ORIGINAL ARTICLE



Pomegranate seed oil extraction by cold pressing, microwave and ultrasound treatments

Aslı Gök¹ · Hilal Uyar¹ · Özge Demir¹

Received: 15 December 2023 / Revised: 14 March 2024 / Accepted: 7 April 2024 © The Author(s) 2024

Abstract

In this study, pomegranate seed oil was extracted by microwave-assisted extraction, ultrasound-assisted extraction, and cold pressing techniques. Dimethyl succinate was the solvent of microwave and ultrasound-assisted extraction methods. The optimum conditions were determined by using response surface methodology (RSM, Design Expert software version 7.0 and Box-Behnken design). The ultrasound-assisted extraction technique was found to be superior to the microwave-assisted extraction technique in terms of extraction efficiency. The maximum extraction efficiency of microwave-assisted extraction was 22.01% under the optimized conditions (liquid/solid ratio, 5/1; time, 3 min; and microwave power, 300 W). The maximum extraction efficiency obtained by ultrasound-assisted extraction was 26.31% under the optimized conditions (liquid/solid ratio, 10/1; pulse duration/pulse interval ratio, 1; temperature 60 °C; and time, 20 min). The extraction efficiencies were compared at the optimum conditions with hexane, which is the most used solvent for pomegranate seed oil extraction, and the difference was insignificant. The results of this study are very important, especially in the field of green chemistry and chemical engineering.

Keywords Pomegranate seed oil · Microwave-assisted extraction · Ultrasound-assisted extraction · Cold pressing

1 Introduction

Pomegranate (*Punica granatum L*.), belonging to Punicaceae's family, has been widely grown in the Mediterranean region. Pomegranate seeds are by-products of the pomegranate juice industry. Since the seeds are rich in essential oils

Highlights

- In this study, pomegranate seed oil was extracted by microwaveassisted extraction, ultrasound-assisted extraction, and cold pressing techniques.
- In this study, dimethyl succinate was used as a green solvent in microwave-assisted extraction and ultrasound-assisted extraction systems, and the efficiencies were compared with hexane at the optimum operating conditions.
- The main goal of this study is to optimize the process conditions for pomegranate seed oil extraction to achieve the highest efficiency.
- Gas chromatography with a flame ionization detector (GC-FID) was used to analyze the oil content in pomegranate seeds.

☑ Özge Demir ozge.demir@iuc.edu.tr

¹ Engineering Faculty, Chemical Engineering Department, Istanbul University-Cerrahpasa, Avcılar 34320, Istanbul, Turkey with several health benefits, they deserve to be used in the food and pharmaceutical industry instead of as animal feed [1]. When extracted by conventional or modern techniques, pomegranate seeds are known to have around 20% oil content by weight, depending on fruit genotypes, cultivation, geographical locations, harvesting, and climate conditions. Pomegranate seed oil (PSO) is a yellow-colored product with a slight odor. It is chiefly used in biodiesel, lubricant, and paint formulations [1]. Later on, its applications have been expanded to cosmetics, pharmaceuticals, and nutrition [2–4]. PSO is rich in essential fatty acids. When the oil content of the pomegranate seed is examined, it is seen that it consists of 65-80% by weight of conjugated fatty acids. [3]. Punicic acid (PA) (also called an omega-5 conjugated linolenic acid) is the most abundant conjugated fatty acid in the pomegranate seed [5]. PA has several health benefits that help to prevent many diseases. PA has antioxidant, antimicrobial, anti-inflammatory, anti-atherosclerotic, anticancer, anti-estrogen, immunomodulatory, nephroprotective, hepatoprotective, and neuroprotective properties [6].

Due to the increasing trend towards natural ingredients in recent years, how these ingredients are produced has begun to be scrutinized, and environmentally friendly production methods have gained importance along with the concept of sustainable environmental management. At this point, researchers are working on new and environmentally friendly solvents and production methods [7-9]. The selection of an accurate method is the most important step in the extraction to obtain the target of interest effectively and efficiently. The most popular methods for oil extraction are microwave-assisted extraction (MAE), ultrasoundassisted extraction (UAE), soxhlet extraction, cold pressing, and supercritical fluid extraction. However, some of these methods can be either time-consuming or costly and might destroy the structure of thermally sensitive compounds. Cold pressing is a mechanical technique that relies on pressure. The cold pressing method is environmentally friendly because it does not require any organic solvent. Besides, cold-pressed oils have better physiochemical properties like low peroxide value, high phenolic content, and good aroma [10]. Even so, the oil efficiency is low. Soxhlet is one of the most used oil extraction methods, but this method is outdated due to the hazardous solvent usage and low oil recovery [11]. Supercritical fluid extraction is a selective, rapid, and clean process. It does not require any solvent; therefore, no residue is left in the oil extract. Nevertheless, the capital and investment costs are the obstacles to further commercialization [12].

MAE is an effective oil extraction method from plant seeds [13–16]. The ion transmission and dipole rotation are the keys to microwave heating. Microwaves are highfrequency electromagnetic waves, the frequency of which changes between 300 MHz and 300 GHz. At this frequency range, the waves are absorbed by the polar oxygen group of water in the plant material. Although seeds are dry, they have moisture, which is the target for microwave heating. When the plant is heated, the moisture evaporates and applies pressure to the cell membrane. Ultimately, the cell membrane explodes, and consequently, bioactive compounds move out of the pores and dissolve in the solvent. MAE is a promising technology because it reduces extraction time and saves energy due to the electromagnetic field conveying heat throughout the material [17]. Factors affecting the selection of the proper solvent are the ability of the solvent to absorb microwave radiation, the interaction between the solvent and the plant, and the solubility of the sample in the solvent. Solvents with high dipole moments with a polar characteristic absorb the microwave energy better and may get heated faster. On the other hand, non-polar solvents having low dipole moments may not heat the sample effectively [18].

UAE is another alternative technique to the traditional ones. Several bioactive compounds have been extracted from plant matrixes by UAE [19–23]. The main advantages of UAE are lower operation temperatures, less solvent consumption, and lower extraction times [5]. These moderate

operating conditions reduce the energy input and protect the structural properties of thermally sensitive bioactive compounds. The mechanical vibrations induced by ultrasound waves create cavitation bubbles during the expansion and contraction cycle. The formed bubbles weaken due to the continued expansion and contraction cycle. Ultimately, the bubbles implode, bringing high energy through heat, pressure, and mechanical shear [24]. The implosion near the solid surface causes plant tissue rupture and particle break-down, releasing intercellular molecules into the solvent [25].

Hexane is chiefly used in vegetable oil extractions due to its apolar characteristic, low vapor pressure, low cost, high stability, and high solvation ability [17, 26–28]. Today, hexane still remains a popular option. However, hexane is produced from non-renewable petroleum products. It is toxic and flammable [29]. Hexane consumption is questioned due to some possible residues in the extract. Because of global concerns about environmental pollution, there is a growing demand to use environmentally friendly solvents. In this context, dimethyl succinate can be considered as a great option. Dimethyl succinate has several properties that make it profitable to use as a solvent instead of hexane. It is less toxic, less flammable, and more biodegradable than hexane [30].

