ORIGINAL ARTICLE



Evaluation of alkaline-based activated carbon from *Leucaena Leucocephala* produced at different activation temperatures for cadmium adsorption

Wan Muhammad Hilmi Wan Ibrahim¹ · Mohd Hazim Mohamad Amini¹ · Nurul Syuhada Sulaiman² · Wan Rasidah Wan Abdul Kadir³

Received: 7 July 2020 / Accepted: 16 November 2020 / Published online: 1 December 2020 © The Author(s) 2020

Abstract

Heavy metal contamination in water is happening worldwide. Adsorption using activated carbon is a common choice for cleaning the wastewater. The drawback of activated carbon is the higher cost of production due to the need for high heat in the process. This work investigated on activated carbon produced from the abundantly available *Leucaena leucocephala* biomass in order to reduce the cost of raw material. The biomass was chemically activated at different activation temperatures. The produced activated carbon was characterized using SEM, FT-IR, surface analyzer, and TGA. Isothermic and thermodynamic studies were done to evaluate the adsorption properties of the activated carbon. It was found out that higher surface area can be obtained using the higher activation temperature. Higher NaOH to carbonized sample ratios also resulted in higher surface area for all activation temperatures, which are $662 \text{ m}^2\text{g}^{-1}$ for 700 °C activation temperature, 735 m²g⁻¹ for 750 °C, and 776 m²g⁻¹ for 800 °C. Isothermic studies showed that all of the activated carbon that is produced from *Leucaena leucocephala* biomass are fit to the Langmuir isotherm, regardless of any activation temperature. Lastly, the thermodynamic study found out the adsorption process is endothermic, reflected by the positive value of ΔH^0 . It can be concluded that *Leucaena leucocephala* is a promising alternative material for producing activated carbon.

Keywords Activated carbon · Leucaena leucocephala · Cadmium · Adsorption · Isothermic · Thermodynamic

Introduction

Heavy metals have been associated with toxic elements. They have been listed in the United States Environmental Protection Agency (USEPA) as a pollutant that needs to be controlled and addressed (Gupta et al. 2002). Any form of metal, mineral salts, and trace elements has a specific function. Some need each other to work optimally. However, some substances will cause harm to the body at a specific threshold with toxic side effects (Rafatullah et al. 2011). It is

- ² School of Industrial Technology, Universiti Sains Malaysia, 18000 George, Penang, Malaysia
- ³ Forest Research Institute Malaysia, 52109 Kepong, Kuala Lumpur, Selangor, Malaysia

even more unfortunate if the presence of the material cannot be accurately confirmed until symptoms arise. The materials or metals often associated with such phenomena are lead, cadmium, and chromium (Singh et al. 2007).

In developing countries, various types of development projects are being implemented and involve varying degrees of environmental disturbance. Because of these developments, various problems have cropped up in the course of these developments. The most significant effect is the impact on human health. When humans are exposed to heavy metals for an extended period, it can cause serious health problems such as cancer (Lee et al. 2018).

Cadmium is one of the dangerous heavy metals utilized by humans for various material industries. It is used in battery manufacturing. It is beneficial, but it could be a danger to human health when it exposes to water (Wu et al. 2016). In industrial settings, cadmium waste flowed into the water and resulted in hazardous exposure to human health. Cadmium is present as a by-product of the filtration of some heavy metals such as lead and zinc. The danger of cadmium



Mohd Hazim Mohamad Amini hazimamini@gmail.com

¹ Faculty of Bio-Engineering and Technology, Universiti Malaysia Kelantan, 17600 Jeli, Kelantan, Malaysia

is that it can enter our bodies through food, and for longterm exposure, it could lead to serious health problems like cancer (Chunhabundit 2016).

The case of cadmium poisoning happened in a few countries around the world. Japan experienced cadmium poisoning in 1974. The pollution is caused by the disposal of lead–zinc mining waste, which contains cadmium into the Jintsu River. Overflowing of polluted rivers flooded the rice fields, which resulted in the absorption of cadmium into rice plants. This has an impact on the content of cadmium in rice, whereby it reaches between 3 and 4 μ g cadmium per kg (Uetani et al. 2006).

