Scientific note: *Varroa jacobsoni* and *V. destructor* on hill and plains strains of *Apis cerana* in southern India

Chet Prasad BHATTA^{1,2}, M. S. REDDY³, Deborah Roan SMITH¹

¹Department of Ecology and Evolutionary Biology, University of Kansas, Haworth Hall, 1200 Sunnyside Avenue, Lawrence, KS 66045, USA

²Department of Biology, Radford University Carilion, Roanoke, VA 24013, USA

³Department of Zoology, Center for Apiculture Studies, Jnana Bharathi, Bengaluru 560 056, India

Received 8 July 2019 – Revised 11 October 2019 – Accepted 10 November 2019

Abstract – Two types of *Apis cerana* are known from India: black or hill bees and yellow or plains bees, which differ in mitochondrial haplotype. We investigated whether black and yellow *A. cerana* host different species or strains of *Varroa*. *Apis cerana* and any associated *Varroa* mites were collected from 21 colonies (5 with *Varroa*), in Bengaluru, India, and surrounding areas. Bees were scored for color and mitochondrial haplotype using sequences of the cytochrome oxidase I and II genes and the intervening leucine tRNA gene and non-coding region. Mites were scored for species and strain using partial sequence of cytochrome oxidase I. We found both *V. destructor* with a haplotype similar to the previously reported Sri Lanka1 and a novel haplotype of *V. jacobsoni*. Contrary to our expectations, we found no association of mite species and bee mitochondrial haplotype.

mitochondrial DNA / host-parasite coevolution / *Apis cerana* / *Varroa* / India / *Varroa jacobsoni* / *Varroa destructor*

Anderson and Trueman (2000) recognized *Varroa destructor* Anderson and Trueman 2000 and *V. jacobsoni* Oudemans, 1904 as separate, cryptic species with broadly allopatric geographic and host ranges. Two genetic strains of *V. destructor* from northeast Asia, the Japan and the Korea strains, have successfully made the transition from the original host, *Apis cerana* Fabricius, 1793 to the novel host *Apis mellifera* Linnaeus, 1758 and become one of the most serious threats to *A. mellifera* beekeeping. *V. destructor*'s impact on *A. mellifera* has led to extensive study of geographically structured genetic variation in both *V. destructor* and *V. jacobsoni*.

Geographic structuring of mtDNA variation in *V. destructor* and *V. jacobsoni* populations is broadly congruent with the geographic structuring of mtDNA variation in *A. cerana* (Anderson and Trueman 2000; Navajas et al. 2010; Smith and Hagen 1996; Smith 2011;

Warrit et al. 2006; Zhou et al. 2004). *V. jacobsoni* parasitizes southern lineages of *A. cerana*, primarily those occupying lands on the Sunda shelf, which were intermittently united to the mainland during Pleistocene glacial periods. This includes Indonesia, peninsular and Bornean Malaysia, peninsular Thailand south of ~11° N, and probably adjacent areas (Smith and Hagen 1996; Smith 2011). *Varroa destructor* is found on lineages of *A. cerana* across most of mainland Asia, including Korea, Japan, China, and Thailand north of ~11° N (Navajas et al. 2010).

India is home to two varieties of *A. cerana*—the yellow or plains bee and the black or hill bee (see, for example, Kapil 1956). These are characterized by differences in coloration, mating flight time (Oldroyd et al. 2006), and mtDNA haplotype (Smith and Hagen 1996). We had an opportunity to investigate *A. cerana* and *Varroa* from Bengaluru (Bangalore), India and surrounding areas March 3–5, 2014, with the goal of determining whether one or both mite species were present, the mites' mitochondrial haplotypes, and whether the hill and plain bees were host to different mites. Both the bee and the mite samples were preserved in 90% ethanol; mites were stored together in

Corresponding author: D. Smith, debsmith@ku.eduR.

Smith, debsmith@ku.edu

Handling Editor: Peter Rosenkranz

one vial with *A. cerana* workers from their host nests. These specimens were loaned to the University of Kansas Snow Entomological Collection by author MSR for purposes of identification.

