

Neurobiology of Arousal and Sleep: Updates and Insights Into Neurological Disorders

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Abstract Brain activation during wakefulness is sustained by multiple arousal systems. Dysfunction of one or more arousal systems is a feature of neurological disorders associated with hypersomnolence and/or sleep-wake cycle disturbance. Narcolepsy, Alzheimer's disease (AD), Parkinson's disease (PD), and traumatic brain injury appear to involve hypocretin (HCT) and possibly histamine insufficiency as a mechanism related to excessive daytime sleepiness. Loss of cholinergic neurons in AD and of dopamine neurons in PD contributes to sleep-wake disturbance. GABAergic neurons in the preoptic hypothalamus and rostral medulla promote sleep through inhibition of arousal systems. Pathology of preoptic sleep regulatory circuits is correlated with sleep disturbance in AD. An unidentified endogenous somnogen that potentiates the actions of gamma-aminobutyric acid (GABA) is present in the cerebrospinal fluid (CSF) of patients with primary hypersomnia.

Descending pathways from the dorsal lateral pons to the ventral medulla and spinal cord are responsible for the inhibition of spinal motoneurons during rapid eye movement (REM) sleep and are implicated in human REM sleep behavior disorder.

Keywords Monoamines · Hypocretin · Orexin · Acetylcholine · Dopamine · Glutamate · Galanin · Preoptic hypothalamus · Basal forebrain · Tuberomammillary nucleus · Locus coeruleus · Laterodorsal tegmentum · Alzheimer's disease · Parkinson's disease · Traumatic brain injury

Introduction

As in all areas of modern neuroscience, rapid progress is being made in understanding the cellular and molecular mechanisms and organization of brain circuits that regulate sleep and wakefulness. It is clear that sleep-wake regulatory mechanisms identified in rodents are highly conserved and that fundamental neurobiological investigations in animal models can inform our understanding sleep disturbances in human disorders. Here we briefly review the current status of knowledge about the functional neuroanatomy and neuropharmacology of sleep and arousal regulation, and attempt to integrate this knowledge with the known neuropathology in some common neurological disorders associated with sleep-wake abnormalities.

Arousal Mechanisms

Generalized electrographic, behavioral, and autonomic activation during waking emerges from the activity of neurochemically specified arousal systems located in the brain

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stem, posterior and lateral hypothalamus, and basal forebrain. These include neurons that express histamine (HA), serotonin (5HT), noradrenalin (NA), acetylcholine (ACH), glutamate (GLU), dopamine (DA), or hypocretin/orexin (HCT). Each of these neurotransmitter/neuropeptide systems participates in specific aspects of cognition, behavior, sensory processing, and/or autonomic control during waking, but here we focus on their collective contribution to general arousal (Table 1).

Arousal systems can impact neocortical activity indirectly through projections to the thalamus, lateral hypothalamus, or basal forebrain, while others send direct projections to the cortex. There are differences in the pattern and intensity of neuronal activity across sleep-waking states among the arousal systems which has implications for potential roles in sleep-wake disturbance associated with neurological disorders.

Monoaminergic Neurons HA neurons in the hypothalamic tuberomammillary nucleus (TMN), 5HT neurons in the dorsal raphe nucleus (DRN), and NA neurons in the locus coeruleus (LC) are characterized by aggregated cell bodies in distinct nuclei with projections to widespread forebrain and brain stem targets [1–3]. All three nuclei target the thalamus, hypothalamus, and basal forebrain and send direct projections to the cortex. Descending projections target visceromotor and somatic motor cranial nerve nuclei.

Monoaminergic neurons also share a “REM off” discharge pattern, characterized by tonic discharge rates during waking, reduced discharge during non-REM sleep, and cessation of activity during REM sleep [4, 5]. Drugs that promote release or inhibit reuptake of one or more monoaminergic systems promote waking. Drugs that antagonize the postsynaptic actions of the monoamines can be sedating [5].

Acetylcholine ACH-containing neurons are localized in the dorsolateral [6] pontomesencephalic reticular formation, including the pedunculopontine tegmental (PPT) and laterodorsal tegmental (LDT) nuclei and in the basal forebrain

[6]. ACH neurons in the LDT/PPT project to the thalamus and hypothalamus, and those in the basal forebrain project to the limbic system and neocortex. Neurons in both groups exhibit higher rates of discharge in both waking and REM than in NREM sleep [6, 7]. ACH neurons in the BF contribute to the generation of gamma oscillations in neocortical circuits during activated behavioral states [6]. In mice expressing the light-sensitive excitatory opsin channelrhodopsin-2 in basal forebrain ACH neurons, light-induced activation during NREM sleep evokes short-latency EEG and behavioral arousal [8].