The main goal of this study is to optimize the process conditions for PSO extraction to achieve the highest efficiency. In this study, dimethyl succinate was used as a solvent in MAE and UAE systems, and the efficiencies were compared with hexane at the optimum operating conditions. For MAE, the optimum operating conditions (liquid/solid ratio, time and microwave power) were determined using Box-Behnken design of response surface methodology (RSM). Similarly, for UAE, the optimum operating conditions (liquid/solid ratio, pulse duration/pulse interval ratio, temperature, and time) were determined by using Box-Behnken design of RSM. Cold pressing is used as a reference method to compare the results obtained by MAE and UAE methods. Gas chromatography with a flame ionization detector (GC-FID) was used to analyze the oil content in pomegranate seeds at the optimum extraction conditions. At the end of the paper, the oil contents and efficiencies were compared in detail with other studies reported in the literature. Here we propose a novel and green approach to obtain essential oils from PSO.

2 Materials and methods

2.1 Pomegranate seed preparation

Pomegranate seeds were purchased from a local market (Istanbul, Turkey). All the seed samples were washed with tap water and rinsed with distilled water to remove dust and impurities. Further, the seed samples were dried naturally in the air. The dried seed samples were stored in polyethylene bags at -4 °C until analysis. Before the experiments, the material was ground with a mixer mill and passed through a 60-mesh sieve to obtain homogeneous particle sizes.

2.2 MAE procedure

PSO microwave-assisted extraction was performed in a microwave laboratory oven (Milestone, NEOS-GR Bergamo, Italy) equipped with a time controller, a temperature sensor, a power regulator, and a circulating water-cooling system. The input microwave power was altered from 200 to 300 W at 2450 MHz frequency. Efforts were made to create a synergistic combination of the process variables namely microwave power, liquid/solid ratio and microwave time. Therefore, the input variables were selected based on the preliminary trial tests. During the treatments, power, temperature and time were controlled by a control panel. In the beginning, 2 g of seeds was added to the flask containing the solvent. Then, the flask was placed on the turntable plate of the microwave oven. The microwave power, time, and solid/liquid ratio entered into the Design-Expert program were determined through preliminary trials. After the extraction procedure, the mixture was centrifuged at 4000 rpm for 20 min to obtain a clear layer of solvent-oil mixture at the top of the tube. After that, the sample was filtered on a 0.45µm filter paper. The solvent was removed from the mixture by lyophilization (Virtis Advantage, SP Scientific, Suffolk, UK). Lyophilization (freeze-drying) is a process in which solvent is sublimated from the product after freezing at a specified temperature and pressure. The oil was stored in a freezer until GC-FID analysis. The extracted oil was characterized, and the oil efficiency was determined according to the formula below. The extraction procedure was repeated thrice, and the mean value was taken for the calculations to minimize measurement errors.

Oil yield(%) = m(oil extracted;mL)/m(pomegranate seed;g) \times 100 (1)

2.3 UAE procedure

Ultrasound was applied with an ultrasonic probe (Sonics VCX500) operating at 25 kHz. First, 2 g of seed powder was placed into the flask and mixed with a specified amount of the solvent, and then the extraction was achieved by applying acoustic energy with an ultrasonic probe. Attempts were made to establish a successful combination of process variables. Thus, the variables were selected after conducting preliminary trial tests. The solid/liquid ratio, pulse duration/ pulse interval ratio, temperature, and time have been determined based on the results of preliminary trials. The intervals were selected based on the possible optimal efficiencies

that the extraction system can provide. The temperature of the ultrasonic-assisted extraction system was monitored through the control panel. After extraction, the mixture was centrifuged and filtered under the same conditions with microwave treatment. After that, the solvent was removed by lyophilization and stored in the freezer until the analysis. The oil efficiency was determined as described above. The extraction procedure was repeated three times to ensure consistency, and the average value was taken for the calculation.

2.4 Cold pressing procedure

Pomegranate seed was pressed by a cold pressing machine (NF 80, Turkey) that operates at room temperature. The extract was filtered and centrifuged at 4000 rpm for 20 min to separate the oil. Then, the oil was stored until analysis. Three independent experiments were done for each analysis, and the average value was taken. Similarly, the oil efficiency was determined according to the formula above.

2.5 GC-FID analysis

The fatty acid composition of extracted oils was determined by GC-FID analysis after deriving fatty acid methyl esters (FAMEs) [26]. The chromatographic analysis was performed on a gas chromatograph (Shimadzu GC 2010 plus) equipped with a flame ionization detector and a fused silica capillary column (20 m×0.18 mm). Air was used as the carrier gas at a 1 mL/min flow rate. The samples were injected in split mode. The split ratio was 1:100; the sample injection volume was 1 µl. All samples were injected in triplicate. The injection port and detector temperature were maintained at 250 °C. The oven temperature program was operated as follows: initial temperature were 80 °C then temperature increase to 160 °C at 40 °C/min speed rate, and increase from 160 to 180 °C at 5 °C/min, finally hold isothermally constant at 185 °C for 15 min. The peaks were identified based on the retention times using commercial reference compounds (Sigma Aldrich, USA).

2.6 Statistical analyses

The Box-Behnken design of response surface methodology (Design Expert software, State-ease Inc., Minneapolis, USA, Version: 7.0) was applied for optimization purposes. RSM is used to achieve maximum response with the minimum number of experiments. RSM also examines the individual and combination effects of the factor variables. The MAE variables were liquid/solid ratio (A), time (B), and microwave power (C), while the response was the extraction yield (Y). The interactions were investigated with 16 runs. The design matrix is presented in Table 1. The UAE variables were liquid/solid ratio (A), pulse duration/pulse interval

 Table 1
 Symbols and levels for MAE optimization

Parameters	Units	Symbols	Levels	6	
Liquid/solid ratio	mL/g	А	5	7.5	10
Time	min	В	2	3	4
Microwave power	W	С	200	250	300

ratio (B), temperature (C), and time (D). The response was the extraction yield (Y). The interactions were investigated with 27 runs. The UAE design matrix is shown in Table 2. The experimental data were fitted to a second-order polynomial equation in Eq. (2).

$$Y = \beta_0 + \sum_{i=1}^{k} \beta_i x_i + \sum_{i=1}^{k-1} \sum_{j=2}^{k} \beta_{ij} x_i x_j + \sum_{i=1}^{k} \beta_{ii} x_i^2 + \varepsilon$$
(2)

Y is the extraction efficiency (%), X_i and X_j are the independent variables. β_o , β_i , β_{ii} and B_{ij} are the regression coefficients. k refers to the number of independent variables, and ε signifies the residual term. The statistical importance of the results was evaluated by the analyses of variance (ANOVA) test.

