**ORIGINAL ARTICLE** 



# Control of the post-infarct immune microenvironment through biotherapeutic and biomaterial-based approaches

Shreya S. Soni<sup>1</sup> · Arielle M. D'Elia<sup>1</sup> · Christopher B. Rodell<sup>1</sup>

Accepted: 3 January 2023 / Published online: 10 February 2023 © The Author(s) 2023

#### Abstract

Ischemic heart failure (IHF) is a leading cause of morbidity and mortality worldwide, for which heart transplantation remains the only definitive treatment. IHF manifests from myocardial infarction (MI) that initiates tissue remodeling processes, mediated by mechanical changes in the tissue (loss of contractility, softening of the myocardium) that are interdependent with cellular mechanisms (cardiomyocyte death, inflammatory response). The early remodeling phase is characterized by robust inflammation that is necessary for tissue debridement and the initiation of repair processes. While later transition toward an immunoregenerative function is desirable, functional reorientation from an inflammatory to reparatory environment is often lacking, trapping the heart in a chronically inflamed state that perpetuates cardiomyocyte death, ventricular dilatation, excess fibrosis, and progressive IHF. Therapies can redirect the immune microenvironment, including biotherapeutic and biomaterial-based approaches. In this review, we outline these existing approaches, with a particular focus on the immunomodulatory effects of therapeutics (small molecule drugs, biomolecules, and cell or cell-derived products). Cardioprotective strategies, often focusing on immunosuppression, have shown promise in pre-clinical and clinical trials. However, immunoregenerative therapies are emerging that often benefit from exacerbating early inflammation. Biomaterials can be used to enhance these therapies as a result of their intrinsic immunomodulatory properties, parallel mechanisms of action (e.g., mechanical restraint), or by enabling cell or tissue-targeted delivery. We further discuss translatability and the continued progress of technologies and procedures that contribute to the bench-to-bedside development of these critically needed treatments.

Keywords Inflammatory disease · Heart failure · Immune modulation · Biomaterials · Biotherapeutics

# Introduction

Cardiovascular disease (CVD) remains among the greatest causes of morbidity and mortality, contributing to nearly 30% of deaths worldwide [1]. CVD includes a broad array of conditions, spanning from congenital heart and vascular defects to acquired diseases such as coronary artery disease, myocardial infarction (MI), arrhythmias, and varying presentations of heart failure (HF). In the western world, HF remains the leading cause of death and is expected to afflict nearly 8 million Americans by 2030. Heart disease affects individuals of all socio-demographic backgrounds, with an age-adjusted prevalence varying between whites (11.0%), blacks (9.7%), Hispanics (7.4%), and Asians (6.1%) [2]. The

Christopher B. Rodell christopher.b.rodell@drexel.edu

<sup>1</sup> School of Biomedical Engineering, Science and Health Systems, Drexel University, Philadelphia, PA 19104, USA variability among populations has been attributed, in part, to healthcare access, the geographic prevalence of risk factors, and genetic background [3]. Nearly 70% of HF cases are ischemic heart failure (IHF), precipitated by either a partial or complete blockage of blood flow to the myocardium and ensuing left ventricular (LV) remodeling [4, 5]. Atherosclerosis often underlies IHF [6], as continual plaque accumulation, fueled by the accumulation of monocytes (Mo) and macrophages (MF) within the lesion, occludes blood flow to result in myocardial ischemia. Plaque rupture is a primary cause of coronary occlusion and MI.

The LV remodeling process that occurs after ischemic injury is characterized by maladaptive geometric and functional changes in the heart, which are rooted both in mechanical and inflammatory effects (Fig. 1) [7]. Mediators of the post-MI immune microenvironment are further detailed in Table 1. In the hours and days post-MI, a cascade of ischemia, necrosis, and loss of myocardial contractility result in early expansion of the infarct. This process is paralleled



Fig.1 Left ventricular (LV) remodeling and progression of the post-MI inflammatory response. **a** The heart undergoes LV remodeling after MI. Initial ischemia (left) results in cardiomyocyte apoptosis and loss of muscle contraction. Within the following days and weeks, softening of the myocardium by ECM degradation and apoptosis in the border zone result in geometric changes to the heart that include infarct thinning and expansion (middle). Over time, global remodeling is characterized by ventricular dilation, cardiac hypertrophy, and valve dysfunction which result in a loss of

by an early pro-inflammatory response, hallmarked by the rapid recruitment of innate immune cells (neutrophils, Mo, and MF) that begins within minutes post-MI and persists for greater than a week. This robust cellular infiltrate is required for tissue debridement and to initiate repair functions [8]. However, the molecular signals responsible for innate immune cell infiltration are largely pro-inflammatory chemokines (e.g., IP-10, MCP-1), damage-associated molecular patterns (DAMPs, including cell and extracellular matrix (ECM) debris, HMGB1, etc.), and neutrophil degranulation itself [9–12]. Corresponding cell-surface receptors detect these signals to drive homing to the injury site, also initiating pro-inflammatory signaling cascades that produce cytokines and chemokines, further promoting leukocyte recruitment to the site of injury [13].

Within the infarct, Mo rapidly differentiate into MF that persist for weeks, expanding the local population by greater than tenfold [14–16]. In this intermediate phase, MF are pleiotropic

heart function that manifest clinically as ischemic heart failure (IHF). Figure adapted from [5]. **b** Following MI, the inflammatory response is incited and can be described in three phases—an early, intermediate, and late response that temporally correlate to the stages of LV remodeling. Innate immune cells (neutrophils, Mo) initially dominate the immune microenvironment, giving way to a wave of inflammatory (M1-like) and later prohealing (M2-like) MF that exist in a heterogeneous and diverse pool of phenotypes. The inflammatory milieu guides later B and T cell response

regulators of the immune microenvironment and mediators of the tissue remodeling process that exhibit a wide spectrum of functional phenotypes, having complementary or even opposing functions. Inflammatory MF are often canonically denoted as conventional (M1-like) cells, regarded as tissue damaging. In contrast, alternatively activated (M2-like) cells are considered a tissue-reparatory phenotype [17]. Post-MI, initial MF populations are predominantly M1-like, contributing substantially to tissue debridement and ECM breakdown [18]. They also produce abundant pro-inflammatory cytokines (IL-1, IL-6, and TNF- $\alpha$ ) that induce cardiomyocyte hypertrophy or apoptosis, recruit additional cell populations that support chronic inflammation, and are clinical predictors of IHF mortality [19]. Resulting cytokine-induced cardiomyocyte death and breakdown of ECM by overexpression of matrix metalloproteinase (MMP) significantly contribute to infarct thinning and expansion. In later periods post-MI, typically around 1 week in mice, the emergence of M2-like MF coincides with

Table 1 Prevalent cell types and signaling markers with their related function in the immune response post-MI

Components	Functions
Immune cells	
Neutrophils	<ul> <li>First responder, rapidly migrate to infarct [50, 51]</li> <li>Produce ROS, pro-inflammatory cytokines (IL-1β, IL-6, TNF-α), and MMPs [50, 51]</li> </ul>
	Degranulation and chemokine production promote further leukocyte infiltration [52]
	Apoptotic neutrophil phagocytosis promotes anti-inflammatory macrophage activation
Monocytes and macrophages	Monocytes traffic to the infarct largely by CCL2-mediated chemotaxis from the spleen and emergency hematopoiesis [24], differentiate into macrophages [6]
	Debride the tissue via proteolysis [53] and clear cellular debris [54] Pro-inflammatory cytokine production induces cardiomyocyte hypertrophy or apoptosis [50, 51]
	Later promote repair, including by stimulation of ECM production and angiogenesis [53, 55]
T cell	
Regulatory	Terminate the pro-inflammatory phase [56] Stimulate fibroblasts [57]
Memory effector	Autoimmune reaction against the myocardium [38]
B cells	Autoantibody production [33]
Signaling markers	
Pro-inflammatory cytokines (IL-1, TNF- $\alpha$ , IL-6, INF- $\gamma$ , etc.)	Initiate innate and adaptive response Promote further recruitment, proliferation, and activation [58, 59]
Anti-inflammatory cytokines (VEGF, TGF-β, IL-10 [55], IL-4, IL-13 [60], etc.)	Transition macrophages from M1-like to M2-like Suppress infiltration of inflammatory cells [57, 60]
Chemokines (CCL2, CCL5, CXCL1 [61], etc.) and chemotactic cues (selectins)	Promote chemotaxis of both immune and non-immune cells [61]

reparatory signals (e.g., PDGF, IL-10, and TGF- $\beta$ ) essential for angiogenesis, cell viability, and collagen production, respectively [20, 21]. MF therefore contribute substantially to LV remodeling and IHF, but is also critical for later tissue repair due to the opposing function of M1- and M2-like phenotypes. As such, modulation of M0/MF populations and phenotype has been widely investigated [22]. Owing to their potential reparatory roles, MF depletion impairs healing and worsens outcome [23]. In contrast, reducing the number of infiltrating Mo, such as by CCR2 blockade, reduces infarct size and supports post-MI recovery [24]. This is likely due to CCR2dependent recruitment of inflammatory Mo subpopulations, while non-classical Mo traffic by alternative means [25].

In the following weeks and months, the transition towards an anti-inflammatory and reparative phase to promote wound healing and scar formation is preferable. However, impediments to this transition often force the myocardium into a chronically inflamed state wherein establishment of chronic inflammation and adaptive immune response play a critical role in the remodeling process. As discussed, activated Mo/MF in the infarct mediate para-inflammation (continued leukocyte recruitment) that drives the formation of a chronic inflammatory milieu [26]. Adaptive immune responses, on the other hand, are relatively specific and mediated predominantly by B and T cells [18, 27-29]. Antigen-presenting cells (dendritic cells (DCs)) and to a lesser extent Mo/MF [30] and potentially neutrophils [31, 32] serve as a critical bridge between the innate and adaptive immune response. B cells are derived from the bone marrow and mature into immunoglobin-secreting plasma cells or memory B cells after encountering an antigen epitope. Evidence is emerging that failures of self-tolerance immune checkpoints can result in auto-antibody production, perpetuating disease progression [33]. In contrast, T cells originate from the thymus and differentiate into effector or memory cells, essential to tissue homeostasis and immune memory. In response to environmental stimuli (cytokine signatures) and presented signals (immune checkpoints, antigens), T cells enact a multitude of inflammatory or anti-inflammation cell programs that impact CVD development and progression [34-36]. Regulatory T cells, though low in numbers following MI, abate the inflammatory response through production of antiinflammatory cytokines [37]. In contrast, development of an adaptive immune response through memory T cells has been shown to coordinate an autoimmune reaction against the myocardium [38]. For a more thorough discussion of adaptive immune response in HF, the reader is referred to recent reviews on this topic [39, 40].

In sum, inflammatory tissue injury underlies LV remodeling and functional declines that manifest as IHF. The process is fundamentally rooted in dysregulation of both the innate and adaptive immune responses that interdependently contribute to LV remodeling, as characterized by cardiomyocyte apoptosis, ventricular dilatation, and myocardial fibrosis that negatively impact heart function [41-45]. Identifying key regulators of these processes is therefore of great importance toward restoring homeostasis and promoting natural injury resolution. To control the post-MI immune microenvironment, therapies have broadly employed the use of cell and bioactive molecule delivery. Cell-based strategies frequently aim to repair or replace the affected tissue. Stem cell therapies, including embryonic (ESC), mesenchymal (MSC), induced pluripotent (iPSC), and others, have been widely explored both as functional tissue replacements and for their immunomodulatory effects [46–48]. However, the delivery of these cells alone is hindered by low cell retention and survival rates, contributing to poor therapeutic efficacy [49] and motivating the use of biomaterial delivery vehicles. Pharmacological approaches include the systemic or local delivery of exogenous cytokines, chemokines, and small molecule drugs. These methods often seek to modulate the hyperinflammatory post-MI environment as a means of cardioprotection or to enhance the body's inherent tissue repair capacity. Such pro-regenerative strategies have shown some recent success, particularly in the use of microRNA and small interference RNA (siRNA) to promote cardiac cell regeneration [46]. The systemic administration of anti-inflammatory drugs, however, often results in chronic immunosuppression and an elevated risk of infection. Biomaterial-based drug delivery systems enable cell- and tissuetargeting strategies to overcome these challenges, while also concentrating therapeutic concentrations at the site of action. Therefore, there is a critical need for effective delivery strategies and sustained release approaches that can instruct the injury resolution process, either by prophylactically intercepting disease progression, reorienting the hyperinflammatory milieu towards a reparatory state, or reversing the deleterious chronic and adaptive immune response.

# Experimental models and assessment of inflammatory pathophysiology

Owing to the multifaceted mechanisms of post-MI remodeling processes, the study of both remodeling events and therapeutic strategies to intercept them require investigation in complex environments. These experimental models should recapitulate necessary aspects of the native injury environment that include, for example, complex cell and matrix composition, signaling pathways, and mechanical forces that underlie disease progression. These contributing factors to the pathophysiology evolve over time through a dynamic discourse, which continues to be studied to better understand disease progression and reveal new targets for intervention. Most often, animal models of disease may best recapitulate these processes, as they afford an intact biological system that is a platform for studying the evolution of disease from initial injury to eventual LV remodeling. These studies frequently benefit from advanced imaging approaches, many of which are likewise applicable for diagnosis and as biomarkers for disease stratification (Fig. 2). Here, we provide a brief overview of pertinent methodologies and techniques to delineate the impact of inflammation on IHF.

## Infarct models

In conducting translational research, it is important to choose models with significant prognostic power. In some cases, reductionist approaches are highly applicable to model aspects of tissue injury for this purpose. Microphysiological systems (MPSs) continue to emerge as a powerful means of generating in vitro models for understanding tissue development and screening experimental therapeutics. Construct design requires careful consideration of chemical, physical, and biological cues included-especially for modeling of mechanically actuating and immunological systems [62, 63]. Three-dimensional (3D) printing and other fabrication techniques using immunocompatible materials, such as collagen, gelatin, and chitosan, have shown great promise in the development of artificial heart systems [64]. For example, Boudou and colleagues engineered cardiac microtissues using microelectromechanical systems to facilitate fundamental understanding of cardiovascular biology, develop model systems in vitro, and potentially replace damaged myocardial tissue in vivo. Varying the mechanical stiffness of the collagen cell matrix increased cardiomyocyte contractility, and electrical stimulation and auxotonic load improved cell alignment and force generation, impacting maturation, structure, and function of myocardial tissue [65]. Jackman et al. similarly employed a cell-forward approach to engineer cardiobundles. This system created 3D cylindrical tissue from rat cardiomyocytes or human stem cell-derived cardiomyocytes. Cardiobundles were able to match contractile force, conduction velocity of action potentials, and cardiomyocyte size to mimic those of adult rat tissue [66]. These and related approaches to in vitro cardiac tissue engineering integrate biochemical, biophysical, and electromechanical stimuli to develop physiologically relevant systems for basic discovery and therapeutic benefit [67-69]. While cardiac MPSs confer the potential to study specific aspects of immune cell function in an appropriate dynamic environment, such as by incorporation of tissue resident MF, these applications have yet to be widely explored [70]. Such models are, however, emerging as a means of understanding and screening for cardiotoxicity in the post-MI environment [71].



While benchtop models of disease are suitable for some purposes, a full model for MI should resemble that of the human disease in terms of etiology and pathophysiology. For in-depth evaluation of tissue remodeling processes and therapeutic efficacy, animal models are often most appropriate. For extensive discussion of relevant animal models of MI and appropriate species selection, the reader is referred to prior reviews [72, 73] and expert guidance by Merry Lindsey and colleagues [74]. Small animal models include mice, rats, guinea pigs, and rabbits, which are cost effective, as well as easy to handle and maintain [75]. However, small animals often significantly deviate from human anatomy which may hinder translational potential [76]. Small animal models are nonetheless critical in the research setting to improve our understanding of disease progression and are an invaluable first line of study to evaluate novel treatment strategies. The first model for ischemic injury was established in a Wistar rat using permanent coronary artery ligation by Pfeffer and colleagues [77], and similar models have since been developed in mice and other species [78]. These methods have been adapted to include ischemia-reperfusion (IR) injury, in which blood flow is temporarily occluded to induce ischemic injury and subsequently unblocked to reinstate blood flow [79]. Recent perspectives discuss the clinical relevance of such permanent occlusion versus IR models [80], highlighting that IR recapitulates best clinical standards of care and may best address cardioprotective therapies. On the other hand, permanent occlusion may better replicate clinical pathophysiology of inflammation-driven LV remodeling and is applicable to examination of wound healing, scar formation, and IHF progression.

Utilizing mouse models for ischemic injury provides other added benefits, such as the potential to understand and characterize pathways at the molecular level. Many genetically engineered mice (GEMs) are available for purchase and can be used to systematically and mechanistically understand mechanisms of IHF [81]. Cell-type specific depletion, such as diphtheria toxin receptor (DTR) mouse models, are readily available and can elucidate the role of discrete immune cell populations by simple administration of the toxin. Conditional transgenic DTR models include FOXP3-DTR for T cell depletion [82], Ly6G-DTR for neutrophil depletion [83], and CD11b-DTR for MF depletion [18, 84], among others [85]. These GEMs are a critical tool to manipulate the immune microenvironment for basic discovery. For example, CD169-DTR and CCL2-DTR mice have been used to selectively deplete tissue resident and inflammatory MF populations, which resulted in divergent effects on further Mo recruitment and LV function [86]. By DTR depletion of CX3CR1 cells (cardiac tissue resident MF) and detailed fate mapping and parabiosis studies, the Epelman group has also revealed a cardioprotective role of resident MF that is not redundant with Mo differentiation in the tissue [87]. Similar to limitations involved with other transgenic animal approaches, DTR mouse models rely on accurate choice of cell-specific promoters and the assumption that cells are defined by single promoter activities. This becomes problematic, especially between closely related immune cells such as MF and Mo [85]. As a result, population depletions are often incomplete, tissue-dependent, and temporary. In addition, repeated diphtheria toxin treatments can result in off-target cell effects, sickening or killing the animal [88, 89]. Although DTR transgenic mice and other GEMs [90] permit a greater biological understanding and develop effective therapies, reasonable caution is warranted with their use.

The purchase or generation of genetically modified mouse models for application-specific purposes is also used in practice. Many studies have made use of genetically modified apolipoprotein E-deficient (Apo $E^{-/-}$ ) mice to study CVD, because they readily reproduce critical aspects of atherosclerosis, the buildup of arterial plaque that is a leading cause of MI, stroke, or angina [91]. Other application-specific models include mice lacking the prostaglandin E2 receptor 4, which has been shown to stimulate cardiomyocyte hypertrophy. After coronary artery ligation to induce MI, the knockout mice showed decreases in hypertrophy, fibrosis, and activation of Stat3, a prominent pro-inflammatory pathway used in T cell maturation relative to wild type [92]. In another example, angiotensin II type IA receptor knockout mice showed decreased levels of TGF-β and fibrosis, which reduced LV remodeling and increased mouse survival [93]. Trib1<sup>-/-</sup> knockout mice, effectively M2-like MF depleted, experienced more frequent cardiac rupture due to reduced collagen fibril formation in the myocardium. However, administration of M2-like MF and exogenous anti-inflammatory cytokines, like IL-4, restored function of the heart, highlighting the imperative role that M2-like MF play in infarct repair [94]. In sum, small animal models are relatively inexpensive to investigate and GEMs, in particular, greatly enhance the ability to mechanistically understand disease progression and modes of therapeutic action, rendering them critical investigational tools in the field.

Large animal models often confer a higher degree of experimental reliability and biological relevance, as the anatomy and timeline of disease progression is more closely aligned with that of humans. Sheep, pigs, dogs, and baboons are commonly employed large animal models of MI. Details of these models and their common limitations have been previously reviewed elsewhere [95, 96]. Notably, these models do face a degree of logistical challenges, including relatively high costs, as well as greater demands for maintenance and care in laboratory settings [97, 98]. From an ethical perspective, the use of larger animals in scientific research has faced societal criticism, contributing to necessary regulatory oversight [98, 99]. Despite these challenges, larger models of disease are often highly desirable because they possess greater similarities to human anatomy. For example, porcine models closely parallel the coronary vasculature, collateral circulation, and metabolic activity of the human heart, making them an often-preferred model of vascular diseases and intervention [100–102]. Sheep and humans likewise share similar cardiac kinetics and healing patterns following myocardial injury [103, 104]. Non-human primates are most closely related to humans due to their genetic homology. They share significant physiological, metabolic, and biochemical similarities, making them the best model for human disease and intervention [105, 106]. These large animal models are well suited to clinical imaging modalities, such as magnetic resonance imaging (MRI) and computed tomography (CT), and are preferred models for the development of medical devices, such as stents and pacemakers [107]. Regarding the translation of immunotherapeutics, specifically, van Hout et al. have performed a meta-analysis of pre-clinical large animal models treated with anti-inflammatory compounds that have failed to translate to successful clinical trials [108]. Treatments generally led to a reduction in infarct size, supporting the concept of anti-inflammatory therapies. However, the association of these effects with timing, sex, and other experimental variables suggests disparities between pre-clinical and clinical study design that underly translational failures. Specifically, the analysis highlighted that the effect size was greatest when therapeutic intervention occurred early (within 4-8 h post-MI) and when studied only in the male sex. Additionally, procedural mortality was increased when the investigators were blinded to the treatment groups. While both pre-clinical and clinical studies may both blind the investigators, such early intervention is not always clinically feasible and studies should investigate the effect of timing to better understand the effective treatment window. Moreover, pre-clinical investigations should better reflect the population and patient demographics, particularly with regard to sex as a biological variable [109, 110] but also in considering the ancestry of cells such as those used for in vitro studies [111]. As discussed in recent reviews, the interdependence of age and sex is a critical factor in the sexual dimorphism of HF cause, disease progression, and response to treatment [112, 113]. This is particularly pertinent to the study of immunotherapeutics, where response may be influenced either by immunosenescence (a decrease in circulating immune cells and disrupted cytokine response) or inflammaging that is characterized by low-grade chronic inflammation. Considering these factors in experimental design will increase the translational value of research as a whole.