Various water treatment techniques were applied, including the adsorption, reverse osmosis membrane, solvent extraction, and high technology ultrafiltration process (Li et al. 2020a). The adsorption method is being the most popular due to lower operation costs and simple implementation (Li et al. 2019b). The activated carbon is a popular choice of material for water treatment. Activated carbon is solid charcoal that has been further processed so that it has better properties as an adsorbent. The ability of activated carbon as an adsorbent is caused by the formation of pores on the charcoal due to the carbonization and activation process (Bansal and Goyal 2005). Properties of activated carbon are affected by the type of raw material and its manufacturing conditions (Ningrum et al. 2008). Due to several factors such as renewability, lower cost, and environmentally friendly, biomassderived activated carbons are chosen by many industrial players (Li et al. 2018). Raw materials that can be used as activated carbon are all carbonaceous, whether derived from plants, animals, or minerals. The materials are various types of wood, rice husks, animal bones, coal, coconut shell, coffee bean skin, and so forth (Subadra et al. 2005).

The primary function of activated carbon in the industry is as a hazardous material adsorbent (Kurniawan 2004). In addition to adsorbents, activated carbon can be used as decolorizer, deodorizer, water purification, and waste treatment (Smíšek and Černý 1970). The advantage of using activated carbon in wastewater processing is that the activated carbon can reduce water contaminants in a large and absorptive capacity. The disadvantage of activated carbon is the price, which is quite expensive and may not be under the purchasing power of the community, resulting in people using non-active carbon. Non-active carbon is carbon that does not undergo the activation process (Alimsyah and Damayanti 2013). However, activated carbon production is still the primary choice as it requires less technological advances compared to the production of the superadsorptive carbon nanotubes while still maintaining high efficiency in hazardous material removal from wastewater (Li et al. 2020b).

There are three activation methods of preparing activated carbon, namely physical activation, chemical activation, and



physiochemical activation (Sulaiman et al. 2018). In this work, activated carbon was produced through the chemical activation method. The chemical activation by alkalis involves solid–solid or solid–liquid reactions, besides the hydroxide reduction and carbon oxidation to generate porous structures in the carbonized material (Adinata et al. 2007). This method involves precursor carbonization in the presence of alkali hydroxides, within a single- or two-step process. The KOH, NaOH, K_2CO_3 , $ZnCl_2$, $FeCl_3$, H_3PO_4 , and H_2SO_4 are among the reagents used, and the activation process needs a lower temperature than the physical activation (Yang and Qiu 2010).

The carbonization or pyrolysis of the raw material was initially done by heating at temperatures between 300 °C and 500 °C to form the char or carbonized material (Wong et al. 2018). The carbonized sample as the precursor was mixed or soaked in alkaline reagent at pre-determined ratio before dried. Dried mixtures were heated at temperature closed above 400 °C in a closed or inert gas-filled atmosphere for specific period to complete the activation process. The examples of raw materials, with their activating chemicals and activating temperatures, are shown in Table 1. The surface area of the resulting activated carbon varies between 770 and 3300 m²g⁻¹.

In the physical activation method, the formation of pores is mainly due to the removal of carbon atoms by gasification. In contrast, the chemical activation works with dehydrogenation of the precursor by the activating agents to form crosslinks between carbons, which later creates the pores on the surface of activated carbon (Liu et al. 2010). Compared to the physical activation, the chemical activation method will usually produce a higher yield (Shi et al. 2010). The chemical activation can also be conducted at a lower temperature compared to physical activation, which reduces energy usage. Other advantages of chemical activation are lesser activation time, the possibility to incorporate suitable functional groups, and production of larger surface area, which are essential for the adsorption process. However, chemical activation requires activating reagents which increase the production price (Danish and Ahmad 2018).

In this work, the *Leucaena leucocephala* biomass was chemically activated at different activation temperatures. *Leucaena leucocephala* is also locally known as Petai Belalang, a tree that is originated from Central America. The Portuguese and the Dutch brought the species to Asian parts in the seventeenth century (Hou et al. 2015). It is a very fast-growing tree which reaches a height of more than 6 m in only two to three years, yielding high biomass productivity at more than 50 ton ha⁻¹ annually (Loaiza et al. 2017). The *Leucaena leucocephala* can be easily cultivated, with most of them wildly grown by themselves. Currently, this tree was only used for animal feed and for burning, which showed lower value usage of the biomass. There are lacks of

 Table 1
 Activated carbon

 produced by chemical activation

References
(Kim et al. 2005)
(Wang et al. 2006)
(Tran et al. 2017)
(Meng et al. 2005)
(Meng et al. 2005)
(Li and Jaroniec 2001)
(Lee et al. 2006)
(Kim et al. 2004)
(Ryoo et al. 2001)
(Gierszal et al. 2006)
(Yang et al. 2004)
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research done on *Leucaena leucocephala* as activated carbon to date. The abundant availability of *Leucaena leucocephala* could be an advantage as a cheap and continuously available raw material for activated carbon production. Later in this work, the produced activated carbons were subjected to characterization and tested for cadmium adsorption. Langmuir and Freundlich isotherms which were commonly used to study the behavior of adsorption were also applied in this work (Li et al. 2019a).