We sampled 21 colonies of *A. cerana*, 5 of which contained mites. We classed bees as black, yellow, or brown (darker yellow) (1) by scoring the amount and distribution of yellow coloration on the first three

Fig. 1 Phylogenetic tree showing that indigenous Indian *Varroa* include both *V. destructor* and *V. jacobsoni*. *V. destructor* and *V. jacobsoni* samples each form well-supported, monophyletic groups. Haplotype SouthIndia1, (*V. destructor* India-Bengaluru MH449978) groups with *V. destructor*, haplotype Bengaluru1 (*V. jacobsoni* India-Bengaluru MH449979) groups with *V. jacobsoni*. Bootstrap consensus tree showing relationships among *V. destructor* and *V. jacobsoni* cytochrome oxidase I partial sequences was constructed using maximum likelihood and the HKY+G model of sequence evolution (Hasegawa et al. 1985) with 500 bootstrap replications, implemented in MEGA7 (Kumar et al. 2016). Branches with less than 80% bootstrap support were collapsed; bootstrap supports of 80% or greater shown above branches. *Varroa rindereri* and *V. underwoodi* were used to root the tree. Each sequence shown is unique and, except for *V. rindereri* and *V. underwoodi*, from *Varroa* collected from *A. cerana*. Duplicate sequences and *V. destructor* collected from *A. mellifera* were not used in tree-building. Sequences are named by species + place of collection + GenBank accession number. The two haplotypes found in this study are shown in larger boldface type.

metasomal tergites of five worker bees per nest (Ruttner 1988; Oldroyd et al. 2006), and (2) sequencing a mitochondrial fragment (comprising a portion of the COI gene, non-coding region, leucine tRNA, and a portion of the COII gene) from one or two workers from nests without or with *Varroa*, respectively (primers as in Hall and Smith 1991; PCR protocol as in Smith and Hagen 1996, Warrit et al. 2006). PCR products were prepared for sequencing using ExoSAP-IT PCR Product Cleanup Reagent as described by the manufacturer (Thermo Fisher Scientific, Waltham, MA, USA). Sequences were compared with gene sequences from known black and yellow Indian bees collected in Bengaluru in August, 1991 (Smith and Hagen 1996).

We found that most of the *A. cerana* colonies carried mtDNA haplotypes characteristic of the black or hill variety: 14 black, 6 yellow, and one abandoned nest with no bees, only free-running mites present. Workers from all 14 nests carrying black or hill variety mtDNA were also visually scored as black; those from the 6 nests carrying yellow or plains mtDNA were scored as yellow or brown.

We used primers V-51F (5'-GTAATTTGTATACA AAGAGGG-3') and V-1400R (5'-CAAT ATCAATAGAAGAATTAGC-3') to amplify a 328-bp fragment of the mitochondrial COI gene of 5–6 *Varroa* from each of the five nests with mites (see Warrit et al. 2006 for PCR conditions). This fragment is located at positions 1888 to 2215 on the complete mitochondrial genome of *V. destructor* (GenBank AJ493124.2; Evans and Lopez 2002). Sequencing and trimming yielded 280 bp of sequence corresponding to the sequences presented by Anderson and Trueman (2000).

To determine if our *Varroa* sequences were novel or had been reported previously, we compared them with *V. destructor* collected from *A. cerana* (AF106896,

AF106900, GQ379068, AF106898, AF106897, AF106901, GQ379067, AF106899), *V. jacobsoni* (GQ387679, DQ061190, DQ061188, AF106906, AF106907, AF106910, AF106910, AF106909, AF106908, AF106905, AF106902, AF106904, AF106903), and Philippine *Varroa* (AF106893, AF106894, AF106895). To assign our *Varroa* sequences to *V. destructor* or *V. jacobsoni*, we constructed a phylogenetic tree using the maximum likelihood method in Mega 7.0 (Kumar et al. 2016) and the best model of substitutions obtained for our data. We used one exemplar of each of the two distinct haplotypes obtained from our Indian mites, representative sequences of *V. destructor* and *V. jacobsoni*, and *V. rindereri* de Guzman and Delfinado-Baker, 1996 (AF107261) and *V. underwoodi* Delfinado-Baker and Aggarwal, 1987 (AF107260) as outgroups to root the tree (Anderson and Trueman 2000; Navajas et al. 2010, Warrit et al. 2006, Zhou et al. 2004). Results are shown in Fig. 1.

Both *V. jacobsoni* and *V. destructor* were present in the *A. cerana* hives we examined. The Indian *V. destructor* haplotype found here (GenBank accession # MH449978) differs by just two bases from the haplotype reported from Sri Lanka (Sri Lanka1; Anderson and Trueman 2000; GenBank Accession AF106896); we call it *V. destructor* haplotype South India1. Our *V. jacobsoni* haplotype (GenBank accession # MH449979) did not match any previously reported haplotypes in Genbank or in the literature; we call it *V. jacobsoni* haplotype Bengaluru1. Table 1 shows the mtDNA haplotypes of the mites in the five *A. cerana* hives with mites (one lacking bees). There appeared to be no correlation of mite species and bee strain; however, our sample of nests carrying mites was very small, and we could not always observe whether the mites

Table 1. *Varroa* species present in colonies of *Apis cerana* in southern India. *Apis cerana* type (black or yellow) and *Varroa* species identifications based on mitochondrial cytochrome oxidase I sequence data. Five or six mites sampled per nest. Sites 1 and 2 were small apiaries in Bengaluru on the Bangalore-Mysore road; site 3 was a roof-top home apiary, Kamakshipalya neighborhood, Bengaluru.