Dopamine DA-containing neurons implicated in arousal are primarily localized in the substantia nigra and the adjacent ventral tegmental area of the midbrain [9]. At the level of the basal ganglia, effects of DA on sleep and arousal are complex, with DA receptors in the nucleus accumbens associated with arousal and DA signaling in the external segment of the globus pallidus implicated in sleep induction [9, 10]. DA-containing wake-active neurons are found in the ventral periaqueductal gray matter, and selective destruction of these DA neurons increases daily sleep time by 20 % [11]. DA is inactivated primarily through reuptake by the DA transporter. Stimulant drugs such as amphetamines and modafinil block the DA transporter and reduce DA reuptake [12], suggesting that the net effect of global elevations in DA signaling is arousal. Several antipsychotic medications block DA receptors and are sedating [5].

DA may play a role in restless legs syndrome (RLS) and the excessive daytime sleepiness associated with RLS. Evidence supporting a role for DA in the pathophysiology of RLS includes improvement in symptoms in response to DA agonist agents and, conversely, worsening RLS symptoms with blockade of DA receptors. However, positron emission tomography (PET) and single-photon emission computed tomography (SPECT) studies imaging DA transporter and receptor densities in human patients with RLS show conflicting results

Table 1 Summary of arousal and sleep regulatory neuronal systems

Neurotransmitter	Location	Neuronal activity		
		Wake	NREM	REM
Histamine	Tuberomammillary nucleus	↑↑↑	↑	↔
Serotonin	Dorsal raphe nucleus	↑↑↑	↑	↔
Noradrenalin	Locus coeruleus	↑↑↑	↑	↔
Hypocretin/orexin	Lateral hypothalamus	↑↑↑	↔	↔
Dopamine	Ventral tegmental area/ventral periaqueductal gray	↑↑↑	↑	?
Acetylcholine	Laterodorsal tementum/basal forebrain	↑↑↑	↑	↑↑↑
Glutamate	Parabrachial nucleus	↑↑↑	↑	↑↑↑
GABA	MnPO/VLPO	↑	↑↑↑	↑↑↑
GABA	Medullary parafacial zone	↑	↑↑↑	?

↑↑↑ high ↑ low ↔ silent ? unknown

[13–17]. Potentially relevant neuroanatomical pathways in RLS could involve the A11 diencephalic DA cell cluster, which sends relays to dorsal root ganglion/sensory afferents in the spinal cord [18, 19].

Glutamate GLU is a widespread excitatory neurotransmitter in the brain, and GLU-containing neurons are found in the pontine and midbrain reticular formation. Specifically, GLU neurons in the parabrachial/precoeruleus (PB/PC) region of the pons have been implicated in the regulation of cortical arousal. Cell-specific lesions of the PB/PC disrupt desynchronized EEG patterns during waking and in response to arousing stimuli [20]. Projections of GLU neurons in the PB/PC to cortically projecting neurons in the basal forebrain are important in mediating PB/PC effects on cortical activation. Similar to ACH, extracellular levels of GLU in the cortex and in the hypothalamus are elevated during waking and REM sleep compared to NREM sleep [21, 22], suggesting that GLU and ACH neurotransmission contributes to brain activation in both states.

Hypocretin/Orexin The HCTs are peptides expressed by neurons in the perifornical lateral hypothalamus. HCT neurons have extensive projections to other hypothalamic nuclei, the limbic system, thalamus, and cortex and to the brain stem and spinal cord [23]. HCT neurons occupy a powerful position among arousal systems in that they target the monoaminergic cell groups, ACH neurons in the brain stem and basal forebrain, and DA neurons in the VTA [24, 25]. Acting through HCT-1 and/or HCT-2 receptors, the neuropeptides exert predominately excitatory effects. Given their central position among the arousal circuits, integrity of the HCT system is critical for consolidated arousal states. Knockout of HCT peptides or HCT receptors in mice yields a narcolepsy-like phenotype that includes fragmented wakefulness [26]. Central or systemic administration of dual antagonists of HCT-1 and HCT-2 receptors has been shown to promote sleep and EEG synchrony in several species [27].