Experimental

Raw material preparation

Wild *Leucaena leucocephala* biomass was collected around Selangor, Malaysia. The sample was left to air dry to prevent difficulties in the size reduction process to a moisture content of below 10%. Chipped samples were then grounded and sieved into particle size between 1 and 2 μ m. Final moisture content was determined using the oven-dry method before kept in an air-tight container for further use (Rafatullah et al. 2011).

Conversion into activated carbon

Carbonization was done by batch using a pyrolyzer with approximately 6 kg of raw *Leucaena leucocephala* biomass for each batch. The activation process was conducted by chemical means. Sodium hydroxide, NaOH, and carbonized *Leucaena leucocephala* biomass were mixed using impregnation ratio NaOH: char of 3:1, 2:1 and 1:1. Approximately 10 g of carbonized *Leucaena leucocephala* was homogenized with a pre-determined amount of NaOH in 100 mL distilled water. Continuous stirring was done for two h before heated in an oven at 105 °C for 24 h (Cazetta et al. 2011). After 24 h, NaOH-impregnated char was activated inside a muffle furnace using temperatures of 700 °C, 750 °C, and 800 °C for 90 min. Continuous nitrogen gas flow of 150 cm³/min into the furnace was ensured until samples were cooled down to prevent oxidation. All of the prepared samples are tabulated in Table 2.

Adsorbent characterization

Raw and activated carbon samples of *Leucaena leucocephala* were sprinkled on separate aluminum stub stickled with glue before analysis. Samples were observed using scanning electromagnetic imaging, SEM (JSM-IT100 JEOL Co.), at suitable magnifications. The surface area of activated carbon produced was determined using a Micromeritics (ASAP 2010) gas adsorption surface analyzer through nitrogen adsorption isotherm at 77°K. Calculation of surface area was done based on Brunauer–Emmett–Teller equation (BET). Surface functional

Table 2 Prepared activated carbon samples

No.	Sample code	Temperature	NaOH: char ratio
1	C3-700	700	3:1
2	C2-700	700	2:1
3	C1-700	700	1:1
4	C3-750	750	3:1
5	C2-750	750	2:1
6	C1-750	750	1:1
7	C3-800	800	3:1
8	C2-800	800	2:1
9	C1-800	800	1:1



groups of the samples were analyzed using Thermo Scientific Nicolet iS10 FTIR Spectrometer. Samples were grounded into a fine powder before scanned between 4000 and 400 cm⁻¹. Thermogravimetric analysis was done using Mettler Toledo Thermogravimetric Analyzer TGA/DSC 1. The sample was heated at a rate of 10 °C/min with a temperature sweep between 30 °C and 900 °C.

Isothermic studies

A stock solution of cadmium of 1000 ppm was prepared from cadmium nitrate powder. Stock solution was diluted into different concentrations of 10, 15, 20, 25, 30, 33, 35, 40, 45, and 50 ppm. Approximately 50 ml of adsorbate was mixed with 0.5 g activated carbon in different conical flasks. Conical flasks were shacked for 160 min, as previously determined in the batch adsorption study on the effect contact time (Wan Ibrahim et al. 2019). Three temperatures were used, including 30 °C, 40 °C, and 50 °C. Mixtures were filtered with filter paper, and filtered solutions were analyzed using inductively coupled plasma optical emission spectrometry (ICP-OES).

Langmuir and Freundlich's model had been applied to evaluate the adsorption behavior of the adsorbate onto the adsorbent. Langmuir isotherm model suggested ideal monolayer adsorption onto a homogenous surface, calculated using Eq. 1:

$$\frac{1}{q_e} = \frac{1}{q_m} + \frac{1}{K_a q_m C_e} \tag{1}$$

where q_e is the amount of adsorbate adsorbed at equilibrium per gram of sample (mg/g), q_m is the saturated amount of adsorbate adsorbed in mg/g, C_e is the equilibrium concentration of cadmium in mg/l, and Ka is the Langmuir adsorption constant in l/mg. A slope of 1/K_aq_m was obtained from 1/q_e versus 1/C_e plot with an intercept at 1/q_m.

