



Plant-mediated bio-synthesis of silver–montmorillonite nanocomposite and antibacterial effects on gram-positive and -negative bacteria

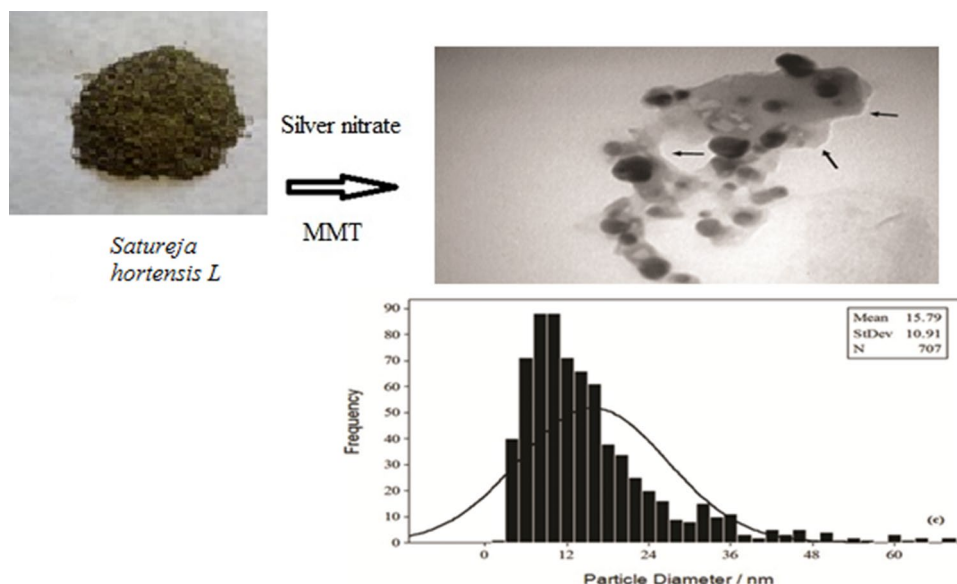
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Abstract

In this project, facile, rapid biosynthesis and characterization of silver–montmorillonite (MMT) nanocomposite is carried out at ambient temperature. Silver nanoparticles (Ag-NPs) were prepared by water extract of the plant [*Satureja hortensis* (L)] as the reducing agent and MMT as interlamellar space for controlling the size of Ag-NPs. MMT was sonicated in the aqueous AgNO_3 solution and Ag^+ ions were reduced to Ag^0 using water extract of *Satureja hortensis* (L). The nanocomposite was evaluated using ultraviolet–visible spectroscopy (UV–Vis), X-ray diffraction (XRD), scanning electron microscopy (SEM) and transmission electron microscopy (TEM). TEM study showed the formations of nanocomposite using water extract of *Satureja hortensis* L in the range of 4.88–26.70 nm and an average particles size of 15.79 nm. In addition, XRD studies indicated that the particles have a face centred cubic (FCC) structure. The nanocomposite showed antibacterial effects against bacteria.

Graphical abstract



Keywords Antibacterial effects · Montmorillonite · Nanocomposite · *Satureja hortensis* L · Transmission electron microscopy

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Introduction

Nanoparticles (NPs) synthesis have recently attracted much attentions; therefore, the synthesis of nano silver (Ag-NPs) has been a subject of great interest in the past several years [1]. Ag-NPs have good electrical, catalytical and medical applications. Because Ag-NPs do not have long life time in aqueous media and may agglomerate fast, to prevent this problem, NPs can be go between the MMT internal spaces or on its external surfaces [2]. So, the Ag-NPs can go onto MMT layers as a desired process to control the particle size. In particular, the interlayer space of MMT can be employed for the synthesis of metal NPs as discussed in our previous works [3–5]. The synthesis of TiO_2 –montmorillonite nanocomposites into the interlayers of montmorillonite has also been successfully carried out [6]. Other research showed that Ag-NPs were prepared by chemical reduction into the interlayer of montmorillonite [7]. Silver–montmorillonite–starch nanocomposites were also synthesized using UV irradiation as the reducing agent, starch as the stabilizer and montmorillonite as a solid support [3]. Synthesis and evaluation of Ag-NPs by UV irradiation to silver nitrate in the interlayer of MMT using no reducing agent or heat process was also reported in other works [8]. Also platinum nanoparticles were also prepared using MMT and catalytic activities were evaluated [9]. The properties of biosynthesized silver nanoparticles were evaluated in other research [10].