3 Results and discussion

3.1 Optimization of MAE conditions

In the MAE optimization study, the examined parameters were liquid/solid ratio, time, and microwave power (see Table 3). The extraction efficiency was analyzed by a second-order polynomial equation, and the data were fitted into Eq. (3) by multiple regression procedure.

$$Y = 12.40 - 1.44A + 2.31B - 2.78C - 1.76AB$$

- 2.90AC - 4.13BC - 2.89A² + 0.34B²
+ 0.057C² - 1.92A²B + 3.48A²C + 0.28AB² (3)

In the above polynomial equation, a positive coefficient implies a positive effect on the response, and a negative coefficient indicates an inverse relationship. The higher the

Table 2 Symbols and levels for UAE optimization

Parameters	Units	Symbols	Level	s	
Liquid/solid ratio	mL/g	A	5	7.5	10
Pulse duration/ pulse interval ratio	-	В	1	3	5
Temperature	°C	С	40	50	60
Time	min	D	20	30	40

Treatment	Liquid/solid ratio (X ₁)	Time $(\min) (X_2)$	Power (Watt) (X ₃)	Yield (%) (Y)
1	7.5	3	250	12.14
2	5	3	300	22.01
3	5	4	250	10.31
4	7.5	4	200	10.05
5	7.5	2	200	8.86
6	5	3	200	9.13
7	7.5	3	250	13.9
8	10	3	200	11.84
9	7.5	3	250	12.25
10	7.5	2	300	13.16
11	10	2	250	14.61
12	7.5	3	250	11.3
13	7.5	4	300	7.32
14	10	4	250	5.94
15	10	3	300	8.2
16	5	2	250	7.4

 Table 3
 MAE results of different treatments

coefficient in the absolute value is, the stronger the effect becomes.

ANOVA test was conducted to evaluate the statistical suitability of the derived model equation. The coefficient of determination value converging to the unity ($R^2 = 0.98$) indicated the good fit of the model to the experimental data. The adequate precision (AP = 16.38) measures the signal to noise ratio, and a ratio value greater than 4 is desirable. The coefficient of variance (% CV = 9.76) value indicated that there are no significant differences between the actual and predicted values. As seen in Table 4, the findings indicate statistically significant differences between the treatments. It is evident that the model is significant, as suggested by the model F value (F = 15.24) and the model probability value (p=0.02). The important variables (p < 0.05) were B and C from the linear coefficients, AB, AC, BC, and A²C from the interaction coefficients, and A² from the quadratic coefficients [6].

The extraction efficiency results of different treatments can be seen in Table 3. The oil efficiency range varied from 5.94 to 22.01%. The maximum efficiency was achieved by Treatment 2, while the minimum efficiency was obtained by Treatment 14. The combined effects of microwave time and liquid/solid ratio can be seen, when the results of treatment 2 and 14 are compared. It can be seen that increasing the liquid/solid ratio together with microwave power facilitated favorable interactions [31]. Thermodynamically, elevating the temperature leads to an increase in the kinetic energy of the molecules, more rapid diffusion of molecules, and thereby enhancing the extraction efficiency [32]. However, keeping the microwave duration excessively long leads to

Table 4 ANOVA results of MAE

Factors	Sum of squares	Degrees of freedom	Mean square	F value	p value
Model	216.47	12	18.04	15.24	0.0229
А	8.29	1	8.29	7.01	0.0772
В	21.34	1	21.34	18.04	0.0239
С	30.80	1	30.80	26.03	0.0146
AB	12.36	1	12.36	10.44	0.0482
AC	33.52	1	33.52	28.33	0.0130
BC	68.23	1	68.23	57.66	0.0047
A^2	33.41	1	33.41	28.23	0.0130
\mathbf{B}^2	0.46	1	0.46	0.39	0.5762
C^2	0.013	1	0.013	0.011	0.9225
A^2B	7.35	1	7.35	6.21	0.0883
A ² C	24.29	1	24.29	20.53	0.0201
AB^2	0.15	1	7.35	6.21	0.0833
Source	Sum of squares				
\mathbb{R}^2	0.9839				
$\mathbf{R_{adj}}^2$	0.9193				
AP	16.389				
% CV	9.76				
SD	1.09				
Lack of fit	7.4				

the degradation of value-added products and unnecessary energy use [7]. Therefore, optimizing process variables is crucial for both process cost and extraction efficiency [25].

Three-dimensional (3D) response surface plots were constructed to study the interactive effects of process variables on oil efficiency. Two of the three variables were changed, while the other was kept constant at the center point to examine the variables' binary effects.

The binary effect of microwave power and treatment time on the oil efficiency is shown in Fig. 1a. Increasing microwave power from 200 to 300 W enhanced the efficiency for 2 and 3 min extraction times. As known, increased microwave power results in higher temperatures in the extraction system. Due to the increment in temperature, the cell structure was damaged more effectively, so extracts could diffuse into the solvent. Moreover, at higher temperatures, the oil solubility increased and solvent viscosity decreased, which promoted the penetration of the solvent into the sample matrix [6]. However, for 4 min duration, increasing microwave power from 200 to 300 W was found to be unfavorable, which is quite likely due to the degradation and volatilization of oil with longer irradiation times [33]. At longer irradiation times, extreme temperatures might have caused severe cellular damage, and thus, impurities may have leached out of the cell and penetrated the solvent, thus decreasing efficiency [34].

Generally, increasing the liquid/solvent ratio promotes the mass transfer between two phases due to the higher concentration gradient, so the efficiency increases. A high amount of solvent can dissolve all the oil. On the other hand, a lower solvent amount would lead to non-uniform distribution and incomplete extraction. Surprisingly, PSO removal efficiency declined sharply as the liquid/solid ratio increased from 5:1 to 10:1 (see Fig. 1b and c). Apparently, excessive solvent usage limited solvent movement, and for this reason, the efficiency decreased. At this point, unnecessary solvent usage is not profitable regarding process economy and waste management [33].

Extraction time is also an important factor affecting oil efficiency. A longer irradiation time can be unfavorable after a point due to the possible degradation of bioactive compounds [33]. The increase in extraction time inversely affected PSO removal efficiency (see Fig. 1a, b). At the end of 4 min, the solvent might have volatilized, which made it difficult to interact with PSO. Besides, the same active compounds might have been absorbed on the sample surface because of the long irradiation time [35].

