First report of sida yellow vein Madurai virus infecting Lisianthus (*Eustoma russellianum*)

U. Premchand¹ · G. M. Santosh¹ · K. S. Shankarappa¹ · M. Mantesh² · V. Venkataravanappa³ · P. Pavankumar⁴ · T. J. Nithin¹ · C. R. Jahir Basha² · C. N. Lakshminarayana Reddy²

Received: 14 September 2023 / Accepted: 28 February 2024 © The Author(s) under exclusive licence to Australasian Plant Pathology Society Inc. 2024

Abstract

Lisianthus (Eustoma russellianum) is one of the emerging cut flower crops in international flower market quickly ranked in top ten cut flowers worldwide. Despite its rising popularity, studies relating to the identification and characterization of viral diseases affecting it are lacking from India. Thus, the present study was focused on identification and characterization of virus in lisianthus plants samples collected from Vensai Floritech, Narasihmanahalli village, Tubagere hobli, Doddaballapur taluk, Bengaluru rural district, Karnataka state, India exhibiting symptoms similar to begomoviruses infections. Association of the begomovirus with sample was confirmed by PCR using begomovirus specific primers which resulted in the expected amplicon (~1.2 kb). Further, whole-genome amplification was done by rolling circle amplification (RCA) for one representative sample (LIS-1). The amplified RCA product was cloned, sequenced and analyzed. The phylogenetic and nucleotide (nt) sequence analysis revealed that the begomovirus associated with lisianthus plants showed the maximum nt identity of 91.0% with sida yellow vein Madurai virus (SIYVMV-TN:OM141480) infecting a weed, Sida cordata, reported from Tamil Nadu, India, which is geographically close to Karnataka. Based on species demarcation criteria for begomoviruses, the collected isolate is identified as a strain of sida yellow vein Madurai virus associated with leaf curl of lisianthus from India and proposed the name "Sida yellow vein Madurai virus -[India:Karnataka:Doddaballap ura:Lisianthus:2023]" and designated as SIYVMV-[IN:Kar:Dod:Lis:23]. Further, recombination analysis revealed a single intra-specific recombination event in the genomic region. Hence, this study provides a one more evidence of expanding host range for begomoviruses in India.

Keywords Characterization \cdot Detection \cdot *Eustoma russellianum* \cdot Lisianthus \cdot India \cdot Phylogeny \cdot Recombination \cdot Sequence demarcation tool

K. S. Shankarappa ksshankarappa@gmail.com

- ¹ Department of Plant Pathology, College of Horticulture, Bengaluru, University of Horticultural Sciences, Bagalkot, 560065, Bengaluru, Karnataka, India
- ² Department of Plant Pathology, College of Agriculture, University of Agricultural Sciences, GKVK,, 560065, Bengaluru, Karnataka, India
- ³ Division of Crop Protection, ICAR-Indian Institute of Horticultural Research, Hessaraghatta Lake PO, 560089, Bengaluru, Karnataka, India
- ⁴ Department of Floriculture and Landscape Architecture, College of Horticulture, 560065, Bengaluru, Karnataka, India

Eustoma russellianum commonly known as lisianthus, is an herbaceous flowering plant that belongs to the Gentianaceae family (Wazir et al. 2014). The popularity of lisianthus is increasing in India, and the area under cultivation is gradually expanding (Bhatia and Sindhu 2019; Lakshmaiah et al. 2019; Rehana and Bala 2022). Lisianthus cultivation faces challenges from various fungal (Bhatia et al. 2020; Wu et al. 2023; Hanagasaki et al. 2023) and viral diseases (Chen et al. 2000; Jan et al. 2003; Beikzadeh et al. 2011; Cohen et al. 2001; Taniguchi et al. 2023) worldwide leading to significant economic losses for growers. There are no reports of viral diseases affecting lisianthus in India. In this study we report the identification and characterization of a begomovirus affecting lisianthus in India.

