

The use of variations in proteomes to predict, prevent, and personalize treatment for clinically nonfunctional pituitary adenomas

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Abstract Pituitary adenomas account for ~10% of intracranial tumors, and they cause the compression of nearby structures and the inappropriate expression of pituitary hormones. Unlike functional pituitary adenomas, nonfunc-

tional (NF) pituitary adenomas account for ~30% of pituitary tumors, and are large enough to cause blindness; because they do not cause any clinical hormone hypersecretion, they are difficult to detect at an early stage; and hypopituitarism results. No effective molecular biomarkers or chemical therapy have been approved for the clinical setting. Because an NF pituitary adenoma is highly heterogeneous, differences in the proteins (the proteome) can distinguish among those heterogeneity structures. The components of a proteome dynamically change as an NF adenoma progresses. Changes in protein expression and protein modifications, individually or in combination, might be biomarkers to predict the disease, monitor the tumor progression, and develop an accurate molecular classification for personalized patient treatment. The modalities of proteomic variation might also be useful in the interventional prevention and personalized treatment of patients to halt the occurrence and progression of NF pituitary adenomas.

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Keywords Clinical nonfunctional pituitary adenoma · Proteomic variation · Predictive diagnosis · Tumor progression monitoring · Interventional prevention · Personalized patient treatment

Abbreviations

ACTH	adrenocorticotrophic hormone
CDK	cyclin-dependent kinase
CDKI	cyclin-dependent kinase inhibitor
CSF	cerebrospinal fluid
DEP	differentially expressed protein
2D DIGE	two-dimensional differential in-gel electrophoresis
2DGE	two-dimensional gel electrophoresis
ERK	extracellular signal-regulated kinase

ESI	electrospray ionization
FSH	follicle-stimulating hormone, or follitropin
GH	growth hormone
IL1	interleukin 1
IPA	Ingenuity Pathway Analysis
JNK	c-Jun N-terminal kinase
LCM	laser-capture microdissection
LH	luteinizing hormone, or lutropin
MALDI	matrix-assisted laser desorption/ionization
MAPK	mitogen-activated protein kinase
MAPKK	MAPK kinase
MAPKKK	MAPKK kinase
M_r	relative molecular mass
MS	mass spectrometry
MS/MS	tandem mass spectrometry
MW	molecular weight
NF	nonfunctional
Nrf2	nuclear factor E2-related factor-2
NTAC	nitrotyrosine affinity column
pI	isoelectric point
PKA	cyclic AMP-dependent kinase
PMF	peptide mass fingerprint
PRL	prolactin
PTM	post-translational modification
RKIP	raf kinase inhibitory protein
RNS	reactive nitrogen species
ROS	reactive oxygen species
SELDI	surface-enhanced laser desorption/ionization
TNF	tumor necrosis factor
TOF	time-of-flight
TSH	thyrotropin-stimulating hormone, or thyrotropin

Introduction

Pituitary adenomas are a common pathological change that account for ~10% of intracranial tumors [1]. They cause significant morbidity through a compression of regional structures and an inappropriate expression of pituitary hormones that disturb the multiple hypothalamic-pituitary-target organ axis systems. Pituitary adenomas are clinically classified as functional (~70%; such as acromegaly and prolactinoma) and nonfunctional (~30%). Unlike functional pituitary adenomas, nonfunctional (NF) pituitary adenomas are typically quite large, do not cause any clinical hormone hypersecretion, and result in hypopituitarism or blindness from regional compression. Moreover, an NF pituitary adenoma is highly heterogeneous in the cell of origin [1]. Therefore, an early-stage diagnosis of NF pituitary adenomas is difficult because there is no hormone hypersecretion and there is no effective chemical therapy for NF pituitary

adenomas. Predictive diagnosis/prevention and early-stage diagnosis/clinical therapy are the crucial aspects needed to conquer a tumor because most of NF pituitary adenomas remain as late-stage diagnosed and neurosurgically/radiation treated. Modern proteomics methods provide insight into the proteome, which is a dynamically changing protein-network system that progresses with disease. Study of the proteomic variations will result in new concepts and models for the more-accurate prediction, prevention, and personalized patient therapy for NF pituitary adenomas. This review article addresses the contribution of tissue-proteomic variations to the interventional prevention and personalized patient treatment, and the contribution of body-fluid proteomic variations to the predictive diagnosis and the assessment of preventive and therapeutic responses.

Pathophysiological basis for the variation of the proteome in an NF pituitary adenoma

Pituitary adenomas are monoclonal in the cell of origin [2, 3], and might arise from any one of five pituitary anterior cell types (Fig. 1) [4, 5]—including corticotrophs [secrete adrenocorticotrophic hormone (ACTH)], somatotrophs [secrete growth hormone (GH)], lactotrophs [secrete prolactin (PRL)], thyrotrophs [secrete thyrotropin-stimulating hormone, or thyrotropin (TSH)], and gonadotrophs [secrete follicle-stimulating hormone, or follitropin (FSH) and luteinizing hormone, or lutropin (LH)]—to produce an imbalance in hormone levels that will disturb the multiple regulatory hypothalamic-anterior pituitary-target organ axes. Each axis performs a range of important physiologic functions in the human body (Fig. 1). Mixed-tumors (co-secrete GH with PRL, TSH, or ACTH) might also arise from single cells. ACTH oversecretion results in Cushing's disease, with features of hypercortisolism; GH hypersecretion leads to acral overgrowth and metabolic dysfunction associated with acromegaly; and PRL oversecretion leads to gonadal failure, secondary infertility, and galactorrhea. More rarely, TSH hypersecretion leads to hyperthyroxinemia and goiter, and hypersecreted GH (or GH subunits) leads to gonadal dysfunction. In contrast, tumors that arise from gonadotroph cells do not efficiently secrete their gene products, and they are usually clinically silent [4]. They are commonly referred to as NF pituitary adenomas, which cannot be diagnosed early because no hormone change is observed. However, NF pituitary adenomas are highly heterogeneous in the cell of origin (Table 1) [1]. Except for a gonadotroph-originated intact LH/FSH adenoma (up to 79% of NF adenomas), immunocytochemical staining of hormones also reveals that some NF pituitary adenomas might arise from a silent corticotroph (ACTH-positive; 8%), silent somatotroph (GH-posi-

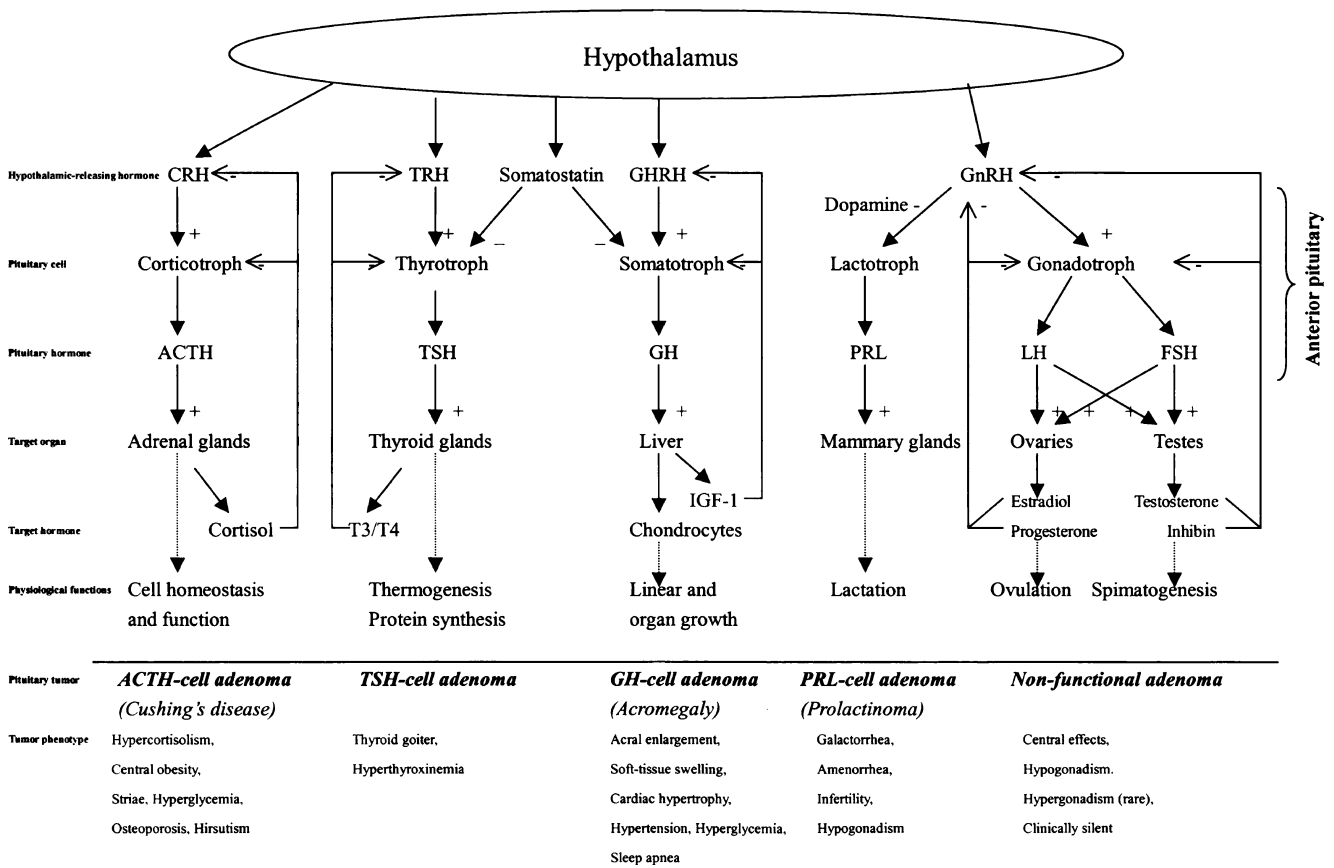


Fig. 1 Scheme of the hypothalamic-anterior pituitary-target organ axis systems and pituitary adenoma pathogenesis [4]. (+)=stimulatory regulation; (-)=inhibitory regulation. Reproduced from Zhan and

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tive; 3%), no hormone-expressed oncocytoma (6%), and null cell (17%).

Those heterogeneous cells of origin of NF pituitary adenomas produce differences in a proteome relative to the control and among different NF pituitary adenoma patients, and that difference in the proteome dynamically changes with tumor progression. The proteomic difference among different types of NF pituitary adenomas, and the dynamic

alteration with tumor progression, both contribute to the prediction, interventional prevention, and personalized patient treatment of NF pituitary adenoma patients.

Targeted organs of proteomic variation that contribute to the prediction, prevention, and personalized treatment of an NF pituitary adenoma

Table 1 Classification of nonfunctional pituitary adenomas by cell of origin

Cell type	Hormone expression	% Nonfunctional tumors
Gonadotrophs	Intact LH/FSH or subunits	40–79
Null cell	None	17
Oncocytoma	None	6
Silent corticotroph	ACTH	8
Silent somatotroph	GH	3

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The pituitary is the most-protected organ in the body, and is the master regulatory gland; several different hypothalamic-pituitary-target organ axes exist. Any metabolic defect in those regulatory systems has an associated pathology. Based on the new concept of prediction, prevention, and personalized patient treatment of an NF pituitary adenoma, pituitary tissues, cerebrospinal fluid (CSF), and blood plasma must be analyzed. Any tissue-proteome variation directly reflects the pathophysiological changes of an NF pituitary adenoma. Those tissue-proteome variations, and the corresponding changes in the biological system, can be developed into an accurate molecular classification, interventional prevention, and personalized patient treatment. In

addition, in an NF pituitary adenoma patient, some secreted proteins and peptides will enter into the CSF and blood circulation. Comparative proteomics can detect those different components (proteins or peptides) in the patient's CSF and blood plasma. Those CSF and blood plasma proteomic and peptidomic variations will lead to the development of an accurate predictive diagnosis and the measurement of interventional prevention and therapy response based on accurate molecular data.

