Morphological and molecular characterization of downy mildew disease caused by *Peronospora variabilis* on *Chenopodium album* in Turkey



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Abstract

White goosefoot (*Chenopodium album*) is a common weed being most frequent in several crops. In April 2018, typical symptoms of downy mildew disease were observed on the leaf surface of *C. album* in pea fields inspected in Hatay province of Turkey. Based on morphological characteristics, molecular sequencing and pathogenicity test, the pathogen was identified as *Peronospora variabilis*. To our knowledge, this is the first confirmed report of downy mildew caused by *P. variabilis* on *C. album* in Eastern Mediterranean region of Turkey.

Keywords Peronospora variabilis · Chenopodium album · Downy mildew

White goosefoot, Chenopodium album (also known as Common lambsquarters) is an important worldwide annual weed being most frequent in several winter vegetable crops such as carrot, parsley, potatoes and cereals (Netland et al. 2001). It is also a typical plant of areas, which are unused for 1 or 2 years (ruderal areas). C. album is also an important plant species due to its medicinal applications and nutritional characteristics (Galwey et al. 1990). Presence of this weed may cause severe reductions in crop yield and, in the most severe cases, complete loss of yield (Conley et al. 2003). Downy mildew is an economically important and widespread disease on C. album. During recent surveys on major fungal disease of pea, we have noticed severe downy mildew symptoms on C. album growing as a weed in several pea fields in Hatay province. Although 20 species of Peronospora have been reported to cause downy mildew on Chenopodium spp., identification and classification of the pathogen remain still uncertain due to its

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taxonomic confusion (Choi et al. 2008). The purpose of this study was to analyse the morphological and molecular characterisation of the causal disease agent of downy mildew observed on *C. album* in Turkey.

During the disease surveys conducted in April 2018, typical symptoms of downy mildew disease were observed on the leaf surface of common weed species *C. album* in 60% of commercial pea fields inspected in Reyhanlı district (36°22' 59.2''N 36°31'29.8"E) of Hatay province of Turkey. Typical disease symptoms included the appearance of chlorotic spots or pink discoloration on the upper leaf surface of infected plants (Fig. 1A, B). These spots later turned necrotic. Dense tan or grey sporulation were frequently observed on the lower leaf surface (Fig. 1C). Leaves densely covered with a lot of conidiophores and conidia ultimately fallen off. When the infection is systemic, the plant showed dwarfing and yellowing. Infection during early development of the plant resulted in heavy defoliation.

The morphological characteristics of pathogen structures on heavily infected young leaves were examined by brightfield and Differential Interference Contrast (DIC) microscope (Olympus BX51, Japan) equipped with epifluorescence optics (Fig. 1D–F). Heavily infected young leaves were cleared in methanol/chloral hydrate and mounted in 50% glycerol as described previously (Soylu and Soylu 2003). Conidia (n = 50) were olivaceous with a greyish tint, broadly ellipsoidal to ellipsoidal, sometimes appearing as obovoid or napiform due to pedicel, greatest width median, 25.0–35.0 µm long, 22.5–27.5 µm wide, with a length/width ratio of 1.2–

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Fig. 1 Typical symptoms and morphological characteristics of downy mildew caused by *Peronospora variabilis* on *Chenopodium album*. Typical chlorotic spots or pink discoloration (arrow) on upper leaf surfaces (**A** and **B**) and dense tan or grey oomycete matrix (arrow)

1.4 (Fig. 1D). Conidiophores (n = 50) emerging through stomata, were up to 10 in a fascicle, colorless, straight to slightly curved 230.0–550.0 µm long; trunk substraight to slightly curved, 110.0–370.0 µm long, basal end not differentiated, rarely bulbous, up to 15.0 µm wide, 11.0–15.0 µm wide at the base, 7.5–10.0 µm wide below the first branch, ultimate branchlets in pairs or single (Fig. 1E). Resting organ (n = 50) were often present and clearly visible as dark brown to yellowish dots on the lower leaf surface; oogonia subglobose to irregular, 32.0–55.0 µm diameter; oospores globose, 20.0– 30.0 µm diameter, golden brown to brown (Fig. 1F).

The morphological characteristics closely resembled those reported for downy mildew disease agent *Peronospora variabilis* (Choi et al. 2008). A dried herbarium of the infected plant was deposited in the Korea University herbarium (KUS-F31532), Seoul, Korea.