3.2 Optimization of UAE conditions

The treatments and the oil efficiency results are written in Table 5. PSO efficiency varied from 6.71 to 26.3%. The maximum oil efficiency (%26.3, Treatment 21) was achieved under the process conditions: 7.5 liquid/solid ratio, 3 pulse interval/pulse duration ratio, 50 °C temperature, and 20 min time. UAE conditions were optimized by

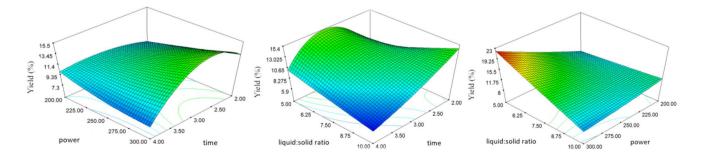


Fig. 1 Binary effects of a power and time, b liquid/solid ratio and time, and c liquid/solid ratio and power on the yield

Table 5UAE results ofdifferent treatments

Treatment	Liquid/solid ratio (X ₁)	The pulse duration/pulse interval ratio (X_2)	Temperature $(C^{\circ})(X_3)$	Time (min) (X ₄)	Yield (%) (Y)
1	7.5	1	50	40	16.7
2	5	5	40	20	17.6
3	10	5	60	20	9.66
4	10	1	60	60	17.18
5	7.5	3	60	40	25.42
6	5	5	60	20	11.39
7	7.5	3	50	40	17.65
8	7.5	3	40	40	12.14
9	5	3	50	40	13.55
10	10	1	40	20	9.77
11	10	5	40	20	14.17
12	7.5	3	50	40	17.54
13	7.5	3	50	60	24.52
14	7.5	3	50	40	17.36
15	7.5	5	50	40	8.17
16	10	1	60	20	26.31
17	5	5	40	60	9.087
18	5	1	60	20	24.57
19	10	5	60	60	14.64
20	5	1	60	60	22.3
21	7.5	3	50	20	26.3
22	10	1	40	60	9.14
23	5	1	40	60	6.71
24	10	5	40	60	20.91
25	10	3	50	40	16.77
26	5	5	60	60	20.13
27	5	1	40	20	11.28

using the design of RSM (see Table 6). A second-order polynomial equation was developed to relate the response and operational variable and written in Eq. (4). The variable D was the most significant factor because it has the highest positive coefficient value in Eq. (4).

$$Y(\%) = 18.29 + 1.61A - 4.27B + 6.64C - 0.89D + 0.23AB$$

- 1.24AC + 0.54AD - 3.71BC + 1.78BD + 0.58CD
- 3.52A² - 6.25B² + 0.098C² + 6.73D² - 0.71ABC
+ 0.90ABD - 1.86ACD + 1.36BCD + 3.66A²B
- 3.67A²C + 0.60A²D - 1.69AB² (4)

The data was analyzed by ANOVA test. The model's F and p values were 22.89 and 0.0039, respectively. Therefore, the model was found to be significant. The model's regression coefficient value (R2 = 0.99) implied that the model is potentially correct. The signal to noise ratio (AP) obtained is 16.52, which indicates an adequate signal;

hence, the model can be used to navigate the design space. The low value of the coefficient of variance (% CV = 8.28) indicated a high degree of precision.

3-D surface plots were used to study the interactive effect of the operating variables on the response. The contour plots were drawn by changing two variables while keeping the other variables constant at the center points.

Obviously, increasing the temperature from 40 to 60 °C enhanced the efficiency (see Fig. 2b, d, and f). With higher temperatures, the mass transfer from solid matrix to bulk liquid is enhanced, likely due to decreased solvent viscosity and density [36]. Nevertheless, a further increase in temperature would adversely affect PSO efficiency, such as that of Rojo-Gutiérrez et al.'s study [6]. They studied the temperature effect for PSO extraction by UAE and reported that the efficiency decreased from 45 to 60 °C. They explained this outcome with the vapor pressure effect. As known, the solvent's vapor pressure directly affects the acoustic cavitation's intensity and formation. At lower temperatures, the

Table 6ANOVA results of UAE

Factors	Sum of squares	Degrees of free- dom	Mean square	F value	p value
Model	920.80	22	41.85	22.89	0.0039
А	5.18	1	5.18	2.84	0.1675
В	36.38	1	36.38	19.90	0.0112
С	88.18	1	88.18	48.23	0.0023
D	1.58	1	1.58	0.87	0.4046
AB	0.82	1	0.82	0.45	0.5386
AC	24.78	1	24.78	13.56	0.0212
AD	4.59	1	4.59	2.51	0.1881
BC	220.57	1	220.57	120.65	0.0004
BD	50.93	1	50.93	27.86	0.0062
CD	5.40	1	5.4	2.95	0.1609
A^2	31.89	1	31.89	17.44	0.0140
\mathbf{B}^2	100.34	1	100.34	54.89	0.0018
C^2	0.025	1	0.025	0.014	0.9128
D^2	116.41	1	116.41	63.67	0.0013
ABC	8	1	8	4.38	0.1046
ABD	12.98	1	12.98	7.1	0.0561
ACD	55.55	1	55.55	30.39	0.0053
BCD	29.41	1	29.41	16.09	0.0160
A^2B	23.82	1	23.82	13.03	0.0226
A ² C	23.95	1	23.95	13.10	0.0224
A ² D	0.64	1	0.64	0.35	0.5864
AB^2	5.08	1	5.08	2.78	0.1708
Source	Sum of squares				
\mathbb{R}^2	0.9921				
R_{adj}^{2}	0.9488				
AP	16.529				
% CV	8.28				
SD	1.35				
Lack of fit	7.7				

solvent has a lower vapor pressure; therefore, less number of bubbles form. Even so, the acoustic cavitation is high, and bubbles collapse with high intensity, which disrupts the cell membrane more effectively. On the other hand, at higher temperatures, the vapor pressure of the solvent increases, and thus, more bubbles form. However, the bubbles collapse with less intensity because of the small pressure difference between the internal and external medium of the bubbles, which limits the cell tissue disruption. In addition to that, Gaulo's study on PSO claims similar results [37]. The main reason why our study revealed the opposite results might be explained by the solvent type difference. Both of the above mentioned studies used hexane as the solvent. At the same temperatures, the vapor pressure of dimethyl succinate and hexane is different; expectedly, it affects the extraction efficiency differently.

The liquid/solid ratio of 7.5 gave higher efficiencies than the ratio of 5 (see Fig. 2a, b, and c), possibly due to the increased driving force for the mass transfer between solid particles and bulk liquid in the case of a higher solvent amount [25]. However, increasing the ratio to 10 caused a sharp decline in efficiency. This might be due to the fact that increasing the contact surface area between the solvent and targeted compounds enabled decreased penetration of solvent into the sample matrix and thus decreased the extraction efficiency [34].

Within the first 20 min, the oil efficiency reached the maximum value, and no significant change was observed up to 60 min. Apparently, the cell membrane was completely damaged, and the solvent sufficiently penetrated the solid matrix in 20 min. The mechanical effect of cavitation, along with the high shear force and local turbulence generated around the bubbles, leads to the disruption of the plant matrix. Impurities may have arisen due to the extended extraction time at the 40th minute. While more oil may have been obtained at the 60th minute, 20-min duration can be determined as an appropriate extraction time, as an extended period may lead to the emergence of impurities along with the oil (see Fig. 2c, e and f) [38].

Regarding the process economy, there is no need to prolong UAE time. Besides, prolonged extraction times would cause more solid matrix degradation, so impurities like pectin, cytosol, gums, and lipid suspends would limit the oil recovery.

The pulse duration/pulse interval ratio is also a significant factor affecting the oil efficiency. The efficiency decreased as the pulse duration/pulse interval ratio increased from 3 to 5. A higher duration to interval ratio means less processing time, and less processing time is not enough to complete the mass transfer. Similar results were obtained by Goulo's study [37].