Pituitary tumor tissues are generally acquired with neurosurgery from patients, whereas control tissues are generally obtained via forensic anatomy from post-mortem cadavers. Therefore, there is some uncontrollable bias among pituitary adenoma tissues and “so-called” control tissue proteome such as sex, age, race, and time of tissue acquisition; and some biochemical changes might occur in post-mortem tissue compared to surgical tissue. Also, an NF pituitary adenoma is monoclonal in origin, and is generally benign. An NF pituitary adenoma tissue contains a relatively pure cell population, whereas control post-mortem pituitary tissues contain different types of cells, including at least corticotrophs, somatotrophs, lactotrophs, thyrotrophs, and gonadotrophs, etc. When the tumor tissues and post-mortem tissues are used to analyze the proteomic variation, another bias is possible. The use of laser-capture microdissection (LCM) to preferentially enrich a specific tumor cell type from tumor tissues and the corresponding normal cells from post-mortem tissues overcomes this bias. Compared to a tissue proteome, those limitations do not exist for the CSF and blood specimens. Also, CSF and blood specimens are much more accessible from patients and controls than the pituitary tissues.

Techniques to measure proteomic variation

Modern proteomics uses protein-separation techniques such as gel and non-gel methods, protein-identification techniques such as mass spectrometry (MS), multiple protein-preparation techniques, and systems biology techniques to quantitatively measure proteomic variation in tissue, CSF, and blood plasma samples from patients and controls.

Gel methods include two-dimensional gel electrophoresis (2DGE) and 2D differential in-gel electrophoresis (2D DIGE). 2DGE-based comparative proteomics is a classical, reliable method to quantify accurately any proteomic variation. Briefly, proteins are arrayed with 2DGE according to their pI and M_r , and are stained. Each 2D gel spot-volume reflects the abundance of the protein contained in that spot. Each individual gel spot-volume is generally normalized with the total spot-volumes of each gel image [6, 7]. 2DGE images from tumors and controls are compared with 2D gel-analysis software to obtain statisti-

cally significant differential 2D gel spot-volume between tumors and controls. The amino acid sequence of a protein in each 2D gel spot is obtained with MS, including peptide mass fingerprint (PMF) (which determines the MW of each tryptic peptide from that protein) and/or tandem mass spectrometry (MS/MS) (which determines the amino acid sequence of each tryptic peptide), and corresponding protein database analysis [8].

2D DIGE is another gel method to quantify the proteomic variation [9–11]. Generally, the tumor and control samples are labeled with different fluorescent dyes such as CyDye 3 and CyDye 5. The CyDye-labeled samples are mixed equally (1:1), and the mixed sample is separated with 2DGE according to pI and M_r . The proteins are visualized, and the image is scanned with a CyDye 3 channel and CyDye 5 channel to form two fluorescent images, which are processed with DeCyder DIGE analysis software or Delta 2D software. Any color difference in a gel spot defines and quantifies the protein content between tumors and controls. The amino acid sequence of tryptic peptides from each protein in each gel spot is determined with MS/MS. 2D DIGE is more useful to analyze precious LCM-enriched samples because 2D DIGE requires less protein amount and fewer gel-analysis procedures relative to the traditional 2DGE-based comparative proteomics.

Non-gel methods mainly include the analysis of trypsin-digests from a complex mixture of proteins with multidimensional chromatography coupled on-line with MS [12, 13]. That strategy generates tryptic peptides that are separated in the first dimension with any one of several different chromatographies, and with reversed-phase chromatography in the second dimension. The separated peptides are analyzed with MS/MS, and the amino acid sequences are analyzed with a database analysis for protein identification. The technique has been used to analyze some human tissue proteomes [14]. For quantification of proteomic variation, that technique must be coupled with stable-isotope labeling prior to multidimensional-chromatography analysis [15]. The light and heavy isotope-labeled peptides were quantified with MS/MS to quantify and identify the protein amount between tumor and control proteomes. A wide variety of stable isotope-labeling techniques have been developed; for example, cysteine-specific tagging such as ICAT [16], lysine-specific tagging [17, 18], and amine-specific tagging such as iTRAQ [19–21]. Moreover, some software has been developed to quantitatively analyze the stable isotope-labeled peptides; for example, XPRESS software for ICAT-labeled peptides. Non-gel methods effectively detect low-abundance proteins, extremely acidic/basic proteins, and low/high- M_r proteins. Although the gel methods are limited by the extremely acidic ($pI < 3.5$ or 4)/basic ($pI > 7.5$ or 8) proteins, extremely high-mass (> 150 kDa) or low-mass (< 10 kDa) proteins, and hydro-

phobic proteins [5], the non-gel techniques are complementary with the gel-based methods, and improve the proteomic coverage of an NF pituitary adenoma proteome.

Systems biology techniques such as pathway analysis help to elucidate each protein variation within a pathway network system. The development of bioinformatics and computation biology underpins systems biology developments. A wide-range of pathway analysis programs has been developed such as Ingenuity Pathway Analysis (IPA) (www.ingenuity.com) and MetaCore (www.genego.com/metacore.php). Pathway analysis has been used to mine significant pathway networks from pituitary adenoma tissue comparative proteomic data and proteomic modification data [22]. The recovery of the tumor-altered pathway system to the normal pathway system would contribute significantly to interventional prevention and effective chemical therapy. Those proteomic variations include variations in protein expression and post-translational modifications (PTMs).

Tissue proteomic variations that contribute to accurate molecular classification, interventional prevention, and personalized patient treatment

A tissue proteome is a systematic and tightly regulated protein-network system that dynamically changes during pathophysiological processes. Multiple endogenous and exogenous factors cause an alteration of certain elements within a proteome relative to controls. Those altered elements include protein differential expression (DEPs), splicing isoforms (such as hormone isoforms), and protein modifications (such as tyrosine nitration and phosphorylation), etc.

A total of 56 DEPs has been identified among NF adenomas and controls with 2DGE comparative proteomics strategies (Table 2) [1]. A large-scale comparative proteomics study was performed on a set of human pituitary samples [1, 5, 23]: controls ($n=8$) versus several different cell-types of NF pituitary adenomas (NF-, $n=3$; LH+, $n=3$; FSH+, $n=3$; FSH+/LH+, $n=3$) and hyperprolactinomas ($n=4$). A total of 93 differential protein-spots (65 decreased spot volumes, 28 increased) were found in NF pituitary adenomas and prolactinomas. Seventy-two spots (50 decreased, 22 increased) that represented 56 DEP's (34 down-regulated, 22 up-regulated) were characterized with MS and database analysis. Functional roles are summarized in Fig. 2. Results indicated that: (i) neuroendocrine-related proteins, including somatotropin, secretagogin, and mu-crystallin homolog, were down-regulated in NF pituitary adenomas and the hyperprolactinoma; (ii) somatotropin existed in at least 17 isoforms that were down-regulated in NF adenomas and the hyperprolactinomas; (iii) prolactin

existed in six isoforms that were down-regulated in NF adenomas, and no-change in the hyperprolactinomas; (iv) immunologic regulation proteins and tumor-related antigen (for example, immunoglobulin, tumor-rejection antigen 1) were down-regulated in NF adenomas; (v) cell proliferation, differentiation, and apoptosis-related proteins were down-regulated in the NF-adenomas and the hyperprolactinomas; (vi) some cell-defense and stress-resistance proteins (for example, phospholipid hydroperoxide glutathione peroxidase, CD59 glycoprotein, and heat-shock 27 kDa protein) were down-regulated in the NF pituitary adenomas; (vii) some metabolic enzyme-related proteins (for example, cytoplasmic isocitrate dehydrogenase [NADP], tryptophan 5-hydroxylase 2, matrix metalloproteinase-9, aldose reductase, lactoylglutathione lyase, acyl-CoA-binding protein, etc.) were up-regulated in the NF pituitary adenomas; and (viii) for cell-signal proteins, some were down-regulated and some were up-regulated in NF adenomas; those cell signals are involved in the complex biological roles in the cell growth, proliferation, differentiation, apoptosis, and death cycles.

The heterogeneity of hormone expression is an important characteristic in an NF pituitary adenoma. Protein isoforms result from protein PTMs, splicing, etc. A comparative proteomics study of NF adenomas demonstrated that two hormones (GH and PRL) had multiple isoforms [1, 23]. Twenty-four 2D gel spots that contained GH were found in control pituitary tissues [24] (Fig. 3). Those hGHs in the 24 2D gel spots were classified into four types of hGH-splicing isoforms, 1–4. The expression proportion of those four isoforms were isoform 1 (87.5 %) > isoform 2 (8.1 %) > isoform 3 (3.3 %) > isoform 4 (1.1 %) (Fig. 4); and a significant statistical difference was found among those isoforms. Not all of isoforms related to NF pituitary adenomas [1]. Somatotropin was significantly down-regulated at the protein and the mRNA levels in the NF pituitary adenoma [1] and in prolactinomas [23]; that finding is consistent with their monoclonal origin [2, 3]—an NF adenoma originated from gonadotrophs, and a prolactinoma from lactotrophs; the GH receptor gene was unchanged. Those data showed that GH hyposecretion in an NF-adenoma results from the hypo-expression of the GH gene. However, the more important finding is that the multiple GH isoforms (17 spots contain somatotropin) were detected [1, 23]; those data cannot be interpreted from transcriptomics data. The down-regulated ratio of the different GH isoforms was different in each cell-type of NF pituitary adenoma relative to the controls. Those data suggested that the proportion of the different GH isoforms changed in each cell-type NF adenoma compared to controls. Other researchers showed that the proportion of a circulating GH isoform significantly changed in pituitary adenomas and other pituitary diseases [25, 26]. The

Table 2 Differentially expressed proteins identified in human nonfunctional pituitary adenoma tissues with 2DGE-based comparative proteomics