Discharge of HCT neurons is highest during waking, declines dramatically during NREM sleep, and remains low during REM sleep [6]. Discharge during waking is phasic, often occurring during vigorous waking movements. Studies in mice and humans suggest that maximal activity of HCT neurons occurs when working for positive reinforcement (mice) and during positive emotions and social interactions (humans) [28, 29].

Neurological Disorders of Arousal

Several clinical neurological disorders exhibiting deficits in central arousal mechanisms are worth mentioning, namely narcolepsy (with and without cataplexy), but also Alzheimer's

disease (AD), Parkinson's disease (PD), and traumatic brain injury (TBI). HCT deficiency or dysfunction may be a common mechanism underlying the inability to maintain normal arousal in these disease states (Table 2).

Patients with narcolepsy experience excessive daytime sleepiness and "sleep attacks." Narcolepsy in humans results from the loss of HCT-producing neurons within the lateral and posterior hypothalami [30]. Recent evidence, including the increase in the incidence of narcolepsy after the H1N1 vaccination in 2010, supports a role for autoimmunity in the destruction of HCT neurons [31, 32]. Of note, in the current International Classification of Sleep Disorders 3rd Edition (ICSD-3), narcolepsy type 1 (formerly known as "narcolepsy with cataplexy" and sometimes referred to as "hypocretin deficiency syndrome") is distinguished from narcolepsy type 2 (formerly known as "narcolepsy without cataplexy") on the basis of HCT deficiency [33]. In addition, the HA system may also be affected in narcolepsy, as HA levels in the CSF may be low in patients with narcolepsy and primary hypersomnia [34]. Two independent groups recently reported an increased number of HA cells in the TMN in post-mortem cases of human narcolepsy, possibly due to a compensatory response to HCT loss and/or related to the process itself causing human narcolepsy [35, 36].

In AD, the most common sleep-related complaints are insomnia, sleep fragmentation, and excessive daytime sleepiness [37]. Neuropathology of the cholinergic basal forebrain and monoaminergic locus coeruleus both feature prominently and early in the time course of AD [38, 39]. Given the roles of ACH and NE in arousal identified from basic neuroscience studies above, both neurotransmitter systems likely contribute to symptoms of excessive daytime sleepiness in AD. Similarly, recent convergent evidence from several different groups has pointed to an important role for HCT in the pathogenesis of AD. Using a transgenic mouse model of AD, HCT infusion into the brain directly increased interstitial levels of soluble amyloid-beta, and administration of the dual HCT receptor antagonist almorexant significantly reduced amyloid-beta plaque burden [40]. Double transgenic mice overexpressing

Table 2 Affected arousal and sleep regulatory neuronal systems in selected neurological disorders

Disease	Neurotransmitter					
	HA	HCT	NA	DA	ACH	GABA
Alzheimer's disease		↑	↓		↓	↓
Parkinson's disease		↓		↓		
Traumatic brain injury	↓	↓				
Narcolepsy type 1	↑	↓				
Primary hypersomnia	↓					↑

ACH acetylcholine, DA dopamine, GABA gamma-aminobutyric acid, HCT hypocretin (orexin), HA histamine, NA noradrenalin

amyloid plaques and with a knockout of the HCT gene showed a significant decrease in amyloid plaques along with an increase in total sleep time [41]. Human studies also support the role of HCT in the progression of AD. Increased cerebrospinal fluid (CSF) levels of HCT significantly predict sleep-wake deterioration in AD patients and are significantly associated with cognitive decline [42•].

In PD, the degeneration of the nigrostriatal DA system is the neuropathological hallmark of the disease. As pharmacological studies of brain DA demonstrate a potent role for this neurotransmitter in regulating generalized brain activation, DA deficiency may contribute to symptoms of excessive daytime sleepiness in PD. Recent studies indicate that HCT deficiency may also contribute to hypersomnia in advanced PD [43–47].