#### **Imaging techniques**

An initial step toward effective therapy includes understanding of disease progression, and a plethora of techniques are available for probing orientation of the immune environment post-MI and assessing functional outcomes. With regard to assessment of LV remodeling, biomedical imaging techniques are widely used in experimental models of CVD and have been recently reviewed elsewhere [114, 115]. These techniques include structural, functional, and biochemical readouts by a variety of techniques that include angiography, echocardiography, MRI, positron emission tomography (PET), CT, and fluorescence imaging. Each of these is a part of the toolbox of techniques available to understand disease progression and therapeutic outcomes (Fig. 2). Through these methods, longitudinal assessment of tissuescale remodeling (e.g., geometry, tissue microstructure) and also cellular processes (e.g., metabolism, enzymatic activity) is made possible, often paralleled by the development and use of molecular imaging probes [116-120]. However, spatiotemporal insights into the behavior of specific immune cell subsets are notoriously difficult to gain from such wholebody imaging or conventional analysis immunological evaluations (flow cytometry, histology).

A better appreciation of the complex relationship between immune cell subsets is often made possible through cell imaging, including intravital microscopy [121]. Direct imaging of cellular processes in disease states can provide fundamental information about cell homing migration, and interactions that are otherwise inaccessible. An array of suitable fluorescent reporter mice and labeling techniques are available [121, 122] and have been used to provide fundamental insights into cardioimmunology both in the healthy and injured heart. For example, Hulsmans and colleagues used a CX3CR1GFP/+ MF reporter mouse to quantify MF abundance in the atrioventricular node and left ventricle, ultimately revealing that MF directly contribute to electrical conduction in the heart via connexin-43-containing gap junctions [123]. Regulatory T cell (Treg) trafficking has been examined in the infarcted myocardium using FoxP3<sup>EGFP</sup> reporter mice, where Treg depletion increased myocardial dilation, upregulated the expression of CCL2, and accelerated MF infiltration. Treg-targeted therapies, owing to their anti-inflammatory properties, could be a promising method for attenuating post-infarct remodeling [124]. The continued use of single cell imaging is well warranted, and continued advancements, such as intravital microscopy in the beating heart [125], will continue to reveal new targets for immune modulation and are likely to become a fundamental tool for assessment of experimental therapeutics.

In addition to the visualization of immune cell subsets at the cell level, gross evaluation of cellular abundance is also useful as a prognostic and diagnostic biomarker of disease [126, 127]. Biomaterial-based contrast agents and molecular probes to assess immune cell populations and their behavior at the tissue scale are in clinical use and continue to be further developed. MRI is a non-invasive imaging tool, widely used due to its minimal radiation exposure and ready use to detect metallic, paramagnetic, and discrete chemical signatures [128]. Superparamagnetic iron oxide (SPIO) and ultrasmall super paramagnetic iron oxide (USPIO) nanoparticles were developed as negative contrast agents for MRI. These nanoparticles are readily uptaken by MF, which make them advantageous for cell mapping in atherosclerotic plaques, infarcted tissues, and solid tumors. Monocrystalline iron oxide nanoparticle-47 (MION-47) and FDA-approved ferumoxytol are similarly able to detect infiltrating MF in atherosclerosis and the infarcted myocardium [129, 130].

Although angiography and CT are universally used for imaging the coronary artery, specifically detecting MF using CT becomes difficult because high concentrations of absorbent biomaterials are required for the X-ray. However, Hyafil and colleagues developed an iodinated nanoparticulate contrast agent, N1177, that could be uptaken in atherosclerotic MF and subsequently imaged with CT in rabbits to determine MF accumulation in the tissue [131]. Additionally, Cormode et al. characterized MF accumulation in atherosclerotic plaque using their developed gold high-density lipoprotein contrast agent for CT in ApoE<sup>-/-</sup> mice [132].</sup>

PET is another form of imaging used widely for cellular tracking. Inflammatory processes, including in the post-MI environment, can be imaged using <sup>18</sup>F-FDG, which is preferentially accumulated in M1-like cells as a result of intracellular transport [133, 134]; concurrent suppression of cardiomyocyte glucose uptake is however necessary to reduce background in cardiac tissues [135]. To combat this, translocator protein (TSPO) may be used to assess inflammatory cell infiltration post-MI, including, for example, <sup>18</sup>F-LW223 to map MFdriven inflammation post-MI [136]. TSPO-PET ligands may be uptaken preferentially by M1-like cells, but likewise accumulate in M2-like MF, neutrophils, Mo, T cells, and B cells to a lesser degree [133]. Using <sup>18</sup>F-FDG in PET and MRI, one study characterized arterial inflammation in atherosclerosis. Results showed that uptake of the compound by MF was significantly higher in plaque-free arterial areas compared to the inside of plaques, suggesting that arterial inflammation does occur in early stages of atherosclerosis [137]. A <sup>68</sup>Ga-NOTAanti-MMR Nb tracer was designed to target mannose receptor on the surface of M2-like MF to demonstrate their abundance and localization in the infarct. Cell mapping with this nanotracer could therefore reveal a better understanding of the resolution of inflammation and predict cardiac remodeling outcomes post-MI (Fig. 3a) [138]. Macrin is a spherical polyglucose (i.e., dextran) nanoparticle, developed by Nahrendorf and colleagues. Using PET for quantitative assessment of cardiac MF, macrin was modified with <sup>64</sup>Cu and used to treat mice, rabbits, and pigs. PET imaging indicated MF accumulation in



◄Fig. 3 Nanomaterials and probes for imaging. a Confocal fluorescence images of <sup>68</sup>Ga-NOTA-anti-MMR Nb, a nanotracer with specificity toward M2-like MF via mannose receptor (MR), uptaken in MF in the infarct zone 7 days post-MI. Figure reproduced from [138]. b PET/MRI of a non-human primate after administration of <sup>18</sup>F-Macroflor over 90 min. The MF-targeted agent is rapidly cleared from circulation by renal excretion (half-life of 21.7 min) to enable subsequent whole-body imaging of MF abundance. Figure reproduced from [141]. c MRI (left) and corresponding histology (right) of infarcted rat hearts before and after injection of the theranostic iron oxide polymer nanocarriers (PP/PS@MIONs), showing MF-targeted accumulation that is further enhanced by application of an external magnetic field (+M). Image reproduced from [143]

the infarcted myocardium in all animals tested [139]. These demonstrate that <sup>64</sup>Cu-macrin serves as an excellent nanotracer for MF, including for applications in cardiovascular medicine and quantitative assessment of tumor-associated MF (TAMs) [140]. The Nahrendorf lab also modified the macrin particle to create <sup>18</sup>F-Macroflor and delivered it to non-human primates, mice, and rabbits, showing enrichment in cardiac and plaque MF (Fig. 3b) [141].

In some instances, these and other imaging probes may be used as a multifunctional tool for simultaneous therapy and diagnostics (i.e., theranostics) [142]. For example, Chen et al. developed a dual targeting theranostic system, PP/PS@ MIONs, composed of magnetic iron oxide nanocubes for visualization via MRI that were enclosed in a zwitterionic copolymer, poly(lactide)-polycarboxybetaine (PLA-PCB, PP). Further surface modification by phosphatidylserine was used to modulate MF phenotypes. MRI showed PP/ PS@MIONS accumulated in significantly greater amounts in infarcted tissue compared to other groups. Treatment also decreased the expression of pro-inflammatory markers, CD86, TNF- $\alpha$ , and IL-1 $\beta$ , and increased that of antiinflammatory markers, CD206, TGF-\beta, and IL-10, in vitro (Fig. 3c) [143] and is an excellent example of theranostic systems that can simultaneously assess and treat underlying pathology. Quite separately, it is interesting to consider that simultaneous image-based evaluation and therapeutic intervention do not have to be administered as a single entity. The presented imaging modalities can additionally serve as companion imaging agents, administered concurrent with therapeutic delivery. Such techniques uniquely enable the real-time quantitative assessment of outcomes. While these techniques have been advanced in the area of cancer immunotherapies [144], their use in cardiovascular medicine has been less well explored.

# **Therapeutic strategies**

While appropriate physiological models and advanced imaging techniques allow for the study of inflammatory processes and their relationship to tissue-level processes, a wide variety of therapeutics (Table 2) allow for direct perturbation of the inflammatory response. Many of these tools are biotherapeutics, comprised of or derived from a biological source. These may include cells, cell-derived products (extracellular vesicles (EVs), antibodies, and cytokines), and even bioactive materials (collagen, decellularized ECM). In contrast, fully synthetic approaches such as small-molecule pharmaceuticals and synthetic biomaterials readily afford scalable production, tunable function, and greater control over batch-to-batch variability.

Therapeutic strategies to address IHF progression can be either cardioprotective (often through immunosuppression) or immunoregenerative. The rationale for immunosuppressive therapies is to (i) attenuate leukocyte-mediated cardiomyocyte apoptosis that contributes to loss of contractility and border zone expansion, (ii) restrain protease activation to limit infract thinning and expansion, (iii) suppress fibrosis, and (iv) prevent secondary cardiac events (e.g., plaque rupture). Reparative strategies are a seemingly natural extension of cardioprotective therapeutics that seek to modulate the phenotype of cells in the infarct environment or selectively recruit progenitor cells to promote angiogenesis or other forms of myocardial regeneration. Here, we review cell-based and pharmaceutical approaches that have been implemented in pre-clinical and clinical studies, while EVs and naturally derived materials are later discussed.

#### Cells

Cell therapies have been widely explored for HF treatment. Stem cell therapies have attracted particular attention, with early studies aimed at functional tissue replacement by cell differentiation. While claims of stem cell transdifferentiation into cardiomyocytes have been disputed [177, 178], they mediate the post-MI environment through a variety of signaling mechanisms to produce abundant growth factors, cytokines, microRNAs, and exosomes that constitute an immunomodulatory secretome [179–181]. It is through these paracrine signals that stem cell therapies modify immune cell recruitment and function [48, 182, 183], offering a means to both reduce detrimental inflammation while simultaneously promoting a switch towards tissue repair processes.

MSCs have become the prevailing cell type for HF treatment because they are pluripotent, genomically stable, and easily harvested from both mouse and human tissue [184–188]. MSCs modulate the inflammatory microenvironment of the myocardium via membrane receptors and a paracrine secretome that affect the migration, apoptosis, and phenotypic polarization of immune cells [48]. The specific interactions that exist between MSCs and immune cells are continuing to come to light. Although studies showing interactions between MSCs and neutrophils are sparse, Kang and colleagues reported a marked increase in neutrophil recruitment after

Therapeutic cargo	Considerations	Examples	References
Cells	Cell sourcing, regulatory approval, and need for GMP processing with appropriate supply chains Potential for in situ recruitment or cell engineering may overcome barriers to production and therapy cost	Endothelial progenitor cells (EPCs) Stem cells MSCs ASCs BMSCs iPSC-derived cardiac progenitor cells CAR T	[145, 146] [147, 148] [149] [150] [151] [152]
Extracellular vesicles and exosomes	Require an abundant cell source for isolation Remain difficult to fully characterize Mechanisms of effect are yet to be better understood	BMSC-derived EPC-derived DC-derived MSC-derived	[153] [154] [155] [156]
DNA/RNA	Diverse structure and function Directly defined function Unstable, require vehicles for delivery Potentially immunostimulatory via TLR activation	miRNA siRNA mRNA siCCR2 siCRMP2	[157–160] [161, 162] [163] [164] [165]
Cytokines and chemokines	Direct biological signals with defined function Suitable for bioconjugation Required dosing may be unknown or difficult to achieve	IFN-γ IL-10 CSF-1 and IL-4 CCL5, CXCL12	[166] [167] [168] [169]
Small molecule drugs	Readily produced at industrial scale and amenable to synthetic modification Potential for oral bioavailability Frequent poor biodistribution May not require intracellular delivery, depending on drug target	Terpines Epinephrine Irbesartan Celastrol 1, 25-Dihydroxyvitamin D3 CCR2 antagonists Pitavastatin Atorvastatin	[170] [147] [171] [172] [173] [174] [175] [176]

 Table 2
 Types of therapeutic cargo to aid in immunomodulation

MSC treatment. In the same study, MSC-conditioned media inhibited neutrophil apoptosis [189]. While neutrophils are crucial in post-MI repair, their overactivation can lead to reactive oxygen species (ROS) production and worsen injury [190, 191]; therefore, further studies are needed to understand these cellular interactions. MSCs have also shown direct effects on MF chemotaxis to the myocardium via signaling molecules such as CCL2, CCL7, and CCL12 [192, 193]. In another example, when MSCs were co-cultured with MF, the culture medium contained lesser amounts of pro-inflammatory markers, such as TNF- $\alpha$ , IL-1 $\beta$ , and IFN- $\gamma$ , and greater amounts of anti-inflammatory cytokines, such as TGF- $\beta$  and IL-10 [194, 195]. Even further, the MSC secretome contains prostaglandin E2 (PGE2), IL-1R $\alpha$ , and TGF- $\beta$ , all of which have been shown to guide M1-like to M2-like MF polarization [194, 196, 197].

MSCs can also mediate the cells of the adaptive immune response. MSCs express inhibitory signaling ligands that bind to complementary receptors on T cells and induce apoptosis, which halts T cell proliferative capacity, downregulates proinflammatory T cell populations, and abates the damaging state of the myocardium after MI [198, 199]. MSCs do not need to be in direct contact with T cells because they possess paracrine factors within their secretome, such as nitric oxide, TGF- $\beta$ , and PGE-2, which prevent T cell proliferation and limit cellular impact on the infarcted heart [200–202]. In addition, culturing T cells with MSCs has resulted in the proliferation of FOXP3<sup>+</sup> Tregs, which are crucial in propagating a reparatory state post-injury [203–205]. Although the known interactions between stem cells and B cells are limited, Che et al. found co-culture with MSCs to suppress B cell differentiation and proliferation [206]. ESCs have been shown to differentiate into M1-like and M2-like MF phenotypes and alter the inflammatory environment accordingly [207]. For example, Kudo and colleagues created an ESC-derived suppressor cell line containing a hybrid M1-like and M2-like MF phenotype that suppressed T cell responses [208]. When directly exposed to ESCs, CD3<sup>+</sup> T cell populations within the myocardium increased, which induced Treg differentiation. However, due to the plasticity of Tregs, the resulting response tended to be heterogenous [209–211].

While stem cell-based therapies have largely focused on anti-inflammatory paracrine effects as mediators of LV remodeling, a critical alternative model of action has recently been proposed. Vagnozzi et al. proposed a comprehensive proinflammatory immunoregenerative hypothesis as the mechanism of therapeutic activity [212]. They compared the effects of locally injecting either mature stem cells or zymosan (a tolllike receptor 2 agonist) into healthy hearts, both of which produced transient accumulation of activated (CCR2<sup>+</sup>, CX3CR1<sup>+</sup>) MF at the site. When applied to the following ischemic injury, both treatments exhibited comparable improvements in cardiac function 2 and 8 weeks post-injection, relative to saline controls. These benefits were lost when mice were immunosuppressed or when MF were depleted, indicating that MF presence and activation were essential, including towards preferential alteration in ECM content and associated mechanical properties. Similar results were observed with injection of non-viable stem cells, ruling out potential paracrine signaling mechanisms. Taken together, these studies indicate that the transient accumulation of activated MF subtypes following immunostimulatory injection improves function of the injured heart by influencing cardiac fibroblasts.

Direct transplantation, including of naturally occurring or engineered immune cells, has also emerged as a cardioprotective treatment for MI. Such adoptive cell transfers using naturally occurring Tregs are an attractive approach [213], which has been leveraged by Sharir and colleagues to influence LV remodeling. The adoptive transfer of Tregs in mice reduced infarct size, attenuated LV remodeling, and improved heart function. Treg depletion using anti-CD25, however, had no effects on cardiac repair [214]. In vitro, Tregs are able to modulate Mo differentiation to a more anti-inflammatory subset. An in vivo myocarditis model was treated with Tregs, which showed cardioprotection against inflammatory damage and fibrosis through Mo modulation [215]. Meng et al. have further explored engineered cell therapies, wherein they induced MSC overexpression of IL-10 using CRISPR. Treatment showed increased IL-10 expression in the heart and decreased inflammatory cell infiltration, pro-inflammatory markers, and cardiac cell apoptosis, all of which improved cardiac recovery [216]. In recent groundbreaking work, the Epstein lab uniquely targeted cardiac fibrosis by the targeted elimination of myocardial fibroblasts, accomplished via adoptive transfer of CAR T cells active against fibroblast activation protein (FAP). Treatment significantly reduced cardiac fibrosis and partially rescued heart function in a mouse model of hypertensive cardiac injury [152].

The studies outlined here provide a basis for the beneficial effects of cell therapy on the post-infarct myocardium. The ability of stem cells to recruit immune cells to the injured area and facilitate modulation of their function is a promising methodology; though, the underpinning mechanisms are continuing to be better understood. Moreover, adoptive cell transfer is an exciting avenue for more targeted and intentional therapeutic outcomes. These cell therapies are at the cutting edge of cardioimmunology, particularly in the case of engineered effector cell types that uniquely enable discrete manipulation of the post-MI immune microenvironment through the targeted depletion of harmful cell types or the selective production of reparatory soluble signals. While such cell therapies are challenged by issues of cell sourcing, in vitro expansion, and need for the maintenance of supply chains in GMP processing, autologous cell therapies are among the fastest expanding markets for immunoncology [217]. Improvements to the in vivo lifetime of these cells and the ability to generate them directly in situ (vide infra, lipid-based nanoparticles) will continue to advance their road to the clinic.

#### **Biomolecules**

Therapeutic biomolecules include a host of cell-derived products, ranging from proteins and antibodies to RNA and peptides. In many instances, these biological signaling molecules may be isolated components of a particular cell population's secretome with well-defined immunological function. Of the biomolecules that are able to modulate the immune system after MI, proteins make up a significant contribution and include growth factors that can contribute to cardiac repair. Fibroblast growth factors (FGFs) facilitate a number of biological processes; Joki et al. injected FGF21 in a murine model post-MI and found that FGF21 exhibited anti-inflammatory properties (decreased TNF- $\alpha$ , IL-6), which attenuated remodeling and cardiomyocyte apoptosis while encouraging blood vessel formation [218]. Vascular endothelial growth factor (VEGF) has been extensively studied for cardiac repair, owing to its role in angiogenesis [219]. Rosano et al. delivered VEGF to the infarct in a rat MI model. The delivery of VEGF reduced collagen deposition, increased systolic function, and promoted microvascularization [220]. Bauza and colleagues investigated the effects of HMGB1, a non-histone chromatin binding protein and pro-inflammatory alarmin, on sheep with acute MI. Results showed that high-dose HMGB1 injection increased Ki67<sup>+</sup> cardiomyocytes and overexpressed VEGF. This was accompanied with enhanced LV ejection fraction and wall thickening [221]. These outcomes are an interesting parallel to the earlier discussed treatments with zymosan, supporting the role of early pro-inflammatory interventions to promote reparatory processes in the injured heart.

Peptide therapeutics first emerged a century ago and have since become widespread in the field due to their low toxicity, high potency, and strong selectivity [222]. The cardioprotective potential of peptide therapeutics in MI has also become a field of interest. Qin et al. employed a glucocorticoid-regulated anti-inflammatory mediator, annexin-A1 (ANX-A1), and demonstrated its ability as a "triple shield" therapy, inhibiting neutrophil infiltration and preserving both cardiomyocyte viability and myocardial contractility [223]. Stromal cell-derived factor (SDF)-1 $\alpha$  has previously been reported to improve vasculogenesis and cardiac function after MI. However, the bulky structure and short halflife are suboptimal for therapeutic use. Therefore, Hiesinger and colleagues developed a minimized peptide analog of SDF-1 $\alpha$  and demonstrated improved ventricular function in a rat model of MI [224]. In an interesting example, a purified leech peptide was able to inhibit MF migration through mechanisms involving JNK and p38 MAPK pathways [225].

Under the umbrella of protein-based drugs, antibodies have also emerged as potential therapeutic strategies. Unfortunately, early clinical trials using antibodies to target glycoprotein receptors on the surfaces of immune cells did not show positive results. For example, Baran and colleagues investigated the efficacy of an anti-CD18 recombinant monoclonal antibody in a double-blind randomized trial. It was found that while the antibody was tolerated, cardiac end points, such as coronary blood flow or infarct size, were not improved [226]. In another example, antibody blockade of the CD11/CD18 integrin receptor was investigated. However, treatment resulted in no reduction of infarct size in patients with acute MI [227]. Pexelizumab, a humanized monoclonal antibody, binds to the C5 component of the complement cascade and has been implicated in apoptosis inhibition and leukocyte infiltration in experimental models [228, 229]. In a clinical trial with over 5000 patients, pexelizumab treatment showed no effects in improving acute MI [230]. Trials targeting interleukins released from activated immune cells also showed limited benefit. Abbate et al. conducted a pilot study using anakinra, a recombinant IL-1 receptor agonist. There were no significant differences between control and treatment groups when comparing the primary endpoint of LV end-systolic volume [231]. IL-6 has been shown to contribute to atherosclerotic plaque destabilization, which leads to MI [232]. Therefore, Kleveland and colleagues employed tocilizumab, a humanized anti-IL-6 receptor antibody, in a clinical trial for MI. Results showed little to no effects on attenuating the acute inflammatory response [233]. In the MRC-ILA Heart Study, an IL-1 receptor agonist was directly injected into patients with acute coronary syndrome. The study concluded that treatment showed some reduction in inflammatory markers; however, further studies would need to be conducted to confirm these findings [234]. The emphasis on cardiac end points only and the lack of elucidating biological mechanisms from these antibody treatments leave the reasoning for failed clinical trials open to interpretation, and the use of later discussed biomaterials to enhance biodistribution and cell targeting of biotherapeutics could be an attractive method to improve clinical outcomes.