ID	Molecules	Fold-change	Description	Location	Function
P49753	ACOT2	5.8	acyl-CoA thioesterase 2	C	enzyme
P15121	AKR1B1	14.6	aldo-keto reductase family 1B1	C	enzyme
P02647	APOA1	-3.2	apolipoprotein A-I	EC	transporter
P06576	ATP5B	5.0	ATP synthase, H ⁺ -transporting, mitochondrial F1 complex, beta polypeptide	C	transporter
P47756	CAPZB	6.5	capping protein (actin filament) muscle Z-line, beta	C	other
CAPZB	CAPZB	6.5	capping protein (actin filament) muscle Z-line, beta	C	other
P13987	CD59	-9.5	CD59 molecule, complement regulatory protein	PM	other
P12110	COL6A2	-14.7	collagen, type VI, alpha 2	EC	other
P14854	COX6B1	9.3	cytochrome c oxidase subunit Vib polypeptide 1 (ubiquitous)	C	enzyme
Q14894	CRYM	-35.3	crystallin, mu	C	enzyme
CSN1S1	CSN1S1 (includes EG:1446)	-17.8	casein alpha s1	EC	other
P07108	DBI	4.3	acyl-Coenzyme A binding protein	C	other
Q9UHL4	DPP7	-9.3	dipeptidyl-peptidase 7	C	peptidase
Q14259	ERH	5.0	enhancer of rudimentary homolog	N	other
P30040	ERP29	4.6	endoplasmic reticulum protein 29	C	transporter
P00742	F10	-83.0	coagulation factor X	EC	peptidase
P06241	FYN	3.9	FYN oncogene related to SRC, FGR, YES	PM	kinase
P31150	GDI1	9.4	GDP dissociation inhibitor 1	C	other
P01241	GH1	-180.6	growth hormone 1	EC	cytokine
P01241	GH1	-13.3	growth hormone 1	EC	cytokine
P01241	GH1	-13.3	growth hormone 1	EC	cytokine
P01241	GH1	-20.2	growth hormone 1	EC	cytokine
P01241	GH1	-14.1	growth hormone 1	EC	cytokine
P01241	GH1	-128.3	growth hormone 1	EC	cytokine
P01241	GH1	-14.8	growth hormone 1	EC	cytokine
P01241	GH1	-48.1	growth hormone 1	EC	cytokine
P01241	GH1	-17.3	growth hormone 1	EC	cytokine
P01241	GH1	-16.9	growth hormone 1	EC	cytokine
P01241	GH1	-88.8	growth hormone 1	EC	cytokine
P01241	GH1	-27.8	growth hormone 1	EC	cytokine
P01241	GH1	-51.2	growth hormone 1	EC	cytokine
P01241	GH1	-32.1	growth hormone 1	EC	cytokine
P01241	GH1	-478.6	growth hormone 1	EC	cytokine
P01241	GH1	-13.4	growth hormone 1	EC	cytokine
P01241	GH1	-161.0	growth hormone 1	EC	cytokine
P01241	GH1	-37.0	growth hormone 1	EC	cytokine
P01242	GH2	-14.1	growth hormone 2	EC	other
Q04760	GLO1	10.2	glyoxalase I	C	enzyme
Q04760	GLO1	8.3	glyoxalase I	C	enzyme
P09471	GNAO1	9.5	G protein, alpha activating activity polypeptide O	PM	enzyme
P29777	GNAO1	9.5	G protein, alpha activating activity polypeptide O	PM	enzyme
P36969	GPX4	-26.1	glutathione peroxidase 4	C	enzyme
P28161	GSTM2	4.2	glutathione S-transferase M2 (muscle)	C	enzyme
HBB	HBB (includes EG:3043)	-3.1	hemoglobin, beta	C	transporter

Table 2 (continued)

ID	Molecules	Fold-change	Description	Location	Function
HBB	HBB (includes EG:3043)	-99.4	hemoglobin, beta	C	transporter
P02081	HBD	-3.1	hemoglobin, delta	C	transporter
P14625	HSP90B1	-11.0	HSP90 beta (Grp94), member 1	C	other
P04792	HSPB1	-5.1	heat shock 27 kDa protein 1	C	other
Q9UJY1	HSPB8	-3.7	heat shock 22 kDa protein 8	C	kinase
O75874	IDH1	8.7	isocitrate dehydrogenase 1 (NADP+), soluble	C	enzyme
P24592	IGFBP6	-20.9	insulin-like growth factor binding protein 6	EC	other
IGLC1	IGLC1	-32.1	immunoglobulin lambda constant 1	C	other
IGLC3	IGLC3	-32.1	immunoglobulin lambda constant 3	NA	other
P08779	KRT16	-28.8	keratin 16 (focal non-epidermolytic palmoplantar keratoderma)	C	other
Q86U44	METTL3	-8.3	methyltransferase like 3	N	enzyme
Q99542	MMP19	3.1	matrix metalloproteinase 19	EC	peptidase
P12524	MYCL1	-17.8	v-myc myelocytomatosis viral oncogene homolog 1, lung carcinoma derived (avian)	N	transcription regulator
O00217	NDUFS8	5.2	NADH-coenzyme Q reductase	C	enzyme
P20774	OGN	-38.0	osteoglycin	EC	growth factor
Q08752	PPID	5.1	peptidylprolyl isomerase D (cyclophilin D)	C	enzyme
Q00007	PPP2R2A	-8.2	protein phosphatase 2 (formerly 2A), regulatory subunit B, alpha isoform	C	phosphatase
P01236	PRL	-99.9	prolactin	EC	cytokine
P01236	PRL	-9.7	prolactin	EC	cytokine
P01236	PRL	-26.2	prolactin	EC	cytokine
P01236	PRL	-20.1	prolactin	EC	cytokine
P01236	PRL	-36.7	prolactin	EC	cytokine
P01236	PRL	-33.6	prolactin	EC	cytokine
O76038	SCGN	-6.6	secretagogin	C	other
Q9UI15	TAGLN3	5.6	transgelin 3	NA	other
P21980	TGM2	-17.1	transglutaminase 2	C	enzyme
Q8IWU9	TPH2	10.6	tryptophan hydroxylase 2	NA	enzyme
O14530	TXNDC9	-11.4	thioredoxin domain containing 9	NA	other
P08670	VIM	5.5	vimentin	C	other
P27348	YWHAQ (includes EG:10971)	-43.5	tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein, theta polypeptide (14-3-3 protein tau)	C	other
Q14584	ZNF266	7.3	zinc finger protein 266	N	other
IL15-S21AA	P40933-2	-2.7	Splice isoform IL15-S21AA of interleukin-15		
P01620		-32.1	Ig kappa chain V-III region SIE		
IGLC2	P01842	-32.1	Ig lambda chain C regions		
P01621		-32.1	Ig Kappa chain V-III region NG9		
P04433		-32.1	Ig kappa chain V-III region VG		
HBB2	P18988	-3.1	Hemoglobin beta-2 chain (PANLE)		
gi 1066765		-176.7	Hemoglobin beta unit variant		
HBB2	P18988	-99.4	Hemoglobin beta-2 chain (PANLE)		
P01935		-99.4	Hemoglobin alpha-3 chain (PANTR)		
P01968		-20.2	Hemoglobin alpha-2 chain (BOSMU)		
HBB2	P18988	-20.2	Hemoglobin beta-2 chain (PANLE)		

C, cytoplasm; EC, extracellular space; PM, plasma membrane; N, Nucleus; NA, unknown. Modified from Zhan and Desiderio [1, 22], with permission from AACR, copyright 2005; and with permission from BioMed Central open access journal, copyright remains with the authors.

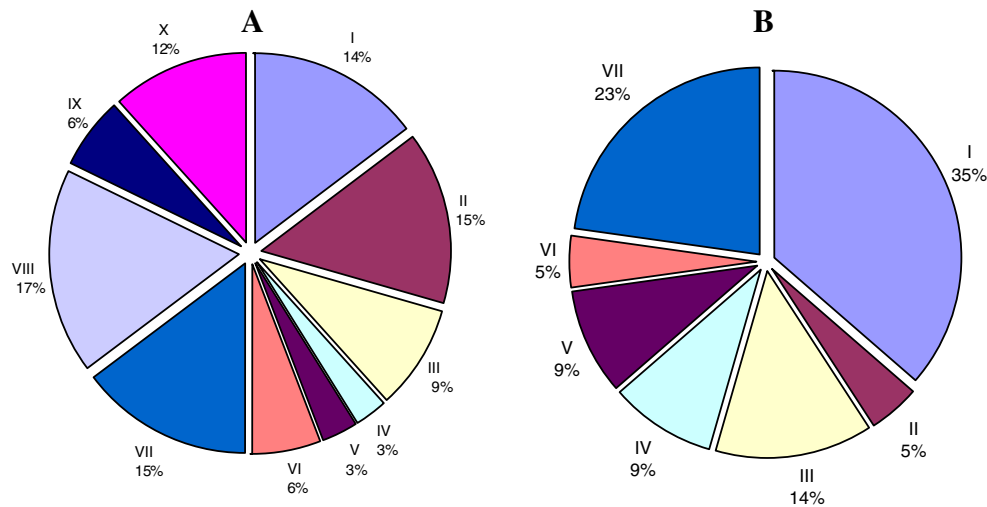


Fig. 2 Functional categories of the 56 DEPs. **A** Down-regulated proteins in pituitary adenoma (34): I. Neuro-endocrine and hormones; II. Cytokine and cellular signal-related proteins; III. Cellular defense and stress resistance; IV. mRNA splicing, transport, or translation-related enzymes; V. DNA-binding proteins; VI. Metabolic enzymes; VII. Immunologic regulation proteins and tumor-related antigen; VIII. Transport proteins; IX. Cell proliferation, differentiation, and

apoptosis-related proteins; X. Others. **B** Up-regulated proteins in pituitary adenomas (22): I. Metabolic enzyme-related proteins; II. Energy metabolism; III. Cellular signal proteins; IV. Cell cycle, cell growth and proliferation proteins; V. Cellular defense response; VI. Protein folding-related protein; and VII. Others. Reproduced from Zhan and Desiderio [5], with permission from Wiley-VCH, copyright 2005

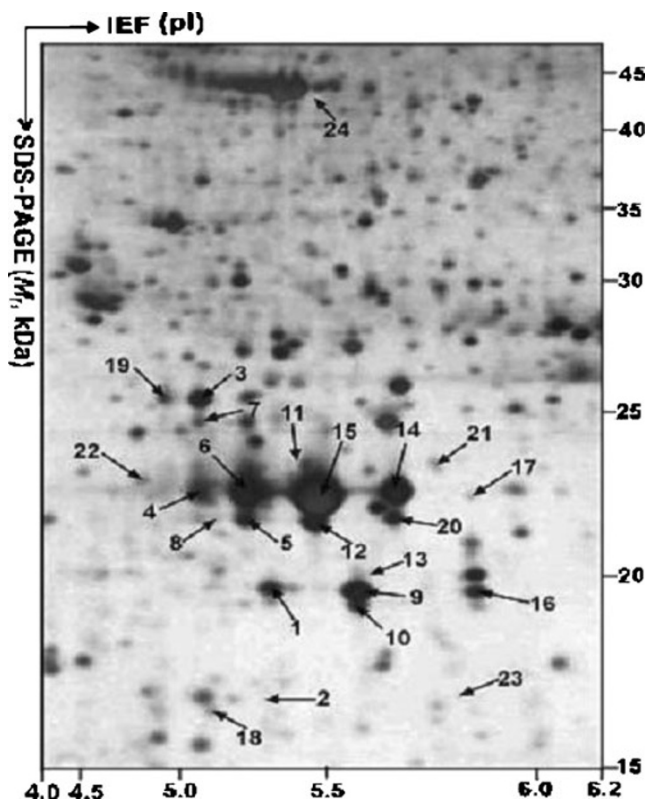


Fig. 3 MS-characterized GH isoforms in a 2-DE map of human pituitary. IEF was performed with an IPG dry strip (18 cm, pH 3–10 nonlinear). SDS-PAGE was performed with a polyacrylamide (12%) resolving gel. The protein in each labeled spot was MS-characterized. Reproduced from Zhan and Desiderio [24], with permission from Wiley-VCH, copyright 2005

proportional change of the different GH isoforms might have an important value in the clinical evaluation of NF adenomas. Recently, studies found that GH isoforms were derived from a variety of splicing variants and PTMs, including phosphorylation [24]. The phosphorylation of endogenous GH in the human pituitary [27] provided new insights into the mechanisms of GH that participate in the neuroendocrine signal pathways.

