



Two New Alkaloids from *Fusarium tricinctum* SYPF 7082, an Endophyte from the Root of *Panax notoginseng*

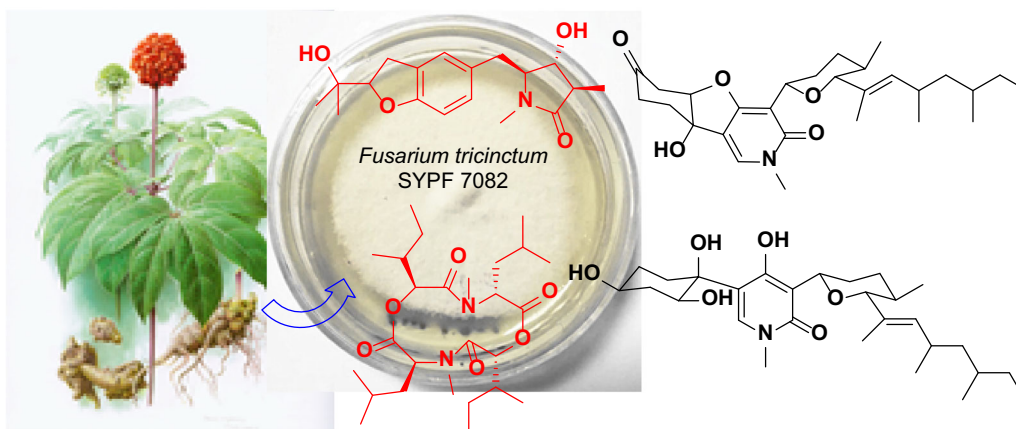
Wen-Jie Sun^{1,2} · Hong-Tao Zhu¹ · Tian-Yuan Zhang³ · Meng-Yue Zhang³ · Dong Wang¹ · Chong-Ren Yang¹ · Yi-Xuan Zhang³ · Ying-Jun Zhang^{1,4} 

Received: 18 April 2018 / Accepted: 28 May 2018 / Published online: 18 June 2018
© The Author(s) 2018

Abstract

Panax notoginseng (Araliaceae) is a famous traditional Chinese medicine mainly cultivated in Yunnan and Guangxi provinces of China. Two new alkaloids, rigidiusculamide E (**1**) and [-(α -oxyisohexanoyl-*N*-methyl-leucyl)₂]- (**2**), together with two known ones, (–)-oxysporidinone (**3**) and (–)-4,6'-anhydrooxysporidinone (**4**) were isolated from the mycelia culture of *Fusarium tricinctum* SYPF 7082, an endophytic fungus obtained from the healthy root of *P. notoginseng*. Their structures were determined on the basis of extensive spectroscopic analyses. Compounds **1–4** were tested for their inhibitory effects against NO production on Murine macrophage cell line, and the new compound **2** showed significant inhibitory activity on NO production with the IC₅₀ value of 18.10 ± 0.16 μ M.

Graphical Abstract



Electronic supplementary material The online version of this article (<https://doi.org/10.1007/s13659-018-0171-0>) contains supplementary material, which is available to authorized users.

✉ Yi-Xuan Zhang
zhangyxzsh@163.com

✉ Ying-Jun Zhang
zhangyj@mail.kib.ac.cn

¹ State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201, People's Republic of China

² University of Chinese Academy of Sciences, Beijing 100049, People's Republic of China

³ Shenyang Pharmaceutical University, Shenyang 110016, People's Republic of China

⁴ Yunnan Key Laboratory of Natural Medicinal Chemistry, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming, People's Republic of China

Keywords *Fusarium tricinctum* SYPF 7082 · Endophytic fungus · Alkaloids · *Panax notoginseng* · Inhibition on NO production