#### Small molecule drugs

Small molecule pharmaceuticals are frontline immunotherapeutics, with applications toward a myriad of chronic immune diseases [235]. Anti-inflammatory therapies have been widely employed in the context of IHF and are a mainstay of current medical management that have been the topic of recent and direct review [236, 237]. Notably, there remains ongoing concern that anti-inflammatory therapies alone may worsen outcomes by inhibiting inflammation-dependent repair mechanisms, including angiogenesis. As these topics have been recently and thoroughly reviewed, only a brief discussion of pro-regenerative immunotherapies is included here.

Resolvins are bioactive lipid mediators that have shown success in inflammatory resolution through interactions with surface receptors on leukocytes [238-241]. Resolvins primarily function to inhibit neutrophil and Mo migration, which can protect tissue against chronic inflammatory injury [242]. In an ApoE<sup>-/-</sup> mouse model, resolvin D2 and maresin 1 treatment prevented atheroprogression by driving MF toward a reparatory phenotype [243]. Treatment with resolvin D2 shows increased myocyte numbers with decreasing levels of TNF- $\alpha$ , granulocyte MF colony-stimulating factor (GM-CSF), and neutrophil migration [244]. In another study, although resolvin E1 reduced expression levels of TNF- $\alpha$  and IFN- $\gamma$ , MF infiltration to the atherosclerotic plaque did not decrease [245]. In a rat model of IR, resolvin E1 reduced leukocyte infiltration 4 h after reperfusion, concurrent with a reduction in infarct size [246]. The same group demonstrated decreased neutrophil infiltration and infarct size in another study centered around MI and depression in rats [247]. In a C57BL/6 J mouse model of coronary artery ligation, resolvin D1 limited neutrophil recruitment in the myocardium, decreased the expression of fibrotic genes, and reduced collagen deposition, all of which ameliorated fibrosis and stabilized the ECM [240].

Statins are promising agents because their anti-inflammatory properties are driven by a plethora of factors. For example, they can inhibit leukocyte migration through decreasing the expression of ICAM-1 and MCP-1 and modulate T cell activity through inhibition of Th1 chemokine receptors [248-250]. In addition, statins can reduce the release of C-reactive peptide, cytokines, chemokines, and adhesion molecules [248]. Shibasaki et al. investigated the effects of pitavastatin in  $ApoE^{-/-}$  mice, finding that arterial inflammation in atherosclerotic plaque was reduced [251]. Simvastatin administration to  $ApoE^{-/-}$  mice decreased the expression of HMGB1, VCAM-1, and MCP-1, in addition to reducing vascular inflammation and atherosclerotic lesions [252]. In one clinical trial, patients were treated with a high dose of atorvastatin and moderate dose of rosuvastatin. Regarding inflammatory activity, both treatments similarly reduced TNF- $\alpha$ and IL-6 [253]. Liu et al. conducted a clinical trial to test if atorvastatin is able to limit inflammation and improve cardiac function after MI. Drug administration demonstrated low levels of C-reactive protein and MMP9, with improvements in LV ejection fraction and heart function [254]. While statins possess antiinflammatory activity, they have also been shown to promote a reparatory M2-like phenotype. In a rat model of MI, atorvastatin administration downregulated pro-inflammatory markers, such as IL-1 $\beta$ , TNF- $\alpha$ , and iNOS, and upregulated anti-inflammatory markers, such as Arg1, indicative of a shift from an M1-like MF phenotype to M2-like [255].

Other small molecule drugs have also shown efficacious results in modulating the inflammatory response post-MI. Cyclophosphamide administration in a rat IR model, for example, resulted in lower rates of leukocyte infiltration and reduced the propensity of ventricular dysfunction [256]. In the COLCOT clinical trial, colchicine, an anti-inflammatory drug targeting MF migration, lowered the risk of adverse cardiac events in post-MI patients [257]. Pyruvate kinase isozyme type M2 (PKM2) is an enzyme in the glycolytic pathway that regulates inflammation in LPS-activated MF [258]. Iminostilbene, a modulator of PKM2, was shown to suppress levels of pro-inflammatory markers, such as IL-1 $\beta$  and IL-6, reduce infiltration of CD86 MF, reduce the phosphorylation of the STAT3 inflammatory pathway, and alleviate cardiomyocyte apoptosis in vitro and in vivo [259].

These and other immunomodulatory drugs are often identified by drug screening processes. For example, Hu et al. performed high-throughput drug screens of approximately 4000 compounds across a variety of drug classes to identify targets for MF modulation [260]. Currently, large drug screenings like this primarily focus on M1-polarizing agents, with limited success in identifying or following up on M2-promoting compounds. More recently, combinatorial drug screens combining immunostimulatory and immunosuppressive drugs or multiphase screens have been used to identify pharmacological promoters of tolerogenic DCs and M2-like MF, respectively [261, 262]. Continued developments in drug screening methods, such as automated drug screening and methods to directly assay for M2-like promotion, will continue to move this field forward. As for antibody-based strategies, clinical trials have shown limited benefits to date and may gain better insights through the addition of secondary outcomes that include assessment of LV remodeling and inflammatory mediators.

# Nanomaterials

While therapeutics alone are clearly efficacious tools for reorientation of the post ischemic inflammatory milieu, their use can be hindered by factors such as suboptimal pharmacokinetics (i.e., rapid blood clearance, non-specific cell and tissue biodistribution) and resulting off-target effects such as systemic toxicity and increased risk of infection [263]. Functional biomaterials have been widely used to address these challenges in cardiac repair [264, 265], which may be composed of either natural or synthetic components (Table 3). Here, we outline biomaterial-based strategies, including systemically or locally administered therapeutic vehicles that have demonstrated utility in modulating the immune response to mitigate impacts toward IHF. However, it is worth noting that biomaterials themselves can have a profound effect on immune cell behaviors (Fig. 4), including via material composition or surface properties [266]. Here, we briefly review these nanoparticle properties in the context of IHF; the following sections focus on the use of these systems as drug carriers, used primarily for systemic administration to enable cell-targeted delivery.

#### **Nanomaterial properties**

In the case of polymeric materials in particular, specific receptormediated interactions with immune cells can occur that are critical to immune engineering and biomaterial design [267]. Hyaluronic acid (HA), a glycosaminoglycan prevalent in the ECM, interacts directly with CD44, CD168 (receptor for HAmediated motility, RHAMM), and toll-like receptors (TLRs). These interactions are critical toward neutrophil recruitment and MF polarization and are highly dependent on polymer molecular weight [268, 269]. Molecular weight of HA has been shown to have varying MF polarization potential and cardioregenerative effects [270, 271]. For example, 50 kDa, 130 kDa, and 170 kDa HA hydrogels were used to treat experimental MI, with 50 kDa HA exhibiting the greatest myocardial regeneration and functional recovery [272]. Wang et al. investigated short-chain HA fragments (6-10 disaccharides) that decreased the inflammatory response caused by neutrophils and facilitated MF polarization to the M2-like phenotype in a mouse MI model [273]. In an

 Table 3
 Material compositions used in both nanotherapeutic and bulk materials

Material composition	Considerations	Examples	References
Natural	Often widely regarded as biocompatible and biodegradable May include native functions, such as sites for cell adhesion or immune modulation by defined receptors Often well suited for modification by bioconjugate reactions For general discussion, see [309, 310]	Cardiac ECM Splenic ECM Hyaluronic acid Collagen Silk protein Alginate Chitosan Silica nanoparticles	[149, 311–314] [315] [151, 156, 316] [65, 317] [274] [153, 155, 168, 318] [150] [157]
Synthetic	Allow for user-defined physical and chemical tunability Require direct synthesis and purification For general discussion, see [319, 320]	PEG-based micelles, hydrogels PLGA nanoparticles Zwitterionic co-polymers Bioactive co-polymers Lipid nanoparticles	[148, 172–174] [171, 175] [143] [169, 321] [161–165]



Fig.4 Diversity of immunotherapeutic nanomaterials. **a** Composition, properties, and therapeutic cargo dictate how nanoparticles interact with immune cells. These aspects enable cell-targeted delivery, receptor-mediated control of cell programs, and influence over downstream effector or suppressor signaling programs. **b** Lipid nanoparticles, synthesized with varying surface

charges, were incubated with human MF in vitro; surface charge positively correlated with cell uptake. Figure reproduced from [283]. c siRNA loaded particle treatment (siCCR2) silences CCR2 to reduce inflammatory Mo infiltration and MF populations compared to the control (siCON) following IR injury. Figure reproduced from [164]

interesting example of proteinaceous hydrogels, Song and colleagues formed an injectable hydrogel from sericin, a silk-derived protein. In a mouse model of MI, the hydrogel downregulated the expression of inflammatory markers, such as TNF- $\alpha$  and CCL2, by suppressing TLR4/NF- $\kappa$ B pathways and ultimately decreased the number of MF in the infarct region by 45.8% [274].

Surface modification of materials is also a useful technique for targeting immune cells and modulating their response. Seminal work in this area of cardiac immune engineering was performed by the lab of Smadar Cohen and used phosphatidylserine-presenting liposomes to mimic apoptotic cell endocytosis, in turn polarizing MF towards an M2-like state [275]. Specific pathways can be ingeniously targeted by such methods, and deoxyribozyme (a DNA enzyme able to silence TNF- $\alpha$ ) was conjugated to gold nanoparticles because DNA structures are easily internalized by nucleated cells [276]. After injection in a mouse model of acute MI, TNF- $\alpha$  levels were knocked down by 50% which better maintained cardiac function [277]. Richart and colleagues created nanoparticles made of apolipoprotein AI reconstituted with phosphatidylcholine (n-apo AI), which resembled high-density lipoprotein particles. Following MI, n-apo AI administration decreased the expression levels of chemokines that facilitate leukocyte recruitment by 60-80%, thus reducing the numbers of neutrophils and Mo in the myocardium and attenuating inflammation [278]. In another example, researchers developed  $\alpha$ -gal epitope nanoparticles to incite recruitment of reparatory MF via activation of complement cascade and corresponding chemotactic cues. This was further confirmed by the repopulation of cardiomyocytes and restoration of normal cardiac structure and contractile function in the mice, suggesting a truly regenerative rather than cardioprotective treatment [279].

Surface chemistry and topography of biomaterials has likewise demonstrated distinct influence on contacting cells [280]. For example, neutrophils secrete greater levels of pro-inflammatory cytokines when in contact with hydrophobic surfaces [281], and surface roughness induces greater neutrophil death and ROS production [282]. MF has been likewise studied in this context, in part due to their prominent role in the foreign body response (FBR). Surface charge can have a substantial role in interactions with MF. For example, the surface modification of polystyrene nanoparticles has been used to demonstrate MF uptake in vitro is directly correlated with the surface zeta potential (Fig. 4b) [283]. Hamlet and colleagues demonstrated that hydrophilic surfaces decreased pro-inflammatory cytokine expression in human and mouse MF [284, 285], and similar outcomes have been noted for DCs [286]. In both in vitro and in vivo studies, increasing stiffness of the substrate is associated with a higher prevalence of the M1-like MF phenotype [287, 288], whereas softer surfaces are associated with a lesser FBR and fibrous encapsulation [289]. MF complexity has shown mixed results regarding surface roughness [290, 291], and recent results suggest that governance by Tregs may be responsible for such behavior in vivo [292]. Nanoparticle shape can also be readily tuned [293], providing unique opportunities to adjust how specific cell types interact with these materials. For example, elongated nanoparticles are preferentially uptaken by neutrophils as compared to other innate immune cells, providing the ability to discretely target these drug carriers [294]. These findings indicate the necessity of designing immunomodulatory devices and delivery systems with material composition and structure in mind. By doing so, it is possible to harness these aspects of cell-material interaction to not only aid in cell-targeted therapeutic delivery but also to improve treatment outcomes by the rationale design of drug carriers that complement or synergize with the action of encapsulated therapeutics.

#### **Polymeric nanoparticles**

Nanoparticles are valuable drug delivery vehicles [295], most frequently used for systemic administration. They are well suited to the encapsulation of various small molecule drugs, RNA, and other biomolecules with dependence upon nanoparticle structure and material selection. Nanoformulation of cargo is particularly useful to improve bioavailability by enhancing drug solubility, preventing rapid renal clearance, and shielding sensitive cargo (e.g., proteins, nucleic acids) from enzymatic degradation [296, 297]. Furthermore, these systems have the potential to target delivery to specific cells or tissues via surface ligand modification, minimizing off-target exposure [298, 299]. Here, we will review the use of polymeric nanoparticles in delivering a variety of cargo, including small molecule drugs, mRNA, and others.

As discussed above, a plethora of immunoactive small molecule pharmaceuticals exist at varying stages of development and exploration towards cardiovascular engineering. As many of these are hydrophobic small molecules, they are amenable to encapsulation in polymeric nanoparticles, such as by nanoprecipitation and emulsion methods. Irbesartan is an angiotension II type I receptor blocker with a PPARy agonist effect. In a murine IR model, PLGA nanoparticles loaded with irbesartan demonstrated inhibited recruitment of inflammatory Mo to the heart, reduced infarct size via PPARy-dependent mechanisms, and improved LV remodeling after 3 weeks [171]. As mentioned, statins have also demonstrated cardioprotective effects. Pitavastatin-loaded nanoparticles were intravenously injected into C57BL/6 mice with permanent coronary ligation. Nanoparticles were uptaken by CD11b<sup>+</sup> Mo/MF and reduced their prevalence in the infarcted heart and spleen, which ultimately attenuated LV remodeling [175]. In another example involving statins, atorvastatin loaded supramolecular copolymers demonstrated cellular drug uptake in MF and an increased ratio of M2-like to M1-like presence by 6.3-fold in an in vitro cholesterol model [176, 300].

#### Lipid-based nanoparticles

Micelles are nanosized spherical vesicles composed of a lipid monolayer. During self-assembly, micelles form a hydrophobic core, which allows for incorporation of hydrophobic drugs [301]. These systems have been widely used to modulate the immune microenvironment post-MI. Allen et al. loaded celastrol, a small molecule immunotherapeutic, into poly(ethylene glycol)b-poly(propylene sulfide) (PEG-b-PPS) micelles. Celastrolloaded micelles reduced secretion of TNF- $\alpha$  in RAW264.7 cells in vitro, and their delivery decreased neutrophil and Mo recruitment to atherosclerotic plaque in  $LDLR^{-/-}$  mice [172]. Wang and colleagues developed PEG-based micelles loaded with a small molecule CCR2 antagonist and surface decorated with an anti-CCR2 antibody for cell targeting. Treatment in a murine MI model significantly decreased the number of Ly6Chigh inflammatory cells compared to the control group, while also reducing infarct size [174, 302].

Liposomes are among the first nanoformulations to be clinically used because of their amphiphilic composition that promotes encapsulation of hydrophobic drugs in the lipid bilayer and hydrophilic drugs in the aqueous cavity [175]. In a rat model of acute MI, intravenous injections of phosphatidylserine-presenting liposomes upregulated the expression of antiinflammatory cytokines, such as TGF- $\beta$  and IL-10, increased the number of anti-inflammatory CD206<sup>+</sup> MF, and decreased the levels of pro-inflammatory markers, such as TNF- $\alpha$  and CD86 [275]. In a similar study that employed the same type of liposomes, researchers were able to upregulate the expression of anti-inflammatory genes, while downregulating the expression of pro-inflammatory genes for infarct repair in vivo [303].

The use of RNA as a therapeutic cannot be understated given the success of the mRNA-based vaccines for the recent COVID-19 pandemic. Because many diseases, like cancer and immune disorders, have discrete genetic targets, delivery of RNA is a feasible strategy for treatment, and various RNA therapeutics have been explored in regenerative medicine [304, 305]. However, the delivery of RNA alone is susceptible to rapid degradation and off-target effects [306]. Encapsulation is therefore useful to safely carry RNA to sites of interest [307]. In targeted applications for IHF prevention, siRNA has been widely explored both in ischemic injury and atherosclerosis. For example, collapsin response mediator protein-2 (CRMP2) was shown to be involved in MF polarization; therefore, Zhou and colleagues loaded siCRMP2 into lipid nanoparticles (LNPs), finding MF polarization from M1-like to M2-like, decreased inflammatory and fibrosis markers, and attenuation of LV remodeling in both WT and  $ApoE^{-/-}$  mice [165]. Courties and colleagues likewise identified high levels of interferon regulatory factor 5 (IFR5) expressed by inflammatory MF after injury; siIRF5 delivery reduced the expression levels of M1-like MF markers, supported

inflammation resolution, and promoted infarct healing [161]. In another example, LNPs carrying siCCR2 were shown to accumulate in splenic phagocytic cells and localized to Mo when administered to mice. This treatment significantly decreased the level of inflammatory Mo and MF in atherosclerotic plaque in the ApoE<sup>-/-</sup> model and reduced infarct size following coronary artery occlusion (Fig. 4c) [164]. In yet another study by the Nahrendorf group, siRNA targeting five different cell adhesion molecules were loaded into a single endothelial cell targeted polymeric nanoparticle. Treatment in the ApoE<sup>-/-</sup> and coronary ligation models attenuated leukocyte recruitment to these sites and improved outcomes [162, 308].

While many RNA delivery strategies have focused on silencing specific targets of interest, the same means can be used for cell and gene therapy to promote immunoregulatory behavior. In an exciting example led by the Epstein lab, earlier discussed methods of CAR T therapy have been recently adapted to in situ cell therapies, eliminating the need for initial cell isolation and adoptive transfer [163]. In this work, mRNA necessary for CAR T reprogramming was encapsulated in CD5-targeted LNPs, enabling the transient in vivo generation of FAP CAR T cells that reduced fibrosis and restored cardiac function after injury. Biomaterial-based strategies such as these are invaluable advances in the field-they hold promise to revolutionize the face of cell and gene therapies by eliminating the time and labor-intensive supply chain required for cell manufacturing. By performing these cell manipulations directly within the body, these therapeutic strategies are reduced to a cost-effective offthe-shelf approach that is more accessible to broad use.

### **Biologically derived nanoparticles**

Within the body, cells release EVs through endosomal pathways and budding from the plasma membrane. These naturally arising nanoparticles contain RNA, proteins, and other soluble or membrane-bound factors that are fundamental to understanding, as well as manipulating intercellular communication [322]. Immune cells continually exchange EVs as part of the dynamic network of communication among the innate and adaptive immune compartments. For example, activated DCs express co-stimulatory CD80 and CD86; their secreted EVs can therefore activate T cells [323]. Furthermore, miRNAloaded EVs transferred from Tregs to Th1 cells have been shown to reduce the Th1-driven inflammatory response [324], and MSC-derived EVs suppress inflammatory MF activation through modulated NF-κB pathway signaling [325]. Cardiacderived cells (CDCs) similarly mediate the polarization from an M1-like to M2-like MF phenotype as well [161, 326, 327]. As such, the injection of CDC-derived exosomes demonstrated an increase in anti-inflammatory gene expression, accompanied by a decrease in pro-inflammatory expression [328].

The mechanisms of these effects in cardiac tissues have been both explored and manipulated for therapeutic benefit. The cardioprotective benefits of MSC-derived exosomes, for example, have been associated with miR-182 content, a potential mediator of MF polarization and TLR4 expression [158]. MSCderived EVs have also been purposefully loaded with exogenous miR-101a to target TGF-B and Wnt signaling and attenuate fibrosis [159]. M2-like MF, which were programmed to secrete miR-148a exosomes, were shown to reduce infarct size and improve cardiac function post-MI in vivo [160]. Wu and colleagues engineered M2-like MF exosomes with hexyl 5-aminolevulinate hydrochloride (HAL), an FDA-approved imaging agent that has been shown to initiate the production of anti-inflammatory compounds. The system exhibited anti-inflammatory capabilities and reduced progression of atherosclerosis [329]. While these examples display the complex role of EVs as well as their potential as tunable immunotherapeutics for CVD, a full understanding of their mechanistic origin and functionality remains lacking and will no doubt contribute to further advances in this exciting area.

#### **Bulk materials and devices**

While nanotherapeutics have emerged as critical drug delivery vehicles for cell-targeted delivery, they rarely allow tissue-specific tropism and therefore do not address issues of off-target drug effects such as systemic immunosuppression. Towards this goal, bulk biomaterials and devices have advanced in a parallel manner and offer additional means of mitigating LV remodeling-such as infarct restraint to prevent infarct or LV dilation. For the development of biomaterial strategies to treat MI, multiple design factors must be considered. These include the intended therapeutic payload, material composition, and overall structure-each of which can influence the immune response either inadvertently or for intended effects. As bulk materials are locally applied interventions, the device structure and route of introduction must also be considered. Frequently employed methodologies are myocardial wraps and patches surgically applied to the epicardial surface (Fig. 5) and injectable biomaterials that may be applied by coronary perfusion, intramyocardial injection, pericardial injection, or other means (Fig. 6). Wherever possible, minimally invasive routes of introduction are preferable to open thoracotomy. These considerations are in large part due to procedural complexity, cost, and associated risks of morbidity and mortality. Moreover, it is increasingly realized that surgical stress profoundly influences the systemic immune environment, with implications in both cancer progression and postoperative cardiovascular events [330-332].