Seven leaf samples from lisianthus plants exhibiting symptoms typical to begomovirus infection such as vein



thickening, leaf curling, and reduction in leaf size, yellow mosaic and stunted growth were collected (designated as LIS-1 to 7 isolates) along with an asymptomatic leaf sample from Vensai Floritech (13°29'00.4"N 77°28'18.5"E) (Bengaluru, India) (Fig. 1). To confirm the causal agent, total genomic DNA was isolated from the lisianthus samples (Lodhi et al. 1994) and subjected to PCR amplification using degenerate primers specific to DNA-A (PAR1c496/ PAL1v1978) and DNA-B (PBL1v2040/PCRc1) as described by Rojas et al. (1993). The amplification yielded PCR amplicon of ~1.2 kb, specific to begomoviruses with respect to DNA-A. However, there was no amplification with the DNA-B specific primers, indicating the virus associated with symptomatic lisianthus plants is a monopartite begomovirus. All the samples were also subjected to PCR for detecting the presence of satellite molecules i.e., betasatellite (Briddon et al. 2002) and alphasatellite (Bull et al. 2003) using specific primers but unsuccessful for amplification of both satellite molecules.

Further, one representative sample (LIS-1 isolate) was used for complete genome amplification by rolling circle DNA amplification (RCA) (Inoue-Nagata et al. 2004). The amplified RCA product (2 μ L) was digested with HindIII restriction enzyme to obtain a monomeric unit of ~2.7 kb. The digested product was cloned into the linearized pUC19 plasmid with the respective sites. The recombinant clones were confirmed by HindIII restriction digestion and sequenced by the primer walking method at Medauxin Pvt. Ltd., Bengaluru, Karnataka, India. The obtained complete genome sequence was assembled using different bioinformatic tools (BioEdit, ClustalX2, and SeaView), and sequence similarities were checked at the NCBI database using BLASTn (http://www.ncbi.nlm.nih.gov).

The complete DNA-A genome sequence of LIS-1 isolate was deposited in NCBI GenBank under the accession number OR371601 and its genome consists of 2754 nt with seven open reading frames (ORFs), which were identified by using the ORF finder tool (http://www.Ncbi.nlm.nih. Gov/gorf/gorf.html). Analysis showed that viral genome (DNA-A) codes for seven potential ORFs, two [V2 and V1 (CP)] on virion sense strand and five [C1 (Rep), C2 (TrAP), C3 (REn), C4 and C5] on complementary sense strand. These two strands are separated by intergenic region (IR), which harbored a predicted stem-loop structure containing nonanucleotide sequence TAATATTAC which constitutes the origin of viral DNA replication (Supplementary Tables 1 and Supplementary Fig. 1). Pairwise nt and amino acid (aa) identities for the LIS-1 isolate from lisianthus were compared with selected begomoviruses sequences retrieved from GenBank using the Sequence Demarcation Tool (SDT) (SDT version 1.2) (http://web.cbio.uct.ac.za/ breinev/) (Muhire et al. 2014). Sequence analysis revealed that the LIS-1 isolate exhibited nt sequence identity of 87.8-91.0 per cent for the complete genome with sida yellow vein Madurai virus (SIYVMV). The highest nt identity (91.0%) indicates that the LIS-1 isolate represents a strain of SIYVMV infecting lisianthus (Brown et al. 2015; Table 1 and Suppl. Table 2). This is further supported by SDT analysis (Fig. 2b).

Based on the ICTV guidelines for begomovirus nomenclature (Fauquet and Stanley 2005), we propose the name "Sida yellow vein Madurai virus-[India:Karnataka:Dodd aballapura:Lisianthus:2023]", abbreviated as SIYVMV-[IN:Kar:Dod:Lis:23], for the isolate.

Amino acid identities of the proteins encoded by the isolate with those encoded by other begomoviruses are listed in Table 1.

A phylogenetic tree generated using the Neighbor-Joining method by MEGA11 (Tamura et al. 2021) revealed that SIYVMV-[IN:Kar:Dod:Lis:23] (OR371601) clustered with sida yellow vein Madurai virus (SIYVMV) infecting sida plants reported from Tamil Nadu, India (Fig. 2a). Recombination analysis of SIYVMV-[IN:Kar:Dod:Lis:23] using RDP 5 (Martin et al. 2021) revealed a single significant