Microwave-assisted extraction achieved a peak extraction efficiency of 22.01% when operating under optimized conditions, including a liquid/solid ratio of 5:1, a 3-min extraction time, and a microwave power of 300 W. Under the specified optimal conditions (liquid/solid ratio of 10:1, pulse duration/ pulse interval ratio of 1, temperature of 60 °C, and a 20-min extraction time), ultrasound-assisted extraction yielded a maximum extraction efficiency of 26.31%. The optimum conditions determined by the program have been confirmed with experimental studies. When experiments were conducted under optimum conditions, it was observed that the yield of microwave-assisted extraction under optimum conditions was 22.5%, while the yield of ultrasonic-assisted extraction was 26.7%. Clearly, the predicted efficiencies by the program under optimum conditions closely matched the experimental results at optimum conditions. Consequently, it was observed that the program is adequate in predicting the yield under optimum conditions [38, 39].

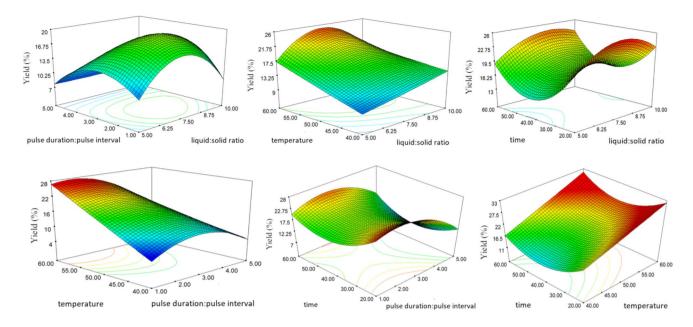


Fig.2 Binary effects of **a** pulse duration/pulse interval ratio and liquid/solid ratio, **b** temperature and liquid/solid ratio, **c** time and liquid/solid ratio, **d** temperature and pulse duration/pulse interval ratio, **e** time and pulse duration/pulse interval ratio, and **f** time and temperature on the yield

3.3 Oil content analysis

The cold pressing method gave 5% oil efficiency. Punicic acid (88.33%) ranked first, followed by oleic acid (3.68%), linoleic acid (3.22%), palmitic acid (2.19%), stearic acid (1.56%), *cis*-11-Eicosenoic acid (0.64%), linolenic acid (0.36%), and lignoceric acid (0.02%).

MAE extraction efficiency with dimethyl succinate was between 5.94 and 22.01%. Punicic acid (87.65–88.15%) ranked first, followed by oleic acid (1.58–3.70%), linoleic acid (3.32–3.64%), palmitic acid (2.20–2.69%), stearic acid (1.55–1.66%), *cis*-11-Eicosenoic acid (0.57–0.64%), linolenic acid (0.30–0.64%), lignoceric acid (0.00–0.04%), and margo oleic acid (0.00–0.04%).

MAE extraction efficiency with hexane was 25.3%. Punicic acid (88.39%) came first, followed by oleic acid (3.55%), linoleic acid (3.27%), palmitic acid (2.19%), stearic acid (1.58%), *cis*-11-Eicosenoic acid (0.64%), linolenic acid (0.37%), and lignoceric acid (0.02%).

UAE extraction efficiency varied from 6.71 to 26.3%. Punicic acid (74.4–88.0%) was the first, followed by linoleic acid (3.4–7.5%), oleic acid (3.6–7.1%), palmitic acid (2.3–5.0%), stearic acid (1.6–3.1%), *cis*-11-Eicosenoic acid (0.6–1.1%), linolenic acid (0.3–0.8%), lignoceric acid (0.08–0.5%), arachidic acid (0.03–0.1%), and heptadecanoic acid (0.0–0.1%).

UAE extraction efficiency with hexane was 31.2%. Punicic acid (75.26%) ranked first, followed by oleic acid (8.23%), linoleic acid (8.03%), palmitic acid (3.94%), stearic acid (2.28%), *cis*-11-Eicosenoic acid (1.47%), linolenic acid (0.72%), and lignoceric acid (0.08%). All the results are written in Table 7.

MAE and UAE extraction procedures were performed with hexane under optimized conditions in order to reveal the extraction ability difference between hexane and dimethyl succinate. The maximum extraction efficiency for MAE was 22.01% under the optimum conditions (liquid/solid ratio of 5, irradiation time of 3 min, and microwave power of 300 W). For UAE, the maximum extraction efficiency was 26.3% under the optimum conditions (liquid/solid ratio, 10; pulse duration/pulse interval ratio, 5; duration, 20 min; and temperature, 60 °C).

Solvents that can dissolve the target of interest and absorb microwave radiation are more suitable for MAE. Hexane recovered more oil than dimethyl succinate from pomegranate seed, which is most likely due to the better dissolving and absorbing ability. Hexane has a higher polarity than dimethyl succinate, directly affecting the microwave radiation absorbing performance. For this reason, it is one of the most studied solvents in the literature [26–28].

 Table 7
 The maximum extraction efficiency obtained by different treatments

Extraction method-solvent	Oil efficiency (%)
MAE-hexane	25.3
MAE-dimethyl succinate	22.01
UAE-hexane	31.2
UAE- dimethyl succinate	26.3
Cold pressing	5

