



“*Candidatus Phytoplasma trifolii*” related strain affecting *Salix babylonica* in Iran

Fatemeh Shahryari¹ · Tohid Allahverdipour¹

Received: 2 May 2018 / Accepted: 12 October 2018 / Published online: 27 October 2018
© Australasian Plant Pathology Society Inc. 2018

Abstract

Over the last few years, in the northwest of Iran, symptoms of witches’ broom have been observed in Babylon willow trees (*Salix babylonica*). Samples from symptomatic and symptomless trees were tested using nested PCR with universal primer pair P1/P7 followed by R16F2n/R2. Phylogenetic and virtual RFLP analysis showed that the phytoplasma associated with willow proliferation is related to the 16SrVI-A subgroup.

Keywords Clover proliferation · Phytoplasma · P1/P7 · R16F2n/R2 · *Salix babylonica*

Salix babylonica, commonly known as Babylon willow, is a medium to large sized deciduous ornamental tree commonly grown in most regions of Iran. During the last few years, symptoms of witches’ broom, little leaf, and yellowing, resembling those often caused by phytoplasmas, have been observed in Babylon willow trees in Zanzan and West Azerbaijan provinces (Fig. 1). A total of 14 samples from both symptomatic (12 samples) and symptomless (2 samples) trees were collected from different areas in Zanzan and West Azerbaijan provinces and transported in cold condition to the laboratory and stored at -20°C until used. Total DNA extraction was performed using the procedure of Murray and Thompson (1980).

The extracted DNA was used as template for PCR with universal primers P1/P7, which amplify a 1784 bp DNA fragment of the phytoplasma 16S/23S rRNA region (Deng and Hiruki 1991; Smart et al. 1996).

Nested-PCR for amplification of the 16S rDNA was conducted using phytoplasma universal primer sets R16F2n/R16R2 (Gundersen and Lee 1996) in the second round. The expected amplicon of 1.2 kb was amplified in all 12 samples from symptomatic trees. Partial sequencing of the 16SrRNA region of four strains from different sampling areas was conducted with universal primer pair R16F2n/R2. The nucleotide sequences were assembled using Bioedit sequence alignment editor, version 7.0.9.0 and compared with available sequences in GenBank using blast alignment. Phylogenetic and molecular analyses were conducted by the neighbor-joining method in MEGA6 software (Tamura et al. 2013).

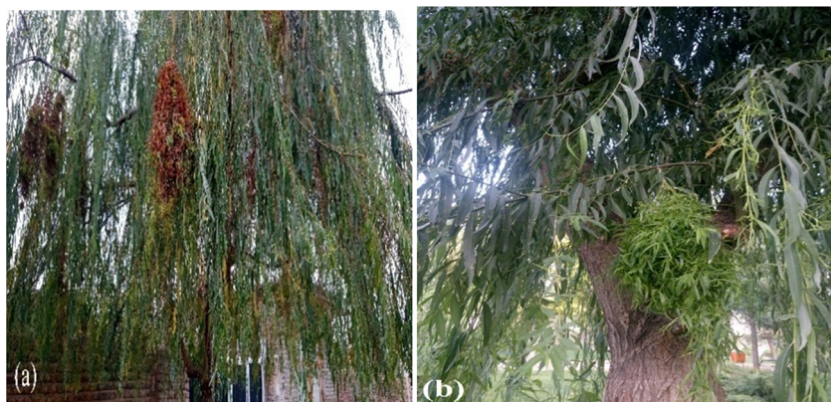
The obtained nucleotide sequences were deposited in the GenBank database under accession numbers MG437258, MG437259, MG437260 and, MG437261. In Blast software, the sequences showed 99% identity with some strains affiliated with ‘*Candidatus Phytoplasma trifolii*’ in GenBank (KX773529,

✉ Fatemeh Shahryari
shahryari@znu.ac.ir

Tohid Allahverdipour
tohid_ts@yahoo.com

¹ Department of Plant Protection, Faculty of Agriculture, University of Zanzan, Zanzan 45371-38791, Iran