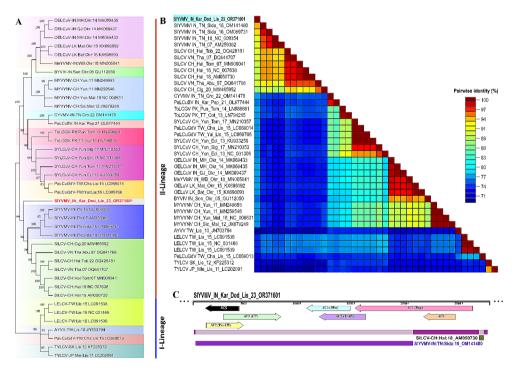
Fig. 1 Lisianthus infected plants showing begomoviral symptoms; (A) Reduction in leaf size, (B) Leaf curling as compare to (C) healthy under field conditions

Table 1 Pairwise nucleotide and amino acid identities between the begomovirus isolated from lishanthus (SIYVMV-[IN:Kar:Dod:Lis:23]) and selected begomoviruses

Begomovi-	Percent nt ider	ntity range (%)	Percent aa ide	entity range (%	b)				
ruses*	DNA-A	IR	AC1**	AC2	AC3	AC4	AC5	AV1	AV2
AYVV	70.6 [1]	56.5 [1]	70.8 [1]	55.5 [1]	64.9 [1]	30.6 [1]	13.2 [1]	72.7 [1]	56.5 [1]
BYVIV	76.5 [1]	53.9 [1]	83.4 [1]	54.5 [1]	67.9 [1]	86.6 ^[1]	14.5 [1]	78.1 [1]	53.9 [1]
CYVMV	78.2 [1]	54.8 [1]	84.0 [1]	64.2 [1]	64.9 [1]	82.5 ^[1]	-	82.8 [1]	54.8 [1]
LELCV	74.5 [3]	55.6 [3]	75.2-75.5 [3]	61.5-62.2 [3]	68.6–69.4 [3]	44.8-45.8 [3]	-	82.8 [3]	55.6 [3]
MeYVMV	77.8 [1]	54.8 [1]	85.7 [1]	53.1 [1]	66.4 [1]	82.5 [1]	14.5 [1]	82.0 [1]	54.7 [1]
MYVYNV	77.9–78.1 ^{[4}]	57.1-58.0 [4]	83.7-84.0 [4]	54.5 [3]	60.4-61.9 [4]	79.4-80.4 [4]	13.6 [2]	82.0-83.2 [4]	54.8-58.0 [4]
OELCuV	77.8–78.2 [5]	54.8-55.6 [5]	84.6–85.9 ^[5]	53.1-55.2 [5]	66.4 [5]	81.4-82.5 [5]	14.5 [3]	89.1-82.0 [5]	54.8-55.6 [5]
PaLCuBV	76.9 [1]	54. 8 [1]	82.6 [1]	61.9 [1]	63.4 [1]	80.0 [1]	15.5 ^[1]	82.8 [1]	54.8 [1]
PaLCuGdV	73.2–77.7 [3]	53.0 [3]	72.7-73.5 [3]	59.2-60.0 [3]	70.9 [3]	45.9 ^[3]	-	72.7-72.7 [3]	53.0 [3]
SiLCV	85.2-87.9 ^[7]	93.0-97.4 ^[7]	80.1-81.8 [7]	86.7-87.4 [7]	94.8–97.7 [7]	44.8-49.5 [7]	-	91.4–94.1 ^[7]	93.0-97.4 ^[7]
SIYVMV	87.8–91.0 ^[4]	34.8–100.0 ^[4]	33.8-83.7 [4]	72.3–95.1 ^[4]	42.5–98.5 ^[4]	45.3–48.4 [4]	-	60.9–94.9 ^[4]	34.8-100.0 ^[4]
SYLCuV	77.5–77.7 ^[4]	56.5 [4]	84.5-85.6 ^[4]	59.7-61.2 [4]	59.7-61.2 [4]	79.4-80.4 [4]	13.6 ^[3]	82.8-84.0 [4]	56.5 [4]
ToLCGV	78.1–78.4 [2]	57.4 [2]	84.0-84.9 [2]	61.2–61.9 ^[2]	61.9 [2]	82.4 [2]	-	82.8-82.2 [2]	57.4 [2]
TYLCV	72.4–72.9 [2]	55.6-56.5 [2]	75.9–76.9 [2]	55.5 [2]	67.1–67.9 [2]	67.0-71.1 [2]	-	66.8 [2]	55.6-56.5 [2]