The Freundlich isotherm assumes heterogeneous, and multilayer adsorption occurred on the adsorbent surface (Çetinkaya et al., 2018), calculated using Eq. 2:

$$\ln q_e = \ln K_F + \frac{1}{n} \ln C_e \tag{2}$$

where K_F is Freundlich equilibrium constant, n is an empirical constant and others, as stated in Eq. 1. A slope of (1/n) was derived from a plot of ln qe vs. ln C_e with an intercept of ln K_F .

Results and discussion

Surface morphology analysis

Figure 1 shows the 150x and 550x magnified SEM images of the raw *Leucaena leucocephala*, carbonized sample, and



the activated carbon. Some minor pores were observed on the raw material's surface, while the carbonization process creates more holes and cavities. The carbonized samples possessed very little tiny holes or pores because of the incomplete decomposition of organic constituents that exist in the carbonaceous precursors. The pores were typically blocked by the carbonization product residue (Rahman and Saad 2003). Pores available on the surfaces of the activated carbons prepared from Leucaena leucocephala were well pronounced with distinct pore walls. They were arranged in a group of honeycombed structures. The carbonization product residue, which blocked the pores on the char, was gasified by the activation process. They were carried away with the exhaust gas leaving most of the pores clear and easily absorbable by the cations. At 2000x magnification, the metals can be seen agglomerated inside the pore of the adsorbent (Fig. 2). Because of well-developed porosities formed by the sodium hydroxide, the adsorption capacity of activated carbon and the surface area on the Leucaena leucocephala has increased.

Surface area

BET surface area of prepared activated carbon is shown in Table 3. Using an activation temperature of 800 °C resulted in a higher surface area, which is 185 m²g⁻¹ for ratio 1:1, 595 m²g⁻¹ for ratio 2:1, and 776 m²g⁻¹ for ratio 3:1. The higher surface area was formed using the highest activation temperature. Higher NaOH to carbonized sample ratios also resulted in higher surface area for all activation temperature, which are 662 m²g⁻¹ for 700 °C activation temperature, 735 m²g⁻¹ for 750 °C, and 776 m²g⁻¹ for 800 °C. This trend was similarly recorded from the previous study by Cazetta et al. (2011), which also used NaOH as the activating agent.

Fourier transform spectroscopy, FT-IR analysis

FT-IR analysis provides information about the molecular structure of the functional groups present on the samples under investigation. Figure 3 shows the infrared spectra of all samples. The raw Leucaena leucocephala biomass showed a higher number of peaks. The C-H group was detected at 2895.87 cm⁻¹ and 826.79 cm⁻¹. The C–N groups were detected at the wave number of 2112.37 cm⁻¹ and 1027.53 cm⁻¹. Imines group was detected at the wave number of 3332.59 cm⁻¹, while the urethanes group was detected at the wave number of 1731.58 cm⁻¹. The infrared spectra of activated carbon with an activation temperature of 700 °C, 750 °C, and 800 °C showed alkyne group at the wave number of 2108.77 cm⁻¹. The O-H group was detected at the wave number of 1400.35 cm⁻¹, and the hydrocarbon group was detected at the wave number of 872.29 cm⁻¹. Details of infrared spectra interpretation for



Fig. 1 SEM images of *Leucaena leucocephala* before carbonization \mathbf{a} , after carbonization \mathbf{b} , and after activation \mathbf{c} at magnification 150x as well as activated carbon at magnification 550x \mathbf{d}



Fig. 2 SEM image of activated carbon after the adsorption process at magnification 2000x

activated carbon are shown in Table 4. There was a lesser

 Table 3
 BET surface area of prepared activated carbon

No.	Sample code	BET Surface area (m^2/g^{-1})
1	C1-700	120
2	C2-700	483
3	C3-700	662
4	C1-750	140
5	C2-750	531
6	C3-750	735
7	C1-800	185
8	C2-800	595
9	C3-800	776

number of functional groups found in the activated carbon compared to raw *Leucaena leucocephala*. During the activation stage, the chemical-activating agent broke many bonds comprise aliphatic and aromatic species in the lignocellulosic precursor (Gerçel et al. 2007).