#### **Externally affixed devices**

Externally affixed myocardial wraps and patches are a form of mechanical stabilization for post-MI treatment, originally intended as a prophylactic strategy to directly prevent infarct

Fig. 5 Epicardial affixed devices such as patches and wraps allow for mechanical stabilization of the infarct and can simultaneously deliver therapeutics or incorporate bioactive materials. a The evolution of epicardial affixed devices initiated with mechanical restraints to prevent LV dilation and has moved to incorporate living tissue constructs and bioactive materials for immunomodulation. b Schematic of the Therapi system, which incorporates a semipermeable membrane in contact with the heart surface and a delivery reservoir, replenishable via an externally accessible refill port. Luciferase-expressing MSCs were loaded before implantation (control) and optionally re-filled (day 4). Figure reproduced from [147]. c Synthesis of PTFU (an ROS scavenger) combined with PTK and PPF is clicked with pro-angiogenic REDV peptides to create a multifunctional macroporous cardiac patch. The cardiac patch is further loaded with rosuvastatin and surgically implanted onto the LV ischemic areas of rat hearts in an acute MI model. In vivo, the patch acts as a ROS scavenger and regulates MF phenotype. Figure reproduced from [321]



CD86/ CD163

expansion and LV dilation. These early biomaterial-based interventions are referred to as ventricular restraint devices (VRDs), and often use a wrap or patch to mechanically restrain the heart [333–335]. One type of VRD arose in the form of a cardiac wrap to enable diastolic reinforcement, called the Cor-Cap Cardiac Support Device (CSD, Acorn Cardiovascular). The CSD is a polyester mesh that fits around the ventricles to reduce wall stress and prevent LV dilation. The device was shown to reduce stress response proteins, attenuate cardiomy-ocyte hypertrophy, and normalize MHC isoforms contributing to improved myocardial kinetics in large animal models

and long-term benefit in clinical trials at 5-year follow-up [336–338]. Progress towards similar devices have included the HeartNet (Paracor Medical, Inc.) [339], which is placed around the heart with an introducer sheath via minithocotomy as well as the quantitative ventricular restraint device (QVR; Polyzen Inc.) that incorporates an inflatable balloon structure and access line for adjusting the heart volume and pressure [340]. Advances in VRDs and related direct cardiac compression devices, as well as their relative advantages and disadvantages, have been the topic of focused review by Naveed et al. [335, 341].

Fig. 6 Injectable materials, such as hydrogels, potentiate minimally invasive and local delivery of therapeutic cargo to the heart via intramyocardial or pericardial injection. a Hydrogels loaded with cells and biologics can be delivered to aid in immunomodulation, while the hydrogels themselves provide needed mechanical restraint of the infarct. b Shearthinning Ad-HA and CD-HA hydrogels including IL-10-loaded NorHA microgels were injected into the border zone of the infarct in a rat MI model. Local delivery of IL-10 decreased CD68+MF after 1 week. Figure reproduced from [167]. c The pericardial space acts as a natural mold for hydrogels to form a cardiac patch in situ and release loaded therapeutics. Pericardial injection of methacrylated HA hydrogels with MSC-derived exosomes in pigs increases exosome retention in the heart and offers a local and minimally invasive delivery approach. Figure reproduced from [151]



Since their conception as VRDs, externally affixed devices have further evolved to incorporate immunotherapeutic payloads, bioactive materials, and living tissue constructs. In particular, the active hydraulic ventricular attaching support system (ASD) device was designed as a multi-purpose device. The ASD is composed of a mesh cover that incorporates silicone tubes, accessible via an external port for injection. The tubes can be used to locally deliver therapeutic drugs [170], as well as to apply altered pressure to tune LV restraint similar to the QVR [342]. Likewise, another device of note is the Therapi system that was designed as an epicardial reservoir, amenable to minimally invasive implantation and refillable loading for the local and sustained presentation of therapeutic cargo [147]. The system is placed in the epicardium, where a subcutaneous catheter port allows for local delivery of therapeutics and a minimally invasive refillable component (Fig. 5b). The delivery of small molecules (epinephrine), macromolecules (dextran and albumin), and cells (MSCs) demonstrated a range of potential cargo. Repeated cell dosing post-MI better maintained heart function, suggestive of the intended therapeutic benefit.

While ventricular restraints (wraps) are more efficacious than infarct stiffening (patches applied to the infarct area alone) [343], cardiac patches are likewise a means of myocardial restraint that can be readily tuned in composition and payload. Patches can be formed from natural (e.g., polysaccharides, decellularized ECM, proteins) or fully synthetic (e.g., PLGA, PLLA, PEG) starting materials, each of which have their own benefits [344–346]. Natural biomaterials often lend themselves towards mimicking the mechanical properties, resorption behavior, and cell interactions of native tissues, whereas synthetic polymers or modifications readily enable tuning of these and other material properties [267, 320, 347]. One promising method for cardiac patch creation incorporates the use of the decellularized ECM because it contains proteins and proteoglycans that can allow for cell attachment and proliferation to facilitate cardiac repair [348, 349]. Sarig et al. investigated the use of a decellularized porcine cardiac ECM patch (pcECM-P), applied to Wistar rats in either the acute or chronic inflammatory phase. The patch induced constructive remodeling, attributed in part to a stark increase in the M2/M1-like MF ratio that was associated with enhanced vascularization and cardiomyocyte differentiation markers [311]. Further work by Ge Zhang and colleagues has used decellularized porcine myocardium slices (dPMS), either as an acellular patch or after seeding with adipose-derived stem cells (ASCs) [149, 312]. At 4 weeks post-MI in rats, acellular dPMS treatment was associated with a more robust MF infiltrate and markedly higher M2/M1-like ratio that was associated with increased vascular density and better preserved fractional shortening. Both rat and pig ASCs were readily able to infiltrate the matrix by seeding in vitro, improving their local retention as compared to direct injection. Such approaches are a promising strategy to encompass infarct restraint, modulation of host immunoregenerative response, and cell therapies within a single approach.

By the inclusion of other cell types, living tissue constructs can also be formed as cardiac muscle patches (CMPs) to treat HF. CMPs can not only assist in ventricular contraction, but also potentially contribute to molecular and electrical signaling. Cardiomyocytes, cardiac vascular cells, and fibroblasts have most commonly been used [350]. However, one study composed a bi-layer cardiac patch from hiPSC-derived cardiomyocytes and a sheet of blood outgrowth endothelial cells (BOECs) and pericytes (PCs). Grafting in a nude rat infarct model attenuated infarct fibrosis and thinning, in part due to microvascular connection between the graft and host tissue [351]. Similarly, Weinberger and Querdel et al. created hiPSCderived cardiomyocyte patches from 3 separate hiPSC lines and studied outcomes in guinea pigs and pigs. In guinea pigs, the patch formed heart muscle, improved electrical function, and increased LV function. When used in pigs, there was successful transplantation and evidence of cardiomyocyte proliferation, suggesting a promising method for post-MI healing [352, 353]. Given the newly established role of cardiac MF in electrical conduction in the heart, it is possible that inclusion of such cell types may further enhance electrical connectivity of CMPs and the native tissue, which has remained an ongoing challenge of the field. Additionally, these fields highlight a need for continuous pharmacological immunosuppression to be used for certain cell sources.

Through further tuning of material composition and cell or drug cargo, cardiac patches can be further developed to directly modulate the post-MI environment [346]. Hosoyama et al. have reported the development of a bilaminar cardiac patch, composed of an elastic hydrodynamic support coupled with aligned electrospun collagen that contained silver (AgNP) or gold (AuNP) for electroconductivity. Interestingly, only AuNPcontaining patches preferentially skewed the M2/M1-like phenotypic ratio towards the creation of a reparatory environment [317]. In another excellent example of synthetic adaptations to develop multifunctional and immunomodulatory patches, elastomeric cardiac patches were prepared to restrict LV remodeling. The patch was composed of polyurethane and unsaturated poly(thioketal) (PTK, as an ROS scavenger), further modified by a pro-angiogenic peptide (REDV) and incorporated rosuvastatin (Fig. 5c). In addition to provision of mechanical stabilization, the multiple precise modifications were intended to suppress early inflammation, foster angiogenesis, and prevent fibrosis throughout the stages of LV remodeling. In a rat model of MI, the multifunctional support effectively increased M2-like MF at the infarct site and downregulated genes associated with IFN-y production and TGF- $\beta$  signaling [321]. While MF modulation remains a mainstay of cardiac immunotherapies, other studies have leveraged an understanding of T cell response. Across CVD and other diseases, the recruitment and differentiation of Tregs is a valuable immunomodulatory mechanism [354, 355]. Ramjee et al. demonstrated that Treg recruitment post-MI is dependent on Hippo signaling, and inflammatory cardiomyopathy and death are therefore exaggerated by disruption of core pathway effectors (YAP/TAZ). Delivery of IFN-y by formation of a hydrogel patch photopolymerized directly onto the epicardial surface rescued Treg infiltration and reversed deleterious inflammation [166]. This approach represents a divergence from conventional approaches. While many methods focus on suppression of the innate immune response including downregulation of IFN-y production, these inflammatory mediators are likewise a valuable tool for regulating the downstream adaptive immune compartment (B and T cells). These studies demonstrate the multitude of methods, spanning from electrical signaling to emerging therapeutic targets, that can be used to modulate post-MI inflammation.

#### **Injectable materials**

Another common form of cardiac therapies is the use of injectable hydrogels, which are an alternative means of mechanical support that enable local and minimally invasive delivery to the heart [334, 356]. Alginate has been widely investigated in pre-clinical large animal models, where introduction can be performed by intracoronary or intramyocardial injection to attenuate LV remodeling [357–360]. This line of work has progressed to clinical trials (AUGMENT-HF; NCT01311791, NCT03082508), where Algisyl-LVR improved clinical outcomes in patients with advanced HF [318]. Additional investigations through the use of synthetically tunable materials have continued to demonstrate through a combination of experimental and computational approaches that supraphysiological material stiffness, hydrogel injection volume, and even injection location can further improve these outcomes [361–366].

As for myocardial wraps and patches, mechanical restraint is a primary mechanism of therapeutic action; however, bioactive roles are also critical. Again, decellularized ECM has become often employed due to its bioactive role [367]. The Christman lab has pioneered this work by developing techniques to process decellularized porcine myocardial ECM into an injectable form, amenable to catheter-based injection [313, 368]. While these and related studies indicate only a moderate effect on MF infiltration and polarization state, an overall effect on inflammatory pathway activation has been noted [314]. A clinical trial arising from this work (Ventrix, Inc.; NCT02305602) indicated moderate increases in clinical outcomes, and no incidence of adverse events was definitively linked to VentriGel injection [314]. Other source tissues have also been explored. Inspired by its immunological function, Liu et al. used hydrogels derived from spleen ECM [315]. At physiological temperatures, the materials self-assembled into a hydrogel that drove an anti-inflammatory MF phenotype that was recapitulated in vivo. Improvements in both lymphangiogenesis and heart function were noted following MI treatment.

Self-assembling hydrogels are also useful as vehicles for therapeutic delivery. Rodell et al. have developed injectable hydrogels based on guest-host associations that can be pre-formed, injected via shear-thinning processes, and rapidly reassembled in the tissue [316]. Secondary covalent crosslinking interactions were used to achieve supraphysiological moduli for mechanical restraint [363, 369]. Alternatively, the shear-thinning materials alone have been used for the delivery of bone marrow cell chemotaxis enhancers and endothelial progenitor cells (EPCs) [145, 146]. Even though these hydrogels can be delivered alone, they are also convenient for cytokine and chemokine delivery. A follow-up study subsequently delivered anti-inflammatory cytokine IL-10 from a supramolecular HA hydrogel/microgel composite in a rat MI model. Delivery of exogenous IL-10 significantly decreased MF infiltration at 1-week post-treatment and improved vascularization and heart function at study endpoint (Fig. 6b) [167]. Alginate hydrogels were similarly able to differentiate blood Mo into M2-like MF by co-delivery of colony-stimulating factor (CSF-1) and IL-4 [168]. The delivery system moderately increased the presence of M2-like (CD68+CD206+) MF near the infarct site and improved cardiac function at day 15. Projahn et al. used thiol-functionalized star-shaped poly(ethylene oxide-stat-propylene oxide) (sP(EOstat-PO)) and linear poly(glycidol) (PG) degradable hydrogels to temporally control the release of two chemokines: one that reduces neutrophil infiltration (Met-CCL5) within the first few

hours and one that stimulates stem cell recruitment (CXCL12 (S4V)) over the course of several weeks post-MI. The delivery of both fast releasing CXCL12 (S4V) and slow releasing Met-CCL5 hydrogels were able to prevent neutrophil migration into the infarcted myocardium, reduce cardiomyocyte apoptosis, and promote vascularization, all of which improved cardiac function after MI [169, 370].

In addition, hydrogels can be an efficacious vehicle for local cell therapy. Typically, the delivery of cell suspensions alone results in poor cell survival, retention, and engraftment (often < 1%) that motivates the need for a biomaterial carrier [371]. Hydrogels can improve cell viability during injection, enhance cell engraftment, and are permeable to allow for oxygen and nutrient diffusion necessary to support continued viability of encapsulated cells in vivo [147, 372]. Liu et al. co-transplanted bone marrow-derived MSCs (BMSCs) with a chitosan hydrogel to increase stem cell retention and modulate the MI immune environment in mice [150]. Application post-MI alleviated the inflammatory response, as reflected by a reduction in TNF- $\alpha$ , IL-6, IL-1 $\beta$ , caspase-11, and caspase-1. It also protected vascular endothelial cells from pyroptosis and attenuated ventricular remodeling. Likewise, Shin et al. encapsulated MSCs in alginate and confined the cells to rat myocardial walls with a PEG hydrogel following MI. The conversion of pro-inflammatory AMP to anti-inflammatory adenosine through MSCs via CD73 reduced initial neutrophil and MF infiltration, prevented ROS formation, and accelerated cardiac repair [148]. It is clear that vehicle-assisted delivery of MSCs holds potential to aid in restoration of cardiac function after MI; though, effects may differ greatly from originally intended direct cardioregeneration.

Incorporating EVs into hydrogels has also become a promising approach, as they can communicate intracellularly and avoid issues often associated with stem cell sourcing and rejection [373]. Lv et al. delivered BMSC-derived EVs from alginate hydrogels [153], finding that the EVs were retained at the infarct site and decreased cardiac fibrosis and cell death, while also promoting M2-like polarization. Similarly, EPC-derived EVs were delivered to the myocardium via shear-thinning guest-host hydrogels, intended to mimic EPC function. Local delivery allowed for sustained release of EPC-derived EVs over 21 days, promoted angiogenesis, and improved ventricular hemodynamic function [154]. In addition to EVs, Zhang et al. revealed the role of dendritic cell-derived exosomes (DEXs) through their delivery in alginate hydrogels [155]. Notably, infiltration of Tregs and M2-like MF was enhanced in the infarct border zone in mice, and DEXs alone were able to induce both Treg and M2-like polarization in vitro. While effects of DEXs alone were short lived, hydrogel-based delivery prolonged the effects and healing capability to create a long-lasting reparatory environment.

Injectable hydrogels are likewise useful as a vehicle for the local delivery of naturally derived or fully synthetic nanoparticles. A recent study used functionalized mesoporous silica nanoparticles (MSNs) instead of EVs to deliver miRNAs, specifically miR-21-5p, thus creating an MSN/miR-21-5p complex [157]. To deliver the MSNs, a pH-responsive injectable hydrogel delivery system was used, which incorporated MSN/ miR-21-5p encapsulation into a hydrogel matrix (Gel@MSN/ miR-21-5p) that allowed for delivery upon acidic stimulation. The MSNs were able to improve angiogenesis following MI, as well as downregulate TLR2 and subsequently NF-kB signaling, thus decreasing TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 proinflammatory cytokine expression. Yi and colleagues have also reported an injectable filamentous hydrogel for low-dose, sustained delivery of anti-inflammatory nanocarriers. The researchers loaded a bioactive form of vitamin D, which inhibits proinflammatory NF-kB, in PEG-b-PPS filomicelles that transition from a cylindrical to spherical morphology to gradually release drug-loaded micelles. After a single subcutaneous treatment in  $ApoE^{-/-}$  mice, high levels of regulatory T cells were observed both in atherosclerotic lesions and distant organs for several weeks [173].

In an exciting approach to create cardiac patches in a minimally invasive manner, methods of intrapericardial hydrogel injection have been recently demonstrated for immunotherapeutic delivery. Studies by Ke Cheng and colleagues have encapsulated MSC-derived exosomes or iPSC-derived cardiac progenitor cells in methacrylated HA hydrogels with subsequent injection into the pericardial space, either by direct injection in mice and rats or by a minimally invasive thoracoscope-guided approach in pigs (Fig. 6c) [151, 156]. In mice, hydrogel injection resulted in retention and prolonged release of the exosomes. At endpoint in rats, metrics of LV dilation, fibrosis, and myocyte survival were improved by EV delivery relative to saline and HA controls, while pig studies established safety of the interventional approach.

Throughout these many forms of bulk materials and devices, multiple opportunities arise. It is apparent that these systems confer a unique opportunity as multifunctional interventions, as they can simultaneously target multiple mechanisms of LV remodeling. This includes provision of mechanical restraint, mitigating infarct thinning and expansion to directly address tissuelevel remodeling processes. As seen for the CorCap device, mechanical restraint in itself may influence the immune environment post-MI. Direct studies to investigate these interaction affects in detail, however, are largely lacking from the literature. When combined with appropriate materials selection or therapeutics, these local delivery strategies can also directly modulate the hyperinflammatory post-MI milieu that drives continued tissue injury. Importantly, therapeutic delivery is concentrated at the site of action. This is a particularly important consideration in immune modulation, where the systemic administration of immunosuppressive drugs exacerbates the risk of infection or septic shock, which has hindered clinical approval [257, 374]. Conversely, the systemic delivery of immunostimulatory drugs produces a widespread interferon response that likewise mitigates their use. In the case of both immunosuppressive and immunostimulatory strategies, local administration may therefore improve outcomes by targeting action towards the site of injury, prolonging the therapeutic window via controlled release, and by overcoming systemic side effects.

# Conclusion

Immunotherapies are redefining the medical management of disease, enabling the most significant improvements in patient outcomes seen in decades for fields such as cancer treatment and regulation of autoimmune disease. These approaches are now finding their way to regenerative medicine, where cardioimmunology is an emerging frontier open to newfound discoveries in the fundamental pathophysiology of disease development and progression [51, 375]. Emerging evidence reveals that LV remodeling and IHF development, in particular, are driven by both initial and persistent inflammation that continue to damage to the heart. This understanding gives rise to distinct modes of treatment, including cardioprotective and cardioregenerative strategies. Cardioprotective strategies developed to date are largely based on immunosuppression, and are therefore a promising prophylactic approach to abating IHF. On the other hand, leaders in the field have only recently recognized that stimulation of the pro-regenerative immune response is a prime target, and a unique opportunity to reverse deleterious remodeling to restore heart function [376]. Toward each of these general approaches, this review has highlighted interesting counterexamples, wherein pro-inflammatory stimuli (e.g., TLR agonists [11, 212, 273], pro-inflammatory cytokines [166]) are effective in preserving heart function. Overall, these studies are in alignment with the general hypothesis that initial inflammation is essential for subsequent healing, such as by provoking angiogenic response [377, 378]. Though, such functions may be tissue and context-specific [379]. For both immunosuppressive and immunostimulatory therapies in the context of LV remodeling, there remain fundamental gaps in knowledge. As discussed, a more thorough understanding of the therapeutic window is needed, as seen for immunosuppressive treatments through meta-analysis [108]. Moreover, it remains to be seen that cardioregenerative therapies can reverse late-stage remodeling. Related to these considerations, there is a dire need to better characterize the link between the innate and adaptive immune response post-MI. While many studies discussed here have focused on mitigating early inflammation, they fail to characterize downstream effectors, such as B and T cells. This is a crucial consideration, given the demonstrated role of the adaptive immunity in IHF development [33, 38] and the divergent role of regulatory, neonatal, and adult T cells in cardiac regeneration [380, 381].

The continued progress towards immunotherapeutic strategies for IHF is likely to have broader implications for the field. For example, the preservation of donor organs for transplant is essential [382-384]. This is particularly evident in heart transplant, where the acute inflammatory response has been implicated as a driver of tissue injury and waning organ function that ultimately render the organ unusable [385]. Application of appropriate immunosuppressive therapeutics in this context is an interesting strategy to preserve function, whereas regenerative approaches could provide an opportunity to rescue heart function post-transplant. For such therapeutic strategies to become accepted, however, they must first be shown to be safe and effective in clinical use before broader applications may be investigated. Biomaterial-based approaches will continue to aid in this endeavor as they are a platform to provide much needed cell or tissue-targeted delivery that can be multiplexed with the biological function of the material itself, including specific cellmaterial interactions and the provision of mechanical restraint to attenuate concurrent tissue remodeling.

Acknowledgements Figure schematics and the graphical abstract were created with BioRender.com.

Author contribution SSS, AMD, and CBR conceived and wrote the manuscript.

Funding This publication was supported by NIH funds (1R35GM147184, CBR).

Availability of data and materials Not applicable.

#### **Declarations**

Ethics approval and consent to participate Not applicable.

Consent for publication The authors give consent for publication.

Conflict of interest The authors declare no competing interests.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

# References

- Roth GA, Mensah GA, Johnson CO, Addolorato G, Ammirati E, Baddour LM, et al. Global burden of cardiovascular diseases and risk factors, 1990–2019: update from the GBD 2019 study. J Am Coll Cardiol. 2020;76(25):2982–3021.
- Virani SS, Alonso A, Aparicio HJ, Benjamin EJ, Bittencourt MS, Callaway CW, et al. Heart disease and stroke statistics—2021 update. Circulation. 2021;143(8).