Patients with TBI of all severity types (even mild TBI, without obvious lesions seen on neuroimaging) frequently complain of excessive daytime sleepiness and the inability to maintain normal levels of arousal. The neuropathology related to sleep-wake disturbances in TBI is just beginning to be explored in both animal and human studies. Given the phenotype of excessive daytime sleepiness and nighttime sleep fragmentation in human TBI which mimics narcolepsy, several studies have examined HCT in TBI. Measured HCT levels in the cerebrospinal fluid (CSF) were low in 95 % of 44 patients within the first 4 days of moderate to severe TBI [48]. A small study of four patients who died 7 to 42 days after severe TBI showed a 27 % reduction in the number of HCT neurons compared to non-TBI controls [49]. Likewise, studies using mouse models of mild [50•] and moderate [51] TBI show decreased brain HCT levels and decreased HCT neuron activation during wakefulness after brain injury. However, deficits in HCT are unlikely to explain all sleep-wake abnormalities, especially in the chronic phase of TBI, as CSF HCT levels for the most part return to baseline after 6 months post-TBI [52]. In support of this, a recent post-mortem human study in 12 patients with severe TBI showed a 41 % loss of histaminergic neurons in the tuberomammillary nucleus, compared to control subjects [53].

These findings implicating ACH, monoamines, DA, and HCT in clinical disorders of arousal such as narcolepsy, AD, PD, and TBI strongly support the basic neuroscience data described above.

Sleep Onset and NREM Sleep-Promoting Mechanisms

Brain mechanisms that promote sleep must be able to facilitate a coordinated suppression of activity in multiple groups of arousal-promoting neurons over a period of seconds to minutes at the wake to sleep transition, and maintain that suppression of activity over the subsequent sleep period lasting

many minutes (rodents) to hours (primates). The onset and maintenance of sleep are accomplished through interactions among three cellular/molecular and neurochemical mechanisms: (1) systems of GABAergic sleep-promoting neurons located in the preoptic hypothalamus and rostral medulla that exert inhibitory control over key arousal systems, (2) production of endogenous sleep regulatory substances during waking that regulate homeostatic sleep drive by targeting both wake- and sleep regulatory circuits in multiple brain regions, and (3) output of the circadian clock in the suprachiasmatic nucleus that functions to promote wakefulness and sleep at different times of day. The complex topic of the circadian control of sleep and its role in sleep disturbance in neurological disorders is beyond the scope of this article, and the reader is referred to recent comprehensive reviews of the subject [54–56].

Preoptic Hypothalamic Neurons Sleep regulatory neurons have been identified by direct neuronal recording during sleep and wakefulness [57] and by sleep-related expression of the protein product of the *c-fos* gene, an anatomical marker of neuronal activity [58]. The functional importance of sleep-active neurons is confirmed by loss of function lesion studies. These approaches have identified two subregions of the preoptic hypothalamus that contain high densities of functionally important sleep-active neurons, the ventrolateral preoptic area (VLPO) [58, 59] and the median preoptic nucleus (MnPO) [60]. VLPO and MnPO neurons exhibit elevated discharge rates during NREM and REM sleep compared to waking (Table 1). MnPO and VLPO neurons are dynamically responsive to changes in homeostatic sleep pressure induced by sleep deprivation [61•].

Sleep-related c-Fos expression is co-localized with GABA in the MnPO [60] and with GABA and galanin in the VLPO [58, 62]. VLPO neurons project heavily to HA neurons in the TMN and to the DRN, LC, and the PPT/LDT [63]. MnPO neurons project to the DRN and LC. MnPO and VLPO neurons that express sleep-related c-Fos project to the HCT neuronal field in the lateral hypothalamus [64]. Sleep-active neurons in VLPO and MnPO exhibit discharge profiles across the wake-NREM-REM cycle that are reciprocal to those of wake-promoting HA, 5HT, NE, and HCT neurons [63]. These findings support the hypothesis that VLPO/MnPO neurons promote sleep through monosynaptic GABAergic inhibition of monoaminergic, cholinergic, and hypocretinergic neurons. GABAergic VLPO neurons are inhibited by ACH, 5HT, and NE, and reciprocal inhibitory interactions between sleep and arousal regulatory neurons are hypothesized to function as a bi-stable sleep-wake switch [65].