Fig. 3 FT-IR spectra for raw Leucaena leucocephala biomass a, samples with activation temperature 700 °C, b 750 °C, and c 800 °C d

Activation tem- perature, °C	Peak, cm ⁻¹	Assigned group	Details
Raw sample	3332.59	Imines	(=N—N); one band, Amine salts
	2895.87	Alkane	C—H stretching, hydrocarbon chromophore
	2112.37	Isocyanides	C-N stretching, Unsaturated nitrogen compounds
	1731.58	Urethanes	Carbonyl stretching, Amides
	1593.48	Primary	N—H stretching, Amines
	1506.01	Aromatic	C-NO2 nitro compounds, Unsaturated nitrogen compounds
	1421.05	Sulfites	S=O stretching, Sulfur compounds
	1316.50	Sulfonamides	S=O stretching, Sulfur compounds
	1238.03	Nitrates	O-NO ₂ , Unsaturated nitrogen compounds
	1027.53	Aliphatic	C—N vibrations, Amines
	826.79	Alkene, trisubstituted	C—H bending, Hydrocarbon chromophore
700	2108.77	Alkyne, monosubstituted	C—C multiple bond stretching
	2084.04	Isocyanides	C-N stretching vibrations, Unsaturated nitrogen compounds
	1400.35	Miscellaneous chromophoric groups	O—H bending and C–O, Phenols
	872.29	Hydrocarbon chromophore	C—H bending, One hydrogen atom
750	2080.10	Isocyanides	C-N stretching vibrations, Unsaturated nitrogen compounds
	2080.03	Isocyanides	C-N stretching vibrations Unsaturated nitrogen compounds
	1980.49	Allene	C—C multiple bond stretching
800	2084.17	Isocyanides	C-N stretching vibrations Unsaturated nitrogen compounds
	2080.03	Isocyanides	C-N stretching vibrations Unsaturated nitrogen compounds
	2079.35	Isocyanides	C-N stretching vibrations Unsaturated nitrogen compounds
	1980.49	Allene	C—C multiple bond stretching

 Table 4
 Detail interpretation of FT-IR spectra

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Thermal gravimetry analysis, TGA

Generally, there are three stages for thermal gravimetry curves, comprising of the first stage as loss of moisture in the adsorbent, second stage 2 for degassing, and final stage is the residue of the adsorbent. Figure 4 shows the thermal gravimetry curves for activated carbon from Leucaena leucocephala biomass. All of the samples showed an initial decrement of weight at about 10%, which was due to loss in moisture and volatiles. Stage 2 of weight reduction between 200 °C and 500 °C corresponds to the degradation of the polymeric material such as cellulose (Amini et al. 2019). All activated carbon products showed significantly less weight loss compared to the raw biomass, as the lignocellulosic components mostly had been degraded through the pyrolyzing and activation process. Even though the activated carbon made using higher NaOH: biomass ratio has the highest surface area, they are a little less thermally stable compared to the lower ratios at all activation temperatures. For the activation temperature 700 °C, the residue for the ratio 1:1 (C1-700) is 80.58%, for 2:1 (C2-700) is 79.41%, and for the ratio 3:1 (C3-700) is 78.66%. The same goes for the activation temperature of 750 °C and 800 °C. Higher activation temperature also reduces the thermal stability of the activated carbon. Sample from the activation temperature 700 °C resulted in higher residual content, which is 80.58%, while the 750 °C activation temperature results in the residual content of 80.46%. The 800 °C activation temperature resulted in lower 74.93% in the residue. This result was similarly reported from the previous study by Le Van and Luong Thi (2014), which showed increasing activation temperature resulted in lower residual content.

Isothermic studies

Langmuir isotherm

Figure 5 shows Langmuir isotherm plots of 1/Qe versus 1/Ce for all samples at 30 °C, 40 °C, and 50 °C. The R² values for the C1-700 were 0.9925, 0.9506, and 0.9728 at 30 °C, 40 °C, and 50 °C, respectively. The constant q_m, which is a measure of the adsorption capacity to form a monolayer, was found to be as high as 196.078 mg/g at 50 °C. Constant K_I, which denotes adsorption energy, for the corresponding adsorption capacity is between 0.227 and 2.029 l/mg. For C2-700, the R² values are 0.9933, 0.9879, and 0.9686 at 30 °C, 40 °C, and 50 °C, respectively. The highest value for constant q_m, which is a measure of the adsorption capacity to form a monolayer, was 95.238 mg/g at 30 °C. Constant K_I, which denotes adsorption energy, for the corresponding adsorption capacity is between 1.280 and 4.440 l/mg. C3-700 resulted in a correlation coefficient of 0.9700, 0.9325, and 0.9776 at 30 °C, 40 °C, and 50 °C, respectively. The highest constant



Fig. 4 Thermal gravimetry curve for samples produced at 700 °C (a), 750 °C (b), and 800 °C (c) activation temperatures

 q_m was 67.114 mg/g at 30 °C. Constant K_L was between 1.317 and 3.667 l/mg.