Ref	Growing region	Extraction method/solvent	Best conditions	Maximum yield (%)
Barizão et al. [26]	Sao Paulo, Brazil	UAE: hexane SE: hexane BDE: chloroform-methanol- water	SE: seed powder, 5 g; time, 4 h; and temperature, 68 °C	SE: 30% UAE: 27.99% BDE: 22.3%
Tian et al. [36]	Xinjiang, China	UAE: n-hexane, ethyl acetate, diethyl ether, acetone, and isopropanol SE: n-hexane SFE: liquid CO ₂	UAE: petroleum ether; ultrasonic power, 140 W; temperature, 40 °C; and liquid/solid ratio, 10 mL/g	UAE: 25.11% SE: 20.50% SFE: 15.72%
Rojo-Gutiérrez et al. [6]	Hidalgo, Spain	MAE: hexane UAE: hexane	MAE: temperature, 90 °C; time, 10 min; and solid/sol- vent ratio, 1:20 g/mL	MAE: 18.38% UAE: 17.64%
Goula et al. [37]	Thessaloniki, Greece	UAE: hexane	UAE: seed particle size, 0.2 mm; temperature, 20 °C; solvent/solid ratio, 20/1 mL/g; amplitude level, 60%; and pulse duration/ pulse interval ratio, 5/15	UAE: 59.8%
Eikani et al. [42]	Fars Province, Iran	SHHE: n-hexane SE: n-hexane Cold pressing	SHHE: temperature, 80 °C; particle size, 0.25 mm; and flow rate, 1 mL/min	SHHE: 22.18% SE: 17.94% Cold pressing: 4.29%
Silva et al. [43]	Saveh, Iran	Aqueous extraction Hexane extraction Cold pressing Hot pressing	Aqueous extraction: water/ solid ratio, 2.2:1.0 mL/g; pH, 5; temperature, 63 °C; and time, 375 min	Aqueous extraction: 19.3% Hexane extraction: 26.8% Cold pressing: 7.0% Hot pressing: 8.6%
Cavdar et al. [44]	Nigde, Turkey	MAE: n-hexane SE: n-hexane Cold pressing	MAE: power, 220 W; liquid/ solid ratio, 10; and extrac- tion time, 5 min	MAE: 35.19% SE: 34.70% Cold pressing: 17.50%
Aruna et al. [45]	Navi Mumbai, India	SE: hexane, petroleum ether, chloroform, chloroform: methanol (2:1 v/v), and ethanol Cold pressing	SE: hexane; time, 8 h; and temperature, 30 °C	SE: 20% Cold pressing: 13.2%
Liu et al. [46]	Lintong, China	SE: n-hexane, petroleum ether MAE: n-hexane, petroleum ether UAE: n-hexane, petroleum ether SubE: n-hexane, petroleum ether Shaking extraction: n-butane	SE: n-hexane; time, 8 h; temperature, 40 °C	 SE: n-hexane, 15.66%; petroleum ether, 14.92% MAE: n-hexane, 12.47%; petroleum ether 11.72% UAE: n-hexane, 13.63%; petroleum ether, 12.82% SubE: n-butane, 14.50% Shaking extraction: n-hexane, 12.10%; petroleum ether, 11.32%
Ahangari et al. [47]	Tehran, Iran	SC-CO ₂ : CO ₂ SC-Propane: Propane SE: n-hexane	SE: extraction time, 1200 min; temperature, 70 °C; and seed amount; 10 g	SE: 22.31% SC-Propane: 17.12% SC-CO ₂ : 13.06%
Natolino et al. [48]	Giulia, Italy	SC-CO ₂ : CO ₂ SE: hexane	SC-CO ₂ : pressure, 320 bar; temperature, 60 °C; and flow rate, 8 kg/h	SC-CO ₂ : 18%
Goula et al. [49]	Thessaloniki, Greece	UAE aqueous enzymatic extraction: cellulase, pecti- nase enzymes Aqueous enzymatic extrac- tion: cellulase, pectinase enzymes	UAE aqueous enzymatic extraction: enzyme type, Peclyve V; temperature, 55 °C; liquid/solid ratio, 6/1 mL/g; enzyme concen- tration, 2% w/w; and time, 2 h	UAE aqueous enzymatic extraction: 18.15% Aqueous enzymatic extraction: 15.33%

Table 8 Literature studies on PSO extraction

The economic aspects are the most important factor affecting the method selection. Long extraction times and increased temperature result in higher costs. In this study, UAE not only gave a higher efficiency than MAE but also operated at moderate conditions while extracting the maximum oil (26.3%). Therefore, UAE seems a better option than MAE for PSO extraction [40, 41].

3.4 Comparison with literature studies

For PSO extraction, the previously applied methods are UAE, MAE, cold pressing, subcritical extraction (SubE), supercritical carbon dioxide extraction (SC-CO₂), supercritical fluid extraction (SFE), superheated hexane extraction (SHHE), bling and dyer extraction (BDE), and shaking extraction (see Table 8). Clearly, the cold pressing technique has relatively low efficiency, which is surely due to the incomplete extraction [42, 43]. Obviously, SE provides high efficiency; however, SE necessitates more time to complete the extraction and requires a high amount of solvent that must be recovered and recycled through evaporation and condensation [26]. Shaking extraction leads to residual solvents and impurities in the extract. MAE, UAE, and SubE are novel techniques. MAE uses microwave heating energy to increase efficiency. Uniform and rapid microwave energy enhances product recovery and effectively shortens extraction time. UAE applies cavitation force, which comes from the acoustic waves. Due to the cavitation force, the cell wall breaks down easily and solvent penetrates into the sample matrix. SFE is also another environmentally friendly technique, but the large-scale application of SFE is limited due to the high operational and equipment cost. SubE is a good choice for the extraction of heat-sensitive products. For the case of SubE, the target of interest is easily removed under a certain pressure by using liquefied subcritical solvent; however the solvent must be removed and evaporated, which results in high cost.

Our study showed similar efficiency results with literature studies on the PSO extraction. The slight differences between results might be due to the treatment variations, extraction conditions, growing regions, climate conditions, and fruit genotypes [33]. Table 8 revealed that hexane is superior to other solvents for all the extraction techniques studied. Cold pressing gave relatively low efficiency [42, 44, 45]. SE was found to be one of the most effective methods in terms of efficiency; however, it has taken long extraction times. Liu et al. [46] extracted PSO with SE method at 8 h. Ahangari et al. [47] extracted PSO with SE technique at 1200 min. It is clear from the results that SE takes far more time and consumes a lot more solvent compared to UAE and MAE techniques. It is clear from the results that MAE and UAE are the most effective methods in terms of efficiency. For UAE, only Barizao et al. (27. 99%) and Goula et al. (59.8%) reported better efficiency results than the results

obtained by this study [26, 37]. However, there is no sharp difference between this study and Barizao et al.'s study, possibly due to the similar operating conditions. Goula et al. achieved 59.8% oil efficiency, which is quite surprising. In case of MAE, only Cavdar et al.'s study [44] achieved better extraction efficiency than this study (35.19% > 22.01%). The difference might be explained by growing region and climate conditions because the optimized conditions are similar with this study. Most importantly, the last three successful studies mentioned above used hexane as the solvent. Hexane is still one of the greatest solvents available today, but according to EU Annex II of the cosmetics regulations, it cannot be used to make cosmetic items. As a result, the finished product needs to be highly purified. On the other hand, DMS, ranked number 1 in the "Environmental Working Group" (EWG) classification, is an eco-friendly solvent that can be used in cosmetics. DMS is more sustainable than hexane, as shown by the study's findings, and the differences in extraction efficiency were not noticeably large. Therefore, for the sake of protecting the environment and public health, DMS can be considered a good substitute for hexane in extracting PSO.

4 Conclusion

After GC-FID analyses, punicic acid was the pomegranate seed's most abundant fatty acid (>80%). UAE method was more effective than MAE method for PSO extraction in terms of oil efficiency. RSM determined the best conditions of MAE and UAE treatments. The highest efficiency was 22.01% in the case of MAE, while the highest efficiency was 26.31% in the case of UAE. For PSO extraction, UAE seems to outperform MAE as the more suitable method. The optimum conditions of MAE were a liquid/solid ratio of 5, irradiation time of 3 min, and microwave power of 300 W. The optimum conditions of UAE were a liquid/solid ratio of 10, pulse duration/pulse interval ratio of 1, 60 °C temperature, and 20 min extraction time. A sustainable strategy has been the focus of this research within the use of dimethyl succinate. Dimethyl succinate is a more environmentally friendly solvent compared to hexane. The results obtained from this study are promising for the use of dimethyl succinate instead of hexane in oil extraction studies. If the oil extraction from pomegranate seeds is to be carried out under the conditions specified in this study, our recommendation is to utilize the ultrasonic-assisted extraction method with dimethyl sulfoxide as the solvent.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s13399-024-05611-4.