Fig. 1 Witches' broom symptoms on Babylon willow trees from northwest of Iran. (a) Symptoms of Phytoplasma in Babylon willow tree, West Azerbaijan province, (b) Zanjan province, Iran



KP864671 and, KP864669). In the resulting phylogenetic tree constructed, based on the 16S rDNA sequences, all phytoplasma strains associated with Babylon willow witches' broom (BWVB) were clustered in the 16SrVI-A subgroup (Fig. 2).

Virtual RFLP of the sequences obtained (1.2 kbp) was performed with 17 typical restriction enzymes, *AluI*, *BamHI*, *BfaI*, *BstUI* (*ThaI*), *DraI*, *EcoRI*, *HaeIII*, *HhaI*, *HinfI*, *HpaI*, *HpaII*, *KpnI*, *MseI*, *RsaI*, *Sau3AI* (*MboI*), *SspI*, and *TaqI*, used for RFLP analyses of phytoplasma 16S rDNA genes (Lee et al. 1998) by iPhyClassifier (<https://plantpathology.ba.ars.usda.gov/cgi-bin/resource/iphyclassifier.cgi>) (Zhao et al. 2009). After in silico restriction digestion a virtual gel electrophoresis image was plotted and captured and putative restriction site maps were

compared with the pattern of strains deposited in GenBank. The restriction patterns revealed that the detected strains in the present study, SB21U, SB22U, SB23Z and SB24Z shared respectively 98.2%, 98.0%, 98.5 and 97.6% similarity with “*Candidatus* Phytoplasma trifolii” reference strain (AY390261) belonging to the 16SrVI-A subgroup. According to the iPhyClassifier online software, SB21U and SB23Z have the closest pattern with reference strain (AY390261) (Fig. 3).

Several phytoplasmas were previously reported from *Salix* trees including ‘*Ca. Phytoplasma asteris*’ (16SrI-C) and ‘*Ca. P. trifolii*’ (16SrVI-A) in China (Wei et al. 2009; Zhang et al. 2012), clover proliferation (16SrVI) and stolbur group in Spain (Alfaro-Fernández et al.

Fig. 2 The neighbor-joining tree generated using 16S rDNA sequences of isolated phytoplasma strains and 23 selected ‘*Candidatus* Phytoplasma species’ from GenBank. The numbers next to nodes are confidence values of bootstrap (1000 replicates). *Acholeplasma laidlawii* was used as the out-group. The scale at the bottom represents genetic distance in nucleotide substitutions per site. *Ca. P. sp.*: ‘*Candidatus* Phytoplasma’ sp.