Rostral Medullary Neurons An older literature examining the behavioral and EEG effects of brain stem transections and lesions provided evidence for sleep-promoting mechanisms in the brain stem as well as the forebrain [66]. Recent findings

have identified a sleep regulatory neuronal group in the rostral medulla of rats and mice, located lateral and dorsal to the facial nerve in the parafacial zone (PZ) [67, 68•]. PZ neurons express c-Fos during sleep but not during waking and sleep-related c-Fos is co-localized with markers for GABA [67]. Excitotoxic lesions of the PZ result in persistent sleep loss [67], and acute pharmacogenetic activation of PZ GABAergic neurons in transgenic mice results in increases in non-REM sleep and suppression of waking and REM sleep [68•]. PZ neurons project rostrally to the PB/PC nuclei [67] that contain arousal-promoting GLU neurons [20]. Optogenetic activation of GABAergic neurons in the PZ evokes GABA-mediated inhibition of PB/PC neurons that project to the basal forebrain [68•]. The ascending circuit of GABAergic PZ neurons evoking sleep-related inhibition of PB/PC neurons that normally excites cortically projecting basal forebrain neurons during waking complements the descending sleep-related inhibition of arousal systems that originates in the MnPO/VLPO.

Sleep Regulatory Substances Homeostatic sleep pressure accumulates slowly during periods of sustained waking and dissipates slowly during subsequent sleep. It has long been hypothesized that accumulation and dissipation of endogenous somnogenic substances during waking and sleep, respectively, underlie the dynamics of sleep homeostasis [69].

Adenosine (ADO) is an inhibitory neuromodulator in the CNS, whose role in sleep is suggested by the potent arousal-producing effects of caffeine, an antagonist of A₁ and A_{2A} ADO receptors [70]. Sleep deprivation is accompanied by elevated ADO levels in the basal forebrain and cortex, followed by a decline during recovery sleep [4, 71]. ADO and its analogs promote sleep after systemic and central administration, and ADO-induced sleep is accompanied by increased EEG slow-wave activity (SWA), as is sleep that follows sleep deprivation [4, 70]. Astrocytes as well as neurons are sources of ADO in the brain. A conditional knockout of the A₁-R gene in mice attenuates EEG SWA in NREM sleep following sleep deprivation [72]. ACH neurons in the basal forebrain are important targets of the sleep-promoting effects of ADO acting on A₁R [73]. Additional arousal systems may be targeted by ADO. Local perfusion of A₁-R agonists in the rat lateral hypothalamus suppresses waking c-Fos expression in HCT neurons [74]. Complementary actions of ADO on the excitatory A_{2A}-R also impact sleep. Central administration of A_{2A}-R agonists promotes sleep and activates VLPO neurons [75]. Administration of A_{2A} agonists into the subarachnoid space ventral to the preoptic area increases sleep and increases c-Fos expression in GABAergic neurons in the MnPO and VLPO [76]. A_{2A} receptors in the shell of the nucleus accumbens have also been implicated in sleep regulation [9].

Cytokines, including interleukin-1 β (IL-1 β) and tumor necrosis factor- α (TNF- α), are sleep promoting [69]. Antagonism of IL-1 β and TNF- α can disrupt normal sleep and impair

homeostatic responses to sleep deprivation [69, 77]. Expression of IL-1 β and TNF- α is elevated in multiple brain regions in response to waking neuronal activity [77]. Cellular mechanisms of cytokine-mediated sleep generation are not completely understood but may involve a combination of arousal system inhibition and activation of preoptic sleep regulatory neurons [78–81].

Neurological Disorders of Sleep and Sleep-Promoting Systems

Several clinical disorders also involve the VLPO, GABA, and galanin systems as described above. We discuss two such examples below for Alzheimer's disease (AD) and primary hypersomnia (Table 2). These examples highlight the fact that sleep-promoting systems identified in animal models are highly conserved and that inhibitory pathways within the hypothalamus may also be relevant to human neurological disorders of sleep and excessive sleepiness.

Patients with AD frequently report significant insomnia and sleep fragmentation. A recent study examining 45 older adults with and without AD found that individuals with more galanin-immunoreactive neurons within the intermediate nucleus (the putative human homologue to VLPO in rodents) exhibited less fragmented sleep [82]. This suggests that GABA/galanin in the intermediate nucleus is important for the consolidation of sleep and that clinical disorders exhibiting sleep fragmentation may involve this neuroanatomical pathway.