Biological systems present in water extract of plants is good candidate to be used for the bio-reduction of Ag^+ ions to Ag-NPs as a green chemical, via a cost-effective and environmentally friendly route [11]. In our previous work, Ag-NPs were successfully prepared with an average size of 15 ± 7.4 nm by the water extract of this plant at ambient temperature without any support [12]. In this project, rapid and eco-friendly bio-reduction method for the synthesizing of Ag-NPs is reported. In this method, controlled and one pot reduction without any hazardous products is utilized. Ag-NPs were prepared into the lamellar space of MMT using water extract of *Satureja hortensis* L in ambient temperature. MMT was used to prevent nanoparticles from aggregation. The antibacterial effect of the nanocomposite was evaluated against *Escherichia coli* (Gram-negative) and *Staphylococcus aureus* (Gram-positive) bacteria by the minimum bactericidal concentration (MBC) method.

Experimental

Materials and methods

Chemicals were purchased from the Merck, Fluka and Aldrich chemical companies. All reagents were prepared as analytical grade. Double distilled water was also used in all experiments. Nicolet 370 FT-IR spectrometer, Philips X-ray diffraction, cam scan MV2300 as scanning electron microscopy (SEM), Philips EM208 transmission electron microscope (TEM) were used in this project. The UV–visible spectra were also obtained using UV Bio–TEK UV–visible spectrophotometer from 300 to 800 nm. The antibacterial tests were characterized using the minimum bactericidal concentration (MBC) method [13, 14].

Preparing the water extract of plant

Fresh plants were collected from Malard region of Iran in September 2017. Silver nitrate (99.80%), used as the Ag^+ source, was purchased from Merck chemical co. and the MMT from Fluka chemical co. Kanamycin (antibiotic) was supplied by Duchefa Biochemie and vancomycin (antibiotic) was obtained from Sigma Aldrich chemical co. Fresh leaves of *Satureja hortensis* L were dried for a week in shade. The *Satureja hortensis* L dried leaves were extracted in water and used for reduction of Ag^+ ions to Ag^0 . For this purpose, 30 g of dried leaves of the plant was added to 100 mL of water and the mixture was heated for 20 min at 40 °C. The obtained water extract was centrifuged at 3000 rpm for 20 min.

Green biosynthesis of silver nanoparticles

0.01 g of clay was added in 50 mL of distilled water in 2 h. Clay solution was poured to 100 mL of 0.01 mM of AgNO_3 solutions to prepare the Ag/MMT nanocomposite. The prepared solution was then added to 40 mL of plant extract at ambient temperature for 48 h as mixing.

Results and discussion

UV–Vis absorption measurements

In Fig. 1, the UV–Vis spectra of Ag-NPs synthesized by the extract of *Satureja hortensis* L, clay and plant extract are illustrated (a–c). The SPR bands for silver particles are shown in the range of 350–420 nm. These peaks are due to the Ag-NPs below 30 nm. The peak near 400 nm is due to the formation of Ag/MMT nanocomposite in nanometer



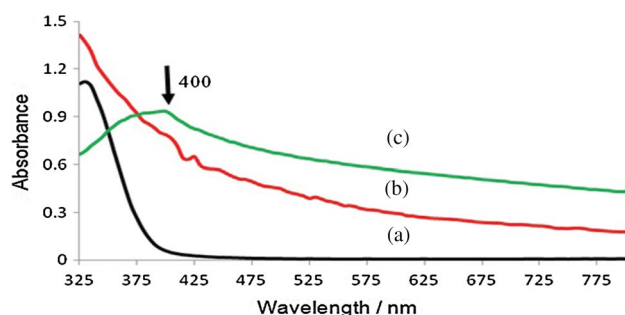


Fig. 1 UV–Vis spectra: **a** plant extract of *Satureja hortensis* L, **b** (MMT) and **c** Ag/MMT nanocomposite suspension in water after 48 h

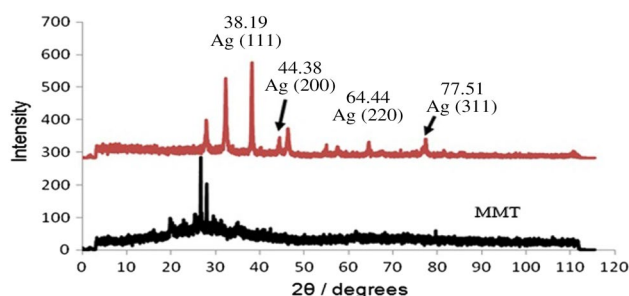


Fig. 2 XRD patterns of nano clay and Ag/MMT nanocomposite synthesized by *Satureja hortensis* L water extract after 48 h

size which is in correlation with TEM results as 15.79 nm [14, 15].