Pathogenicity test was performed twice by inoculating the leaves of 10 healthy plant seedlings (3-weeks-old). Conidia were collected from fresh sporulating source plant leaves,

consisting of conidiophores, conidia and oospores on the corresponding lower leaf surfaces (**C**). Microscopic examination of *P. variabilis* showing (**D**) conidia, (**E**) ultimate branchlets on conidiophore, (**F**) oogonia (arrow) and oospore (os) in the infected *C. album* leaf. Bar = 20 μ m

dispersed into sterilised water with a drop of 0.1% Tween20 and mixed to produce a conidial suspension of 10^6 conidia/ml. Seedlings were inoculated with conidial suspension by placing several droplets onto lower leaf surfaces. Five noninoculated plants served as controls. Inoculated and noninoculated plants were maintained where they covered in a growth chamber at 20 °C/15 °C for 12 h day/night cycle and 95% relative humidity. Typical symptoms of downy mildew developed on the lower surfaces of inoculated leaves after 15 days. Non-inoculated plants remained symptomless. The pathogen present on the inoculated plants was morphologically identical to the original one observed on the diseased plants, fulfilling Koch's postulates.

To confirm the morphological identification, genomic DNA was extracted from conidiophores and conidia formed on leaves of infected host plants. The internal transcribed spacer (ITS) rDNA was amplified with primers ITS1-O and LR-0 and cytochrome oxidase II (*cox2*) mtDNA with primers *cox2*-F and *cox2*-RC4 (Choi et al. 2015). GenBank BLASTn

search revealed that the ITS and *cox*2 sequences of the Turkish isolate DMCa1 (Accession No. MK394004 and MK408662, respectively) were identical to ones of *P. variabilis* parasitic on *C. album* (EF614959 and KJ654199) and on *C. quinoa* (KF269542 and KF269666, respectively).

Based on morphological characteristics, molecular sequencing and pathogenicity test, the pathogen was identified as *P. variabilis*. Although disease agent was previously listed to occur on *C. album* in Ankara province (Gobelez 1962), to our knowledge, this is the first confirmation of downy mildew caused by *P. variabilis* on *C. album* in the Eastern Mediterranean region of Turkey. Disease agent was previously reported on *Chenopodium* spp. from different geographic origins including Europe, America and Asia (Choi et al. 2008; Choi et al. 2010; Farr and Rossman 2019).

References

Choi YJ, Denchev CM, Shin HD (2008) Morphological and molecular analyses support the existence of host-specific *Peronospora* species infecting *Chenopodium*. Mycopathologia 165:155–164

- Choi YJ, Danielsen S, Lübeck M, Hong SB, Delhey R, Shin HD (2010) Morphological and molecular characterization of the causal agent of downy mildew on quinoa (*Chenopodium quinoa*). Mycopathologia 169:403–412
- Choi YJ, Beakes G, Glockling S, Kruse J, Nam B, Nigrelli L, Ploch S, Shin HD, Shivas RG, Telle S, Voglmayr H, Thines M (2015) Towards a universal barcode of oomycetes–a comparison of the *cox1* and *cox2* loci. Mol Ecol Resour 15:1275–1288
- Conley SP, Stoltenberg DE, Boerboom CM, Binning LK (2003) Predicting soybean yield loss in giant foxtail (*Setaria faberi*) and common lambsquarters (*Chenopodium album*) communities. Weed Sci 51:402–407
- Farr DF, Rossman AY (2019) Systematic mycology and microbiology laboratory, ARS, USDA. http://nt.ars-grin.gov/fungaldatabases/ Accessed 13 December 2019
- Galwey NW, Leakey CLA, Price KR, Fenwick GR (1990) Chemical composition and nutritional characteristics of quinoa (*Chenopodium quinoa* Willd.). Food Sci Nutr 42F:245–261
- Gobelez M (1962) La mycoflore de Turkue I. Mycopathol Mycol Appl 19:296–314
- Netland J, Dutton LC, Greaves MP, Baldwin M, Vurro M, Evidente A, Einhorn G, Scheepens PC, French LW (2001) Biological control of *Chenopodium album* L. in Europe. Biocontrol 46:175–196
- Soylu EM, Soylu S (2003) Light and electron microscopy of the compatible interaction between Arabidopsis and the downy mildew pathogen *Peronospora parasitica*. J Phytopathol 151:300–306