- Forouzanfar MH, Afshin A, Alexander LT, Anderson HR, Bhutta ZA, Biryukov S, et al. Global, regional, and national comparative risk assessment of 79 behavioural, environmental and occupational, and metabolic risks or clusters of risks, 1990–2015: a systematic analysis for the Global Burden of Disease Study 2015. The lancet. 2016;388(10053):1659–724.
- GóMez AM, Guatimosim S, Dilly KW, Vassort G, Lederer WJ. Heart failure after myocardial infarction. Circulation. 2001;104(6):688–93.
- 5. Jessup M, Brozena S. Heart failure. N Engl J Med. 2003;348(20):2007–18.
- Libby P. The changing landscape of atherosclerosis. Nature. 2021;592(7855):524–33.
- Anzai T. Post-infarction inflammation and left ventricular remodeling. 2013;77(3):580–7.
- Kologrivova I, Shtatolkina M, Suslova T, Ryabov V. Cells of the immune system in cardiac remodeling: main players in resolution of inflammation and repair after myocardial infarction. Front Immunol. 2021;12: 664457.
- Cavalera M, G Frangogiannis N. Targeting the chemokines in cardiac repair. Current pharmaceutical design. 2014;20(12):1971–9.
- Silvis MJ, Kaffka genaamd Dengler SE, Odille CA, Mishra M, Van der Kaaij NP, Doevendans PA, et al. Damage-associated molecular patterns in myocardial infarction and heart transplantation: the road to translational success. Front Immunol. 2020;11:599511.
- Foglio E, Pellegrini L, Russo MA, Limana F. HMGB1-mediated activation of the inflammatory-reparative response following myocardial infarction. Cells. 2022;11(2):216.
- Ma Y, Yabluchanskiy A, Lindsey ML. Neutrophil roles in left ventricular remodeling following myocardial infarction. Fibrogenesis Tissue Repair. 2013;6(1):1–10.
- Newton K, Dixit VM. Signaling in innate immunity and inflammation. Cold Spring Harb Perspect Biol. 2012;4(3):a006049-a.
- Nahrendorf M, Swirski FK, Aikawa E, Stangenberg L, Wurdinger T, Figueiredo JL, et al. The healing myocardium sequentially mobilizes two monocyte subsets with divergent and complementary functions. J Exp Med. 2007;204(12):3037–47.
- Leuschner F, Rauch PJ, Ueno T, Gorbatov R, Marinelli B, Lee WW, et al. Rapid monocyte kinetics in acute myocardial infarction are sustained by extramedullary monocytopoiesis. J Exp Med. 2012;209(1):123–37.
- Dewald O, Ren G, Duerr GD, Zoerlein M, Klemm C, Gersch C, et al. Of mice and dogs: species-specific differences in the inflammatory response following myocardial infarction. Am J Pathol. 2004;164(2):665–77.
- 17. Sica A, Mantovani A. Macrophage plasticity and polarization: in vivo veritas. J Clin Investig. 2012;122(3):787–95.
- Heidt T, Courties G, Dutta P, Sager HB, Sebas M, Iwamoto Y, et al. Differential contribution of monocytes to heart macrophages in steadystate and after myocardial infarction. Circ Res. 2014;115(2):284–95.
- Bozkurt B, Mann DL, Deswal A. Biomarkers of inflammation in heart failure. Heart Fail Rev. 2010;15(4):331–41.
- Frangogiannis NG. The inflammatory response in myocardial injury, repair, and remodelling. Nat Rev Cardiol. 2014;11(5):255–65.
- De Couto G. Macrophages in cardiac repair: environmental cues and therapeutic strategies. Exp Mol Med. 2019;51(12):1–10.
- Italiani P, Boraschi D. From monocytes to M1/M2 macrophages: phenotypical vs. functional differentiation. Front Immunol. 2014;5.
- van Amerongen MJ, Harmsen MC, van Rooijen N, Petersen AH, van Luyn MJ. Macrophage depletion impairs wound healing and increases left ventricular remodeling after myocardial injury in mice. Am J Pathol. 2007;170(3):818–29.
- Honold L, Nahrendorf M. Resident and monocyte-derived macrophages in cardiovascular disease. Circ Res. 2018;122(1):113–27.
- 25. Peet C, Ivetic A, Bromage DI, Shah AM. Cardiac monocytes and macrophages after myocardial infarction. Cardiovasc Res. 2020;116(6):1101–12.

- Medzhitov R. Origin and physiological roles of inflammation. Nature. 2008;454(7203):428–35.
- Bansal SS, Ismahil MA, Goel M, Patel B, Hamid T, Rokosh G, et al. Activated T lymphocytes are essential drivers of pathological remodeling in ischemic heart failure. Circ Heart Fail. 2017;10(3):e003688.
- EpeIman S, Liu PP, Mann DL. Role of innate and adaptive immune mechanisms in cardiac injury and repair. Nat Rev Immunol. 2015;15(2):117–29.
- Adamo L, Rocha-Resende C, Prabhu SD, Mann DL. Reappraising the role of inflammation in heart failure. Nat Rev Cardiol. 2020:1–17.
- Jakubzick C, Gautier EL, Gibbings SL, Sojka DK, Schlitzer A, Johnson TE, et al. Minimal differentiation of classical monocytes as they survey steady-state tissues and transport antigen to lymph nodes. Immunity. 2013;39(3):599–610.
- Mysore V, Cullere X, Mears J, Rosetti F, Okubo K, Liew PX, et al. FcγR engagement reprograms neutrophils into antigen cross-presenting cells that elicit acquired anti-tumor immunity. Nat Commun. 2021;12(1):1–23.
- Zhao T, Jiang Q, Li W, Wang Y, Zou Y, Chai X, et al. Antigenpresenting cell-like neutrophils foster T cell response in hyperlipidemic patients and atherosclerotic mice. Front Immunol. 2022;13.
- 33. Keppner L, Heinrichs M, Rieckmann M, Demengeot J, Frantz S, Hofmann U, et al. Antibodies aggravate the development of ischemic heart failure. Am J Physiol Heart Circ Physiol. 2018;315(5):H1358–67.
- Kumar BV, Connors TJ, Farber DL. Human T cell development, localization, and function throughout life. Immunity. 2018;48(2):202–13.
- Hosokawa H, Rothenberg EV. How transcription factors drive choice of the T cell fate. Nat Rev Immunol. 2021;21(3):162–76.
- Simons KH, de Jong A, Jukema JW, de Vries MR, Arens R, Quax PH. T cell co-stimulation and co-inhibition in cardiovascular disease: a double-edged sword. Nat Rev Cardiol. 2019;16(6):325–43.
- Latet SC, Hoymans VY, Van Herck PL, Vrints CJ. The cellular immune system in the post-myocardial infarction repair process. Int J Cardiol. 2015;179:240–7.
- Bansal SS, Ismahil MA, Goel M, Patel B, Hamid T, Rokosh G, et al. Activated T lymphocytes are essential drivers of pathological remodeling in ischemic heart failure. Circ Heart Fail. 2017;10(3): e003688.
- Santos-Zas I, Lemarié J, Tedgui A, Ait-Oufella H. Adaptive immune responses contribute to post-ischemic cardiac remodeling. Front Cardiovasc Med. 2019;5:198.
- Mallat Z, Binder CJ. The why and how of adaptive immune responses in ischemic cardiovascular disease. Nature Cardiovascular Research. 2022;1(5):431–44.
- Pluijmert NJ, Atsma DE, Quax PHA. Post-ischemic myocardial inflammatory response: a complex and dynamic process susceptible to immunomodulatory therapies. Front Cardiovasc Med. 2021;8: 647785.
- 42. Liu J, Wang H, Li J. Inflammation and inflammatory cells in myocardial infarction and reperfusion injury: a double-edged sword. Clin Med Insights: Cardiol. 2016;10:CMC.S33164.
- Prabhu SD, Frangogiannis NG. The biological basis for cardiac repair after myocardial infarction. Circ Res. 2016;119(1):91–112.
- Anzai T. Post-infarction inflammation and left ventricular remodeling: a double-edged sword. Circ J. 2013;77(3):580–7.
- Sutton MGSJ, Sharpe N. Left ventricular remodeling after myocardial infarction. Circulation. 2000;101(25):2981–8.
- Bar A, Cohen S. Inducing endogenous cardiac regeneration: can biomaterials connect the dots? Front Bioeng Biotechnol. 2020;8:126.
- Moore JB, Wysoczynski M. Immunomodulatory effects of cell therapy after myocardial infarction. J Clin Cell Immunol. 2021;3(2):85–90.
- Wagner MJ, Khan M, Mohsin S. Healing the broken heart; the immunomodulatory effects of stem cell therapy. Front Immunol. 2020;11:639.
- Vasu S, Zhou J, Chen J, Johnston PV, Kim D-H. Biomaterials-based approaches for cardiac regeneration. Korean Circ J. 2021;51(12):943.

- Daseke MJ, Valerio FM, Kalusche WJ, Ma Y, DeLeon-Pennell KY, Lindsey ML. Neutrophil proteome shifts over the myocardial infarction time continuum. Basic Res Cardiol. 2019;114(5):37.
- Swirski FK, Nahrendorf M. Cardioimmunology: the immune system in cardiac homeostasis and disease. Nat Rev Immunol. 2018;18(12):733–44.
- Zhang N, Aiyasiding X, Li W-J, Liao H-H, Tang Q-Z. Neutrophil degranulation and myocardial infarction. Cell Comm Signal. 2022;20(1).
- Yap J, Cabrera-Fuentes HA, Irei J, Hausenloy DJ, Boisvert WA. Role of macrophages in cardioprotection. Int J Mol Sci. 2019; 20(10):2474.
- Shapouri-Moghaddam A, Mohammadian S, Vazini H, Taghadosi M, Esmaeili SA, Mardani F, et al. Macrophage plasticity, polarization, and function in health and disease. J Cell Physiol. 2018;233(9):6425–40.
- Poller WC, Nahrendorf M, Swirski FK. Hematopoiesis and cardiovascular disease. Circ Res. 2020;126(8):1061–85.
- Santos-Zas I, Lemarié J, Tedgui A, Ait-Oufella H. Adaptive immune responses contribute to post-ischemic cardiac remodeling. Front Cardiovasc Med. 2018;5:198.
- Hofmann U, Frantz S. Role of lymphocytes in myocardial injury, healing, and remodeling after myocardial infarction. Circ Res. 2015;116(2):354–67.
- Nian M, Lee P, Khaper N, Liu P. Inflammatory Cytokines and postmyocardial infarction remodeling. Circ Res. 2004;94(12):1543–53.
- Hanna A, Frangogiannis NG. Inflammatory cytokines and chemokines as therapeutic targets in heart failure. Cardiovasc Drugs Ther. 2020;34(6):849–63.
- Viola A, Munari F, Sánchez-Rodríguez R, Scolaro T, Castegna A. The metabolic signature of macrophage responses. Front Immunol. 2019;10:1462.
- Soehnlein O, Drechsler M, Döring Y, Lievens D, Hartwig H, Kemmerich K, et al. Distinct functions of chemokine receptor axes in the atherogenic mobilization and recruitment of classical monocytes. EMBO Mol Med. 2013;5(3):471–81.
- Hammel JH, Zatorski JM, Cook SR, Pompano RR, Munson JM. Engineering in vitro immune-competent tissue models for testing and evaluation of therapeutics. Adv Drug Deliv Rev. 2022;182: 114111.
- Ogle BM, Bursac N, Domian I, Huang NF, Menasché P, Murry CE, et al. Distilling complexity to advance cardiac tissue engineering. Sci Transl Med. 2016;8(342):342ps13.
- Qasim M, Haq F, Kang MH, Kim JH. 3D printing approaches for cardiac tissue engineering and role of immune modulation in tissue regeneration</p&gt. Int J Nanomedicine. 2019;14:1311–33.</li>
- Boudou T, Legant WR, Mu A, Borochin MA, Thavandiran N, Radisic M, et al. A microfabricated platform to measure and manipulate the mechanics of engineered cardiac microtissues. Tissue Eng Part A. 2012;18(9–10):910–9.
- Jackman CP, Carlson AL, Bursac N. Dynamic culture yields engineered myocardium with near-adult functional output. Biomaterials. 2016;111:66–79.
- Thomas D, Choi S, Alamana C, Parker KK, Wu JC. Cellular and engineered organoids for cardiovascular models. Circ Res. 2022;130(12):1780–802.
- Mohr E, Thum T, Bär C. Accelerating cardiovascular research: recent advances in translational 2D and 3D heart models. Eur J Heart Fail. 2022.
- Cho S, Discher DE, Leong KW, Vunjak-Novakovic G, Wu JC. Challenges and opportunities for the next generation of cardiovascular tissue engineering. Nat Methods. 2022;19(9):1064–71.
- Suku M, Forrester L, Biggs M, Monaghan MG. Resident macrophages and their potential in cardiac tissue engineering. Tissue Eng Part B Rev. 2022;28(3):579–91.
- Richards DJ, Li Y, Kerr CM, Yao J, Beeson GC, Coyle RC, et al. Human cardiac organoids for the modelling of myocardial infarction and drug cardiotoxicity. Nat Biomed Eng. 2020;4(4):446–62.

Kumar M, Kasala ER, Bodduluru LN, Dahiya V, Sharma D, Kumar V, et al. Animal models of myocardial infarction: Mainstay in clinical translation. Regul Toxicol Pharmacol. 2016;76:221–30.

 Martin TP, MacDonald EA, Elbassioni AAM, O'Toole D, Zaeri AAI, Nicklin SA, et al. Preclinical models of myocardial infarction: from mechanism to translation. Br J Pharmacol. 2022;179(5):770–91.

- Lindsey ML, Bolli R, Canty JM Jr, Du XJ, Frangogiannis NG, Frantz S, et al. Guidelines for experimental models of myocardial ischemia and infarction. Am J Physiol Heart Circ Physiol. 2018;314(4):H812–38.
- Recchia FA, Lionetti V. Animal models of dilated cardiomyopathy for translational research. Vet Res Commun. 2007;31(Suppl 1):35–41.
- Ciszek B, Skubiszewska D, Ratajska A. The anatomy of the cardiac veins in mice. J Anat. 2007;211(1):53–63.
- Pfeffer MA, Pfeffer JM, Fishbein MC, Fletcher PJ, Spadaro J, Kloner RA, et al. Myocardial infarct size and ventricular function in rats. Circ Res. 1979;44(4):503–12.
- Wang J, Bo H, Meng X, Wu Y, Bao Y, Li Y. A simple and fast experimental model of myocardial infarction in the mouse. Tex Heart Inst J. 2006;33(3):290–3.
- Chen T, Vunjak-Novakovic G. In vitro models of ischemia-reperfusion injury. Regen Eng Transl Med. 2018;4(3):142–53.
- Lindsey ML, de Castro Brás LE, DeLeon-Pennell KY, Frangogiannis NG, Halade GV, O'Meara CC, et al. Reperfused vs. nonreperfused myocardial infarction: when to use which model. Am J Physiol Heart Circ Physiol. 2021;321(1):H208-h13.
- 81. Yutzey KE, Robbins J. Principles of genetic murine models for cardiac disease. Circulation. 2007;115(6):792–9.
- Bansal SS, Ismahil MA, Goel M, Zhou G, Rokosh G, Hamid T, et al. Dysfunctional and proinflammatory regulatory T-lymphocytes are essential for adverse cardiac remodeling in ischemic cardiomyopathy. Circulation. 2019;139(2):206–21.
- Chao T, Lu L, Zhang L, Huang R, Liu Z, Zhou B, et al. An inducible model for specific neutrophil depletion by diphtheria toxin in mice. Sci China Life Sci. 2021;64(8):1227–35.
- 84. Stoneman V, Braganza D, Figg N, Mercer J, Lang R, Goddard M, et al. Monocyte/macrophage suppression in CD11b diphtheria toxin receptor transgenic mice differentially affects atherogenesis and established plaques. Circ Res. 2007;100(6):884–93.
- Ruedl C, Jung S. DTR-mediated conditional cell ablation—progress and challenges. Eur J Immunol. 2018;48(7):1114–9.
- Bajpai G, Bredemeyer A, Li W, Zaitsev K, Koenig AL, Lokshina I, et al. Tissue resident CCR2– and CCR2+ cardiac macrophages differentially orchestrate monocyte recruitment and fate specification following myocardial injury. Circ Res. 2019;124(2):263–78.
- Dick SA, Macklin JA, Nejat S, Momen A, Clemente-Casares X, Althagafi MG, et al. Self-renewing resident cardiac macrophages limit adverse remodeling following myocardial infarction. Nat Immunol. 2019;20(1):29–39.
- Männ L, Kochupurakkal N, Martin C, Verjans E, Klingberg A, Sody S, et al. CD11c.DTR mice develop a fatal fulminant myocarditis after local or systemic treatment with diphtheria toxin. Eur J Immunol. 2016;46(8):2028–42.
- Zaft T, Sapoznikov A, Krauthgamer R, Littman DR, Jung S. CD11chigh dendritic cell ablation impairs lymphopenia-driven proliferation of naive and memory CD8+ T cells. J Immunol. 2005;175(10):6428–35.
- Ramachandran A, Jha S, Lefer D. pathophysiology of myocardial reperfusion injury: the role of genetically engineered mouse models. Vet Pathol. 2008;45(5):698–706.
- Lo Sasso G, Schlage WK, Boué S, Veljkovic E, Peitsch MC, Hoeng J. The Apoe-/- mouse model: a suitable model to study cardiovascular and respiratory diseases in the context of cigarette smoke exposure and harm reduction. J Trans Med. 2016;14(1).
- Qian J-Y, Harding P, Liu Y, Shesely E, Yang X-P, Lapointe MC. Reduced cardiac remodeling and function in cardiac-specific EP 4 receptor knockout mice with myocardial infarction. Hypertension. 2008;51(2):560–6.

- Harada K, Sugaya T, Murakami K, Yazaki Y, Komuro I. Angiotensin II Type 1A Receptor knockout mice display less left ventricular remodeling and improved survival after myocardial infarction. Circulation. 1999;100(20):2093–9.
- Shiraishi M, Shintani Y, Shintani Y, Ishida H, Saba R, Yamaguchi A, et al. Alternatively activated macrophages determine repair of the infarcted adult murine heart. J Clin Investig. 2016;126(6):2151–66.
- Lukács E, Magyari B, Tóth L, Petrási Z, Repa I, Koller A, et al. Overview of large animal myocardial infarction models (review). Acta Physiol Hung. 2012;99(4):365–81.
- Shin HS, Shin HH, Shudo Y. Current status and limitations of myocardial infarction large animal models in cardiovascular translational research. Front Bioeng Biotechnol. 2021;9: 673683.
- Freedman LP, Cockburn IM, Simcoe TS. The economics of reproducibility in preclinical research. PLoS Biol. 2015;13(6):e1002165.
- Camacho P, Fan H, Liu Z, He J-Q. Large mammalian animal models of heart disease. J Cardiovasc Dev Dis. 2016;3(4):30.
- 99. National Research Council Committee on Scientific, Humane Issues in the Use of Random Source Dogs and Cats in Research. The National Academies Collection: Reports funded by National Institutes of Health. Scientific and humane issues in the Use of random source dogs and cats in research. Washington (DC): National Academies Press (US). Copyright © 2009. National Academy of Sciences. 2009.
- Schook LB, Collares TV, Hu W, Liang Y, Rodrigues FM, Rund LA, et al. A genetic porcine model of cancer. PLoS ONE. 2015;10(7): e0128864.
- 101. De Nisco G, Chiastra C, Hartman EMJ, Hoogendoorn A, Daemen J, Calò K, et al. Comparison of swine and human computational hemodynamics models for the study of coronary atherosclerosis. Front Bioeng Biotechnol. 2021;9.
- 102. Maxwell MP, Hearse DJ, Yellon DM. Species variation in the coronary collateral circulation during regional myocardial ischaemia: a critical determinant of the rate of evolution and extent of myocardial infarction. Cardiovasc Res. 1987;21(10):737–46.
- 103. Milani-Nejad N, Janssen PM. Small and large animal models in cardiac contraction research: advantages and disadvantages. Pharmacol Ther. 2014;141(3):235–49.
- Lelovas PP, Kostomitsopoulos NG, Xanthos TT. A comparative anatomic and physiologic overview of the porcine heart. J Am Assoc Lab Anim Sci. 2014;53(5):432–8.
- 105. Contamin H, Rioufol G, Bettinger T, Helbert A, Portier KG, Lepage OM, et al. A minimally-invasive closed chest myocardial occlusionreperfusion model in rhesus monkeys (Macaca mulatta): monitoring by contrast-enhanced ultrasound imaging. Int J Cardiovasc Imaging. 2012;28(3):531–42.
- Shen YT. Primate models for cardiovascular drug research and development. Curr Opin Investig Drugs. 2010;11(9):1025–9.
- 107. Ribitsch I, Baptista PM, Lange-Consiglio A, Melotti L, Patruno M, Jenner F, et al. Large animal models in regenerative medicine and tissue engineering: to do or not to do. Front Bioeng Biotechnol. 2020;8:972.
- 108. van Hout GPJ, Jansen of Lorkeers SJ, Wever KE, Sena ES, Kouwenberg LHJA, van Solinge WW, et al. Translational failure of anti-inflammatory compounds for myocardial infarction: a metaanalysis of large animal models. Cardio Res. 2015;109(2):240–8.
- Arnegard ME, Whitten LA, Hunter C, Clayton JA. Sex as a biological variable: a 5-year progress report and call to action. J Womens Health. 2020;29(6):858–64.
- Lindsey ML, Carter JR, Ripplinger CM, Kassiri Z, Hansell Keehan K, Brunt KR, et al. Sex still matters in cardiovascular research. MD: American Physiological Society Rockville; 2022.
- Moore E, Allen JB, Mulligan CJ, Wayne EC. Ancestry of cells must be considered in bioengineering. Nat Rev Mater. 2021;1–3.
- 112. De Kort B, Koch S, Wissing T, Krebber M, Bouten C, Smits A. Immuno-regenerative biomaterials for in situ cardiovascular tissue engineering–do patient characteristics warrant precision engineering? Adv Drug Deliv Rev. 2021;178: 113960.