Nest	Collection site	Nest condition	<i>A. cerana</i> haplotype	<i>Varroa destructor</i>	<i>Varroa jacobsoni</i>
2014-11	Site 1	Adult workers and drone brood present	Black	0	5
2014-12	Site 2	Adult workers and drone brood present	Yellow	1	4
2014-19	Site 3	No workers; drone brood present	Yellow	4	1
2014-20	Site 3	Adult workers and drone brood present	Yellow	5	1
2014-21	Site 3	No bees present; mites on comb	No bees	5	0

were successfully reproducing in the nests from which they were collected. Thus, though we can say that both *V. destructor* and *V. jacobsoni* with a novel COI sequence are present on *A. cerana* in India, we cannot yet determine if the two *Varroa* species are selective regarding which type of *A. cerana* they reproduce on.

ACKNOWLEDGEMENTS

This work was supported in part by USDA grant 2010-65104-20533 to Olav Rueppell and DRS. DNA sequencing was carried out at the Idaho State University Molecular Research Core Facility, Idaho State University, Pocatello, ID, USA. We thank Dr. Annupamaa for her invaluable help in making collections.

AUTHORS' CONTRIBUTIONS

DRS conceived this research and carried out collections; CPB and DRS carried out laboratory analyses of the bee and mite specimens; MSR coordinated travel and collecting in and around Bengaluru; DRS and CPB wrote the paper and participated in the revisions of it. All authors read and approved the final manuscript.

COMPLIANCE WITH ETHICAL STANDARDS

Conflict of interest The authors declare that they have no conflict of interest.

REFERENCES

- Anderson, D.L., Trueman, J.W.H. (2000) *Varroa jacobsoni* (Acari: Varroidae) is more than one species. Exp. Appl. Acarol. 24:165-189.
- Evans, J.D., Lopez, D.L. (2002) Complete mitochondrial DNA sequence of the important honey bee pest, *Varroa destructor* (Acari: Varroidae). Exp. Appl. Acarol. 27: 69-78. <https://doi.org/10.1023/A:1021574306010>
- Hall, H.G., Smith, D.R. (1991) Distinguishing African and European honeybee matrilineages using amplified mitochondrial DNA. PNAS USA 88:4548-4552.
- Hasegawa, M., Kishino, H., Yano, T. (1985) Dating the human-ape split by a molecular clock of mitochondrial DNA. J. of Mol. Evol. 22:160-174.
- Kapil, R.P. (1956) Variation in biometrical characters of the Indian honey-bee (*Apis indica* F.), Indian J. Entomol. 18:440-457.
- Kumar, S., Stecher, G., Tamura, K. (2016) MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol. Biol. Evol. 33:1870-1874. doi:<https://doi.org/10.1093/molbev/msw054>
- Navajas, M., Anderson, D.L., de Guzman, L.I., Huang, Z.Y., Clement, J., Zhou, T., Le Conte, Y. (2010) New Asian types of *Varroa destructor*: a potential new threat for world apiculture. Apidologie 41:181-193. doi:<https://doi.org/10.1051/apido/2009068>
- Oldroyd, B.P., Reddy, M.S., Chapman, N.C., Thompson, G.J., Beekman, M. (2006) Evidence for reproductive isolation between two colour morphs of cavity nesting honey bees (*Apis*) in south India. Insectes Sociaux 53:428-434. doi:<https://doi.org/10.1007/s00040-005-0889-2>
- Ruttner, F. (1988) Biogeography and taxonomy of honeybees. Springer-Verlag, Berlin.
- Smith, D.R. (2011) Asian honey bees and mitochondrial DNA. In: Hepburn HR, Radloff SE (eds) Honeybees of Asia. Springer Verlag, Berlin, pp 69-93.
- Smith, D.R., Hagen, R.H. (1996) The biogeography of *Apis cerana* as revealed by mitochondrial DNA sequence data. J. Kans. Entomol. Soc. 69:294-310.
- Warrit, N., Smith, D.R., Lekprayoon, C. (2006) Genetic subpopulations of *Varroa* mites and their *Apis cerana* hosts in Thailand. Apidologie 37:19-30. doi:<https://doi.org/10.1051/apido:2005051>
- Zhou, T., Anderson, D.L., Huang, Z.Y., Huang, S.X., Yao, J., Ken, T., Zhang, Q.W. (2004) Identification of *Varroa* mites (Acari: Varroidae) infesting *Apis cerana* and *Apis mellifera* in China. Apidologie 35:645-654. doi:<https://doi.org/10.1051/apido:2004059>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.