1 Introduction

Panax notoginseng (Burk.) F. H. Chen (Araliaceae), known as Sanqi or Tianqi in China, is a famous traditional Chinese medicine [1], with a broad spectrum of pharmacological effects, e.g., anti-atherosclerotic [2], hemostatic and wound healing [3], antioxidant [4], anti-inflammatory [5], hypoglycemic and anti-hyperlipidemia [6], neuroprotective [7], and anti-tumor [8] activities. The plant has been cultivated and domesticated for approximately 400 years, mainly in Yunnan and Guangxi provinces, China. Continuous cultivation of *P. notoginseng* in the same field will lead it to be attacked vulnerably by various soil-borne pathogens, like fungi, bacteria and nematodes [9]. The rhizospheric and endophytic fungal communities are considered not only of vital importance for plant health and soil fertility, but also to have positive effects on plant resistance to diseases and insects. These factors might be useful for the biological control of continuous cropping of *P. notoginseng* [10].

Fusarium species, a group of filamentous fungi with a number of plant pathogens in it [11], are widely distributed in soil, plants and plant-products. The secondary metabolites of which could be great resources for finding new compounds with a variety of biological activities [12]. For example, previous studies on *F. tricinctum* led to the identification of neosolanol monoacetate and visoltricin from the strains of field-loss peanuts [13] and wheat kernels [14], and tricinoic acid and tricindiol, enniatins and fusarielins, and fusartricin from the endophytic fungi from *Rumex hymenosepalus* [15], *Aristolochia paucinervis* [16, 17], and *Salicornia bigelovii* [18], respectively.

During the research on the formation mechanism of continuous cropping obstacles of *P. notoginseng*, two new alkaloids, rigidiusculamide E (**1**) and [$-(\alpha$ -oxyisohexanoyl-*N*-methyl-leucyl) $_2$ -] (**2**), together with two known ones (**3** and **4**) were identified from the mycelia culture of *F. tricinctum* SYPF 7082, an endophytic fungus isolated from the healthy root of *P. notoginseng*. Their structures were determined by extensive spectroscopic analyses. Moreover, the inhibitory activities of compounds **1–4** against NO production in Murine macrophage cell line were evaluated. This paper describes the isolation, structure elucidation and results of bioassay.

2 Results and discussion

The EtOAc extract of the mycelia culture of *F. tricinctum* SYPF 7082, isolated from the root of *P. notoginseng* was applied to repeated column chromatography (CC) over MCI-gel CHP20P and silica gel, followed with semi-preparative HPLC, to afford four alkaloids (**1–4**) (Fig. 1). Two of them, **1** and **2** were new compounds.

Rigidiusculamide E (**1**), a colorless oil, had a molecular formula of C₁₈H₂₅NO₄ on the basis of HRESIMS (m/z 342.1673 [M+Na]⁺, calcd. 342.1676) and NMR data (Table S1), requiring seven degrees of unsaturation. The IR spectrum showed the presence of hydroxyl group (3419 cm⁻¹), amide (1669 cm⁻¹) and benzene ring (1492 and 1442 cm⁻¹). The ¹³C NMR and DEPT data of **1** exhibited 18 carbon resonances assignable to four methyls (δ_C 8.6, 24.4, 26.6, 27.9), two methylenes (δ_C 30.9, 32.5), four methines (δ_C 42.7, 64.5, 69.1, 90.0), one oxygenated quaternary carbon (δ_C 72.0), one carboxylic carbon (δ_C 176.2), and six aromatic carbons [δ_C 109.2 (CH), 126.0 (CH), 128.9 (CH), 128.0 (C), 129.2 (C), 158.6 (C)] arising from a tri-substituted benzene ring. The ¹H NMR spectrum displayed the existence of three singlet [δ_H 1.15, 1.28, 2.80 (each s)] and one doublet (δ_H 1.13, d, J = 7.2 Hz) methyls, and a set of aromatic protons [δ_H 6.66 (1H, d, J = 8.4 Hz), 6.98 (1H, d, J = 8.4 Hz), and 7.06 (1H, s)] from an ABX coupled system (Table 1). These NMR features are closely related to those of rigidiusculamide D, an alkaloid reported previously from *Albonectria rigidiuscula* [19]. However, instead of the oxygenated quaternary C-3 (δ_C 75.1, qC) in rigidiusculamide D, an aliphatic methine (δ_C 42.7, CH) was present in **1**, suggesting that compound **1** was an analog of rigidiusculamide D without oxygen-substitution at C-3 position.