Patients with primary hypersomnia have excessive daytime sleepiness in the absence of other known causes of sleepiness. A recent study in 32 hypersomnolent patients identified a naturally occurring substance in human CSF which augmented inhibitory GABA signaling *in vitro*—though the exact substance is still yet to be determined. Furthermore, flumazenil (a GABAergic benzodiazepine receptor antagonist) normalized vigilance in a subset of these patients [83•]. The same group has also studied the macrolide antibiotic clarithromycin in enhancing vigilance in hypersomnia patients, presumably via pharmacological antagonism of GABA-A receptors. A retrospective review of clarithromycin treatment in 53 patients with this GABA-related hypersomnia showed 64 % reported improvement in daytime sleepiness [84]. These findings suggest that GABA may play a role in the promotion of sleep in pathological hypersomnia. However, as mentioned in the “[Neurological Disorders of Arousal](#)” section, patients with primary hypersomnia can also have low levels of CSF histamine, although this remains somewhat controversial [34, 85]. Therefore, the pathophysiology of hypersomnia is likely to be complex and multifactorial in nature.

REM Sleep

Muscle Atonia The core circuits that generate the REM sleep state are located in the brain stem. The most striking manifestation of REM sleep is the coincidence of widespread brain activation with somatic muscle atonia. Several of the waking neuronal arousal systems are silent during REM sleep, with activation of wake-REM active GLU and ACH neurons supporting brain activation during REM sleep. At the level of motor neurons in the spinal cord, REM sleep muscle atonia is the result of GABA and glycine-mediated postsynaptic inhibition, indicated by motoneuron hyperpolarization and episodic inhibitory postsynaptic potentials (IPSPs). Sources of inhibition are spinally projecting GABA/glycine neurons in the ventromedial medulla and glycinergic spinal interneurons. Hyperpolarization and episodic IPSPs have been recently described in hypoglossal motoneurons during REM sleep [86]. Disfacilitation of brain stem and spinal cord motoneurons contributes to REM sleep atonia due to loss of excitatory inputs from 5HT, NA, and HCT neurons. REM sleep atonia is periodically interrupted by phasic muscle twitches that often accompany bursts of rapid eye movements. Phasic twitches are associated with GLU-mediated excitatory postsynaptic potentials in motoneurons and reflect activation of central motor systems during REM sleep.

REM Sleep-Generating Mechanisms An older literature, based largely on electrophysiological and neuropharmacological studies in cats, emphasized ACH neurons in the PPT/LDT and their reciprocal inhibitory interactions with monoaminergic neurons as being critical for the switching between non-REM to REM sleep [70]. Recent findings in rodents emphasize interactions among GABAergic and glutamatergic neurons in the caudal midbrain and pons. A population of “REM on” neurons (i.e., neurons with elevated discharge during REM sleep compared to waking and NREM sleep) is located ventral to the LC [7, 87, 88]. This part of the rostral pons has been variously called subcoeruleus nucleus, pontine inhibitory area, and peri-LC α , but the sublateralodorsal tegmental nucleus (SLD) is currently most widely used. In addition to the SLD, expression of c-Fos during augmented REM sleep in rats has identified REM on neurons in the precoeruleus region and medial parabrachial nucleus [89, 90]. REM off GABAergic neurons in the ventrolateral periaqueductal gray (vIPAG) and lateral pontine tegmentum (LPT) inhibit the activity of REM on neurons in the SLD [91]. One model of NREM-REM switching involves reciprocal inhibitory interactions among GABAergic vIPAG/LPT neurons and GABAergic REM on neurons in the SLD [65]. The ventral SLD contains glutamatergic REM on neurons that project to GABA/glycine neurons in the ventromedial medulla and spinal cord and regulates muscle atonia of REM sleep [88, 91]. Cell-specific lesions or inhibition of the vIPAG/LPT increases the amount of REM

sleep and produce cataplexy-like periods of atonia during wakefulness, and SLD lesions and medial medulla lesions can result in REM sleep without atonia [88]. Another set of GLU neurons in the PB/PC nuclei project to the basal forebrain and regulate EEG phenomena of REM sleep. Current evidence suggests that most ACH neurons in the LDT/PPT are wake/REM active [7] and, through ascending projections, promote thalamocortical desynchrony during REM sleep. ACH in the LDT/PPT and monoaminergic neurons in the TMN, DRN, and LC appear to be modulatory of the core REM-generating circuitry, with ACH signaling facilitating REM sleep and monoaminergic neuronal activation suppressing REM sleep.