Prolactin is another important pituitary hormone. Six prolactin isoforms were detected with proteomics [1, 23]. Similar to GH, each prolactin isoform was down-regulated in each cell-type NF adenoma, with a different down-regulated ratio relative to controls. There was no significant expression change of the prolactin gene at the protein and

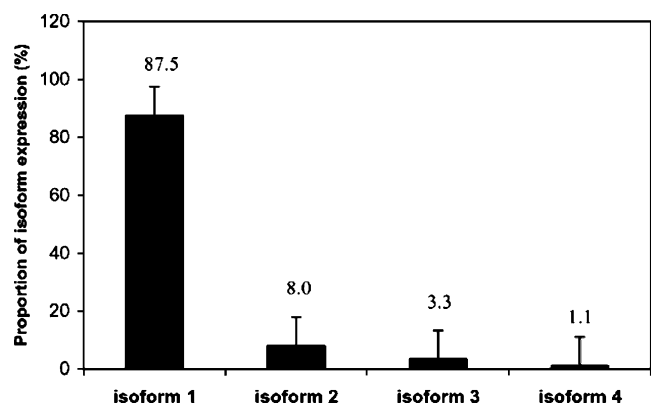


Fig. 4 Expression proportion of the four splicing isoforms of GH in human pituitary glands. Reproduced from Zhan and Desiderio [24], with permission from Wiley-VCH, copyright 2005

mRNA levels in the prolactinoma relative to controls; that finding is consistent with a prolactinoma's monoclonal origin. However, the proportion of those six prolactin isoforms changed in NF adenomas and prolactinomas compared to controls.

Other researchers showed that glycosylation is an important prolactin modification [28] that produces different isoforms; glycosylation of human prolactin might down-regulate its hormone bioactivity and promote its metabolic clearance [29]. Other studies showed that the main variant of prolactin was the non-glycosylated form of PRL in human prolactinomas [30]; that finding is consistent with the result that some prolactin isoforms were down-regulated in prolactinomas compared to controls [23]. Thus, it has been speculated that the prolactin isoform in those down-regulated spots was possibly a glycosylated prolactin. Therefore, further detailed study of the different isoforms and PTMs of GH and prolactin could lead to new insights into the clinical importance of pituitary hormones in the formation of an NF adenoma. The ratio of the change of each isoform in a human pituitary tissue has value for clinical research.

Except for protein-expression variation and splicing isoforms, protein PTMs are an important source of proteome variation that is associated with different pathophysiological processes. Also, the degree and the sites of the PTMs could dynamically change with pathophysiological processes.

Oxidative/nitrative stresses are involved in a variety of tumorigenesis mechanisms. DEP analyses [1] demonstrated that many reactive-oxygen species (ROS)-related proteins were changed in human pituitary adenomas, including heat shock protein, cytochrome P450, GST, and phospholipid hydroperoxide glutathione peroxidase (Table 2). An important oxidative stress-related modification—tyrosine nitration—was studied in an NF adenoma [31] and a post-mortem control [32, 33]. 2DGE-based nitrotyrosine Western blot and MS/MS identified eight nitroproteins from a pituitary control (Table 3) [32, 33]. Those control nitroproteins are involved in the transforming growth factor beta 1 (TFGB1) and actin cellular skeleton signaling networks (Fig. 5a), which function in gene expression, cellular development, and connective-tissue development [22]. The nitroproteins in that network include SNAP91, FCAR, actin, PRKG2, STC1, PAQR3, and PSMA2. Nitrotyrosine affinity column (NTAC)-based MS/MS identified nine nitroproteins, and three non-nitrated proteins that interact with nitroproteins, from a NF adenoma [31] (Table 3). Those NF adenoma nitroproteins and their complexes are involved in the tumor necrosis factor (TNF) and interleukin 1 (IL1) signaling networks (Fig. 5b), which function in cancer, cell cycle, and reproductive-system disease [22]. The nitroproteins in that network include ARHGAP5, PRKAR1B,

PSMA2, IL1F6, and RHPN2. The non-nitrated proteins that interact with nitroproteins include IRAK2, GRIP2, and ubiquitin. Three nitroprotein-protein complexes were identified: the nitrated proteasome-ubiquitin complex, the nitrated beta-subunit of PKA complex, and the nitrated IL1F6-IL1 receptor-IL1 receptor-associated kinase-like 2 (IL1F6-IL1R-IRAK2) complex. Two pathway networks (Fig. 5 a and b) include a same beta-estradiol signal pathway, which indicates that hormone metabolism involves the control pituitary and pituitary adenoma. This result is consistent with the fact that NO participates in pituitary hormone metabolism in normal physiology, and that a tumor interferes with hormone metabolism. Based on the documented functions of those identified proteins, each nitroprotein and nitroprotein-protein complex was rationalized into the corresponding functional system (Fig. 6) [31]. The nitrated CENT β 1 and the nitrated PKAR1 β are involved in the cAMP-dependent protein-kinase signal pathway. The nitrated RHOGAP5 and the nitrated raphilin 2 are involved in the GTPase signal pathway. IRAK-2 in the IL1-R complex and the nitrated IL1-F6 are involved in the cytokine system. The nitrated ZFP432 is involved in transcription regulatory systems. The nitrated S1P lyase 1 participates in sphingolipid metabolism to regulate cell proliferation, survival, and cell death [34] as well as the immune system [35, 36]. The nitrated LIRA4 might be involved in the immune system. The nitrated proteasome-ubiquitin complex is an important enzymatic complex that is involved in the intracellular nonlysosomal proteolytic pathway [37, 38]. These data demonstrate that those nitroproteins and nitroprotein-protein complexes might participate in several different and very important functional pathways in human pituitary tumor cells.

Four pathway-network systems, including mitochondrial dysfunction, oxidative stress, cell-cycle dysregulation, and MAPK-signal abnormality for the NF adenoma, were generated from those DEP and nitroprotein data [22]. Those four pathway-network systems have been described in detail [22]. (i) Mitochondrial dysfunctions underlie a broad spectrum of human diseases [39, 40] that includes cancer [41, 42], neurodegenerative diseases [43], cardiovascular diseases [44], diabetes mellitus [45], and inflammatory diseases [46, 47]. Notable differences in the structure and function of mitochondria appear between cancer and normal cells [41, 42]. Mitochondrial dysfunctions have been proposed as a cause of cancer [48], are a biomarker for the early detection of cancer, and are a therapeutic target for cancer. Figure 7 shows the canonical pathway of mitochondrial dysfunctions [22]. Mitochondrial dysfunction could be confirmed with a mitochondrial morphological change and the increased number of mitochondria in a human pituitary tumor [49–52]. Mitochondria are emerging as biomarker targets for the early detection of,

Table 3 Nitroproteins and non-nitrated proteins identified in a pituitary adenoma [31], and control tissue [32, 33]

Pituitary adenoma		Pituitary control	
Protein name	nY site	Protein name	nY site
<i>Nitrated protein:</i>			
Rho-GTPase-activating 5 [Q13017] (ARHGAP5)	Y ⁵⁵⁰	Synaptosomal-associated protein (SNAP91)	Y ²³⁷
Leukocyte immunoglobulin-like receptor A4 [P59901]	Y ⁴⁰⁴	Ig alpha Fc receptor [P24071] (FCAR)	Y ²²³
Zinc finger protein 432 [O94892]	Y ⁴¹	Actin [P03996] (ACTA2, ACTG2, ACTC1)	Y ²⁹⁶
PKA beta regulatory subunit [P31321] (PRKAR1B)	Y ²⁰	PKG 2 [Q13237] (PRKG2)	Y ³⁵⁴
Sphingosine-1-phosphate lyase 1 [O95470]	Y ³⁵⁶ , Y ³⁶⁶	Mitochondrial co-chaperone protein HscB [Q8IWL3]	Y ¹²⁸
Centaurin beta 1 [Q15027]	Y ⁴⁸⁵	Stanniocalcin 1 [P52823] (STC1)	Y ¹⁵⁹
Proteasome subunit alpha type 2 [P25787] (PSMA2)	Y ²²⁸	Proteasome subunit alpha type 2 (PSMA2)	Y ²²⁸
Interleukin 1 family member 6 [Q9UHA7] (IL1F6)	Y ⁹⁶	Progesterin and adipoQ receptor family member III [Q6TCH7] (PAQR3)	Y ³³
Rhopilin 2 [Q8IUC4] (RHPN2)	Y ²⁵⁸		
<i>Nitroprotein-interacting protein</i>			
Interleukin-1 receptor-associated kinase-like 2 (IRAK-2) [O43187] (IRAK2)			
Glutamate receptor interacting protein 2 [Q9C0E4] (GRIP2)			
Ubiquitin [P62988] (UBB or UBC)			

Modified from Zhan and Desiderio [31–33], with permission from Elsevier, copyright 2004, 2006, and 2007.

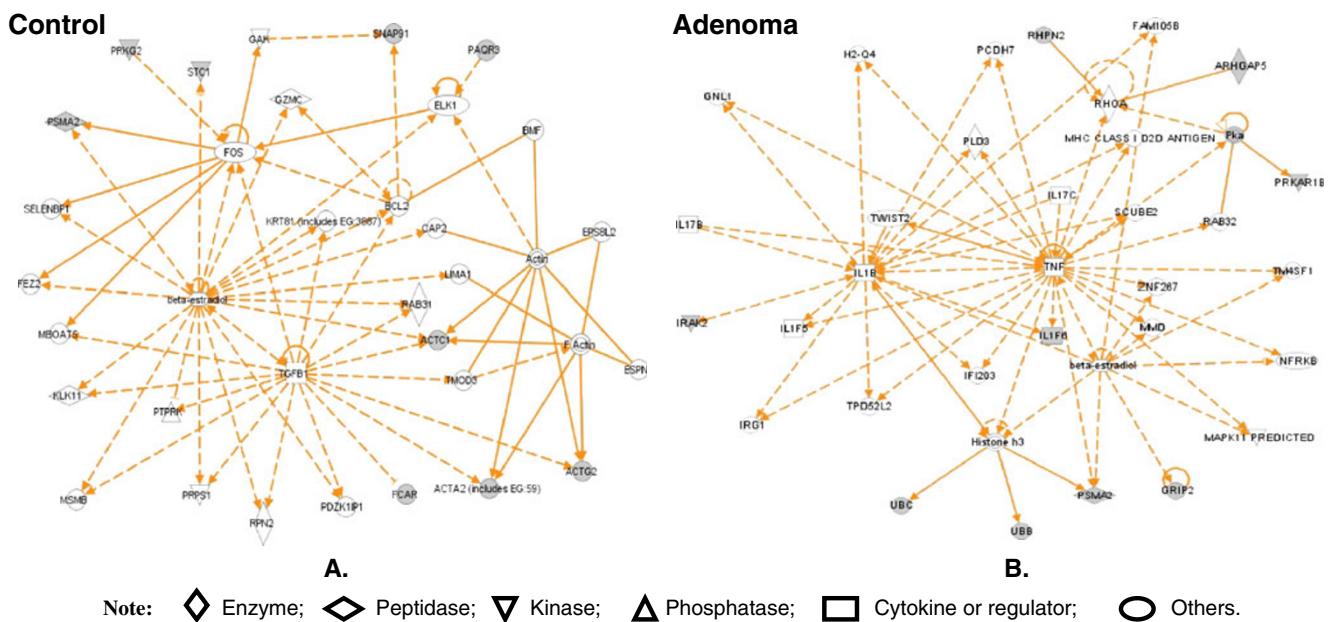


Fig. 5 Significant signaling pathway networks that are involved in human pituitary adenoma nitroproteomic data. Network A derives from control nitroproteomic data, and functions in gene expression, cellular development, and connective-tissue development and function. A gray node denotes an identified nitroprotein. Network B derives from adenoma nitroproteomic data, and functions in cancer, cell cycle, and reproductive-system disease. A gray node denotes an identified nitroprotein, or a protein that interacts with nitroproteins. An

orange solid-edge denotes a direct relationship between two nodes (molecules: proteins; genes). An orange nonsolid-edge denotes an indirect relationship between two nodes (molecules: proteins; genes). The various shapes of nodes denote the different functions. A curved line means intracellular translocation; a curved arrow means extracellular translocation. Reproduced from Zhan and Desiderio [22], with permission from BioMed Central open access journal; copyright remains with the authors