Acknowledgements The authors gratefully acknowledge the financial support received from BAP (Project No: FYL-34834) from Istanbul University-Cerrahpasa.

Author contributions Aslı Gök: Conceptualization, Investigation, Writing-Original draft.

Hilal Uyar: Optimization study, Methodology, Formal analyses. Özge Demir: Investigation, Writing-Original draft.

Funding Open access funding provided by the Scientific and Technological Research Council of Türkiye (TÜBİTAK).

Data availability All data and materials are available in this manuscript.

Declarations

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Ethical approval There was no ethical approval required.

Competing interests The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Mohagheghi M, Rezaei K, Labbafi M, Ebrahimzadeh SM (2011) Pomegranate seed oil as a functional ingredient in beverages. Eur J Lipid Sci Technol 113:730–736. https://doi.org/10.1002/ejlt. 201000334
- Boroushaki MT, Mollazadeh H, Afshari AR (2016) Pomegranate seed oil: a comprehensive review on its therapeutic effects. Int J Pharm Sci Res 7:430. https://doi.org/10.13040/IJPSR.0975-8232. 7(2).430-42
- Goula AM, Adamopoulos KG (2012) A method for pomegranate seed application in food industries: seed oil encapsulation. Food Bioprod Process 90:639–652. https://doi.org/10.1016/j.fbp.2012.06.001
- Vroegrijk IOCM, van Diepen JA, van den Berg S et al (2011) Pomegranate seed oil, a rich source of punicic acid, prevents diet-induced obesity and insulin resistance in mice. Food Chem Toxicol 49:1426–1430. https://doi.org/10.1016/j.fct.2011.03.037
- Paul A, Radhakrishnan M (2020) Pomegranate seed oil in food industry: extraction, characterization, and applications. Trends Food Sci Technol 105:273–283. https://doi.org/10.1016/j.tifs. 2020.09.014
- Rojo-Gutiérrez E, Carrasco-Molinar O, Tirado-Gallegos JM et al (2021) Evaluation of green extraction processes, lipid composition and antioxidant activity of pomegranate seed oil. J Food Meas Charact 15:2098–2107. https://doi.org/10.1007/ s11694-020-00804-7
- Lalikoglu M, Ince E (2021) The synthesis of benzyl acetate as a fragrance and flavoring agent with green solvents. J Cosmet Sci 72:362–378

- Gökdemir B, Baylan N, Çehreli S (2020) Application of a novel ionic liquid as an alternative green solvent for the extraction of curcumin from turmeric with response surface methodology: determination and optimization study. Anal Lett 53:2111–2121. https://doi.org/10.1080/00032719.2020.1730394
- Doldolova K, Bener M, Lalikoğlu M et al (2021) Optimization and modeling of microwave-assisted extraction of curcumin and antioxidant compounds from turmeric by using natural deep eutectic solvents. Food Chem 353:129337. https://doi.org/10.1016/j.foodc hem.2021.129337
- Khoddami A, Bin Y, Man C, Roberts TH (2014) Physico-chemical properties and fatty acid profile of seed oils from pomegranate (Punica granatum L.) extracted by cold pressing. Eur J Lipid Sci Technol 116:553–562. https://doi.org/10.1002/ejlt.201300416
- Rasul MG (2018) Conventional extraction methods use in medicinal plants, their advantages and disadvantages. Int J Basic Sci Appl Comput 2:10–14
- Esfandiari N (2015) Production of micro and nano particles of pharmaceutical by supercritical carbon dioxide. J Supercrit Fluids 100:129–141. https://doi.org/10.1016/j.supflu.2014.12.028
- Zhang DY, Yao XH, Luo M et al (2016) Optimization of negative pressure cavitation-microwave assisted extraction of yellow horn seed oil and its application on the biodiesel production. Fuel 166:67–72. https://doi.org/10.1016/j.fuel.2015.10.022
- Zhong J, Wang Y, Yang R et al (2018) The application of ultrasound and microwave to increase oil extraction from Moringa oleifera seeds. Ind Crops Prod 120:1–10. https://doi.org/10.1016/j. indcrop.2018.04.028
- Ye M, Zhou H, Hao J et al (2021) Microwave pretreatment on microstructure, characteristic compounds and oxidative stability of Camellia seeds. Ind Crops Prod 161:113193. https://doi.org/10. 1016/j.indcrop.2020.113193
- Gumaling RP, Agusan JR, Ellacer NVCR et al (2018) Increased bio-oil yield from Swietenia macrophylla seeds through microwave pretreatment and ultrasonic-assisted solvent extraction. Sustain Environ Res 28:430–437. https://doi.org/10.1016/j.serj.2018. 06.003
- Đurđević S, Milovanović S, Šavikin K et al (2017) Improvement of supercritical CO2 and n-hexane extraction of wild growing pomegranate seed oil by microwave pretreatment. Ind Crops Prod 104:21–27. https://doi.org/10.1016/j.indcrop.2017.04.024
- Rodríguez AM, Prieto P, De La Hoz A et al (2015) Influence of polarity and activation energy in microwave-assisted organic synthesis (MAOS). ChemistryOpen 4:308–317. https://doi.org/ 10.1002/open.201402123
- Hernández-Santos B, Rodríguez-Miranda J, Herman-Lara E et al (2016) Effect of oil extraction assisted by ultrasound on the physicochemical properties and fatty acid profile of pumpkin seed oil (Cucurbita pepo). Ultrason Sonochem 31:429–436. https://doi. org/10.1016/j.ultsonch.2016.01.029
- Perrier A, Delsart C, Boussetta N et al (2017) Effect of ultrasound and green solvents addition on the oil extraction efficiency from rapeseed flakes. Ultrason Sonochem 39:58–65. https://doi.org/10. 1016/j.ultsonch.2017.04.003
- Panadare DC, Gondaliya A, Rathod VK (2020) Comparative study of ultrasonic pretreatment and ultrasound assisted three phase partitioning for extraction of custard apple seed oil. Ultrason Sonochem 61:104821. https://doi.org/10.1016/j.ultsonch.2019.104821
- Ma Y, Wu X, Zhao L et al (2019) Comparison of the compounds and characteristics of pepper seed oil by pressure-assisted, ultrasound-assisted and conventional solvent extraction. Innov Food Sci Emerg Technol 54:78–86. https://doi.org/10.1016/j.ifset.2019. 03.011
- 23. Tang W, Wang B, Wang M, Wang M (2021) Ultrasound-assisted extraction of Osmanthus fragrans fruit oil and evaluation of its fatty acid composition, physicochemical properties and

antioxidant activity. J Appl Res Med Aromat Plants 25:100331. https://doi.org/10.1016/j.jarmap.2021.100331