Hypothalamic Modulation of REM Sleep Descending inputs from sleep and arousal regulatory hypothalamic neuronal systems are sources of modulatory control of REM sleep circuits. The HCT peptides have REM-suppressing effects. HCT receptor antagonists augment REM sleep [27]. Optogenetic activation of HCT neurons during either NREM or REM sleep evokes waking [92]. HCT neurons target key nodes in brain stem REM sleep circuitry including neurons in the vIPAG/LPT, DRN, and LC. Activation of HCT neurons during waking suppresses manifestations of REM sleep, and REM-generating circuits are disinhibited during NREM sleep when HCT neuronal activity is minimal.

Neurons expressing the inhibitory peptide melanin-concentrating hormone (MCH) are localized in the lateral hypothalamus, zona incerta, and dorsomedial hypothalamus [93]. Included among the targets of MCH neurons are LC, DRN, and vIPAG. MCH neurons also express GABA. MCH neurons are nearly silent during waking, seldom discharge during NREM sleep, and are most active during REM sleep [94]. Expression of c-Fos in MCH neurons is increased during REM-enriched sleep following prolonged REM sleep deprivation [95]. Optogenetic activation of MCH neurons during NREM sleep significantly increases the probability of transitions to REM sleep [96]. Mechanisms of MCH REM sleep enhancement involve disinhibition of SLD REM on neurons via MCH/GABA inhibition of REM off neurons in the TMN, vIPAG, DRN, and LC [93, 96].

GABAergic neurons in the VLPO target many of the REM off neuronal populations that impact the SLD, including the vIPAG, DRN, and LC [63]. The majority of VLPO neurons exhibit elevated discharge during both NREM and REM sleep, and a population of REM on neurons is located in the dorsal extended VLPO [57, 61•]. Similar to MCH neurons, inhibitory effects of VLPO neurons on the REM off neuronal populations facilitate REM sleep onset and maintenance through disinhibition of the SLD.

Neurological Disorders of REM Sleep

Several clinical disorders are associated with REM sleep behavior disorder (RBD), which involves loss of REM sleep muscle atonia. The most common neurological disorders include the alpha-synucleinopathies, such as PD and “Parkinson’s plus” syndromes like dementia with Lewy bodies (DLB) and multiple systems atrophy. The neuropathology in these synucleinopathies largely supports findings in animal models that suggest the SLD and other brain stem regions are critical for normal REM sleep atonia.

Neuroimaging studies show that damage within or near the pontine tegmental regions (which includes the SLD) is associated with RBD symptoms [97–99]. A recent imaging study in 36 human subjects with PD showed that the degree of neuromelanin signal (a surrogate marker for neuronal integrity) in the SLD strongly predicted REM sleep without atonia in PD patients with RBD, but not in PD patients without RBD [100]. Furthermore, pathological neurodegeneration characteristic of PD and Parkinson’s plus syndromes (i.e., Lewy bodies, gliosis, neuronal loss) is frequently observed in or near the subcoeruleus, gigantocellular reticular nucleus, and pedunculopontine nucleus [101, 102]. These findings support data from animal models implicating the SLD and other brain stem regions in RBD.

Conclusions

Basic research using animal models of arousal and sleep mechanisms continues to identify novel neuroanatomical circuits controlling sleep and wakefulness in human clinical disorders. Arousal mechanisms include histamine (HA), serotonin (5HT), noradrenaline (NA), acetylcholine (ACH), glutamate (GLU), dopamine (DA), and hypocretin/orexin (HCT) as neurotransmitters acting within the basal forebrain, hypothalamus, and brain stem. Clinical disorders of arousal including narcolepsy, Alzheimer’s disease, Parkinson’s disease, and traumatic brain injury appear to involve HCT and possibly histamine as common mechanisms related to excessive daytime sleepiness. NREM sleep-promoting pathways largely involve GABA and galanin in the VLPO and MnPO and show striking similarity between phenotypes in rodent models and human disorders such as Alzheimer’s disease and primary hypersomnia. REM sleep-promoting pathways, especially those involving the SLD of the pons, also show parallels between rodent models of REM muscle atonia and human RBD associated with Parkinson’s disease and other synucleinopathies. In conclusion, our understanding of the basic neurobiology of arousal and sleep-promoting systems continues to evolve with the use of animal models to inform further studies on human clinical sleep disorders and vice versa.

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Compliance with Ethics Guidelines

Conflict of Interest Miranda M. Lim and Ronald Szymusiak declare that they have no conflict of interest.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

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