The value of the correlation coefficient for C1-750 was 0.9702, 0.9804, and 0.9588 at 30 °C, 40 °C, and 50 °C, respectively. The highest q_m was 89.256 mg/g at 50 °C. Constant K_L is between 0.684 and 1.261 l/mg. C2-750 showed the value of correlation coefficient of 0.9850, 0.9813, and 0.9752 at 30 °C, 40 °C, and 50 °C, respectively. The highest constant q_m is 46.729 mg/g at 50 °C, and the constant K_L was between 3.262 and 9.804 l/mg. C3-750 showed a correlation coefficient of 0.9884, 0.9774,





Fig. 5 Langmuir plot of 1/Qe vs 1/Ce for samples C1-700 (1), C2-700 (2), C3-700 (3), C1-750 (4), C2-750 (5), C3-750 (6), C1-800 (7), C2-800 (8), and C3-800 (9) at 30 °C, 40 °C, and 50 °C

and 0.9892 at 30 °C, 40 °C, and 50 °C, respectively. The highest q_m was 54.645 mg/g at 50 °C, while the constant K_I was between 0.436 and 12.071 l/mg.

The R² values for C1-800 are 0.9627, 0.9609, and 0.9873 at 30 °C, 40 °C, and 50 °C, respectively. The maximum value of constant q_m was 70.423 mg/g at 50 °C, and the constant K_L was between 2.600 and 3.627 l/mg. The R² values for C2-800 are 0.9855, 0.9783, and 0.9707 at 30 °C, 40 °C, and 50 °C, respectively. The constant q_m , which is a measure of the adsorption capacity to form a monolayer, is reported to be as high as 53.191 mg/g at 50 °C. Constant K_L showed values between 4.500 and 8.133 l/mg. For C3-800, the R² values are 0.9671, 0.9655, and 0.9542 at 30 °C, 40 °C, and 50 °C. Constant K_L was valued between 0.589 and 8.231 l/mg.



The plot of linearized equations for Freundlich for the adsorption of cadmium is shown in Fig. 6. The Freundlich model is applicable for non-ideal sorption on heterogeneous surfaces and multilayer sorption processes. The correlation coefficients for the adsorption of cadmium ions for C1-700 were 0.8700, 0.8662, and 0.9454 for 30 °C, 40 °C, and 50 °C, respectively. C2-700 showed 0.9672, 0.9893, and 0.8870, while C3-700 showed 0.7956, 0.8222, and 0.9082 for the correlation coefficient, 30 °C, 40 °C, and 50 °C, respectively.

The values of 1/n were obtained from the slope of the plotted graph ln qe versus ln Ce. The n value lying between 1 and 10 indicates a favorable condition for adsorption. The C1-700 sample showed 1/n values of 1.3812, 1.3320,





Fig. 6 Linear regression analysis of Freundlich isotherm for samples C1-700 (1), C2-700 (2), C3-700 (3), C1-750 (4), C2-750 (5), C3-750 (6), C1-800 (7), C2-800 (8), and C3-800 (9) at 30 °C, 40 °C, and 50 °C

and 2.1336 for 30 °C, 40 °C, and 50 °C, respectively. The C2-700 showed 1/n values of 2.2217, 1.3890, and 1.1202 for 30 °C, 40 °C, and 50 °C, respectively, while C3-700 showed 1.4739, 1.1640, and 1.3659 for 30 °C, 40 °C, and 50 °C, respectively. The value of 1/n for C1-750 was 1.5144, 1.9205, and 1.8936 for 30 °C, 40 °C, and 50 °C, respectively. For C2-750, the values were 1.6614, 1.7862, and 1.8036 for 30 °C, 40 °C, and 50 °C, respectively, while C3-750 showed 1/n values of 1.3186, 1.4263, and 2.0352 for 30 °C, 40 °C, and 50 °C, respectively. For C1-800, the value of 1/n was 1.7987, 1.2863, and 2.3551 for 30 °C, 40 °C, and 50 °C, respectively. The C2-800 recorded 2.1227, 1.5924, and 1.6174 for 30 °C, 40 °C, and 50 °C, respectively, and C3-800 showed 0.9878, 1.0455, and 2.3296 for 30 °C, 40 °C, and °C, respectively. All of the samples showed favorable condition for adsorption except for C3-800 at 30 °C. However, the value is very closed to a favorable region at 0.9878.