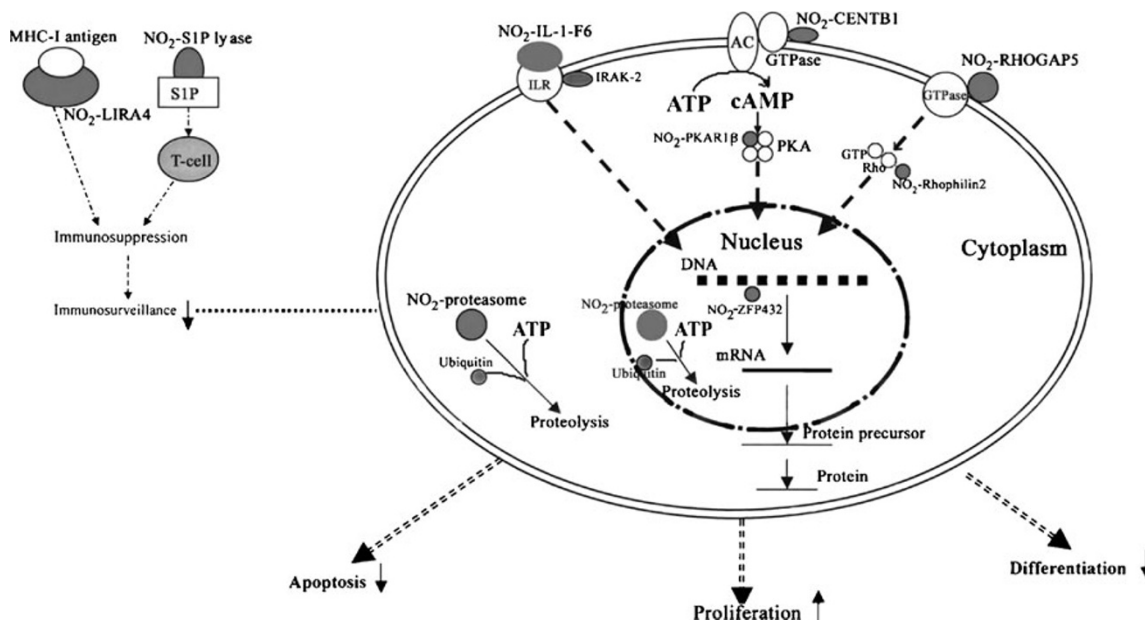


Fig. 6 Experimental data-based model of nitroproteins and their functions in human nonfunctional pituitary adenomas. NO₂⁻, nitroprotein. Reproduced from Zhan and Desiderio [31], with permission from Elsevier, copyright 2006

Mitochondrial dysfunction

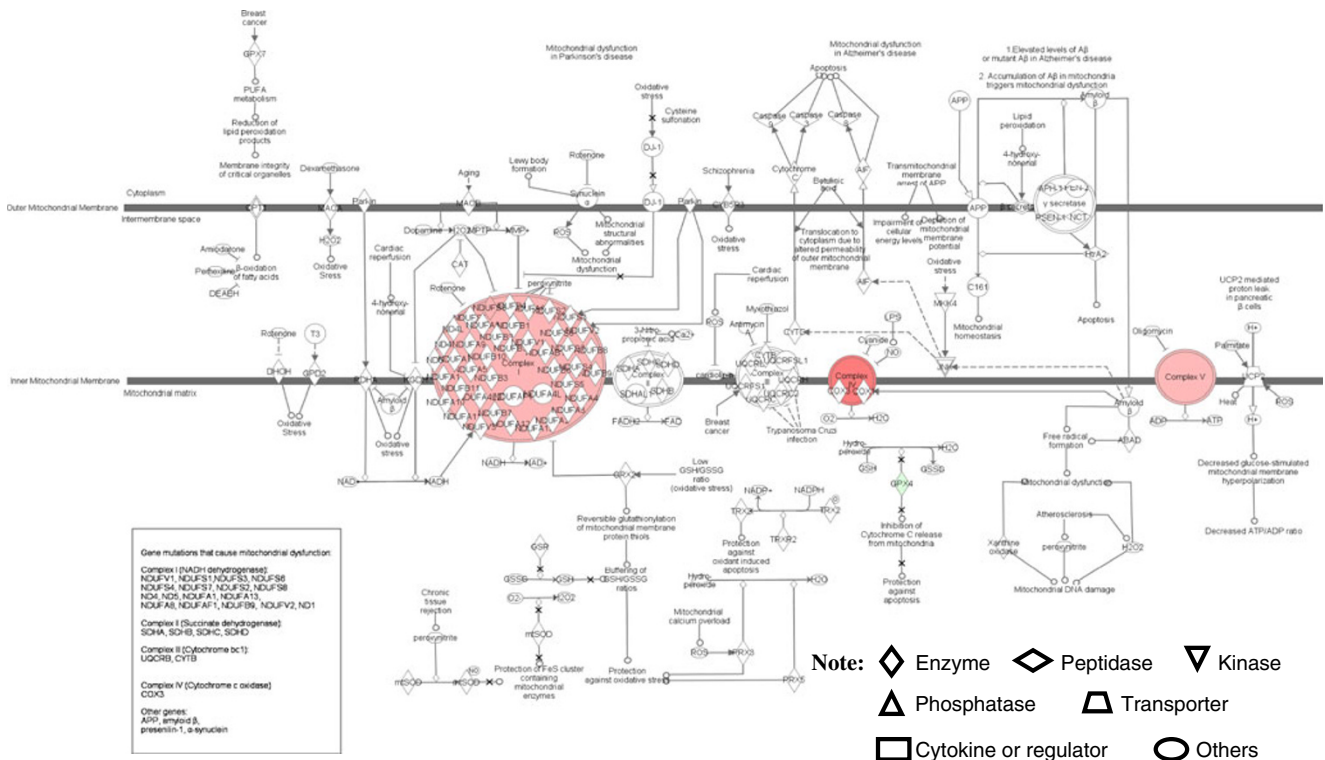


Fig. 7 Mitochondrial dysfunctional pathway network. Red label= upregulated, green label=downregulated. The various shapes of nodes denote the different functions. A duplicated shape means this node contains multiple components. An arrow denotes the pathway

direction. A line with a small circle denotes a biological result. Reproduced from Zhan and Desiderio [22], with permission from BioMed Central open access journal; copyright remains with the authors

and for novel therapeutic targets in, cancer [48, 53–56]. (ii) Oxidative stress occurs when the balance is disturbed between an upload of free-radical/reactive oxygen/nitrogen species (ROS/RNS) from *in vivo* formation and from *in vitro* environmental carcinogens, and the capability of endogenous antioxidant defense mechanisms to remove those reactive species [57, 58]. Many studies have indicated that ROS and RNS are involved in pituitary adenoma pathology [59–61]. A nitroproteomics study [31] discovered nine tyrosine-nitrated proteins in human pituitary adenoma tissues, and each tyrosine nitration site is located within an important protein domain to alter protein functions. However, with the formation of ROS and RNS, the *in vivo* antioxidative mechanism is also initiated against ROS/RNS [62]. Pivotal to the antioxidant response is the transcription factor Nrf2 (nuclear factor-E2-related factor-2). DEP and nitroproteomic data clearly reveal that the Nrf2-mediated oxidative-stress response pathway system (Fig. 8) is involved in an NF pituitary adenoma. Any decrease in the capability of this antioxidant protective system could increase the susceptibility to oxidative stress, tumor inflammation, carcinogen toxicity, and tumorigenesis. Therefore, the oxidative stress-antioxidative stress-response system could be the novel target to develop effective therapeutic agents that could be used for human pituitary adenomas [63, 64]. (iii) Cell-cycle dysregulation is involved in an NF pituitary adenoma. The basic biological characteristics of tumor cells are the unrestricted proliferation and growth that are controlled by the dysregulated cell-cycle. The cell cycle is regulated by a cyclically operating biochemical system that includes cyclins, cyclin-dependent kinases (CDK), and their inhibitors (CDKI) [65]. Studies have demonstrated that the ectopic expression of cyclin D and the overexpression of Cyclins A, B, and E occur in an NF pituitary adenoma to regulate different phases of the cell cycle, and to accelerate the progression of the cell-cycle [65]. The details on cell-cycle dysregulation in a human pituitary adenoma have been reviewed [66–69]. The pathway analysis of NF pituitary adenoma nitroproteomic data and DEP data clearly revealed the cell-cycle G2/M DNA damage checkpoint-regulation pathway (Fig. 9) in NF adenomas. The important cell-cycle regulator 14-3-3 protein was down-regulated (44-fold) in NF adenomas compared to controls (Table 2). Moreover, nitroproteomic data demonstrate that a nitrated proteasome could interfere with the functions of the ubiquitin-proteasome system in the regulation of the cell cycle. The proteasome inhibitors can induce apoptosis in GH-and prolactin-secreting rat pituitary tumor cells through a blocking of the cell cycle at the G2/M transition [70]. (iv) MAPK-signaling abnormality operates in an NF pituitary adenoma. MAPK-signaling pathways link the extracellular signal stimuli to functional cellular processes such as

Fig. 8 NRF2-mediated oxidative-stress response pathway. Red label= upregulated, green label=downregulated. The various shapes of nodes denote the different functions. A duplicated shape means this node contains multiple components. An arrow denotes the pathway direction. A line with a small circle denotes a biological result. Reproduced from Zhan and Desiderio [22], with permission from BioMed Central open access journal; copyright remains with the authors

growth, proliferation, migration, and apoptosis. The basic MAPK pathway is stimulus (mitogens, growth factors, cytokines, stress, etc.) → G-protein (Ras, Rac, Cdc42, Rho) → MAPKKK (Raf, Tpl2, MEKK, MLK, TAK, ASK, TAO) → MAPKK (MEK) → MAPK (ERK, JNK, P38) to generate responses (proliferation, differentiation, apoptosis, and migration). ERKs (extracellular signal-regulated kinases), JNKs (c-Jun N-terminal kinases), and p38-MAPKs are the three main subfamilies of MAPKs. The details of MAPK-signaling pathways in cancer have been reviewed [71–73]. The MAPK pathways are emerging as potential therapeutic targets for cancer [74, 75], and the development of inhibitors of MAPK pathways has a growing importance in cancer therapy. The pathway analyses of NF adenoma proteomic data clearly demonstrate that MAPK-signaling pathways are involved in pituitary tumorigenesis. DEP proteomic data show that Ras, ERK, JNK, p38-MAPK, Akt, TNF, TGFβ1, MAPK, and NFκB are the key nodes in their pathway networks, and that ERK/MAPK signaling (Fig. 10) is the significant canonical pathway in adenomas. The nitroproteomic data of NF adenomas show that TNF and IL1B are the key nodes in their pathway networks, and that p38-MAPK signaling (Fig. 11) is the significant canonical pathway that participates in an oxidative-stress response in an adenoma. Moreover, the PKA type I beta regulatory subunit is nitrated in human pituitary adenomas (Fig. 6; Table 3), and tyrosine nitration occurs within the dimerization region [31]; those nitrations could interfere with dimerization and affect PKA activity to suppress Raf activity. Recent studies demonstrate that an overexpression of B-Raf mRNA and protein is a feature of NF adenomas; that overactivity highlights an overactivity of the Ras-B-Raf-MAP kinase pathway to promote pituitary tumorigenesis [76], and that the low levels of Raf kinase inhibitory protein (RKIP) in a GH-pituitary adenoma correlate with a poor clinical response to somatostatin analog therapy because RKIP can bind to and inhibit Raf1 kinase to attenuate MAPK signaling [77].