The structure of **1** was further confirmed by 2D NMR experiments. In the ¹H-¹H COSY spectrum, three partial structures of -C₍₁₄₎H₃-C₍₃₎H-C₍₄₎H(O)-C₍₅₎H(N)-C₍₆₎H₂-, -C₍₁₁₎H=C₍₁₂₎H- and -C₍₁₅₎H₂-C₍₁₆₎HO- were observed. The HMBC correlations from H₂-15 (δ_H 3.10) to C-8 (δ_C 126.0), C-9 (δ_C 128.0), and C-10 (δ_C 158.6), and from H-16 (δ_H 4.53) to C-10 indicated the presence of dihydrobenzofuran ring. Moreover, HMBC correlations from the *N*-methyl protons at δ_H 2.80 to C-2 (δ_C 176.2) and C-5 (δ_C 64.5), from H₃-14 (δ_H 1.13), H-3 (δ_H 2.35) and H-4 (δ_H 3.94) to C-2 revealed the existence of 3-methylpyrrolidin-2-one moiety. Other HMBC correlations (Fig. 2) from H₂-6 (δ_H 2.84) to C-4 (δ_C 69.1), C-5 (δ_C 64.5), C-7 (δ_C 129.2), C-8 (δ_C 126.0), and C-12 (δ_C 128.9) confirmed the planar structure of **1** as shown in Fig. 1, with 4-hydroxy-1,3-

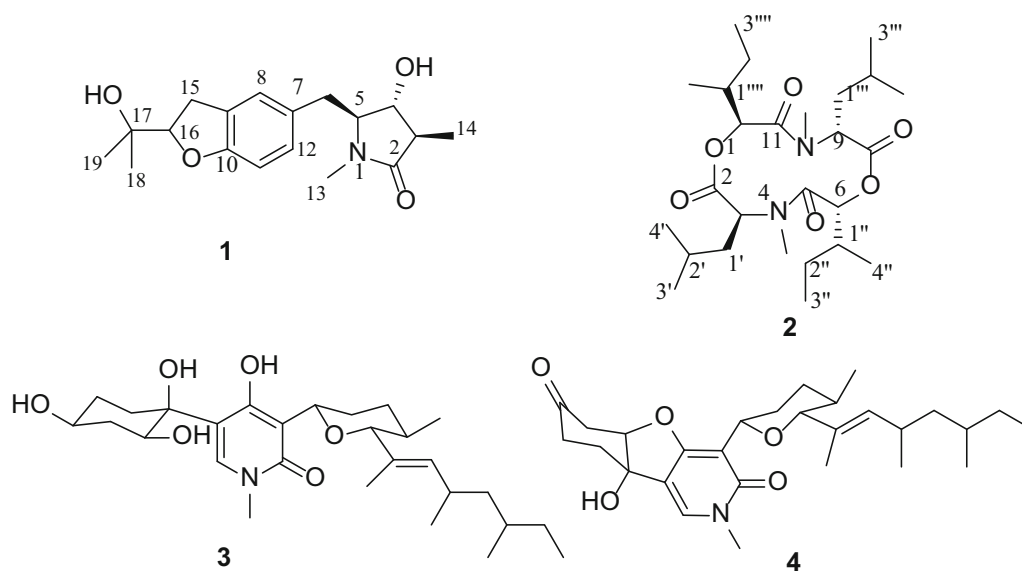


Fig. 1 Structures of compounds **1–4** from *F. tricinatum* SYPF 7082

dimethylpyrrolidin-2-one ring and a dihydrobenzofuran in molecule.