- 114. Wei C, Wang Y, Li P, Fu Q. Molecular nanoprobe for diagnosis of cardiovascular diseases. Adv Trans Med. 2022;61–76.
- 115. Sosnovik DE, Scherrer-Crosbie M. Biomedical imaging in experimental models of cardiovascular disease. Circ Res. 2022;130(12):1851–68.
- 116. Pilla G, Levack M, Mcgarvey J, Hwuang E, Zsido G, Gorman J, et al. Alterations in intracardiac flow patterns affect mitral leaflets dynamics in a model of ischemic mitral regurgitation. Cardiovasc Eng Technol. 2021;12(6):640–50.
- Sosnovik DE, Wang R, Dai G, Reese TG, Wedeen VJ. Diffusion MR tractography of the heart. J Cardiovasc Magn Reson. 2009;11(1):1–15.
- Pumphrey A, Yang Z, Ye S, Powell DK, Thalman S, Watt DS, et al. Advanced cardiac chemical exchange saturation transfer (cardioCEST) MRI for in vivo cell tracking and metabolic imaging. NMR Biomed. 2016;29(1):74–83.
- 119. Su H, Spinale FG, Dobrucki LW, Song J, Hua J, Sweterlitsch S, et al. Noninvasive targeted imaging of matrix metalloproteinase activation in a murine model of postinfarction remodeling. Circulation. 2005;112(20):3157–67.
- Curley D, Lavin Plaza B, Shah AM, Botnar RM. Molecular imaging of cardiac remodelling after myocardial infarction. Basic Res Cardiol. 2018;113(2):1–18.
- Pittet MJ, Garris CS, Arlauckas SP, Weissleder R. Recording the wild lives of immune cells. Science Immunol. 2018;3(27):eaaq0491.
- 122. Rodell CB, Koch PD, Weissleder R. Screening for new macrophage therapeutics Theranostics. 2019;9(25):7714.
- Hulsmans M, Clauss S, Xiao L, Aguirre AD, King KR, Hanley A, et al. Macrophages facilitate electrical conduction in the heart. Cell. 2017;169(3):510-22.e20.
- 124. Saxena A, Dobaczewski M, Rai V, Haque Z, Chen W, Li N, et al. Regulatory T cells are recruited in the infarcted mouse myocardium and may modulate fibroblast phenotype and function. Am J Physiol Heart Circ Physiol. 2014;307(8):H1233–42.
- 125. Vinegoni C, Aguirre AD, Lee S, Weissleder R. Imaging the beating heart in the mouse using intravital microscopy techniques. Nat Protoc. 2015;10(11):1802–19.
- Tang X. Tumor-associated macrophages as potential diagnostic and prognostic biomarkers in breast cancer. Cancer Lett. 2013;332(1):3–10.
- 127. DeBerge M, Shah SJ, Wilsbacher L, Thorp EB. Macrophages in heart failure with reduced versus preserved ejection fraction. Trends Mol Med. 2019;25(4):328–40.
- Liu W, Frank JA. Detection and quantification of magnetically labeled cells by cellular MRI. Eur J Radiol. 2009;70(2):258–64.
- 129. Morishige K, Kacher DF, Libby P, Josephson L, Ganz P, Weissleder R, et al. High-resolution magnetic resonance imaging enhanced with superparamagnetic nanoparticles measures macrophage burden in atherosclerosis. Circulation. 2010;122(17):1707–15.
- 130. Yilmaz A, Dengler MA, van der Kuip H, Yildiz H, Rösch S, Klumpp S, et al. Imaging of myocardial infarction using ultrasmall superparamagnetic iron oxide nanoparticles: a human study using a multi-parametric cardiovascular magnetic resonance imaging approach. Eur Heart J. 2013;34(6):462–75.
- 131. Hyafil F, Cornily J-C, Feig JE, Gordon R, Vucic E, Amirbekian V, et al. Noninvasive detection of macrophages using a nanoparticulate contrast agent for computed tomography. Nat Med. 2007;13(5):636–41.
- Cormode DP, Roessl E, Thran A, Skajaa T, Gordon RE, Schlomka JP, et al. Atherosclerotic plaque composition: analysis with multicolor CT and targeted gold nanoparticles. Radiology. 2010;256(3):774–82.
- Borchert T, Beitar L, Langer LBN, Polyak A, Wester HJ, Ross TL, et al. Dissecting the target leukocyte subpopulations of clinically relevant inflammation radiopharmaceuticals. J Nucl Cardiol. 2021;28(4):1636–45.

- 134. Satomi T, Ogawa M, Mori I, Ishino S, Kubo K, Magata Y, et al. Comparison of contrast agents for atherosclerosis imaging using cultured macrophages: FDG versus ultrasmall superparamagnetic iron oxide. J Nucl Med. 2013;54(6):999–1004.
- Wilk B, Wisenberg G, Dharmakumar R, Thiessen JD, Goldhawk DE, Prato FS. Hybrid PET/MR imaging in myocardial inflammation post-myocardial infarction. J Nucl Cardiol. 2020;27(6):2083–99.
- 136. Macaskill MG, Stadulyte A, Williams L, Morgan TEF, Sloan NL, Alcaide-Corral CJ, et al. Quantification of macrophage-driven inflammation during myocardial infarction with 18F-LW223, a novel TSPO radiotracer with binding independent of the rs6971 human polymorphism. J Nucl Med. 2021;62(4):536–44.
- 137. Fernández-Friera L, Fuster V, López-Melgar B, Oliva B, Sánchez-González J, Macías A, et al. Vascular inflammation in subclinical atherosclerosis detected by hybrid PET/MRI. J Am Coll Cardiol. 2019;73(12):1371–82.
- 138. Varasteh Z, Braeuer M, Mohanta S, Steinsiek A-L, Habenicht A, Omidvari N, et al. In vivo visualization of M2 macrophages in the myocardium after myocardial infarction (MI) using (68) Ga-NOTA-Anti-MMR Nb: targeting mannose receptor (MR, CD206) on M2 macrophages. Front Cardio Med. 2022;9:889963.
- 139. Nahrendorf M, Hoyer FF, Meerwaldt AE, Van Leent MMT, Senders ML, Calcagno C, et al. Imaging cardiovascular and lung macrophages with the positron emission tomography sensor 64Cu-Macrin in mice, rabbits, and pigs. Circ Cardio Imag. 2020;13(10).
- Kim H-Y, Li R, Ng TSC, Courties G, Rodell CB, Prytyskach M, et al. Quantitative imaging of tumor-associated macrophages and their response to therapy using 64Cu-labeled macrin. ACS Nano. 2018;12(12):12015–29.
- 141. Keliher EJ, Ye Y-X, Wojtkiewicz GR, Aguirre AD, Tricot B, Senders ML, et al. Polyglucose nanoparticles with renal elimination and macrophage avidity facilitate PET imaging in ischaemic heart disease. Nat Commun. 2017;8(1):14064.
- 142. Bejarano J, Navarro-Marquez M, Morales-Zavala F, Morales JO, Garcia-Carvajal I, Araya-Fuentes E, et al. Nanoparticles for diagnosis and therapy of atherosclerosis and myocardial infarction: evolution toward prospective theranostic approaches. Theranostics. 2018;8(17):4710.
- 143. Chen J, Yang J, Liu R, Qiao C, Lu Z, Shi Y, et al. Dual-targeting theranostic system with mimicking apoptosis to promote myocardial infarction repair via modulation of macrophages. Theranostics. 2017;7(17):4149–67.
- 144. Miller MA, Gadde S, Pfirschke C, Engblom C, Sprachman MM, Kohler RH, et al. Predicting therapeutic nanomedicine efficacy using a companion magnetic resonance imaging nanoparticle. Sci Trans Med. 2015;7(314):314ra183–314ra183.
- 145. Klyachkin YM, Idris A, Rodell CB, Tripathi H, Ye S, Nagareddy P, et al. Cathelicidin related antimicrobial peptide (CRAMP) enhances bone marrow cell retention and attenuates cardiac dysfunction in a mouse model of myocardial infarction. Stem cell reviews and reports. 2018;14(5):702–14.
- 146. Gaffey AC, Chen MH, Venkataraman CM, Trubelja A, Rodell CB, Dinh PV, et al. Injectable shear-thinning hydrogels used to deliver endothelial progenitor cells, enhance cell engraftment, and improve ischemic myocardium. J Thorac Cardiovasc Surg. 2015;150(5):1268–76.
- 147. Whyte W, Roche ET, Varela CE, Mendez K, Islam S, O'Neill H, et al. Sustained release of targeted cardiac therapy with a replenishable implanted epicardial reservoir. Nat Biomed Eng. 2018;2(6):416–28.
- 148. Shin EY, Wang L, Zemskova M, Deppen J, Xu K, Strobel F, et al. Adenosine production by biomaterial-supported mesenchymal stromal cells reduces the innate inflammatory response in myocardial ischemia/reperfusion injury. J Am Heart Assoc. 2018;7(2): e006949.

- Shah M, Kc P, Zhang G. In vivo assessment of decellularized porcine myocardial slice as an acellular cardiac patch. ACS Appl Mater Interfaces. 2019;11(27):23893–900.
- 150. Liu Y, Li P, Qiao C, Wu T, Sun X, Wen M, et al. Chitosan hydrogel enhances the therapeutic efficacy of bone marrow-derived mesenchymal stem cells for myocardial infarction by alleviating vascular endothelial cell pyroptosis. J Cardiovasc Pharmacol. 2020;75(1):75–83.
- 151. Zhu D, Li Z, Huang K, Caranasos TG, Rossi JS, Cheng K. Minimally invasive delivery of therapeutic agents by hydrogel injection into the pericardial cavity for cardiac repair. Nat Commun. 2021;12(1):1412.
- 152. Aghajanian H, Kimura T, Rurik JG, Hancock AS, Leibowitz MS, Li L, et al. Targeting cardiac fibrosis with engineered T cells. Nature. 2019;573(7774):430–3.
- Lv K, Li Q, Zhang L, Wang Y, Zhong Z, Zhao J, et al. Incorporation of small extracellular vesicles in sodium alginate hydrogel as a novel therapeutic strategy for myocardial infarction. Theranostics. 2019;9(24):7403–16.
- 154. Chen CW, Wang LL, Zaman S, Gordon J, Arisi MF, Venkataraman CM, et al. Sustained release of endothelial progenitor cell-derived extracellular vesicles from shear-thinning hydrogels improves angiogenesis and promotes function after myocardial infarction. Cardiovasc Res. 2018;114(7):1029–40.
- 155. Zhang Y, Cai Z, Shen Y, Lu Q, Gao W, Zhong X, et al. Hydrogelload exosomes derived from dendritic cells improve cardiac function via Treg cells and the polarization of macrophages following myocardial infarction. Journal of Nanobiotechnology. 2021;19(1):271.
- 156. Cheng G, Zhu D, Huang K, Caranasos TG. Minimally invasive delivery of a hydrogel-based exosome patch to prevent heart failure. J Mol Cell Cardiol. 2022;169:113–21.
- 157. Li Y, Chen X, Jin R, Chen L, Dang M, Cao H, et al. Injectable hydrogel with MSNs/microRNA-21–5p delivery enables both immunomodification and enhanced angiogenesis for myocardial infarction therapy in pigs. Sci Adv. 2021;7(9):eabd6740.
- Zhao J, Li X, Hu J, Chen F, Qiao S, Sun X, et al. Mesenchymal stromal cell-derived exosomes attenuate myocardial ischaemiareperfusion injury through miR-182-regulated macrophage polarization. Cardiovasc Res. 2019;115(7):1205–16.
- 159. Wang J, Lee CJ, Deci MB, Jasiewicz N, Verma A, Canty JM, et al. MiR-101a loaded extracellular nanovesicles as bioactive carriers for cardiac repair. Nanomedicine. 2020;27: 102201.
- 160. Dai Y, Wang S, Chang S, Ren D, Shali S, Li C, et al. M2 macrophage-derived exosomes carry microRNA-148a to alleviate myocardial ischemia/reperfusion injury via inhibiting TXNIP and the TLR4/NF-κB/NLRP3 inflammasome signaling pathway. J Mol Cell Cardiol. 2020;142:65–79.
- 161. Courties G, Heidt T, Sebas M, Iwamoto Y, Jeon D, Truelove J, et al. In vivo silencing of the transcription factor IRF5 reprograms the macrophage phenotype and improves infarct healing. J Am Coll Cardiol. 2014;63(15):1556–66.
- 162. Sager HB, Dutta P, Dahlman JE, Hulsmans M, Courties G, Sun Y, et al. RNAi targeting multiple cell adhesion molecules reduces immune cell recruitment and vascular inflammation after myo-cardial infarction. Sci Transl Med. 2016;8(342):342ra80.
- 163. Rurik JG, Tombácz I, Yadegari A, Méndez Fernández PO, Shewale SV, Li L, et al. CAR T cells produced in vivo to treat cardiac injury. Science. 2022;375(6576):91–6.
- Leuschner F, Dutta P, Gorbatov R, Novobrantseva TI, Donahoe JS, Courties G, et al. Therapeutic siRNA silencing in inflammatory monocytes in mice. Nat Biotechnol. 2011;29(11):1005–10.
- 165. Zhou L-S, Zhao G-L, Liu Q, Jiang S-C, Wang Y, Zhang D-M. Silencing collapsin response mediator protein-2 reprograms macrophage phenotype and improves infarct healing in experimental myocardial infarction model. J Inflamm. 2015;12(1):11.
- 🖄 Springer

- 166. Ramjee V, Li D, Manderfield LJ, Liu F, Engleka KA, Aghajanian H, et al. Epicardial YAP/TAZ orchestrate an immunosuppressive response following myocardial infarction. J Clin Investig. 2017;127(3):899–911.
- Chen MH, Chung JJ, Mealy JE, Zaman S, Li EC, Arisi MF, et al. Injectable supramolecular hydrogel/microgel composites for therapeutic delivery. Macromol Biosci. 2019;19(1):e1800248-n/a.
- Bloise N, Rountree I, Polucha C, Montagna G, Visai L, Coulombe KLK, et al. Engineering immunomodulatory biomaterials for regenerating the infarcted myocardium. Front Bioeng Biotechnol. 2020;8.
- 169. Projahn D, Simsekyilmaz S, Singh S, Kanzler I, Kramp BK, Langer M, et al. Controlled intramyocardial release of engineered chemokines by biodegradable hydrogels as a treatment approach of myocardial infarction. J Cell Mol Med. 2014;18(5):790–800.
- 170. Naveed M, Wenhua L, Gang W, Mohammad IS, Abbas M, Liao X, et al. A novel ventricular restraint device (ASD) repetitively deliver Salvia miltiorrhiza to epicardium have good curative effects in heart failure management. Biomed Pharmacother. 2017;95:701–10.
- 171. Nakano Y, Matoba T, Tokutome M, Funamoto D, Katsuki S, Ikeda G, et al. Nanoparticle-mediated delivery of irbesartan induces cardioprotection from myocardial ischemia-reperfusion injury by antagonizing monocyte-mediated inflammation. Sci Rep. 2016;6:29601.
- 172. Allen SD, Liu YG, Kim T, Bobbala S, Yi S, Zhang X, et al. Celastrolloaded PEG-b-PPS nanocarriers as an anti-inflammatory treatment for atherosclerosis. Biomater Sci. 2019;7(2):657–68.
- 173. Yi S, Karabin NB, Zhu J, Bobbala S, Lyu H, Li S, et al. An injectable hydrogel platform for sustained delivery of anti-inflammatory nanocarriers and induction of regulatory T cells in atherosclerosis. Front Bioeng Biotechnol. 2020;8:542.
- 174. Wang J, Seo MJ, Deci MB, Weil BR, Canty JM, Nguyen J. Effect of CCR2 inhibitor-loaded lipid micelles on inflammatory cell migration and cardiac function after myocardial infarction. Int J Nanomedicine. 2018;13:6441–51.
- 175. Mao Y, Koga JI, Tokutome M, Matoba T, Ikeda G, Nakano K, et al. Nanoparticle-mediated delivery of pitavastatin to monocytes/ macrophages inhibits left ventricular remodeling after acute myocardial infarction by inhibiting monocyte-mediated inflammation. Int Heart J. 2017;58(4):615–23.
- 176. Zhang Q, He J, Xu F, Huang X, Wang Y, Zhang W, et al. Supramolecular copolymer modified statin-loaded discoidal rHDLs for atherosclerotic anti-inflammatory therapy by cholesterol efflux and M2 macrophage polarization. Biomater Sci. 2021;9(18):6153–68.
- 177. Grinnemo KH, Månsson-Broberg A, Leblanc K, Corbascio M, Wärdell E, Siddiqui AJ, et al. Human mesenchymal stem cells do not differentiate into cardiomyocytes in a cardiac ischemic xenomodel. Ann Med. 2006;38(2):144–53.
- 178. Murry CE, Soonpaa MH, Reinecke H, Nakajima H, Nakajima HO, Rubart M, et al. Haematopoietic stem cells do not transdifferentiate into cardiac myocytes in myocardial infarcts. Nature. 2004;428(6983):664–8.
- Sanganalmath SK, Bolli R. Cell therapy for heart failure: a comprehensive overview of experimental and clinical studies, current challenges, and future directions. Circ Res. 2013;113(6):810–34.
- Le Blanc K, Mougiakakos D. Multipotent mesenchymal stromal cells and the innate immune system. Nat Rev Immunol. 2012;12(5):383–96.
- Cao W, Cao K, Cao J, Wang Y, Shi Y. Mesenchymal stem cells and adaptive immune responses. Immunol Lett. 2015;168(2):147–53.
- Pandey AC, Lancaster JJ, Harris DT, Goldman S, Juneman E. Cellular therapeutics for heart failure: focus on mesenchymal stem cells. Stem Cells Int. 2017;2017:1–12.
- van den Akker F, Deddens JC, Doevendans PA, Sluijter JPG. Cardiac stem cell therapy to modulate inflammation upon myocardial infarction. Biochim Biophys Acta Gen Subj. 2013;1830(2):2449–58.
- Ullah I, Subbarao RB, Rho GJ. Human mesenchymal stem cells current trends and future prospective. Biosci Rep. 2015;35(2).

- 185. Hare JM, DiFede DL, Rieger AC, Florea V, Landin AM, El-Khorazaty J, et al. Randomized comparison of allogeneic versus autologous mesenchymal stem cells for nonischemic dilated cardiomyopathy. J Am Coll Cardiol. 2017;69(5):526–37.
- Fisher SA, Zhang H, Doree C, Mathur A, Martin-Rendon E. Stem cell treatment for acute myocardial infarction. Cochrane Database Sys Rev. 2015(9).
- 187. Li M, Wu H, Yuan Y, Hu B, Gu N. Recent fabrications and applications of cardiac patch in myocardial infarction treatment. VIEW. 2022;3(2):20200153.
- White SJ, Chong JJH. Mesenchymal stem cells in cardiac repair: effects on myocytes, vasculature, and fibroblasts. Clin Ther. 2020;42(10):1880–91.
- 189. Kang Y, Nasr M, Guo Y, Uchida S, Weirick T, Li H, et al. Administration of cardiac mesenchymal cells modulates innate immunity in the acute phase of myocardial infarction in mice. Sci Rep. 2020;10(1):14754.
- 190. Frangogiannis NG. Regulation of the inflammatory response in cardiac repair. Circ Res. 2012;110(1):159–73.
- 191. Frangogiannis NG. Inflammation in cardiac injury, repair and regeneration. Curr Opin Cardiol. 2015;30(3):240–5.
- 192. Guilloton F, Caron G, Ménard C, Pangault C, Amé-Thomas P, Dulong J, et al. Mesenchymal stromal cells orchestrate follicular lymphoma cell niche through the CCL2-dependent recruitment and polarization of monocytes. Blood. 2012;119(11):2556–67.
- 193. Ren G, Zhao X, Wang Y, Zhang X, Chen X, Xu C, et al. CCR2dependent recruitment of macrophages by tumor-educated mesenchymal stromal cells promotes tumor development and is mimicked by TNFα. Cell Stem Cell. 2012;11(6):812–24.
- 194. Maggini J, Mirkin G, Bognanni I, Holmberg J, Piazzón IM, Nepomnaschy I, et al. Mouse bone marrow-derived mesenchymal stromal cells turn activated macrophages into a regulatory-like profile. PLoS ONE. 2010;5(2): e9252.
- 195. Dayan V, Yannarelli G, Billia F, Filomeno P, Wang XH, Davies JE, et al. Mesenchymal stromal cells mediate a switch to alternatively activated monocytes/macrophages after acute myocardial infarction. Basic Res Cardiol. 2011;106(6):1299–310.
- 196. Vasandan AB, Jahnavi S, Shashank C, Prasad P, Kumar A, Prasanna SJ. Human Mesenchymal stem cells program macrophage plasticity by altering their metabolic status via a PGE2-dependent mechanism. Sci Rep. 2016;6(1):38308.
- 197. Luz-Crawford P, Djouad F, Toupet K, Bony C, Franquesa M, Hoogduijn MJ, et al. Mesenchymal stem cell-derived interleukin 1 receptor antagonist promotes macrophage polarization and inhibits B cell differentiation. Stem Cells. 2016;34(2):483–92.
- 198. Akiyama K, Chen C, Wang D, Xu X, Qu C, Yamaza T, et al. Mesenchymal-stem-cell-induced immunoregulation involves FAS-ligand-/FAS-mediated T cell apoptosis. Cell Stem Cell. 2012;10(5):544–55.
- Lee RH, Yoon N, Reneau JC, Prockop DJ. Preactivation of human MSCs with TNF-α enhances tumor-suppressive activity. Cell Stem Cell. 2012;11(6):825–35.
- 200. Ren G, Zhang L, Zhao X, Xu G, Zhang Y, Roberts AI, et al. Mesenchymal stem cell-mediated immunosuppression occurs via concerted action of chemokines and nitric oxide. Cell Stem Cell. 2008;2(2):141–50.
- 201. Xu G, Zhang Y, Zhang L, Ren G, Shi Y. Bone marrow stromal cells induce apoptosis of lymphoma cells in the presence of IFNgamma and TNF by producing nitric oxide. Biochem Biophys Res Commun. 2008;375(4):666–70.
- 202. Krampera M, Glennie S, Dyson J, Scott D, Laylor R, Simpson E, et al. Bone marrow mesenchymal stem cells inhibit the response of naive and memory antigen-specific T cells to their cognate peptide. Blood. 2003;101(9):3722–9.