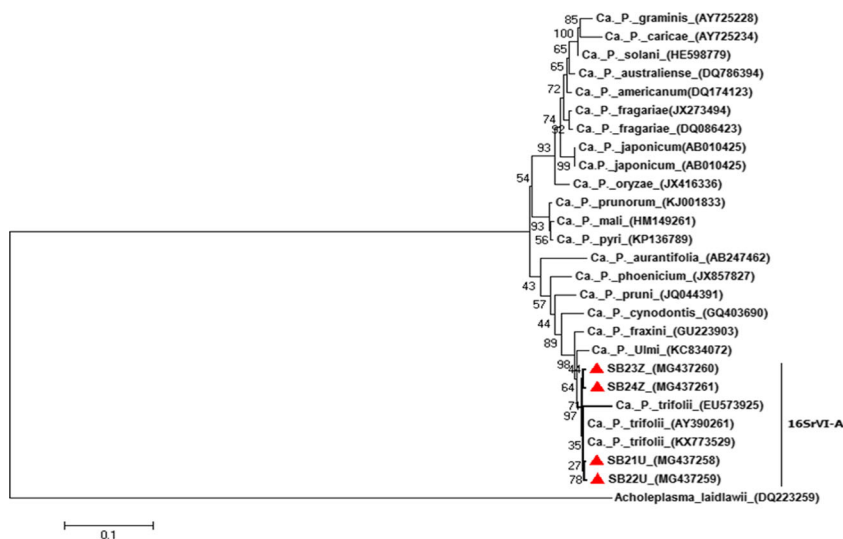
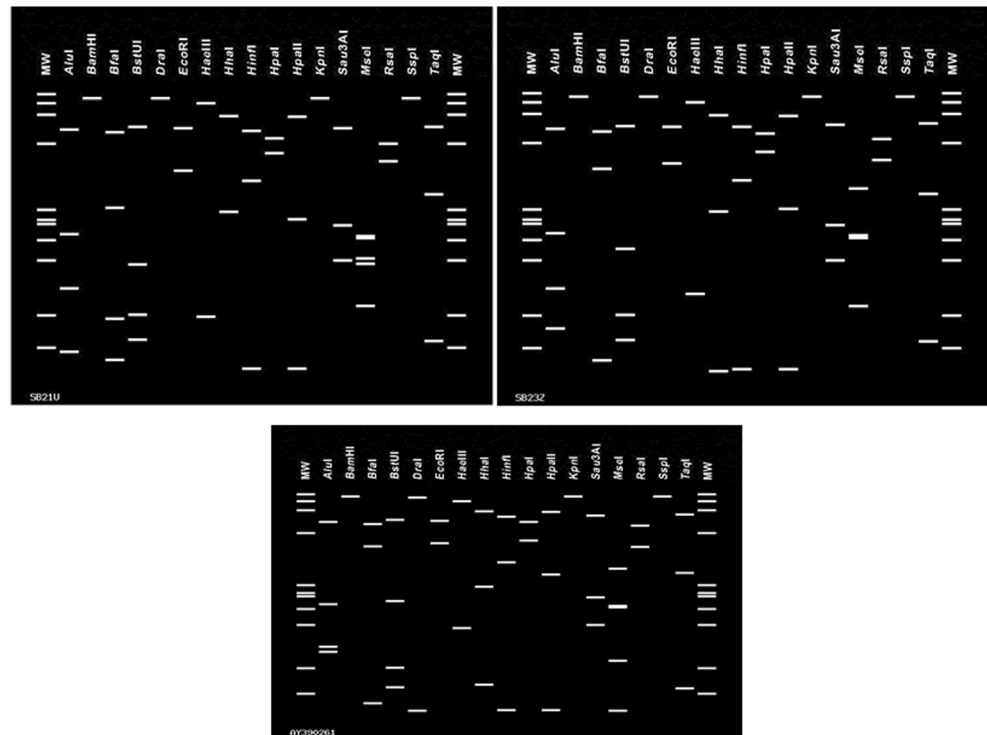


Fig. 3 Virtual RFLP patterns from in silico digestion of about 1200 bp nucleotide sequence of 16S rRNA gene of phytoplasmas associated with Babylon willow witches' broom (SB21U and SB23Z), and reference strain ('*Ca. P. trifolii*'; AY390261) using 17 restriction enzymes. (*AluI*, *BamHI*, *BfaI*, *BstUI* (*ThaI*), *DraI*, *EcoRI*, *HaeIII*, *HhaI*, *HinfI*, *HpaI*, *HpaII*, *KpnI*, *Sau3AI* (*MboI*), *MseI*, *RsaI*, *SspI*, and *TaqI*) in iPhyClassifier software. MW: Φ X174 DNA-*HaeIII* digested size marker



2011; Khadhair and Hiruki 1995), '*Ca. P. phoenicium*' (16SrIX) and *Ca. P. solani*' (16SrXII) in Iran (Ghayeb Zamhari 2017; Ghayeb Zamharir and Taheri 2017). Clover proliferation phytoplasma group (16SrVI) causes diseases in numerous plant species in different geographic areas worldwide (Raj et al. 2009; Choueiri et al. 2007; Salehi et al. 2008). However, to the authors' knowledge, there is no previous report of the 16SrVI phytoplasma group associated with Babylon willow witches' broom from Iran.