In the ROESY spectrum of **1**, correlations of H-3 with H-5 (δ_{H} 3.52, m), and of H-4 with H-6a (δ_{H} 2.95, dd, $J = 13.2, 4.2$ Hz) and H-6b indicated that H₃-14 and H₂-6 were at the same side, while H-3 and H-5 were on the opposite orientation of the 4-hydroxy-3-methylpyrrolidin-2-one ring (Fig. 2), thereby established the relative configurations of **1**. On the basis of the above evidence, the structure of **1** was deduced as shown.

$[-(\alpha\text{-Oxyisohexanoyl-}N\text{-methyl-leucyl})_2\text{-}]$ (**2**), obtained as colorless crystal, had a molecular formula of C₂₆H₄₆N₂O₆, deduced from the HRESIMS (m/z 505.3247 [M + Na]⁺, calcd. 505.3248), with five degrees of unsaturation. The IR spectrum showed the presence of carboxyl ester (1758 cm⁻¹) and amide (1657 cm⁻¹) groups. The ¹³C NMR and DEPT spectra of **2** exhibited 13 carbon resonances, arising from five methyls (δ_{C} 11.5, 16.7, 22.6, 23.6, 30.8), two methylenes (δ_{C} 26.2, 40.9), four methines (δ_{C} 26.4, 38.3, 58.0, 83.8), and two carboxylic carbons (δ_{C} 171.1, 172.2). The ¹H NMR spectrum displayed the existence of one singlet (δ_{H} 3.02, s), two doublet [δ_{H} 1.05, 1.08 (each d, $J = 6.0$ Hz)] and one triplet (δ_{H} 0.96, t, $J = 7.5$ Hz) methyls, and two oxymethines [δ_{H} 4.84 (dd, $J = 15.0, 7.2$ Hz); 5.01 (d, $J = 9.6$ Hz)] (Table 1). The above-mentioned data accounted for all the ¹H and ¹³C NMR resonances and the molecular formula suggested that **2** had a symmetrical structure. The ¹H-¹H COSY spectrum showed the existence of two partial structures, -CHO-CH-(CH₃)-CH₂-CH₃ and -CHN-CH₂-CH-(CH₃)₂ (Fig. 3). In the HMBC spectrum of **2**, correlations from *N*-methyl proton (δ_{H} 3.02) to C-3 (δ_{C} 83.8) and C-5 (δ_{C} 172.2), from H-3 (δ_{H} 5.01) to C-2, C-5 (δ_{C} 172.2), C-1' (δ_{C} 40.9) and

C-2' (δ_{C} 26.4), from H-6 (δ_{H} 4.84) to C-5, C-8 (δ_{C} 171.1), *N*-methyl (δ_{C} 30.8), C-1'' (δ_{C} 38.3) and C-2'' (δ_{C} 26.2) (Fig. 3), established the fragment structures of *N*-methyl-leucyl and α -oxyisohexanoyl moieties, and the gross structure of **2** when considering of its symmetrical structure. The ROESY correlations of *N*₄-CH₃ with H-1', H-6 and H-9 indicated that these protons on the same face of the cyclodipeptide ring, thereby established the relative configurations of **2** (Fig. 3). Therefore, the structure of **2** was determined as shown.

The known compounds **3** and **4** were identified to be (-)-oxysporidinone (**3**) [20] and (-)-4,6'-anhydrooxysporidinone (**4**) [21] by comparing their spectroscopic data with literature values. Both of them were isolated for the first time from *F. tricinatum*.

The inhibitory activities of compounds **1–4** against NO production on Murine macrophage cell line were evaluated by Griess assay [22]. Compound **2** showed inhibition of NO production with the IC₅₀ value of 18.10 ± 0.16 μM, while compounds **1**, **3** and **4** were inactive at the concentration of 25 μM.