- English K, Wood KJ. Mesenchymal stromal cells in transplantation rejection and tolerance. Cold Spring Harb Perspect Med. 2013;3(5):a015560-a.
- Prevosto C, Zancolli M, Canevali P, Zocchi MR, Poggi A. Generation of CD4+ or CD8+ regulatory T cells upon mesenchymal stem cell-lymphocyte interaction. Haematologica. 2007;92(7):881–8.
- 205. English K, Ryan JM, Tobin L, Murphy MJ, Barry FP, Mahon BP. Cell contact, prostaglandin E(2) and transforming growth factor beta 1 play non-redundant roles in human mesenchymal stem cell induction of CD4+CD25(High) forkhead box P3+ regulatory T cells. Clin Exp Immunol. 2009;156(1):149–60.
- Che N, Li X, Zhou S, Liu R, Shi D, Lu L, et al. Umbilical cord mesenchymal stem cells suppress B-cell proliferation and differentiation. Cell Immunol. 2012;274(1):46–53.
- 207. Mosser DM, Edwards JP. Exploring the full spectrum of macrophage activation. Nat Rev Immunol. 2008;8(12):958–69.
- 208. Kudo H, Wada H, Sasaki H, Tsuji H, Otsuka R, Baghdadi M, et al. Induction of macrophage-like immunosuppressive cells from mouse ES cells that contribute to prolong allogeneic graft survival. PLoS ONE. 2014;9(10): e111826.
- 209. Wada H, Kojo S, Kusama C, Okamoto N, Sato Y, Ishizuka B, et al. Successful differentiation to T cells, but unsuccessful B-cell generation, from B-cell-derived induced pluripotent stem cells. Int Immunol. 2011;23(1):65–74.
- Kofidis T, deBruin JL, Tanaka M, Zwierzchoniewska M, Weissman I, Fedoseyeva E, et al. They are not stealthy in the heart: embryonic stem cells trigger cell infiltration, humoral and T-lymphocyte-based host immune response. Eur J Cardiothorac Surg. 2005;28(3):461–6.
- 211. Swijnenburg RJ, Tanaka M, Vogel H, Baker J, Kofidis T, Gunawan F, et al. Embryonic stem cell immunogenicity increases upon differentiation after transplantation into ischemic myocardium. Circulation. 2005;112(9 Suppl):1166–72.
- Vagnozzi RJ, Maillet M, Sargent MA, Khalil H, Johansen AKZ, Schwanekamp JA, et al. An acute immune response underlies the benefit of cardiac stem cell therapy. Nature. 2020;577(7790):405–9.
- Abou-El-Enein M, Volk HD, Reinke P. Clinical development of cell therapies: setting the stage for academic success. Clin Pharmacol Ther. 2017;101(1):35–8.
- Sharir R, Semo J, Shimoni S, Ben-Mordechai T, Landa-Rouben N, Maysel-Auslender S, et al. Experimental myocardial infarction induces altered regulatory t cell hemostasis, and adoptive transfer attenuates subsequent remodeling. PLoS ONE. 2014;9(12): e113653.
- Pappritz K, Savvatis K, Miteva K, Kerim B, Dong F, Fechner H, et al. Immunomodulation by adoptive regulatory T-cell transfer improves Coxsackievirus B3-induced myocarditis. FASEB J. 2018;32(11):6066–78.
- 216. Meng X, Zheng M, Yu M, Bai W, Zuo L, Bu X, et al. Transplantation of CRISPRa system engineered IL10-overexpressing bone marrowderived mesenchymal stem cells for the treatment of myocardial infarction in diabetic mice. J Biol Eng. 2019;13(1).
- 217. Nash A, Aghlara-Fotovat S, Hernandez A, Scull C, Veiseh O. Clinical translation of immunomodulatory therapeutics. Adv Drug Deliv Rev. 2021;176: 113896.
- 218. Joki Y, Ohashi K, Yuasa D, Shibata R, Ito M, Matsuo K, et al. FGF21 attenuates pathological myocardial remodeling following myocardial infarction through the adiponectin-dependent mechanism. Biochem Biophys Res Commun. 2015;459(1):124–30.
- Formiga FR, Tamayo E, Simón-Yarza T, Pelacho B, Prósper F, Blanco-Prieto MJ. Angiogenic therapy for cardiac repair based on protein delivery systems. Heart Fail Rev. 2012;17(3):449–73.
- 220. Rosano JM, Cheheltani R, Wang B, Vora H, Kiani MF, Crabbe DL. Targeted delivery of VEGF after a myocardial infarction reduces collagen deposition and improves cardiac function. Cardiovasc Eng Technol. 2012;3(2):237–47.

- 221. Bauzá MdR, Giménez CS, Locatelli P, De Lorenzi A, Hnatiuk A, Capogrossi MC, et al. High-dose intramyocardial HMGB1 induces long-term cardioprotection in sheep with myocardial infarction. Drug Deliv Trans Res. 2019;9(5):935–44.
- 222. Muttenthaler M, King GF, Adams DJ, Alewood PF. Trends in peptide drug discovery. Nat Rev Drug Discovery. 2021;20(4):309–25.
- 223. Qin C, Yang YH, May L, Gao X, Stewart AG, Tu Y, et al. Cardioprotective potential of annexin-A1 mimetics in myocardial infarction. Pharmacol Ther. 2015;148:47–65.
- 224. Hiesinger W, Frederick JR, Atluri P, McCormick RC, Marotta N, Muenzer JR, et al. Spliced stromal cell-derived factor-1α analog stimulates endothelial progenitor cell migration and improves cardiac function in a dose-dependent manner after myocardial infarction. J Thorac Cardiovasc Surg. 2010;140(5):1174–80.
- 225. Hu B, Xu L, Li Y, Bai X, Xing M, Cao Q, et al. A peptide inhibitor of macrophage migration in atherosclerosis purified from the leech Whitmania pigra. J Ethnopharmacol. 2020;254: 112723.
- 226. Baran KW, Nguyen M, McKendall GR, Lambrew CT, Dykstra G, Palmeri ST, et al. Double-blind, randomized trial of an anti-CD18 antibody in conjunction with recombinant tissue plasminogen activator for acute myocardial infarction: limitation of myocardial infarction following thrombolysis in acute myocardial infarction (LIMIT AMI) study. Circulation. 2001;104(23):2778–83.
- 227. Faxon DP, Gibbons RJ, Chronos NA, Gurbel PA, Sheehan F. The effect of blockade of the CD11/CD18 integrin receptor on infarct size in patients with acute myocardial infarction treated with direct angioplasty: the results of the HALT-MI study. J Am Coll Cardiol. 2002;40(7):1199–204.
- 228. Vakeva AP, Agah A, Rollins SA, Matis LA, Li L, Stahl GL. Myocardial infarction and apoptosis after myocardial ischemia and reperfusion: role of the terminal complement components and inhibition by anti-C5 therapy. Circulation. 1998;97(22):2259–67.
- Thomas TC, Rollins SA, Rother RP, Giannoni MA, Hartman SL, Elliott EA, et al. Inhibition of complement activity by humanized anti-C5 antibody and single-chain Fv. Mol Immunol. 1996;33(17–18):1389–401.
- Armstrong PW, Granger CB, Adams PX, Hamm C, Holmes D Jr, O'Neill WW, et al. Pexelizumab for acute ST-elevation myocardial infarction in patients undergoing primary percutaneous coronary intervention: a randomized controlled trial. JAMA. 2007;297(1):43–51.
- 231. Abbate A, Van Tassell BW, Biondi-Zoccai G, Kontos MC, Grizzard JD, Spillman DW, et al. Effects of interleukin-1 blockade with anakinra on adverse cardiac remodeling and heart failure after acute myocardial infarction [from the Virginia Commonwealth University-Anakinra Remodeling Trial (2) (VCU-ART2) pilot study]. Am J Cardiol. 2013;111(10):1394–400.
- 232. Yudkin JS, Kumari M, Humphries SE, Mohamed-Ali V. Inflammation, obesity, stress and coronary heart disease: is interleukin-6 the link? Atherosclerosis. 2000;148(2):209–14.
- 233. Kleveland O, Kunszt G, Bratlie M, Ueland T, Broch K, Holte E, et al. Effect of a single dose of the interleukin-6 receptor antagonist tocilizumab on inflammation and troponin T release in patients with non-ST-elevation myocardial infarction: a double-blind, randomized, placebo-controlled phase 2 trial. Eur Heart J. 2016;37(30):2406–13.
- 234. Morton AC, Rothman AM, Greenwood JP, Gunn J, Chase A, Clarke B, et al. The effect of interleukin-1 receptor antagonist therapy on markers of inflammation in non-ST elevation acute coronary syndromes: the MRC-ILA Heart Study. Eur Heart J. 2015;36(6):377–84.
- Li P, Zheng Y, Chen X. Drugs for autoimmune inflammatory diseases: from small molecule compounds to anti-TNF biologics. Front Pharmacol. 2017;8:460.
- 236. Panahi M, Vadgama N, Kuganesan M, Ng FS, Sattler S. Immunopharmacology of post-myocardial infarction and heart failure medications. J Clin Med. 2018;7(11):403.

- Huang S, Frangogiannis NG. Anti-inflammatory therapies in myocardial infarction: failures, hopes and challenges. Br J Pharmacol. 2018;175(9):1377–400.
- Narasimha DU. Lipoxins, resolvins, protectins, maresins and nitrolipids, and their clinical implications with specific reference to cancer: part I. Clinical Lipidology. 2013;8(4):437–63.
- Ji R-R, Xu Z-Z, Strichartz G, Serhan CN. Emerging roles of resolvins in the resolution of inflammation and pain. Trends Neurosci. 2011;34(11):599–609.
- Kain V, Ingle KA, Colas RA, Dalli J, Prabhu SD, Serhan CN, et al. Resolvin D1 activates the inflammation resolving response at splenic and ventricular site following myocardial infarction leading to improved ventricular function. J Mol Cell Cardiol. 2015;84:24–35.
- 241. Lee CH. Resolvins as new fascinating drug candidates for inflammatory diseases. Arch Pharm Res. 2012;35(1):3–7.
- 242. Serhan CN, Hong S, Gronert K, Colgan SP, Devchand PR, Mirick G, et al. Resolvins: a family of bioactive products of omega-3 fatty acid transformation circuits initiated by aspirin treatment that counter proinflammation signals. J Exp Med. 2002;196(8):1025–37.
- 243. Viola JR, Lemnitzer P, Jansen Y, Csaba G, Winter C, Neideck C, et al. Resolving lipid mediators maresin 1 and resolvin D2 prevent atheroprogression in mice. Circ Res. 2016;119(9):1030–8.
- Zhang MJ, Sansbury BE, Hellmann J, Baker JF, Guo L, Parmer CM, et al. Resolvin D2 enhances postischemic revascularization while resolving inflammation. Circulation. 2016;134(9):666–80.
- Salic K, Morrison MC, Verschuren L, Wielinga PY, Wu L, Kleemann R, et al. Resolvin E1 attenuates atherosclerosis in absence of cholesterol-lowering effects and on top of atorvastatin. Atherosclerosis. 2016;250:158–65.
- 246. Keyes KT, Ye Y, Lin Y, Zhang C, Perez-Polo JR, Gjorstrup P, et al. Resolvin E1 protects the rat heart against reperfusion injury. Am J Physiol Heart Circ Physiol. 2010;299(1):H153–64.
- 247. Gilbert K, Bernier J, Godbout R, Rousseau G. Resolvin D1, a metabolite of omega-3 polyunsaturated fatty acid, decreases post-myocardial infarct depression. Mar Drugs. 2014;12(11): 5396–407.
- Corsonello A, Garasto S, Abbatecola AM, Rose G, Passarino G, Mazzei B, et al. Targeting inflammation to slow or delay functional decline: where are we? Biogerontology. 2010;11(5):603–14.
- Jougasaki M, Ichiki T, Takenoshita Y, Setoguchi M. Statins suppress interleukin-6-induced monocyte chemo-attractant protein-1 by inhibiting Janus kinase/signal transducers and activators of transcription pathways in human vascular endothelial cells. Br J Pharmacol. 2010;159(6):1294–303.
- 250. Singh P, Kohr D, Kaps M, Blaes F. Influence of statins on MHC class I expression. Ann N Y Acad Sci. 2009;1173:746–51.
- 251. Shibasaki M, Wang J-G, Figueiredo J-L, New SEP, Quillard T, Goettsch C, et al. Pitavastatin reduces inflammation in atherosclerotic plaques in apolipoprotein e-deficient mice with late stage renal disease. PLoS ONE. 2015;10(9): e0138047.
- 252. Liu M, Yu Y, Jiang H, Zhang L, Zhang P-P, Yu P, et al. Simvastatin suppresses vascular inflammation and atherosclerosis in ApoE–/– mice by downregulating the HMGB1-RAGE axis. Acta Pharmacol Sin. 2013;34(6):830–6.
- 253. Aydin MU, Aygul N, Altunkeser BB, Unlu A, Taner A. Comparative effects of high-dose atorvastatin versus moderate-dose rosuvastatin on lipid parameters, oxidized-LDL and inflammatory markers in ST elevation myocardial infarction. Atherosclerosis. 2015;239(2):439–43.
- 254. Liu HL, Yang Y, Yang SL, Luo JP, Li H, Jing LM, et al. Administration of a loading dose of atorvastatin before percutaneous coronary intervention prevents inflammation and reduces myocardial injury in STEMI patients: a randomized clinical study. Clin Ther. 2013;35(3):261–72.
- 255. Yang N, Cheng W, Hu H, Xue M, Li X, Wang Y, et al. Atorvastatin attenuates sympathetic hyperinnervation together with the

augmentation of M2 macrophages in rats postmyocardial infarction. Cardiovasc Ther. 2016;34(4):234-44.

- Zhu J, Qiu Y, Wang Q, Zhu Y, Hu S, Zheng L, et al. Low dose cyclophosphamide rescues myocardial function from ischemiareperfusion in rats. Eur J Cardiothorac Surg. 2008;34(3):661–6.
- 257. Tardif J-C, Kouz S, Waters DD, Bertrand OF, Diaz R, Maggioni AP, et al. Efficacy and safety of low-dose colchicine after myocardial infarction. N Engl J Med. 2019;381(26):2497–505.
- Alves-Filho JC, Pålsson-McDermott EM. Pyruvate kinase M2: a potential target for regulating inflammation. Front Immunol. 2016;7:145.
- 259. Lu S, Tian Y, Luo Y, Xu X, Ge W, Sun G, et al. Iminostilbene, a novel small-molecule modulator of PKM2, suppresses macrophage inflammation in myocardial ischemia-reperfusion injury. J Adv Res. 2021;29:83–94.
- 260. Hu G, Su Y, Kang BH, Fan Z, Dong T, Brown DR, et al. Highthroughput phenotypic screen and transcriptional analysis identify new compounds and targets for macrophage reprogramming. Nat Comm. 2021;12(1).
- Deak P, Knight HR, Esser-Kahn A. Robust tolerogenic dendritic cells via push/pull pairing of toll-like-receptor agonists and immunomodulators reduces EAE. Biomaterials. 2022;286: 121571.
- 262. Soni SS, D'Elia AM, Alsasa A, Cho S, Tylek T, O'Brien EM, et al. Sustained release of drug-loaded nanoparticles from injectable hydrogels enables long-term control of macrophage phenotype. Biomater Sci. 2022.
- Patra JK, Das G, Fraceto LF, Campos EVR, Rodriguez-Torres MDP, Acosta-Torres LS, et al. Nano based drug delivery systems: recent developments and future prospects. J Nanobiotechnol. 2018;16(1).
- Liu C, Fan Z, He D, Chen H, Zhang S, Guo S, et al. Designer functional nanomedicine for myocardial repair by regulating the inflammatory microenvironment. Pharmaceutics. 2022;14(4):758.
- Tariq U, Gupta M, Pathak S, Patil R, Dohare A, Misra SK. Role of biomaterials in cardiac repair and regeneration: therapeutic intervention for myocardial infarction. ACS Biomater Sci Eng. 2022;8(8):3271–98.
- Deng J, Wang J, Shi J, Li H, Lu M, Fan Z, et al. Tailoring the physicochemical properties of nanomaterials for immunomodulation. Adv Drug Deliv Rev. 2022;180: 114039.
- 267. Soni SS, Rodell CB. Polymeric materials for immune engineering: molecular interaction to biomaterial design. Acta Biomaterialia. 2021.
- Rayahin JE, Buhrman JS, Zhang Y, Koh TJ, Gemeinhart RA. High and low molecular weight hyaluronic acid differentially influence macrophage activation. ACS Biomater Sci Eng. 2015;1(7):481–93.
- 269. Alstergren P, Zhu B, Glougauer M, Mak TW, Ellen RP, Sodek J. Polarization and directed migration of murine neutrophils is dependent on cell surface expression of CD44. Cell Immunol. 2004;231(1–2):146–57.
- Chen CH, Wang SS, Wei EI, Chu TY, Hsieh PC. Hyaluronan enhances bone marrow cell therapy for myocardial repair after infarction. Mol Ther. 2013;21(3):670–9.
- 271. Bonafè F, Govoni M, Giordano E, Caldarera CM, Guarnieri C, Muscari C. Hyaluronan and cardiac regeneration. J Biomed Sci. 2014;21(1).
- 272. Yoon SJ, Hong S, Fang YH, Song M, Son KH, Son HS, et al. Differential regeneration of myocardial infarction depending on the progression of disease and the composition of biomimetic hydrogel. J Biosci Bioeng. 2014;118(4):461–8.
- 273. Wang N, Liu C, Wang X, He T, Li L, Liang X, et al. Hyaluronic acid oligosaccharides improve myocardial function reconstruction and angiogenesis against myocardial infarction by regulation of macrophages. Theranostics. 2019;9(7):1980–92.
- Song Y, Zhang C, Zhang J, Sun N, Huang K, Li H, et al. An injectable silk sericin hydrogel promotes cardiac functional recovery after ischemic myocardial infarction. Acta Biomater. 2016;41:210–23.

- Harel-Adar T, Mordechai TB, Amsalem Y, Feinberg MS, Leor J, Cohen S. Modulation of cardiac macrophages by phosphatidylserine-presenting liposomes improves infarct repair. Proc Natl Acad Sci. 2011;108(5):1827–32.
- Cutler JI, Auyeung E, Mirkin CA. Spherical nucleic acids. J Am Chem Soc. 2012;134(3):1376–91.
- 277. Somasuntharam I, Yehl K, Carroll SL, Maxwell JT, Martinez MD, Che P-L, et al. Knockdown of TNF-α by DNAzyme gold nanoparticles as an anti-inflammatory therapy for myocardial infarction. Biomaterials. 2016;83:12–22.
- Richart AL, Reddy M, Khalaji M, Natoli AL, Heywood SE, Siebel AL, et al. Apo AI nanoparticles delivered post myocardial infarction moderate inflammation. Circ Res. 2020;127(11):1422–36.
- 279. Galili U, Zhu Z, Chen J, Goldufsky JW, Schaer GL. Near complete repair after myocardial infarction in adult mice by altering the inflammatory response with intramyocardial injection of α-Gal nanoparticles. Front Cardiovasc Med. 2021;8: 719160.
- Abaricia JO, Farzad N, Heath TJ, Simmons J, Morandini L, Olivares-Navarrete R. Control of innate immune response by biomaterial surface topography, energy, and stiffness. Acta Biomater. 2021; 133:58–73.
- 281. Abaricia JO, Shah AH, Musselman RM, Olivares-Navarrete R. Hydrophilic titanium surfaces reduce neutrophil inflammatory response and NETosis. Biomater Sci. 2020;8(8):2289–99.
- 282. Ley K. M1 means kill; M2 means heal. J Immunol. 2017;199(7):2191–3.
- Jeon S, Clavadetscher J, Lee D-K, Chankeshwara S, Bradley M, Cho W-S. Surface charge-dependent cellular uptake of polystyrene nanoparticles. Nanomaterials. 2018;8(12):1028.
- Alfarsi MA, Hamlet SM, Ivanovski S. Titanium surface hydrophilicity modulates the human macrophage inflammatory cytokine response. J Biomed Mater Res A. 2014;102(1):60–7.
- 285. Hamlet S, Alfarsi M, George R, Ivanovski S. The effect of hydrophilic titanium surface modification on macrophage inflammatory cytokine gene expression. Clin Oral Implants Res. 2012;23(5):584–90.
- Vasilijić S, Savić D, Vasilev S, Vucević D, Gasić S, Majstorović I, et al. Dendritic cells acquire tolerogenic properties at the site of sterile granulomatous inflammation. Cell Immunol. 2005;233(2):148–57.
- 287. Blakney AK, Swartzlander MD, Bryant SJ. Student award winner in the undergraduate category for the society of biomaterials 9th World Biomaterials Congress, Chengdu, China, June 1–5, 2012. J Biomed Mater Res Part A. 2012;100A(6):1375–86.
- Dutta B, Goswami R, Rahaman SO. TRPV4 plays a role in matrix stiffness-induced macrophage polarization. Front Immunol. 2020;11:570195.
- Noskovicova N, Schuster R, van Putten S, Ezzo M, Koehler A, Boo S, et al. Suppression of the fibrotic encapsulation of silicone implants by inhibiting the mechanical activation of pro-fibrotic TGF-β. Nat Biomed Eng. 2021;5(12):1437–56.
- 290. Barth KA, Waterfield JD, Brunette DM. The effect of surface roughness on RAW 264.7 macrophage phenotype. J Biomed Mater Res A. 2013;101(9):2679–88.
- 291. Refai AK, Textor M, Brunette DM, Waterfield JD. Effect of titanium surface topography on macrophage activation and secretion of proinflammatory cytokines and chemokines. J Biomed Mater Res A. 2004;70(2):194–205.
- 292. Doloff JC, Veiseh O, de Mezerville R, Sforza M, Perry TA, Haupt J, et al. The surface topography of silicone breast implants mediates the foreign body response in mice, rabbits and humans. Nat Biomed Eng. 2021;5(10):1115–30.
- 293. Meyer RA, Green JJ. Shaping the future of nanomedicine: anisotropy in polymeric nanoparticle design. Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology. 2016;8(2):191–207.