Those NF pituitary adenoma-related tissue proteome data demonstrate clearly that crucial elements in the proteome-network system occur as variations in the NF adenoma pathology, offer systematic concept to treat NF pituitary adenoma patients, and could improve patient healthcare. From a prospective viewpoint, those altered crucial elements in the NF pituitary adenoma tissue proteomic network system could have three main uses: (i)

NRF2-mediated oxidative-stress response

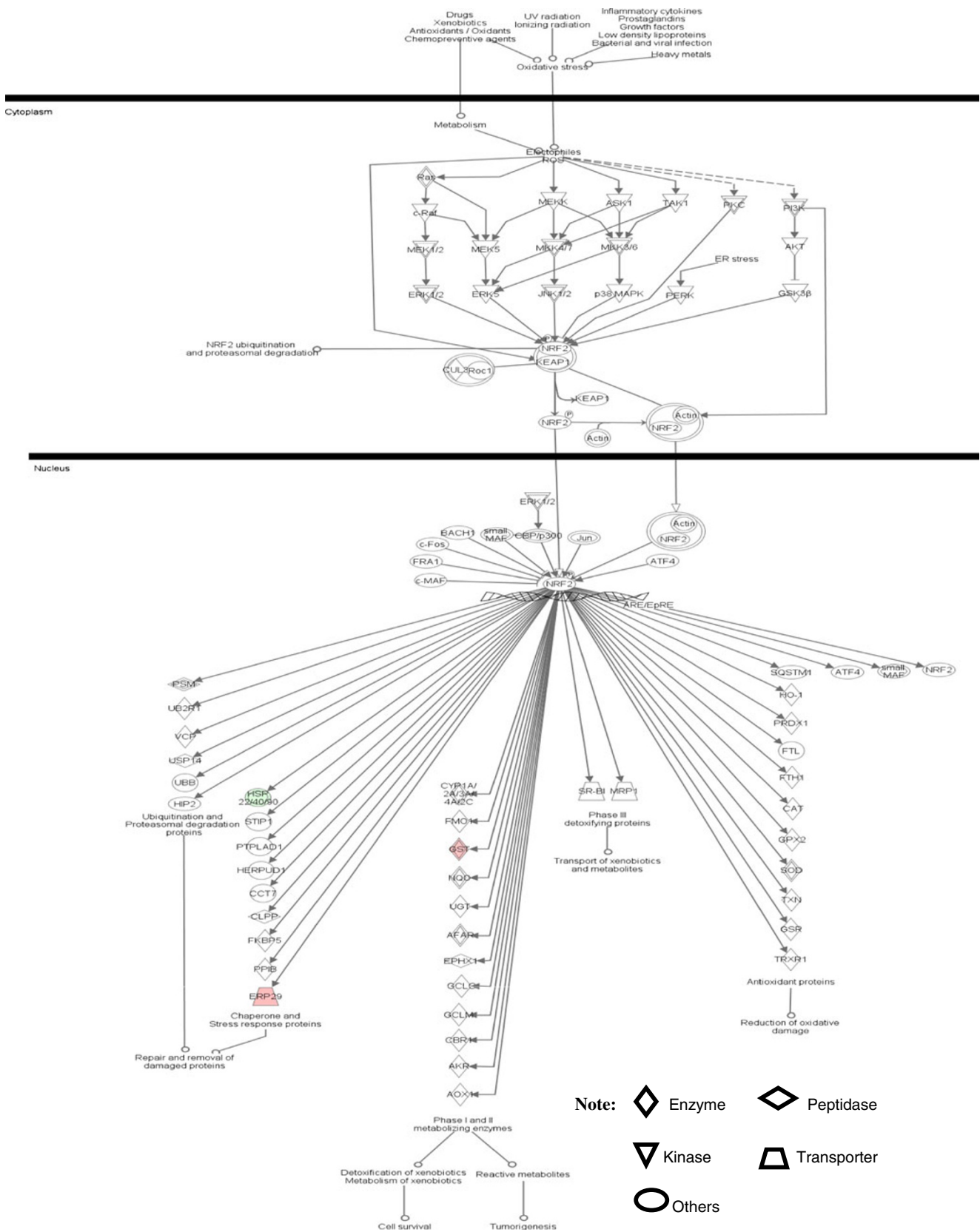
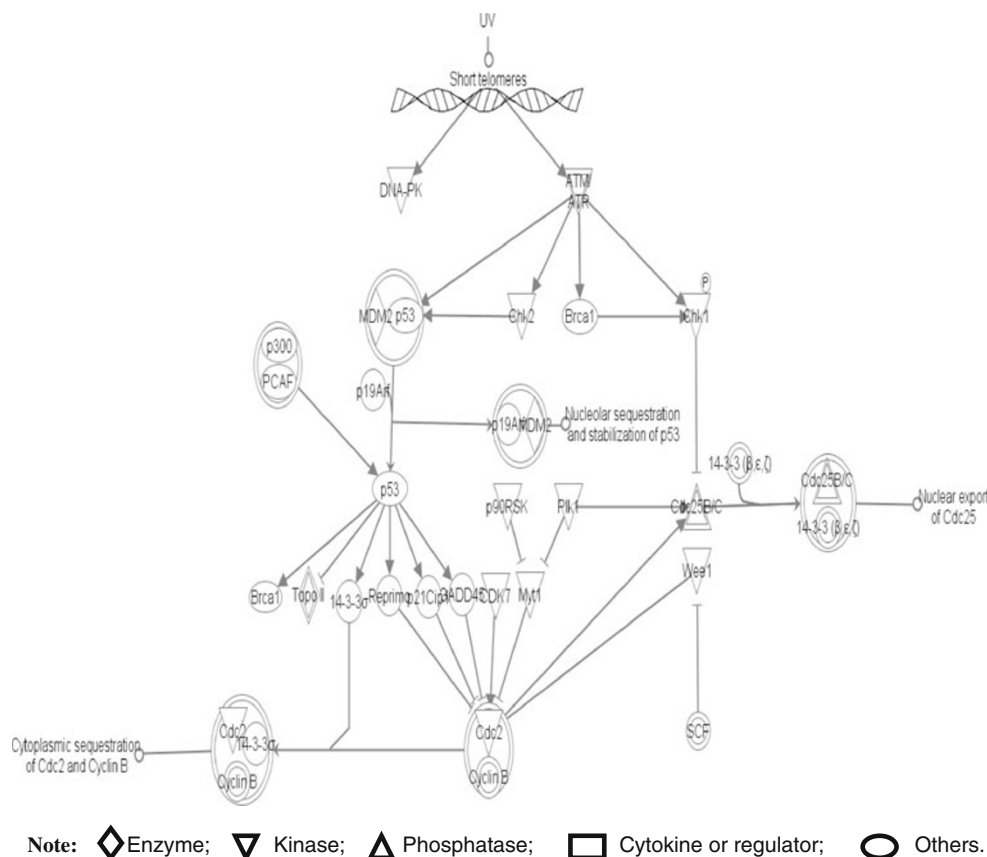


Fig. 9 Pathway component for cell cycle G2/M DNA damage checkpoint regulation. The various shapes of nodes denote the different functions. A duplicated shape means this node contains multiple components. An arrow denotes the pathway direction. A line with a small circle denotes a biological result. Reproduced from Zhan and Desiderio [22], with permission from BioMed Central open access journal; copyright remains with the authors

Cell cycle G2/M DNA damage checkpoint regulation



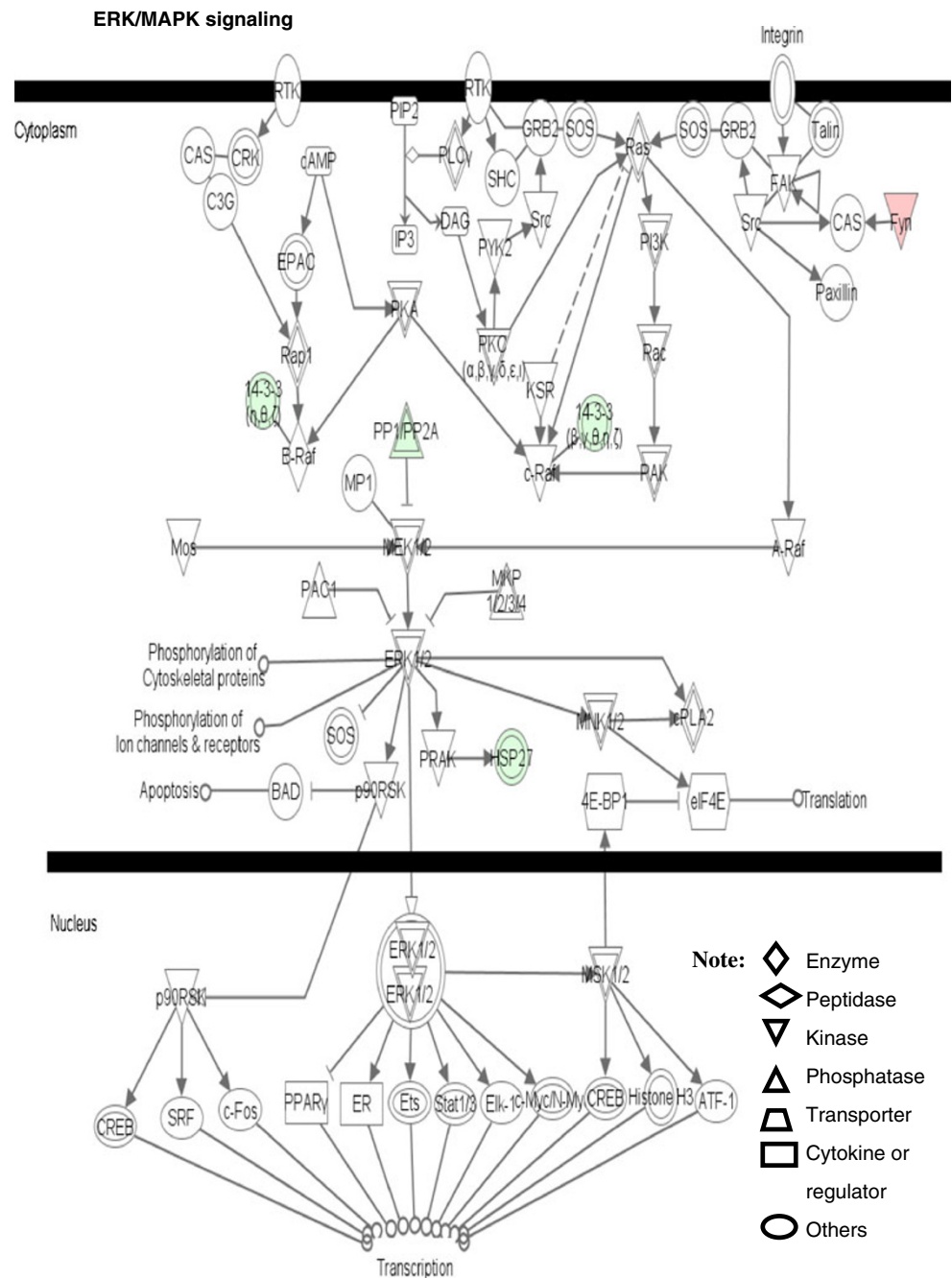
The accurate molecular classification of NF pituitary adenoma, which is necessary for an NF pituitary adenoma, because an NF pituitary adenoma is highly heterogeneous (Table 1). (ii) Interventional prevention of NF pituitary adenomas, which will be an important strategy to eliminate NF adenoma in the pre-clinical stage for population healthcare. Tissue quantitative and comparative proteomics can identify the crucial protein elements unique to an NF pituitary adenoma in the proteome-network systems. From those crucial protein elements, an accurate pharmacological intervention could be developed. The high-risk population that will be identified with body fluid (serum; CSF) biomarkers will benefit from those drugs to recover their proteome to the normal status, and to eliminate NF pituitary adenomas at the pre-clinical stage, which will significantly benefit the pituitary adenoma populations and reduce social economical load. (iii) Personalized proteomic therapy for an NF pituitary adenoma patient. The proteome differs among different type of NF pituitary adenomas, and dynamically alters with disease pathophysiological processes. With the development of proteomic techniques, computation biology, and system biology, the complete proteomic databases unique to each cell-type of NF pituitary adenomas, and different pathophysiological stages, will be accurately established. The individualized unique proteomic elements