Note: Numbers indicated in parenthesis are total sequences retrieved from databases for comparisons. *Ageratum yellow vein virus (AYVV); Bhendi yellow vein India virus (BYVIV); Croton yellow vein leaf curl virus (CYVMV); Lisianthus enation leaf curl virus (LELCV); Malvastrum yellow vein Yunnan virus (MeYVMV); Mesta yellow vein mosaic virus (MYVYNV); Okra enation leaf curl virus (OELCuV); Papaya leaf curl Bagalkote virus isolate (PaLCuBV); Papaya leaf curl Guandong virus (PaLCuGdV); Sida leaf curl virus (SiLCV); Sida yellow vein Madurai virus (SIYVMV); Synedrella leaf curl virus (SYLCuV); Tomato leaf curl Gujarat virus (ToLCGV); Tomato yellow leaf curl virus (TYLCV). **Genes are indicated as AV1:Coat protein (CP), AV2: Pre coat protein, AC1:Replication-associated protein (Rep), AC2:Transcriptional activator protein (TrAP) and AC3:Replication enhancer protein (REn). The products encoded by ORFs AC4 and AC5 have yet to be named

Fig. 2 (A) Phylogenic tree based on the complete nucleotide sequence of SIYVMV isolate from lisianthus (IN:Kar:Dod:Lis:23;OR371601) and other selected begomoviruses. (B) Nucleotide sequence identity matrix obtained from the same dataset using Sequence Demarcation Tool. and (C) recombination breakpoint analysis of SIYVMV (OR371601) isolate with other selected begomoviruses isolates reported across the world



intra-specific recombination event occurring at the 14th to 2087th nt position in the DNA-A genome of SIYVMV-[IN:Kar:Dod:Lis:23]. This recombinant fragment descends from okra enation leaf curl virus (OELCuV: MK069435) and sida yellow vein Madurai virus (SIYVMV: OM141480) as the major and minor parents, respectively (Table 2; Fig. 2c). This report provides evidence of the association of recombinant begomovirus with lisianthus for the first time in India. Furthermore, symptomatology, characterization, phylogeny and recombination analysis indicated that leaf curl disease of lisianthus is caused by a strain of SIYVMV.

Table 2 Recombination analysis of the begomovirus isolate from lisianthus (SIY)	gomovirus i	solate from	lisianthus (S	[IYVMV-[IN:Kar:Dod:Lis:23]).	3]).							
Begomovirus		Break p	Break point (nt)	Recombination parents			-d	<i>p</i> -value				
	Event	Begin End	End	Major parent	Minor parent	RDP	RDP Geneconv	Bootscan MaxChi Chimera SiScan 3Seq	MaxChi	Chimera	SiScan	3Seq
SIYVMV-[IN:Kar:Dod:Lis:23] (OR371601)	1	14	2087	OELCuV-[IN:MH:Okr:14] MK069435	SIYVMV-[IN:TN:Sida:18] 4.40E-19 1.47E-21 OM141480	4.40E-19	1.47E-21	NS*	1.37E-20	1.37E-20 1.63E-03 3.19E-57 1.88E-46	3.19E-57	1.88E-46
*Non significant value												

online Supplementary Information The version contains supplementary material available at https://doi.org/10.1007/s13314-024-00532-7.

Acknowledgements We are thankful to Government of Karnataka for funding Rashtriya Krishi Vikas Yojana (RKVY) project to University of Horticultural Sciences, Bagalkot, India.

Declarations

Accession number OR371601.

Ethical approval This article does not contain any studies with human participants performed by any of the authors.

Conflict of interest The authors declare that they have no competing interests.