3 Experimental Section

3.1 General Experimental Procedures

Optical rotations were measured on a HORIBA SEPA-300 high-sensitive polarimeter, and UV spectra were recorded on a Shimadzu UV2401A ultraviolet–visible spectrophotometer. Infrared spectroscopy (IR) spectra were obtained on a Bio-Rad FTS-135 series spectrometer. HRESIMS data were obtained using API QSTAR Pular-1 spectrometer. ¹H

Table 1 ^1H (600 MHz) and ^{13}C (150 MHz) NMR spectroscopic data for compounds **1–2** (in CDCl_3 , δ in ppm and J in Hz)

No.	1		No.	2	
	δ_{C}	δ_{H}		δ_{C}	δ_{H}
2	176.2, C		2	171.1, C	
3	42.7, CH	2.35, m	3	83.8, CH	5.01, d (9.6)
4	69.1, CH	3.94, dd (9.0, 4.8)	4	30.8, N-CH ₃	3.02, s
5	64.5, CH	3.52, m	5	172.2, C	
6	32.5, CH ₂	2.95, dd (13.2, 4.2) 2.84, dd (13.2, 4.2)	6	58.0, CH	4.84, dd (15.0, 7.2)
7	129.2, C		8	171.1, C	
8	126.0, CH	7.06, s	9	83.8, CH	5.01, d (9.6)
9	128.0, C		10	30.8, N-CH ₃	3.02, s
10	158.6, C		11	172.2, C	
11	109.2, CH	6.66, d (8.4)	12	58.0, CH	4.84, dd (15.0, 7.2)
12	128.9, CH	6.98, d (8.4)	1'	40.9, CH ₂	1.94, m 1.59, m
13	27.9, N-CH ₃	2.80, s	2'	26.4, CH	1.58, m
14	8.6, CH ₃	1.13, d (7.2)	3'	22.6, CH ₃	1.05, d (6.0)
15	30.9, CH ₂	3.10, dd (15.7, 9.0) 3.24, dd (15.7, 9.0)	4'	23.6, CH ₃	1.05, d (6.0)
16	90.0, CH	4.53, t (9.6)	1''	38.3, CH	1.90, m
17	72.0, C		2''	26.2, CH ₂	1.25, m 1.48, m
18	26.6, CH ₃	1.28, s	3''	11.5, CH ₃	0.96, t (7.5)
19	24.4, CH ₃	1.15, s	4''	16.7, CH ₃	1.08, d (6.0)
			1'''	40.9, CH ₂	1.94, m 1.59, m
			2'''	26.4, CH	1.58, m
			3'''	22.6, CH ₃	1.05, d (6.0)
			4'''	23.6, CH ₃	1.05, d (6.0)
			1''''	38.3, CH	1.90, m
			2''''	26.2, CH ₂	1.25, m 1.48, m
			3''''	11.5, CH ₃	0.96, t (7.5)
			4''''	16.7, CH ₃	1.08, d (6.0)

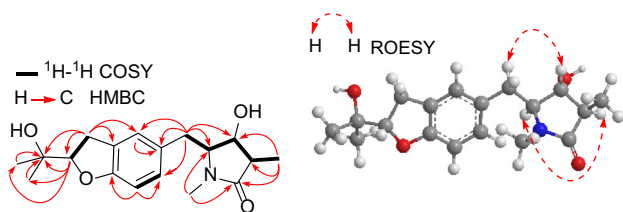


Fig. 2 Key ^1H - ^1H COSY, HMBC and ROESY correlations of **1**

and ^{13}C NMR spectra were acquired with Bruker DRX-600 spectrometer, using CDCl_3 as solvent and TMS as an internal standard. Chemical shifts were reported in units of δ (ppm) and coupling constants (J) were expressed in Hz. Column chromatography (CC) were carried out over silica