X-ray diffraction (XRD) patterns

In Fig. 2 the XRD patterns of the pure montmorillonite and the synthesized Ag/MMT nanocomposite are shown. In the Ag/MMT nanocomposite, the XRD peaks at 2θ of 38.19° , 44.38° , 64.44° and 77.51° correspond to (111), (200), (220) and (311) planes of the face centered cubic (FCC) of nanocomposite, respectively. The mean particle sizes of nanocomposites were characterized by the Debye–Scherer Eq. 1:

$$d = \frac{K\lambda}{\beta \cos \theta} \quad (1)$$

The average sizes of nanocomposites were calculated to be about 16 nm.

SEM and TEM observations

SEM image of Ag/MMT nanocomposite, prepared by green reduction of AgNO_3 in montmorillonite using water extract of *Satureja hortensis* L at room temperature, is shown in Fig. 3. SEM images were used to analyze the morphology of Ag/MMT nanocomposite. The shape of the

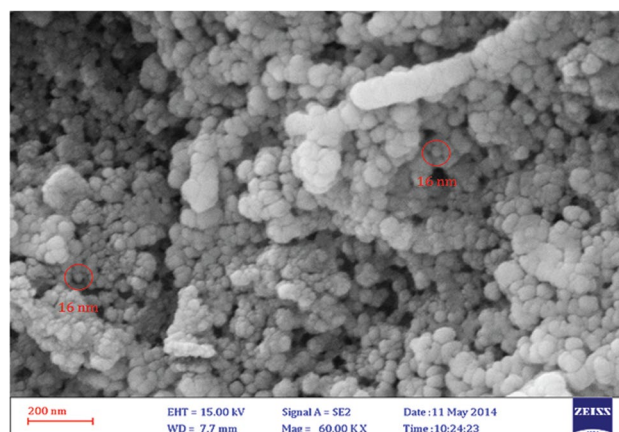


Fig. 3 SEM images of Ag/MMT nanocomposite

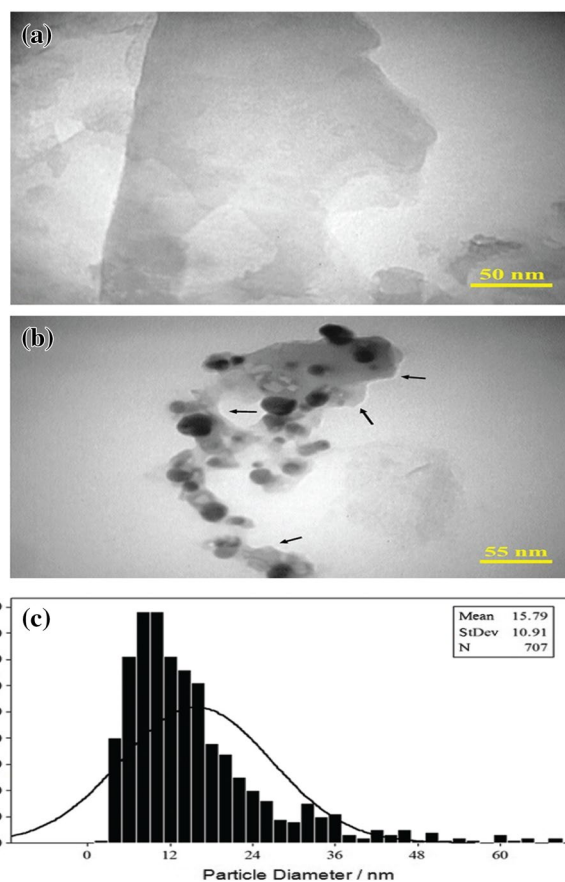


Fig. 4 TEM image: **a** montmorillonite, **b** Ag/MMT nanocomposite and **c** corresponding size of Ag/MMT nanocomposite after 48 h

particles was found to be spherical and their average size was about 16 nm. Because of the structure of clay and the influence of silver ions onto these layers, the data confirm the formation of small silver NPs.