References

- Beikzadeh N, Peters D, Hassani-Mehraban A (2011) First report of Moroccan pepper virus on lisianthus in Iran and worldwide. Plant Dis 95(11):1485-1485
- Bhatia R, Sindhu SS (2019) Vegetative propagation of lisianthus genotypes through stem cuttings: a viable alternative to seed propagation. Indian J Hortic 76(4):714-720
- Bhatia R, Dey SS, Rajkumar R (2020) Lisianthus:new cut flower crop for mid himalayan region. Indian Hortic 65(5):16-19
- Briddon RW, Bull SE, Mansoor S, Amin I, Markham PG (2002) Universal primers for the PCR-mediated amplification of DNA β: a molecule associated with some monopartite begomoviruses. Mol Biotechnol 20:315-318
- Brown JK, Zerbini FM, Navas-Castillo J, Moriones E, Ramos-Sobrinho R, Silva JC, Fiallo-Olivé E, Briddon RW, Hernández-Zepeda C, Idris A, Malathi VG (2015) Revision of Begomovirus taxonomy based on pairwise sequence comparisons. Arch Virol 160:1593-1619
- Bull SE, Briddon RW, Markham PG (2003) Universal primers for the PCR-mediated amplification of DNA 1: a satellite-like molecule associated with begomovirus-DNA ß complexes. Mol Biotechnol 23:83-86
- Chen CC, Chen YK, Hsu HT (2000) Characterization of a virus infecting lisianthus. Plant Dis 84(5):506-509
- Cohen J, Lapidot M, Loebenstein G, Gera A (2001) First report of sweet potato sunken vein virus occurring in Lisianthus. Plant Dis 85(6):679-769
- Fauquet CM, Stanley J (2005) Revising the way we conceive and name viruses below the species level: a review of geminivirus taxonomy calls for new standardized isolate descriptors. Arch Virol 150:2151-2179
- Hanagasaki T, Ajitomi A, Miwa E, Kiyuna T (2023) Field survey of Fusarium stem rot of lisianthus (Eustoma grandiflorum) cultivated in Okinawa, Japan. J Plant Prot Res 1427-4345
- Inoue-Nagata AK, Albuquerque LC, Rocha WB, Nagata T (2004) A simple method for cloning the complete begomovirus genome using the bacteriophage φ 29 DNA polymerase. J Virol Methods 116(2):209-211
- Jan FJ, Chen CC, Hsu HT (2003) Identification of tomato mosaic virus infection in lisianthus in Taiwan. Plant Dis 87(12):1537-1541
- Lakshmaiah K, Subramanian S, Ganga M, Jeyakumar P (2019) Optimization of pinching and GA3 application to improve growth and

flowering of lisianthus (*Eustoma grandiflorum*). J Pharmacogn Phytochem 8(6):614–616

- Lodhi MA, Ye GN, Weeden NF, Reisch BI (1994) A simple and efficient method for DNA extraction from grapevine cultivars and *Vitis* species. Plant Mol Bio Rep 12:6–13
- Martin DP, Varsani A, Roumagnac P, Botha G, Maslamoney S, Schwab T, Kelz Z, Kumar V, Murrell B (2021) RDP5: a computer program for analyzing recombination in, and removing signals of recombination from, nucleotide sequence datasets. Virus Evol 7(1):veaa087
- Muhire BM, Varsani A, Martin DP (2014) SDT: a virus classification tool based on pairwise sequence alignment and identity calculation. PLoS ONE 9(9):e108277
- Rehana S, Bala M (2022) Under exploited ornamental crops: treasure for floriculture industry. Ann Hortic 15(1):43–55
- Rojas MR, Gilbertson RL, Maxwell DP (1993) Use of degenerate primers in the polymerase chain reaction to detect whitefly-transmitted geminiviruses. Plant Dis 77(4):340

- Tamura K, Stecher G, Kumar S (2021) MEGA11: Molecular Evolutionary Genetics Analysis Version 11. Mol Biol Evol 38(7):3022–3027
- Taniguchi M, Sekine KT, Koeda S (2023) Lisianthus enation leaf curl virus, a begomovirus new to Japan, is more virulent than the prevalent tomato yellow leaf curl virus in Ty-gene-mediated resistant tomato cultivars. J Gen Plant Pathol 89(1):35–46
- Wazir JS (2014) Evaluation of eustoma/lisianthus cultivars for assessing their suitability as prominent new cut flower crop under mid hill conditions of HP. Int J Agric Sci Vet Med 2(1):105–110
- Wu CC, Shen YM, Teng YC, Chung WH (2023) First report of lisianthus wilt caused by *Fusarium oxysporum* f. sp. Eustomae Taiwan Crop Prot 29:106298

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.