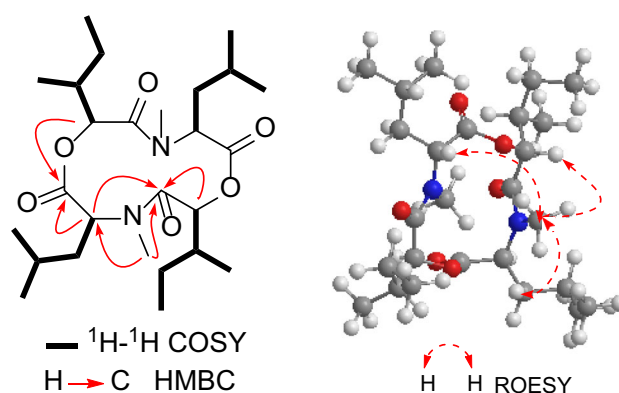


Fig. 3 Key ^1H - ^1H COSY, HMBC and ROESY correlations of **2**

gel (200–300 mesh, Qingdao Haiyang Chemical Co., Ltd., Qingdao, China) and MCI-gel CHP20P (75–100 μm , Mitsubishi Chemical Co. Ltd., Tokyo, Japan). An Agilent series 1260 (Agilent Technologies) were used for semi-preparative HPLC with an Agilent ZORBAX SB-C18 column (5 μm , 250 \times 9.4 mm), with flowing rate of 3 mL/min.

3.2 Fungal material

The fungal strain used in this work was isolated from the healthy root of *P. notoginseng*, which was collected from Wen-Shan district, Yunnan province of China (104°19'17.2"/23°31'48.9"). The RNA sequence data derived from this strain has been submitted and deposited in GenBank with the accession number MG930027. BLAST search results revealed that the isolate belongs to the genus *Fusarium* and had a close relationship (99% identity) with *Fusarium tricinctum* (KR071697). A voucher specimen (SYPF 7082) has been deposited at the School of Life Science and Biopharmaceutics, Shenyang Pharmaceutical University.

3.3 Fermentation and Isolation

The strain of *Fusarium* sp. SYPF7082 was cultivated on potato dextrose agar (PDA) at 25 °C for seven days. Fermentation was carried out in 300 Erlenmeyer flasks (250 mL) each containing 90 g rice. Sterile water (100 mL) was added to each flask, and the contents were autoclaved at 121 °C for 30 min. After cooling down to room temperature, each flask was inoculated with 20.0 mL of the spore and incubated at 25 °C for 40 days.

The fermented rice substrate was extracted repeatedly with EtOAc (3 \times 50 L), and the organic solvent was completely evaporated under vacuum to afford the crude extract (579 g). The crude extract was then suspended into water (3 L) and partitioned with *n*-hexane (3 \times 3 L) and EtOAc (3 \times 3 L), successively. The EtOAc fraction (61 g) was subjected to CC over MCI-gel CHP20P, eluted with gradient mixture of MeOH and H₂O (10:90–100:0, v/v), to give 11 fractions (Fr.1–Fr.11). Fr.10 (656 mg) was separated by silica gel CC, eluting with CHCl₃–MeOH (100:1–20:1) to give five sub-fractions (Fr.10-1–Fr.10-5). Fr.10-2 (101 mg) was purified by semi-preparative HPLC (MeCN–H₂O, 32: 68, v/v) to afford **3** (7.0 mg, t_{R} = 12.7 min) and **4** (23 mg, t_{R} = 21.6 min). Fr.10-3 (68 mg) was subjected to semi-preparative HPLC (MeCN–H₂O, 18: 82, v/v) to afford **2** (2.1 mg, t_{R} = 19.9 min). Fr.10-4 (96 mg) was applied to semi-preparative HPLC (MeCN–H₂O, 28: 72, v/v) to afford **1** (3.4 mg, t_{R} = 15.4 min).