- 294. Safari H, Kelley WJ, Saito E, Kaczorowski N, Carethers L, Shea LD, et al. Neutrophils preferentially phagocytose elongated particles—an opportunity for selective targeting in acute inflammatory diseases. Science Adv. 2020;6(24):eaba1474.
- Rizvi SAA, Saleh AM. Applications of nanoparticle systems in drug delivery technology. Saudi Pharm J. 2018;26(1):64–70.
- Vo TN, Kasper FK, Mikos AG. Strategies for controlled delivery of growth factors and cells for bone regeneration. Adv Drug Deliv Rev. 2012;64(12):1292–309.
- 297. Kohane DS. Microparticles and nanoparticles for drug delivery. Biotechnol Bioeng. 2007;96(2):203–9.
- 298. Liu R, Kay BK, Jiang S, Chen S. Nanoparticle delivery: targeting and nonspecific binding. MRS Bull. 2009;34(6):432–40.
- 299. Friedman AD, Claypool SE, Liu R. The smart targeting of nanoparticles. Curr Pharm Des. 2013;19(35):6315–29.
- 300. Zhang Q, He J, Xu F, Huang X, Wang Y, Zhang W, et al. Supramolecular copolymer modified statin-loaded discoidal rHDLs for atherosclerotic anti-inflammatory therapy by cholesterol efflux and M2 macrophage polarization. Biomater Sci. 2021;9(18):6153–68.
- Katsuki S, Matoba T, Koga JI, Nakano K, Egashira K. Antiinflammatory nanomedicine for cardiovascular disease. Front Cardiovasc Med. 2017;4:87.
- Bulbake U, Doppalapudi S, Kommineni N, Khan W. Liposomal formulations in clinical use: an updated review. Pharmaceutics. 2017;9(4):12.
- Ruvinov E, Harel-Adar T, Cohen S. Bioengineering the infarcted heart by applying bio-inspired materials. J Cardiovasc Transl Res. 2011;4(5):559–74.
- Beavers KR, Nelson CE, Duvall CL. MiRNA inhibition in tissue engineering and regenerative medicine. Adv Drug Deliv Rev. 2015;88:123–37.
- Jo J-I, Gao J-Q, Tabata Y. Biomaterial-based delivery systems of nucleic acid for regenerative research and regenerative therapy. Regen Therap. 2019;11:123–30.
- Paunovska K, Loughrey D, Dahlman JE. Drug delivery systems for RNA therapeutics. Nat Rev Genet. 2022;23(5):265–80.
- 307. Li B, Zhang X, Dong Y. Nanoscale platforms for messenger RNA delivery. Wiley Interdiscip Rev Nanomed Nanobiotechnol. 2019;11(2):e1530-e.
- 308. Han J, Kim YS, Lim M-Y, Kim HY, Kong S, Kang M, et al. Dual roles of graphene oxide to attenuate inflammation and elicit timely polarization of macrophage phenotypes for cardiac repair. ACS Nano. 2018;12(2):1959–77.
- 309. Brovold M, Almeida JI, Pla-Palacín I, Sainz-Arnal P, Sánchez-Romero N, Rivas JJ, et al. Naturally-derived biomaterials for tissue engineering applications. Adv Exp Med Biol. 2018;1077:421–49.
- Muir VG, Burdick JA. Chemically modified biopolymers for the formation of biomedical hydrogels. Chem Rev. 2020;121(18):10908–49.
- Sarig U, Sarig H, de-Berardinis E, Chaw S-Y, Nguyen EBV, Ramanujam VS, et al. Natural myocardial ECM patch drives cardiac progenitor based restoration even after scarring. Acta Biomaterialia. 2016;44:209–20.
- Shah M, Kc P, Copeland KM, Liao J, Zhang G. A thin layer of decellularized porcine myocardium for cell delivery. Sci Rep. 2018;8(1):16206.
- 313. Singelyn JM, Sundaramurthy P, Johnson TD, Schup-Magoffin PJ, Hu DP, Faulk DM, et al. Catheter-deliverable hydrogel derived from decellularized ventricular extracellular matrix increases endogenous cardiomyocytes and preserves cardiac function postmyocardial infarction. J Am Coll Cardiol. 2012;59(8):751–63.
- 314. Traverse JH, Henry TD, Dib N, Patel AN, Pepine C, Schaer GL, et al. First-in-man study of a cardiac extracellular matrix hydrogel in early and late myocardial infarction patients. JACC Basic Transl Sci. 2019;4(6):659–69.

- 315. Liu G, Li L, Li Y, Huo D, Wu Y, Cheng P, et al. Enhancement of cardiac function with spleen-specific hydrogel via improving the immune microenvironment after myocardial infarction. J Biomater Tissue Eng. 2017;7(6):458–68.
- Rodell CB, Kaminski AL, Burdick JA. Rational design of network properties in guest-host assembled and shear-thinning hyaluronic acid hydrogels. Biomacromol. 2013;14(11):4125–34.
- 317. Hosoyama K, Ahumada M, McTiernan CD, Davis DR, Variola F, Ruel M, et al. Nanoengineered electroconductive collagen-based cardiac patch for infarcted myocardium repair. ACS Appl Mater Interfaces. 2018;10(51):44668–77.
- 318. Mann DL, Lee RJ, Coats AJ, Neagoe G, Dragomir D, Pusineri E, et al. One-year follow-up results from AUGMENT-HF: a multicentre randomized controlled clinical trial of the efficacy of left ventricular augmentation with Algisyl in the treatment of heart failure. Eur J Heart Fail. 2016;18(3):314–25.
- Reddy MSB, Ponnamma D, Choudhary R, Sadasivuni KK. A comparative review of natural and synthetic biopolymer composite scaffolds. Polymers (Basel). 2021;13(7).
- Spicer CD. Hydrogel scaffolds for tissue engineering: the importance of polymer choice. Polym Chem. 2020;11(2):184–219.
- 321. Yao Y, Li A, Wang S, Lu Y, Xie J, Zhang H, et al. Multifunctional elastomer cardiac patches for preventing left ventricle remodeling after myocardial infarction in vivo. Biomaterials. 2022;282: 121382.
- Maas SLN, Breakefield XO, Weaver AM. Extracellular vesicles: unique intercellular delivery vehicles. Trends Cell Biol. 2017;27(3):172–88.
- 323. Segura E, Nicco C, Lombard B, Véron P, Raposo G, Batteux F, et al. ICAM-1 on exosomes from mature dendritic cells is critical for efficient naive T-cell priming. Blood. 2005;106(1):216–23.
- 324. Okoye IS, Coomes SM, Pelly VS, Czieso S, Papayannopoulos V, Tolmachova T, et al. MicroRNA-containing T-regulatory-cellderived exosomes suppress pathogenic T helper 1 cells. Immunity. 2014;41(1):89–103.
- 325. Phinney DG, Di Giuseppe M, Njah J, Sala E, Shiva S, St Croix CM, et al. Mesenchymal stem cells use extracellular vesicles to outsource mitophagy and shuttle microRNAs. Nat Commun. 2015;6:8472.
- 326. Barile L, Milano G, Vassalli G. Beneficial effects of exosomes secreted by cardiac-derived progenitor cells and other cell types in myocardial ischemia. Stem Cell Investig. 2017;4:93.
- 327. Ben-Mordechai T, Palevski D, Glucksam-Galnoy Y, Elron-Gross I, Margalit R, Leor J. Targeting macrophage subsets for infarct repair. J Cardiovasc Pharmacol Ther. 2015;20(1):36–51.
- de Couto G, Gallet R, Cambier L, Jaghatspanyan E, Makkar N, Dawkins JF, et al. Exosomal microRNA transfer into macrophages mediates cellular postconditioning. Circulation. 2017;136(2):200–14.
- 329. Wu G, Zhang J, Zhao Q, Zhuang W, Ding J, Zhang C, et al. Molecularly engineered macrophage-derived exosomes with inflammation tropism and intrinsic heme biosynthesis for atherosclerosis treatment. Angew Chem Int Ed Engl. 2020;59(10):4068–74.
- Onuma AE, Zhang H, Gil L, Huang H, Tsung A. Surgical stress promotes tumor progression: a focus on the impact of the immune response. J Clin Med. 2020;9(12):4096.
- 331. Ackland G, Abbott T, Cain D, Edwards M, Sultan P, Karmali S, et al. Preoperative systemic inflammation and perioperative myocardial injury: prospective observational multicentre cohort study of patients undergoing non-cardiac surgery. Br J Anaesth. 2019;122(2):180–7.
- 332. Squiccimarro E, Labriola C, Malvindi PG, Margari V, Guida P, Visicchio G, et al. Prevalence and clinical impact of systemic inflammatory reaction after cardiac surgery. J Cardiothorac Vasc Anesth. 2019;33(6):1682–90.
- 333. Lee LS, Ghanta RK, Mokashi SA, Coelho-Filho O, Kwong RY, Bolman RM 3rd, et al. Ventricular restraint therapy for heart failure: the right ventricle is different from the left ventricle. J Thorac Cardiovasc Surg. 2010;139(4):1012–8.

- Gorman RC, Jackson BM, Burdick JA, Gorman JH. Infarct restraint to limit adverse ventricular remodeling. J Cardiovasc Transl Res. 2011;4(1):73–81.
- 335. Naveed M, Mohammad IS, Xue L, Khan S, Gang W, Cao Y, et al. The promising future of ventricular restraint therapy for the management of end-stage heart failure. Biomed Pharmacother. 2018;99:25–32.
- 336. Sabbah HN, Sharov VG, Gupta RC, Mishra S, Rastogi S, Undrovinas AI, et al. Reversal of chronic molecular and cellular abnormalities due to heart failure by passive mechanical ventricular containment. Circ Res. 2003;93(11):1095–101.
- 337. Pilla JJ, Blom AS, Brockman DJ, Bowen F, Yuan Q, Giammarco J, et al. Ventricular constraint using the acorn cardiac support device reduces myocardial akinetic area in an ovine model of acute infarction. Circulation. 2002;106(12\_suppl\_1):I-207-I-11.
- 338. Mann DL, Kubo SH, Sabbah HN, Starling RC, Jessup M, Oh JK, et al. Beneficial effects of the CorCap cardiac support device: five-year results from the Acorn Trial. J Thorac Cardiovasc Surg. 2012;143(5):1036–42.
- 339. Klodell CT Jr, Aranda JM Jr, McGiffin DC, Rayburn BK, Sun B, Abraham WT, et al. Worldwide surgical experience with the Paracor HeartNet cardiac restraint device. J Thorac Cardiovasc Surg. 2008;135(1):188–95.
- 340. Ghanta RK, Lee LS, Umakanthan R, Laurence RG, Fox JA, Bolman RM III, et al. Real-time adjustment of ventricular restraint therapy in heart failure. Eur J Cardiothorac Surg. 2008;34(6):1136–40.
- 341. Naveed M, Han L, Khan GJ, Yasmeen S, Mikrani R, Abbas M, et al. Cardio-supportive devices (VRD & DCC device) and patches for advanced heart failure: A review, summary of state of the art and future directions. Biomed Pharmacother. 2018;102:41–54.
- 342. Nawaz W, Naveed M, Zhang J, Noreen S, Saeed M, Sembatya KR, et al. Cardioprotective effect of silicon-built restraint device (ASD), for left ventricular remodeling in rat heart failure model. J Mater Sci Mater Med. 2022;33(5):42.
- 343. Enomoto Y, Gorman JH III, Moainie SL, Jackson BM, Parish LM, Plappert T, et al. Early ventricular restraint after myocardial infarction: extent of the wrap determines the outcome of remodeling. Ann Thorac Surg. 2005;79(3):881–7.
- 344. Mei X, Cheng K. Recent development in therapeutic cardiac patches. Front Cardio Med. 2020;7:610364.
- 345. Vasu S, Zhou J, Chen J, Johnston PV, Kim D-H. Biomaterials-based approaches for cardiac regeneration. Korean Circ J. 2021;51(12):943–60.
- Perez-Estenaga I, Prosper F, Pelacho B. Allogeneic mesenchymal stem cells and biomaterials: the perfect match for cardiac repair? Int J Mol Sci. 2018;19(10):3236.
- 347. Gomes M, Azevedo H, Malafaya P, Silva S, Oliveira J, Silva G, et al. Natural polymers in tissue engineering applications. Tissue engineering: Elsevier; 2008. p. 145–92.
- 348. Kyburz KA, Anseth KS. Synthetic mimics of the extracellular matrix: how simple is complex enough? Ann Biomed Eng. 2015;43(3):489–500.
- Bejleri D, Davis ME. Decellularized extracellular matrix materials for cardiac repair and regeneration. Adv Healthc Mater. 2019;8(5): e1801217.
- Wang L, Serpooshan V, Zhang J. Engineering human cardiac muscle patch constructs for prevention of post-infarction LV remodeling. Front Cardio Med. 2021;8:621781.
- 351. Schaefer JA, Guzman PA, Riemenschneider SB, Kamp TJ, Tranquillo RT. A cardiac patch from aligned microvessel and cardiomyocyte patches. J Tissue Eng Regen Med. 2018;12(2):546–56.
- 352. Querdel E, Reinsch M, Castro L, Köse D, Bähr A, Reich S, et al. Human engineered heart tissue patches remuscularize the injured heart in a dose-dependent manner. Circulation (New York, NY). 2021;143(20):1991–2006.

- 353. Weinberger F, Breckwoldt K, Pecha S, Kelly A, Geertz B, Starbatty J, et al. Cardiac repair in guinea pigs with human engineered heart tissue from induced pluripotent stem cells. Sci Trans Med. 2016;8(363):363ra148.
- 354. Sharma M, Schlegel MP, Afonso MS, Brown EJ, Rahman K, Weinstock A, et al. Regulatory T cells license macrophage proresolving functions during atherosclerosis regression. Circ Res. 2020;127(3):335–53.
- 355. Bentley ER, Little SR. Local delivery strategies to restore immune homeostasis in the context of inflammation. Adv Drug Deliv Rev. 2021;178: 113971.
- Tous E, Purcell B, Ifkovits JL, Burdick JA. Injectable acellular hydrogels for cardiac repair. J Cardiovasc Transl Res. 2011;4(5):528–42.
- 357. Leor J, Tuvia S, Guetta V, Manczur F, Castel D, Willenz U, et al. Intracoronary injection of in situ forming alginate hydrogel reverses left ventricular remodeling after myocardial infarction in swine. J Am Coll Cardiol. 2009;54(11):1014–23.
- 358. Sack KL, Aliotta E, Choy JS, Ennis DB, Davies NH, Franz T, et al. Intra-myocardial alginate hydrogel injection acts as a left ventricular mid-wall constraint in swine. Acta Biomater. 2020;111:170–80.
- 359. Sabbah HN, Wang M, Gupta RC, Rastogi S, Ilsar I, Sabbah MS, et al. Augmentation of left ventricular wall thickness with alginate hydrogel implants improves left ventricular function and prevents progressive remodeling in dogs with chronic heart failure. JACC: Heart Fail. 2013;1(3):252–8.
- Choy JS, Leng S, Acevedo-Bolton G, Shaul S, Fu L, Guo X, et al. Efficacy of intramyocardial injection of Algisyl-LVR for the treatment of ischemic heart failure in swine. Int J Cardiol. 2018;255:129–35.
- 361. Ifkovits JL, Tous E, Minakawa M, Morita M, Robb JD, Koomalsingh KJ, et al. Injectable hydrogel properties influence infarct expansion and extent of postinfarction left ventricular remodeling in an ovine model. Proceedings of the National Academy of Sciences PNAS. 2010;107(25):11507–12.
- 362. Dorsey SM, McGarvey JR, Wang H, Nikou A, Arama L, Koomalsingh KJ, et al. MRI evaluation of injectable hyaluronic acid-based hydrogel therapy to limit ventricular remodeling after myocardial infarction. Biomaterials. 2015;69:65–75.
- 363. Rodell CB, Lee ME, Wang H, Takebayashi S, Takayama T, Kawamura T, et al. Injectable shear-thinning hydrogels for minimally invasive delivery to infarcted myocardium to limit left ventricular remodeling. Circulation: Cardio Interv. 2016;9(10):e004058.
- Ghanta RK, Pugazenthi A, Zhao Y, Sylvester C, Wall MJ Jr, Mazur RA, et al. Influence of supraphysiologic biomaterial stiffness on ventricular mechanics and myocardial infarct reinforcement. Acta Biomater. 2022;149:30–9.
- 365. Wang H, Rodell CB, Lee ME, Dusaj NN, Gorman JH III, Burdick JA, et al. Computational sensitivity investigation of hydrogel injection characteristics for myocardial support. J Biomech. 2017;64:231–5.
- 366. Li DS, Avazmohammadi R, Rodell CB, Hsu EW, Burdick JA, Gorman JH III, et al. How hydrogel inclusions modulate the local mechanical response in early and fully formed post-infarcted myocardium. Acta Biomater. 2020;114:296–306.
- 367. Kc P, Hong Y, Zhang G. Cardiac tissue-derived extracellular matrix scaffolds for myocardial repair: advantages and challenges. Regen Biomater. 2019;6(4):185–99.
- 368. Wassenaar JW, Gaetani R, Garcia JJ, Braden RL, Luo CG, Huang D, et al. Evidence for mechanisms underlying the functional benefits of a myocardial matrix hydrogel for post-MI treatment. J Am Coll Cardiol. 2016;67(9):1074–86.
- 369. Rodell CB, MacArthur JW Jr, Dorsey SM, Wade RJ, Wang LL, Woo YJ, et al. Shear-thinning supramolecular hydrogels with secondary autonomous covalent crosslinking to modulate viscoelastic properties in vivo. Adv Func Mater. 2015;25(4): 636–44.

- Groll J, Singh S, Albrecht K, Moeller M. Biocompatible and degradable nanogels via oxidation reactions of synthetic thiomers in inverse miniemulsion. J Polym Sci, Part A: Polym Chem. 2009;47(20):5543–9.
- 371. Li J, Hu S, Zhu D, Huang K, Mei X, López de Juan Abad B, et al. All Roads lead to Rome (the Heart): cell retention and outcomes from various delivery routes of cell therapy products to the heart. J Am Heart Assoc. 2021;10(8):e020402-e.
- Improving viability of stem cells during syringe needle flow through the design of hydrogel cell carriers. Tissue Eng Part A. 2012;18(7–8):806–15.
- 373. Xie Y, Guan Q, Guo J, Chen Y, Yin Y, Han X. Hydrogels for exosome delivery in biomedical applications. Gels. 2022;8(6).
- 374. Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease. N Engl J Med. 2017;377(12):1119–31.
- Lazzerini PE, Hamilton RM, Boutjdir M. Cardioimmunology: Inflammation and Immunity in Cardiovascular Disease. Frontiers in cardiovascular medicine. 2019;6:181.
- 376. Steffens S, Van Linthout S, Sluijter J, Tocchetti G, Thum T, Madonna R. Stimulating pro-reparative immune responses to prevent adverse cardiac remodelling: Consensus Document from the joint 2019 meeting of the ESC Working Groups of Cellular Biology of the heart and Myocardial Function. Cardio Res. 2020.
- 377. Li M, Carpio DF, Zheng Y, Bruzzo P, Singh V, Ouaaz F, et al. An essential role of the NF-κB/Toll-like receptor pathway in induction of inflammatory and tissue-repair gene expression by necrotic cells. J Immunol. 2001;166(12):7128–35.

- Spiller KL, Anfang RR, Spiller KJ, Ng J, Nakazawa KR, Daulton JW, et al. The role of macrophage phenotype in vascularization of tissue engineering scaffolds. Biomaterials. 2014;35(15):4477–88.
- Eming SA, Krieg T, Davidson JM. Inflammation in wound repair: molecular and cellular mechanisms. J Investig Dermatol. 2007;127(3):514–25.
- Dolejsi T, Delgobo M, Schuetz T, Tortola L, Heinze KG, Hofmann U, et al. Adult T-cells impair neonatal cardiac regeneration. Eur Heart J. 2022;43(28):2698–709.
- Li J, Yang KY, Tam RCY, Chan VW, Lan HY, Hori S, et al. Regulatory T-cells regulate neonatal heart regeneration by potentiating cardiomyocyte proliferation in a paracrine manner. Theranostics. 2019;9(15):4324.
- Berendsen TA, Bruinsma BG, Puts CF, Saeidi N, Usta OB, Uygun BE, et al. Supercooling enables long-term transplantation survival following 4 days of liver preservation. Nat Med. 2014;20(7):790–3.
- 383. Clavien P-A, Dutkowski P, Mueller M, Eshmuminov D, Bautista Borrego L, Weber A, et al. Transplantation of a human liver following 3 days of ex situ normothermic preservation. Nature Biotechnol. 2022:1–7.
- O'Neill JD, Guenthart BA, Kim J, Chicotka S, Queen D, Fung K, et al. Cross-circulation for extracorporeal support and recovery of the lung. Nat Biomed Eng. 2017;1(3):1–15.
- 385. Hatami S, White CW, Qi X, Buchko M, Ondrus M, Kinnear A, et al. Immunity and stress responses are induced during ex situ heart perfusion. Circ Heart Fail. 2020;13(6):e006552.

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.