References

- Alfaro-Fernández A, Abad-Campos P, Hernández-Llopis D, Serrano-Fernandez A, Font-San-Ambrosio MI (2011) Detection of stolbur phytoplasma in willow in Spain. *Bull Insectology* 64(Supplement): S111–S112.
- Choueiri E, Salar P, Jreijiri F, El Zammar S, Massaad R, Abdul-Nour H, Foissac X (2007) Occurrence and distribution of '*Candidatus* Phytoplasma trifolii' associated with diseases of solanaceous crops in Lebanon. *Eur J Plant Pathol* 118:411–416
- Deng S, Hiruki C (1991) Amplification of 16S rRNA genes from culturable and nonculturable Mollicutes. *J Microbiol Methods* 14: 53–61
- Ghayeb Zamhari M (2017) First report of a '*Candidatus* Phytoplasma phoenicium'-related strain (16Sr IX) associated with *Salix* witches' broom in Iran. *New Dis Rep* 35:37–37
- Ghayeb Zamharir M, Taheri P (2017) '*Candidatus* Phytoplasma solani' related strain associated with Babylon willow witches' broom in central provinces of Iran. *Australas Plant Dis Notes* 12. <https://doi.org/10.1007/s13314-017-0268-z>
- Gundersen DE, Lee IM (1996) Ultrasensitive detection of phytoplasmas by nested-PCR assays using two universal primer pairs. *Phytopathol Mediterr* 35:144–151
- Khadhair AH, Hiruki C (1995) The molecular genetic relatedness of willow witches'-broom phytoplasma to the clover proliferation group. *Proc Jpn Acad Series B* 71:145–147
- Lee IM, Gundersen-Rindal DE, Davis RE, Bartoszyk IM (1998) Revised classification scheme of phytoplasmas based on RFLP analyses of 16S rRNA and ribosomal protein gene sequences. *Int J Syst Evol Microbiol* 48:1153–1169
- Murray MG, Thompson WF (1980) Rapid isolation of high molecular weight plant DNA. *Nucleic Acids Res* 8:4321–4326
- Raj SK, Snehi SK, Kumar S, Khan MS (2009) First finding of '*Candidatus* Phytoplasma trifolii' (16SrVI group) associated with little leaf disease of *Datura innoxia* in India. *Plant Pathol* 58:791–791
- Salehi M, Izadpanah K, Siampour M (2008) First report of '*Candidatus* Phytoplasma trifolii'-related strain associated with safflower phyllody disease in Iran. *Plant Dis* 92:649–649
- Smart CD, Schneider B, Blomquist CL, Guerra LJ, Harrison NA, Ahrens U, Lorenz K-H, Seemüller E, Kirkpatrick BC (1996) Phytoplasma-specific PCR primers based on sequences of the 16S–23S rRNA spacer region. *Appl Environ Microbiol* 62:2988–2993
- Tamura K, Stecher G, Peterson D, Filipinski A, Kumar S (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol* 30:2725–2729
- Wei T, Wu YF, Wu KK, Hou W, Li YR (2009) First report of a 16SrI-C group phytoplasma associated with a yellows-type disease affecting willow plants in China. *Plant Dis* 93:197–197
- Zhang L, Li Z, Du C, Fu Z, Wu Y (2012) Detection and identification of group 16SrVI phytoplasma in willows in China. *J Phytopathol* 160: 755–757
- Zhao Y, Wei W, Lee M, Shao J, Suo X, Davis RE (2009) Construction of an interactive online phytoplasma classification tool, iPhyClassifier, and its application in analysis of the peach X-disease phytoplasma group (16SrIII). *Int J Syst Evol Microbiol* 59:2582–2593