Table 5 shows the different R^2 values for both isotherms, Langmuir and Freundlich, for the adsorption of cadmium.

The R^2 value for samples with an activation temperature of 700 °C shows that the Langmuir model has a value of correlation coefficient close to 1 and more fit to the Langmuir isotherm instead of Freundlich. All of the activated carbon that is produced from *Leucaena leucocephala* biomass are fit to the Langmuir isotherm for all activation temperature. This concludes that the activated carbon produced is doing monolayer cadmium adsorption onto the surface (Sulaiman et al. 2010).

Thermodynamic analysis

The adsorption characteristic of cadmium ions onto chemical activated carbon made from *Leucaena leucocephala* biomass had been studied in the thermodynamic parameters. Three parameters were studied which were the ΔG° as Gibb's free energy change, kJ/mole, ΔH° as enthalpy change, J/mol, and ΔS° as entropy change, J/K/mol. The linearized equation is as in Eqs. 3, 4;



$$G^{o} = -RT\ln\left(K_{L}\right) \tag{3}$$

$$G^o = H^o - TS^o \tag{4}$$

The R is 8.314 J/K/mol, which is universal gas constant, and T value is the temperature in the Kelvin scale. K_L was described as a value of Langmuir constant. Figure 7 presents the slopes of the linear lines, which were either positive or negative depending on the adsorbate–adsorbent systems. It indicates the adsorption process, which is endothermic in nature or exothermic. The calculated value of ΔH° , ΔS° , and ΔG° for sorption of cadmium onto *Leucaena leucocephala* activated carbon is listed in Table 6.

The negative values for the Gibbs free energy change, ΔG , indicate that the adsorption process for the cadmium was feasible and spontaneous. The increasing in ΔG values with increase in temperature showed the favorable adsorption was at higher temperatures. The result revealed the ΔH^{o} values obtained for the adsorption of cadmium onto Leucaena leucocephala activated carbon is positive for the C1-700. C2-700 and C3-700 show the negative values of ΔH^{o} . The positive value of ΔH^{o} indicates the endothermic nature of adsorbent, whereas the negative value of ΔH° reflects the exothermic nature of the adsorbent interactions. The adsorptions of cadmium ions on all of the produced adsorbents are endothermic. This result was consistent for activated carbon made using an activation temperature of 750 °C and 800 °C. A rise in temperature would increase the rate of diffusion of the cadmium ions across the external boundary layer into the pores of the adsorbent particle, due to the decrement of the viscosity of the solution. Other possibilities are the amplification of pore size distribution and increment of the active surface sites with the increase in temperature. Most of the samples showed positive values of ΔS , showing the increment of an orderliness between the adsorbate and the adsorbent molecules (Koyuncu and Kul 2019).



Fig. 7 Thermodynamic graph for samples C1-700 (1), C2-700 (2), C3-700 (3), C1-750 (4), C2-750 (5), C3-750 (6), C1-800 (7), C2-800 (8), and C3-800 (9)

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Table 5 Langmuir and Freundlich isotherm models for the adsorption of cadmium at all activation temperatures