The formation of Ag/MMT nanocomposite suspension was confirmed by the TEM image. Figure 4a, b show the TEM images of montmorillonite and nanocomposite, respectively, and Fig. 4c shows the size distribution of the Ag/MMT nanocomposite suspension. TEM image and the size distribution of Ag/MMT nanocomposite suspension show the mean diameter and 15.79 ± 10.91 nm standard deviation of the nanocomposite for particles prepared in the montmorillonite layers. In addition, the TEM image confirmed the presence of a layer surrounding the nanocomposite, which prevented the agglomeration of the particles. These layer surroundings are depicted in the figure with black arrows.

FT-IR spectroscopy

In Fig. 5 the FT-IR spectra of clay, plant water extract and nanocomposite are shown. In MMT, the vibration band at 3634 cm^{-1} corresponds to O–H stretching and the peak at 3440 cm^{-1} is related to the interlayered O–H stretching. The interactions between MMT and Ag-NPs are indicated by the peak at 3440 cm^{-1} . This peak is for the interaction between OH groups of clay layers and the positive charge of the silver

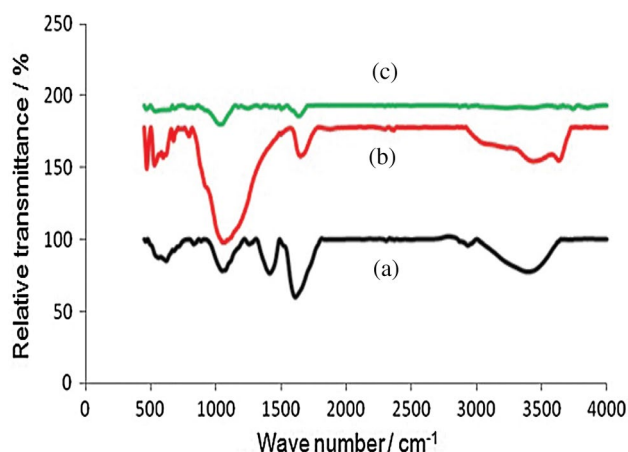


Fig. 5 FT-IR spectra: **a** plant extracts of *Satureja hortensis* L., **b** montmorillonite and **c** Ag/MMT nanocomposite after 48 h

NPs [9]. Also FT-IR was used to evaluate the biomolecules are potentially present in extract and can act as reducing and capping of the Ag-NPs, which can be attributed to the biomolecules consisting –OH functional groups.

Antibacterial tests

The antibacterial tests of Ag/MMT nanocomposite (which is purified by centrifugation) and Kanamycin and Vancomycin antibiotics against *Escherichia coli* and *Staphylococcus aureus* were successfully carried out [15]. The results showed that the optimum concentration was as shown in Table 1.

Conclusions

In summary, Ag/MMT nanocomposites were successfully obtained by a rapid, eco-friendly and biological process without using any toxic chemicals as the reducing agent at ambient temperature and their antibacterial properties were evaluated. The size of nanocomposite particles was between about 15 ± 7.4 nm. Ag/MMT nanocomposite was also confirmed by UV–Visible absorption spectra, TEM, SEM images. The TEM image showed that the Ag/MMT nanocomposites were of spherical shape and X-ray result confirmed that the obtained Ag/MMT nanocomposites have a face centered cubic crystal structure. By size and shape controlling with MMT as inter layer, the prepared nanocomposite has good shape as spherical and prevented the agglomeration; so the antibacterial effects are improved. The antibacterial tests of Ag/MMT nanocomposites on *Escherichia coli* (gram-negative) and *Staphylococcus aureus* (gram-positive) bacteria by the minimum bactericidal concentration (MBC) method were successfully carried out and the results showed that Ag/MMT nanocomposite had strong antibacterial properties in comparison with kanamycin and vancomycin.

Table 1 Values of concentrations of antibiotics and nanocomposite against two bacteria

Compound	Variable		
	(MBC) dilution	Initial concentration ($\mu\text{g mL}^{-1}$)	Effective concentration ($\mu\text{g mL}^{-1}$)
Kanamycin against <i>Escherichia coli</i> bacteria	1:32	1000	31.25
Vancomycin antibiotic against <i>Staphylococcus aureus</i> bacteria	1:64	256	4
Silver–montmorillonite nanocomposite against <i>Escherichia coli</i> bacteria	1: 4096	792	0.193
Silver–montmorillonite nanocomposite against <i>Staphylococcus aureus</i> bacteria	1:4096	792	0.193



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