Rigidiusculamide E (**1**): colorless oil; $[\alpha]_{\text{D}}^{25}$ –61.8 (*c* 0.03, MeOH); UV (MeOH) λ_{max} nm (log ϵ): 471 (2.15), 362 (2.28), 286 (3.42), 228 (3.90), 203 (4.45); IR (KBr) ν_{max} cm⁻¹: 3419, 2973, 2930, 1669, 1492, 1380, 1247, 1180; ¹H and ¹³C NMR (CDCl₃): see Table 1; Positive ESIMS: m/z 342 [M+Na]⁺.

[-(α -Oxyisohexanoyl-*N*-methyl-leucyl)₂-] (**2**): colorless crystal; $[\alpha]_{\text{D}}^{25}$ 8.8 (*c* 0.02, MeOH); UV (MeOH) λ_{max} nm (log ϵ): 292 (3.20), 205 (4.51); IR (KBr) ν_{max} cm⁻¹: 2963, 2932, 2878, 1758, 1657, 1456, 1370, 1177, 1144; ¹H and ¹³C NMR (CDCl₃): see Table 1; Positive ESIMS: m/z 505 [M+Na]⁺.

3.4 The Nitric Oxide Production in RAW264.7 Macrophages

Murine macrophage cell line RAW264.7 was obtained from Cell Bank of Chinese Academy of Sciences (Beijing, People's Republic of China). RAW264.7 cells were seeded in 96-well cell culture plates (1.5 \times 10⁵ cells/well) and treated with serial dilutions of the compounds with a maximum concentration of 25 μM in triplicate, followed by stimulation with 1 $\mu\text{g}/\text{mL}$ LPS (Sigma, St. Louis, MO, USA) for 18 h. Nitric oxide production in the supernatant was assessed by Griess reagents (Reagent A & Reagent B, respectively, Sigma) [22]. The absorbance at 570 nm was measured with a microplate reader (Thermo, Waltham, MA, USA). *N*^G-Methyl-L-arginine acetate salt (L-NMMA, Sigma), a well-known nitric oxide synthase (NOS) inhibitor, was used as a positive control (half maximal inhibitory concentration IC₅₀ = 39.41 \pm 2.43 μM) [23]. All the compounds were prepared as stock solutions in DMSO. The viability of RAW264.7 cells was evaluated by the MTS assay simultaneously to exclude the interference of the cytotoxicity of the test compounds.

4 Conclusions

Two new alkaloids, rigidiusculamide E (**1**) and [-(α -oxyisohexanoyl-*N*-methyl-leucyl)₂-] (**2**), together with two known ones, (-)-oxysporidinone (**3**) and (-)-4,6'-anhydrooxysporidinone (**4**), were identified from *F. tricinctum* SYPF 7082, an endo-phytic fungus isolated from the root of *Panax notoginseng*. All of them were obtained from *F. tricinctum* for the first time. The new compound **2** showed inhibition of NO production in Murine macrophage cell line with the IC₅₀ value of 18.10 \pm 0.16 μM .

Acknowledgements The authors are grateful to the staffs of the analytical and bioactivity screening groups at State Key Laboratory of Phytochemistry and Plant Resources in West China, KIB, CAS, for measuring the spectroscopic data and anti-inflammatory cytotoxicities,

respectively. This work was supported by the Major Science and Technique Programs in Yunnan Province (2016ZF001-001, 2017IB038), the Science and Technology Planning Project of Yunnan Province (2013FC008, 2015IC017) and the National Science and Technology Major Project of China (2018ZX09735001-002-002).

Compliance with ethical standards

Conflicts of interest The authors declare that there are no conflicts of interest.