Sample	Tempera-	Langmuir Isotherm			Freundlich Isoth	erm	
	ture, °C	The maximum monolayer adsorption capacity	Langmuir constant	Correlation Coefficient	Affinity factor	Freundlich exponent	Correlation Coefficient
		$q_{max} (mg/g)$	K _L (l/mg)	\mathbf{R}^2	K _F	l/n	\mathbb{R}^2
C1-700	30	35.714	2.029	0.9925	3.6621	1.3812	0.8700
	40	41.667	1.086	0.9506	3.3951	1.3320	0.8662
	50	196.078	0.227	0.9728	4.0562	2.1336	0.9454
C2-700	30	95.238	1.280	0.9933	4.9962	2.2217	0.9672
	40	23.810	2.877	0.9879	3.3954	1.3890	0.9893
	50	24.752	4.440	0.9686	3.5172	1.1202	0.8870
C3-700	30	67.114	3.239	0.9700	4.6100	1.4739	0.7956
	40	37.037	1.317	0.9325	3.3128	1.1640	0.8222
	50	37.879	3.667	0.9776	4.0386	1.3659	0.9082
C1-750	30	71.429	1.261	0.9702	4.1160	1.5144	0.8520
	40	76.293	0.684	0.9804	3.9681	1.9205	0.9435
	50	89.256	0.982	0.9588	4.3979	1.8936	0.9285
C2-750	30	27.548	9.804	0.9850	4.7149	1.6614	0.9944
	40	36.496	3.262	0.9813	4.3202	1.7862	0.9599
	50	46.729	3.397	0.9752	4.6583	1.8036	0.9287
C3-750	30	29.586	12.071	0.9884	4.4273	1.3186	0.9521
	40	29.851	0.893	0.9774	2.9684	1.4263	0.9548
	50	54.645	0.436	0.9892	3.2605	2.0352	0.9804
C1-800	30	54.054	3.627	0.9627	4.8448	1.7987	0.9401
	40	64.013	2.600	0.9609	4.2177	1.2863	0.7513
	50	70.423	3.302	0.9873	5.7397	2.3551	0.9701
C2-800	30	40.984	8.133	0.9855	5.6249	2.1227	0.9703
	40	50.505	4.500	0.9783	4.7040	1.5924	0.9000
	50	53.191	5.371	0.9707	4.8970	1.6174	0.8874
C3-800	30	31.153	8.231	0.9671	3.8763	0.9878	0.8265
	40	31.447	5.483	0.9655	3.8045	1.0455	0.7990
	50	188.67	0.589	0.9542	5.1123	2.3296	0.9003

Conclusion

The *Leucaena leucocephala* biomass had been converted into activated carbon by chemical mean. Analysis of BET found out that higher surface area can be obtained using the higher activation temperature. Another finding from this work suggests that higher NaOH to carbonized sample ratios will create a larger surface area for all activation temperatures, which are 662 m²g⁻¹ for 700 °C activation temperature, 735 m²g⁻¹ for 750 °C, and 776 m²g⁻¹ for 800 °C. The FT-IR analysis detected a lesser number of functional groups in the activated carbon compared to raw *Leucaena leucocephala* due to deterioration effect of the heating process. The thermogravimetry analysis results concluded that even though the activated carbon made using higher NaOH: biomass ratio has the highest surface area, they are a little less thermally stable compared to the lower ratios at all activation temperatures. The activated carbon that is produced from *Leucaena leucocephala* biomass is fit to the Langmuir isotherm, regardless of any activation temperature, showing that the activated carbon produced is doing monolayer cadmium adsorption onto the surface. Also, the thermodynamic study revealed that the adsorption process is endothermic, reflected by the positive value of ΔH° .



 Table 6
 Thermodynamic parameters of cadmium adsorption for all samples

Sample (°C)	Tempera- ture (K)	ΔG^{o} (kJ/mole)	ΔH^{o} (kJ/mole)	$\Delta S^{o} (kJ/mol/K)$
C1-700	303	-9.007	68.656	0.2544
	313	-9.706		
	323	-14.175		
C2-700	303	-11.478	-55.419	-0.1468
	313	-8.249		
	323	-8.617		
C3-700	303	-10.597	-23.530	-0.0434
	313	-9.399		
	323	-9.759		
C1-750	303	-10.754	15.674	0.0651
	313	-11.279		
	323	-12.062		
C2-750	303	-8.353	21.507	0.0986
	313	-9.361		
	323	-10.324		
C3-750	303	-8.533	24.696	0.1088
	313	-8.838		
	323	-10.744		
C1-800	303	-10.051	10.791	0.0689
	313	-10.823		
	323	-11.425		
C2-800	303	-9.354	10.671	0.0662
	313	-10.206		
	323	-10.672		
C3-800	303	-8.663	72.479	0.2653
	313	-8.973		
	323	-14.072		

Acknowledgement The authors acknowledged the Ministry of Education, Malaysia, for MyMaster scholarship to Wan Muhammad Hilmi bin Wan Ibrahim and Universiti Malaysia Kelantan for Short Term Grant (R/SGJP/A08.00/01046A/001/2015/000242) awarded to Mohd Hazim Mohamad Amini.

Financial Disclosure statement The author(s) received no specific funding for this work

Compliance with ethical standards

Conflict of interest The authors declared no conflict of interest and are mutually agreed on the publication of this work.

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