will be determined for each patient. Toward those personalized proteomic elements, one could apply protein-engineering techniques to correct for those altered proteomic elements to the normal status in the proteome-network system in order to realize personalized patient treatment and to halt disease progression, which will provide a highly efficacious therapy, and will reduce the patient costs compared to traditional therapy strategy.

Body-fluid proteomic and peptidomic variations that potentially will contribute to the predictive diagnosis and the measurement of interventional prevention and therapy response

CSF and blood serum are two important body fluids that are used for pituitary adenoma studies. CSF around the brain and spinal cord contains peptides (several hundred) and proteins (about 1000) that are important for brain physiology [78], and is an excellent source of peptides and proteins that are secreted during brain-tumor development, and also potentially relevant to brain pathology. Serum is another important biological fluid that is an excellent source of peptides and proteins that are metabolized during pituitary-tumor pathological processes. Serum proteomes and pepti-

Fig. 10 ERK/MAPK signaling pathway. Red label=upregulated, green label=downregulated. The various shapes of nodes denote the different functions. A duplicated shape means this node contains multiple components. An arrow denotes the pathway direction. A line with a small circle denotes a biological result. Reproduced from Zhan and Desiderio [22], with permission from BioMed Central open access journal; copyright remains with the authors

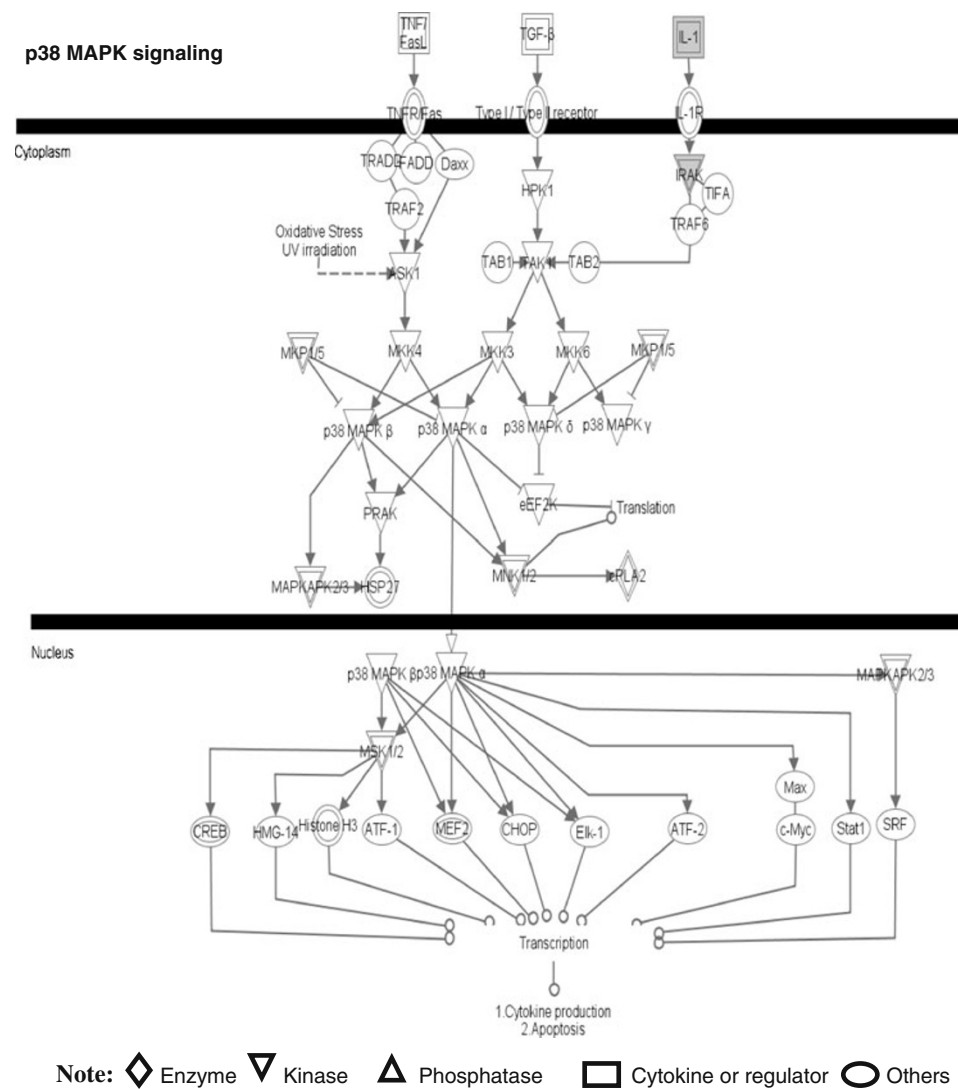


domes contain resident hemostatic proteins, immunoglobulins, cytokines, hormones, secreted proteins, exogenous proteins, peptides; infection indicators; and also tissue-leakage proteins such as those released from damaged or dying cells [79]. Therefore, blood serum contains accurate molecular information on the physiological state of all tissues/organs in the body, and changes with disease status. Monitoring the changes in the peptide and protein composition of an NF pituitary adenoma patient’s CSF and serum can be used as a sensitive and accurate molecular indicator of tumor pathology. Also, CSF and blood specimens are easily accessible from patients and controls, with minimal

invasion. Moreover, CSF and blood specimens that are from different pathological stages can be obtained at different time-points. Therefore, CSF and blood serum/plasma are excellent targets to develop early-stage diagnostic markers, and to develop the assessment indices to measure the interventional prevention and therapy response for NF pituitary adenomas.

We state here an important paradigm shift: in the future, multiple-parameter pattern-diagnostic markers and assessment indices of prevention and therapy response will replace the single-parameter biomarker for the prediction, prevention, and personalized treatment of NF pituitary

Fig. 11 The p38 MAPK signaling pathway. A gray label denotes an identified protein. The various shapes of nodes denote the different functions. A duplicated shape means this node contains multiple components. An arrow denotes the pathway direction. A line with a small circle denotes a biological result. Reproduced from Zhan and Desiderio [22], with permission from BioMed Central open access journal; copyright remains with the authors



adenomas. The reason for that paradigm shift is that a traditional single-molecule biomarker is based on an unrealistic assumption that an increase in the amount of a single compound can unambiguously specify a disease. However, the reality is that NF pituitary adenomas are characterized by a heterogeneity among patients; pituitary adenomas are initiated by numerous factors, and cause a range of different molecular changes. For example, a significant elevation of blood GH and prolactin has been useful to predict and diagnose acromegaly and prolactinoma, respectively. However, a large number of acromegaly and prolactinoma patients are still detected at an advanced stage. For the NF pituitary adenomas, no blood hormone levels are elevated, almost all NF pituitary adenomas are diagnosed at a late stage, and no molecular indices measure the therapy response and prognosis. Multiple-parameter biomarkers from serum or CSF will resolve these problems in the prediction, prevention, and personalized treatment assessment of an NF pituitary adenoma.

Several promising multiparameter strategies from serum or CSF proteomes and peptidomes might meet those requirements, and include protein/peptide pattern diagnostics, protein microarrays, and an antibody microarray.

Serum/CSF protein/peptide pattern diagnostics This diagnostic is an MS-based technique; namely, use the pattern variation of MS peaks to diagnose disease. The concept behind pattern diagnostics is that the CSF/blood serum proteome/peptidome reflects the pituitary tumor tissue and organ pathology that cause patterns of body-fluid protein/peptide changes that have diagnostic potential without even knowing the identities of the individual proteins/peptides [79]. MS can be used to detect that pattern of peaks that correspond to the intact proteins or peptides, and that peak pattern can be used to discriminate among different stages of tumor from controls. The prediction diagnosis and prognostic monitoring becomes the comparison of the pattern or “signature” of the proteins or peptides. A

serum-protein pattern has been used to predict and assess laryngeal squamous cell carcinoma [80] and ovarian cancer [81]. Surface-enhanced laser desorption/ionization time-of-flight mass spectrometry (SELDI-TOF-MS) has been used in this protein-pattern recognition [80, 81]. Serum or CSF samples from tumor and controls were individually applied to a SELDI chip. The retained proteins on the chip were subsequently ionized (MALDI) and detected with TOF-MS. Corresponding bioinformatics software was used to compare the MS spectra and to determine the differential pattern of peaks to discriminate tumors from controls. This approach has been used to identify a “signature” protein pattern within a “training set” of SELDI-TOF MS spectra that contained 50 female ovarian cancer sera and 50 female control sera [81]. The identified “signature” protein pattern was used to identify ovarian cancer in individuals within a new independent “validation set” that contained 50 female ovarian cancer sera and 66 female control sera, with a positive predictive value of 94%, sensitivity of 100%, and specificity of 95% [81].