- Briones-Labarca V, Plaza-Morales M, Giovagnoli-Vicuña C, Jamett F (2015) High hydrostatic pressure and ultrasound extractions of antioxidant compounds, sulforaphane and fatty acids from Chilean papaya (Vasconcellea pubescens) seeds: effects of extraction conditions and methods. LWT-Food Sci Technol 60:525–534. https://doi.org/10.1016/j.lwt.2014.07.057
- Rashid R, Mohd Wani S, Manzoor S et al (2022) Green extraction of bioactive compounds from apple pomace by ultrasound assisted natural deep eutectic solvent extraction: optimisation, comparison and bioactivity. Food Chem 398:133871. https://doi.org/10.1016/j. foodchem.2022.133871
- Barizão ÉO, Boeing JS, Martins AC et al (2015) Application of response surface methodology for the optimization of ultrasoundassisted extraction of pomegranate (Punica granatum L.) seed oil. Food Anal Mehods 8:2392–2400. https://doi.org/10.1007/ s12161-015-0135-5
- Moncada J, Tamayo JA, Cardona CA (2016) Techno-economic and environmental assessment of essential oil extraction from Oregano (Origanum vulgare) and Rosemary (Rosmarinus officinalis) in Colombia. J Clean Prod 112:172–181. https://doi.org/10. 1016/j.jclepro.2015.09.067
- Koubaa M, Mhemdi H, Barba FJ et al (2017) Seed oil extraction from red prickly pear using hexane and supercritical CO2: assessment of phenolic compound composition, antioxidant and antibacterial activities. J Sci Food Agric 97:613–620. https://doi. org/10.1002/jsfa.7774
- Chakraborty S, Weindorf DC, Li B et al (2014) Analysis of petroleum contaminated soils by spectral modeling and pure response profile recovery of n-hexane. Environ Pollut 190:10–18. https:// doi.org/10.1016/j.envpol.2014.03.005
- Wan X, Ren D, Liu Y et al (2018) Facile synthesis of dimethyl succinate via esterification of succinic anhydride over ZnO in methanol. ACS Sustain Chem Eng 6:2969–2975. https://doi.org/ 10.1021/acssuschemeng.7b02598
- Ozturk B, Parkinson C, Gonzalez-Miquel M (2018) Extraction of polyphenolic antioxidants from orange peel waste using deep eutectic solvents. Sep Purif Technol 206:1–13. https://doi.org/10. 1016/j.seppur.2018.05.052
- Wan Mahmood WMA, Lorwirachsutee A, Theodoropoulos C, Gonzalez-Miquel M (2019) Polyol-based deep eutectic solvents for extraction of natural polyphenolic antioxidants from Chlorella vulgaris. ACS Sustain Chem Eng 7:5018–5026. https://doi.org/10. 1021/acssuschemeng.8b05642
- Hu B, Zhou K, Liu Y et al (2018) Optimization of microwaveassisted extraction of oil from tiger nut (Cyperus esculentus L.) and its quality evaluation. Ind Crops Prod 115:290–297. https:// doi.org/10.1016/j.indcrop.2018.02.034
- Zheng B, Yuan Y, Xiang J et al (2022) Green extraction of phenolic compounds from foxtail millet bran by ultrasonic-assisted deep eutectic solvent extraction: optimization, comparison and bioactivities. Lwt 154:112740. https://doi.org/10.1016/j.lwt.2021. 112740
- Ibrahim NA, Zaini MAA (2018) Microwave-assisted solvent extraction of castor oil from castor seeds. Chin J Chem Eng 26:2516–2522. https://doi.org/10.1016/j.cjche.2018.07.009
- Tian Y, Xu Z, Zheng B, Martin Lo Y (2013) Optimization of ultrasonic-assisted extraction of pomegranate (Punica granatum L.) seed oil. Ultrason Sonochem 20:202–208. https://doi.org/10. 1016/j.ultsonch.2012.07.010
- Goula AM (2013) Ultrasound-assisted extraction of pomegranate seed oil - kinetic modeling. J Food Eng 117:492–498. https://doi. org/10.1016/j.jfoodeng.2012.10.009

- Fu X, Wang D, Belwal T et al (2021) Sonication-synergistic natural deep eutectic solvent as a green and efficient approach for extraction of phenolic compounds from peels of Carya cathayensis Sarg. Food Chem 355:129557. https://doi.org/10.1016/j.foodc hem.2021.129577
- Alsaud N, Shahbaz K, Farid M (2021) Application of deep eutectic solvents in the extraction of polyphenolic antioxidants from New Zealand Manuka leaves (Leptospermum Scoparium): optimization and antioxidant activity. J Mol Liq 337:116385. https:// doi.org/10.1016/j.molliq.2021.116385
- Khajebishak Y, Payahoo L, Alivand M, Alipour B (2019) Punicic acid: a potential compound of pomegranate seed oil in Type 2 diabetes mellitus management. J Cell Physiol 234:2112–2120. https://doi.org/10.1002/jcp.27556
- 41. Pereira De Melo IL, Bonifácio E, De Carvalho T, Mancini-Filho J (2014) Pomegranate seed oil (Punica granatum L.): a source of punicic acid (conjugated α -linolenic acid). J Hum Nutr Food Sci 2:1–11
- Eikani MH, Golmohammad F, Homami SS (2012) Extraction of pomegranate (Punica granatum L.) seed oil using superheated hexane. Food Bioprod Process 90:32–36. https://doi.org/10. 1016/j.fbp.2011.01.002
- 43. Silva LDO, Ranquine LG, Monteiro M, Torres AG (2019) Pomegranate (Punica granatum L.) seed oil enriched with conjugated linolenic acid (cLnA), phenolic compounds and tocopherols: improved extraction of a specialty oil by supercritical CO2. J Supercrit Fluids 147:126–137. https://doi.org/10.1016/j.supflu. 2019.02.019
- Çavdar HK, Yanik DK, Gok U, Gogus F (2017) Optimisation of microwave-assisted extraction of pomegranate (Punica granatum L.) seed oil and evaluation of its physicochemical and bioactive properties. Food Technol Biotechnol 55:86–94. https://doi.org/10. 17113/ftb.55.01.17.4638
- Aruna P, Manohar B, Singh RP (2018) Processing of pomegranate seed waste and mass transfer studies of extraction of pomegranate seed oil. J Food Process Preserv 42:1–11. https://doi.org/10.1111/ jfpp.13609
- 46. Liu N, Ren G, Faiza M et al (2022) Comparison of conventional and green extraction methods on oil yield, physicochemical properties, and lipid compositions of pomegranate seed oil. J Food Compos Anal 114:104747. https://doi.org/10.1016/j.jfca.2022. 104747
- Ahangari B, Sargolzaei J (2012) Extraction of pomegranate seed oil using subcritical propane and supercritical carbon dioxide. Theor Found Chem Eng 46:258–265. https://doi.org/10.1134/ S0040579512030013
- Natolino A, Da Porto C (2019) Supercritical carbon dioxide extraction of pomegranate (Punica granatum L.)seed oil: kinetic modelling and solubility evaluation. J Supercrit Fluids 151:30–39. https://doi.org/10.1016/j.supflu.2019.05.002
- Goula AM, Papatheodorou A, Karasavva S, Kaderides K (2018) Ultrasound-assisted aqueous enzymatic extraction of oil from pomegranate seeds. Waste Biomass Valorization 9:1–11. https:// doi.org/10.1007/s12649-016-9740-9

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.