conductance regulator that are associated with pancreatic sufficiency. EMBO J. 1995;14(5):876–83.
- Welsh MJ, Smith AE. Molecular mechanisms of CFTR chloride channel dysfunction in cystic fibrosis. Cell. 1993;73(7):1251–4.
- Comer DM, Ennis M, McDowell C, Beattie D, Rendall J, Hall V, et al. Clinical phenotype of cystic fibrosis patients with the G551D mutation. QJM. 2009;102(11):793–8.
- 17. McKone EF, Emerson SS, Edwards KL, Aitken ML. Effect of genotype on phenotype and mortality in cystic fibrosis: a retrospective cohort study. Lancet. 2003;361(9370):1671–6.

- Knapp EA, Fink AK, Goss CH, Sewall A, Ostrenga J, Dowd C, et al. The Cystic Fibrosis Foundation Patient Registry. Design and methods of a national observational disease registry. Ann Am Thorac Soc. 2016;13(7):1173–9.
- 19. Quanjer PH, Stanojevic S, Cole TJ, Baur X, Hall GL, Culver BH, et al. Multi-ethnic reference values for spirometry for the 3–95-yr age range: the global lung function 2012 equations. Eur Respir J. 2012;40(6):1324–43.
- Hankinson JL, Odencrantz JR, Fedan KB. Spirometric reference values from a sample of the general U.S. population. Am J Respir Crit Care Med. 1999;159(1):179–87.
- Wang X, Dockery D, Wypij D, Fay M, Ferris BJ. Pulmonary function between 6 and 18 years of age. Pediatr Pulmonol. 1993;15(2):75–88.
- 22. Leung GJ, Cho TJ, Kovesi T, Hamid JS, Radhakrishnan D. Variation in lung function and nutritional decline in cystic fibrosis by genotype: an analysis of the Canadian cystic fibrosis registry. J Cyst Fibros. 2020;19(2):255–61.
- 23. Kalydeco prescribing information. Boston, MA: Vertex Pharmaceuticals Incorporated; Revised 9/2020.
- 24. Symdeko prescribing information. Boston, MA: Vertex Pharmaceuticals Incorporated; Revised 12/2020.
- 25. Trikafta prescribing information. Boston, MA: Vertex Pharmaceuticals Incorporated; Revised 12/2020.
- Massie RJH, Poplawski N, Wilcken B, Goldblatt J, Byrnes CA, Robertson C. Intron-8 polythymidine sequence in Australasian individuals with CF mutations *R117H* and *R117C*. Eur Respir J. 2001;17(6):1195–200.