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

1. T. Wang, R. Guo, G. Zhou, X. Zhou, Z. Kou, F. Sui, C. Li, L. Tang, Z. Wang, *J. Ethnopharmacol.* **188**, 234–258 (2016)
2. J.S. Fan, D.N. Liu, G. Huang, Z.Z. Xu, Y. Jia, H.G. Zhang, X.H. Li, F.T. He, *J. Ethnopharmacol.* **142**, 732 (2012)
3. C.M. White, C. Fan, J. Song, J.P. Tsikouris, M. Chow, *Pharmacotherapy.* **21**, 773 (2001)
4. Y. Zhang, L.F. Han, K.J. Sakah, Z.Z. Wu, L.L. Liu, K. Agyemang, X.M. Gao, T. Wang, *Molecules* **18**, 10352–10366 (2013)
5. S.H. Chang, Y. Choi, J.A. Park, D.S. Jung, J. Shin, J.H. Yang, S.Y. Ko, S.W. Kim, J.K. Kim, *Clin. Nutr.* **26**, 785–791 (2007)
6. Z.H. Chen, J. Li, J. Liu, Y. Zhao, P. Zhang, M.X. Zhang, L. Zhang, *Am. J. Chin. Med.* **36**, 939–951 (2008)
7. D. Jia, Y. Deng, J. Gao, X. Liu, J. Chu, Y. Shu, *Int. J. Biol. Macromol.* **63**, 177–180 (2013)
8. N.W. He, Y. Zhao, L. Guo, J. Shang, X.B. Yang, *J. Med. Food* **15**, 350–359 (2012)
9. J. Xie, Y.Y. Wu, T.Y. Zhang, M.Y. Zhang, W.W. Zhu, E.A. Gullen, Z.J. Wang, Y.C. Cheng, Y.X. Zhang, *RSC Adv.* **7**, 38100–38109 (2017)
10. Y. Tan, Y. Cui, H. Li, A. Kuang, X. Li, Y. Wei, X. Ji, *Microbiol. Res.* **194**, 10–19 (2017)
11. X.W. Zhang, D. Zhang, Zhiwu Shengli Xuebao/*Plant. Physiol. J.* **49**, 201–216 (2013)
12. M. Solfrizzo, A. Visconti, *J. Chromatogr. A* **730**, 69 (1996)
13. J.A. Lansden, R.J. Cole, J.W. Dörner, R.H. Cox, H.G. Cutler, J.D. Clark, *J. Agric. Food. Chem.* **26**, 242–244 (1978)
14. A. Visconti, M. Solfrizzo, *J. Agric. Food. Chem.* **42**, 195–199 (1994)
15. B.P. Bashyal, A.A. Leslie, *Gunatilaka. Nat. Prod. Rep.* **24**, 349 (2010)
16. J.P. Wang, A. Debbab, C.F. Hemphill, P. Proksch, *Z. Naturforsch. C* **68**, 223–230 (2013)
17. C.F. Hemphill, P. Surechatchaiyan, M.U. Kassack, M.U. Kassack, R.S. Orfali, W. Lin, G. Daletos, P. Proksch, *J. Antibiot.* **70**, 726–732 (2017)
18. J. Zhang, D. Liu, H. Wang, T. Liu, Z. Xin, *Eur. Food Res. Technol.* **240**, 805–814 (2015)
19. J. Li, S. Liu, S. Niu, W. Zhuang, Y. Che, *Pyrrolidinones from the ascomycete fungus *Albonectria rigidiuscula*.* *J. Nat. Prod.* **72**, 2184 (2009)
20. Q.X. Wang, S.F. Li, F. Zhao, H.Q. Dai, L. Bao, R. Ding, H. Gao, L.X. Zhang, H.A. Wen, H.W. Liu, *Fitoterapia* **82**, 777–781 (2011)
21. J. Zhan, A.M. Burns, M.X. Liu, S.H. Faeth, A.A. Gunatilaka, *J. Nat. Prod.* **70**, 227–232 (2007)
22. V.M. Dirsch, H. Stuppner, A.M. Vollmar, *Planta Med.* **64**, 423–426 (1998)
23. D.W. Reif, S.A. McCreedy, *Arch. Biochem. Biophys.* **320**, 170–176 (1995)