However, even though SELDI-TOF MS was the most commonly used instrument for this protein-pattern recognition, some problems remain to be resolved for the SELDI method: (i) What is the total amount (moles) of ionizable groups (positive, negative) due to Lys, Arg, Asp, Glu, etc. in the biological samples? (ii) What is the total number (moles) of ionizable groups on the SELDI sample probe surface? (iii) What is the ratio of (i)/(ii) in the SELDI experiments? (iv) Does the sample exceed, and/or chemically modify, the chromatography capability of the SELDI probe surface? (v) SELDI does not allow a direct identification of the discriminatory proteins and suffers from low reproducibility, sensitivity, and accuracy [82–86]. To overcome some of those problems, Dekker et al. used MALDI-TOF-MS to replace SELDI-TOF-MS; the body-fluid samples were first digested with trypsin, and the tryptic peptide mixtures were analyzed with MALDI-TOF-MS to obtain the tryptic-peptide profile for each sample to diagnose leptomeningeal metastases in breast cancer patients with an accuracy of 77%, a sensitivity of 79%, and a specificity of 76% [82, 87].

Thempst et al. developed a signature serum-peptide pattern to predict metastatic thyroid carcinomas [88, 89]. The plasma peptides were isolated with magnetic beads, and were analyzed with a MALDI-MS scan to obtain MS spectra. The pattern of peaks that corresponded to the serum peptides were identified in a “training setting”; subsequently, the serum-peptide pattern was used to discriminate the tumors from controls in an independent “validating setting”. This serum-peptidome pattern was used to distinguish metastatic thyroid carcinomas from cancer-free controls with 95% sensitivity and 95% specificity (binomial confidence intervals, 75.1–99.9%) [89].

CSF protein pattern and peptide patterns are also under development to diagnose CNS system disease with a similar strategy [78, 82]. Amino acid sequence data are needed in those studies.

The serum/CSF protein/peptide pattern-recognition approach is emerging as a promising method for cancer prediction and diagnosis, and also offers the promise to measure the interventional prevention and therapeutic response. However, one should clearly realize that the standardized common serum/CSF-sample handling procedure [90] and the large-scale samples analyzed [79] are essential, and that this pattern-recognition method would have more potential in combination with other genetic factor analysis [79], and with amino acid sequence data.

Protein/antibody microarray This technique is derived from DNA-microarray technologies without the use of MS to detect any proteomic variation. It is another promising approach for prediction diagnosis and for the measurement of interventional prevention and therapeutic response. Briefly, a protein microarray immobilizes thousands of protein-detecting features on a microchip. This microchip is incubated with a biological sample to detect any proteomic pattern variation in the disease relative to controls. The common protein array refers to an antibody array that hosts different antibodies such as monoclonal, polyclonal, and antibody fragments to detect their respective antigens from human blood samples. An example is the cytokine-antibody array to detect inflammation-related disease [91]. Another type of protein microarray is to directly immobilize multiple functional proteins onto a microchip to detect protein-protein, protein-ligand, enzyme-substance, protein-DNA interactions, and PTMs, which is also called a “functional protein array” [92–94]. An example is the whole-proteome array from yeast that contains 5,000 purified proteins immobilized onto a glass microscope slide [95]. With the prospective to identify the serum/CSF protein/peptide patterns of NF pituitary adenoma, those signature protein/peptides or their antibodies could be immobilized onto a microchip for the predictive diagnosis and the measurement of preventive and therapeutic response for NF pituitary adenoma patients.

Conclusions

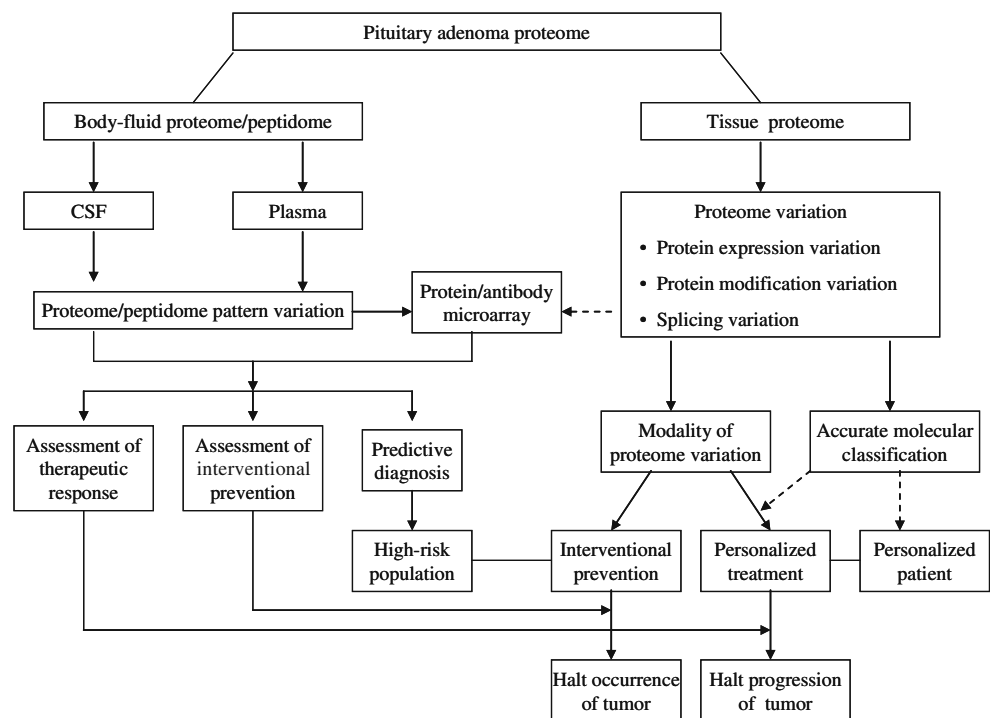
The traditional assessment of the level of one serum-hormone is difficult to detect clinical NF pituitary adenomas; furthermore, no effective chemical therapy exists to treat an NF pituitary adenoma. An NF pituitary adenoma contains highly heterogeneous cell types that are the distinguishing pathophysiological basis that causes the

proteomic variation. In combination with multiple endogenous and exogenous factors, that proteome variation is the basis for personalized patient treatment. A more accurate multiple-parameter global systematic strategy will replace the limited single-factor strategy in the prediction, prevention, and personalized treatment of NF pituitary adenomas. The global-proteomic strategy offers the promise for the multiple-parameter assessment for the predictive diagnosis, measurement of preventive and therapeutic response, and personalized patient treatment. The diagram of the use of proteomic variations to predict, prevent, and personalize treatment of NF adenomas is shown (Fig. 12). A tissue proteomic-variation study would lead to the accurate molecular classification of an NF adenoma, and the modality of proteomic variation for interventional prevention and halting the tumor progression. The serum/CSF proteomic/peptidomic variation would lead to serum/CSF protein/peptide pattern diagnostics, and the protein/antibody microarray for the predictive diagnosis and measurement of interventional prevention and personalized therapeutic response. Briefly, a tumor-tissue proteomic-variation study would lead to an accurate molecular classification that will contribute to identification of personalized patients. Modalities of the altered tissue-proteome of a personalized patient will realize the personalized treatment of NF pituitary adenomas. This therapeutic response will be assessed with body-fluid (serum and/or CSF) proteomic/peptidomic variations to determine whether tumor progression is inhibited or halted. The high-risk population that might develop an NF pituitary adenoma will

be determined with a predictive diagnosis strategy—body-fluid (serum and/or CSF) proteomic/peptidomic variations. Modalities of the altered tissue-proteome of the high-risk population will realize the interventional prevention. This preventive response will be assessed with body-fluid (serum and/or CSF) proteomic/peptidomic variations to determine the preventive effects and whether tumor occurrence is halted. In summary, any proteomic variation is tightly associated with NF pituitary adenoma progression. The detection and modalities of proteomic variation will serve an effective role in the prediction, prevention, and personalized treatment of NF pituitary adenoma patients.

Proteomic variations offer numerous opportunities to predict, prevent, and provide a personalized treatment (PPPM) for NF pituitary adenomas. Currently, whole tissue-proteome variations (protein expression; hormone isoforms; modifications) are studied in NF pituitary adenomas, and the significant signal pathway networks associated with NF pituitary adenomas are mined from those whole tissue-proteome variation data. Four important pathway-network systems, including mitochondrial dysfunction, oxidative stress, cell-cycle dysregulation, and MAPK-signal abnormality, for the NF adenoma are identified. The follow-up studies of those tissue proteome data would contribute to the accurate molecular classification for the diagnosis of personalized patients and their personalized treatment. However, some problems remain to be resolved towards that PPPM goal: (i) the complete tissue-proteomic databases unique to single-cell type pituitary adenomas, and

Fig. 12 Diagram of the use of proteomic variations to predict, prevent, and personalize treatment for NF pituitary adenomas



different pathophysiological stages, must be accurately established towards the development of individualized unique proteomic elements for each patient. Recently, laser-captured single-cell type pituitary prolactinoma cells have been used for proteomics analysis [96]. (ii) The complete body-fluid (serum; CSF) proteomic/peptidomic databases unique to single-cell type pituitary adenomas, and different pathophysiological stages, must be accurately established. For body-fluid proteomic/peptidomic databases, a large-scale of normal population is needed towards the development of the predictive diagnostic pattern. (iii) The novel techniques such as protein-engineering techniques must be developed to correct for those altered personalized proteomic elements to the normal status in the proteome-network system towards an interventional prevention in the high-risk population and for the personalized treatment of NF pituitary adenomas. (iv) The comprehensive computation model system needs to be developed to integrate all proteomic data, personal data, clinical data, and other genetic data towards the facile and accurate predictive diagnosis of the high-risk population and an accurate assessment of preventive and therapeutic responses.

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References

- Moreno CS, Evans CO, Zhan X, Okor M, Desiderio DM, Oyesiku NM. Novel molecular signaling in human clinically non-functional pituitary adenomas identified by gene expression profiling and proteomic analyses. *Cancer Res.* 2005;65:10214–22.
- Herman V, Fagin J, Gonsky R, Kovacs K, Melmed S. Clonal origin of pituitary adenomas. *J Clin Endocrinol Metab.* 1990;71:1427–33.
- Alexander JM, Biler BM, Bikkal H, Zervas NT, Arnold A, Klibanski A. Clinically nonfunctioning pituitary tumors are monoclonal in origin. *J Clin Invest.* 1990;86:336–40.
- Melmed S. Mechanisms for pituitary tumorigenesis: the plastic pituitary. *J Clin Invest.* 2003;112:1603–16018.
- Zhan X, Desiderio DM. Comparative proteomics analysis of human pituitary adenomas: current status and future perspectives. *Mass Spectrom Rev.* 2005;24:783–813.
- Zhan X, Desiderio DM. Differences in the spatial and quantitative reproducibility between two second-dimension gel electrophoresis systems. *Electrophoresis.* 2003;24:1834–46.
- Zhan X, Desiderio DM. Spot volume versus amount of protein loaded onto a gel. A detailed, statistical comparison of two gel electrophoresis systems. *Electrophoresis.* 2003;24:1818–33.
- Zhan X, Desiderio DM. A reference map of a pituitary adenoma proteome. *Proteomics.* 2003;3:699–713.
- Unlu M, Morgan ME, Minden JS. Difference gel electrophoresis: a single gel method for detecting changes in protein extracts. *Electrophoresis.* 1997;18:2071–7.
- Tonge R, Shaw J, Middleton B, Rowlinson E, Currie I, Davison M. Validation and development of fluorescence two-dimensional differential gel electrophoresis proteomics technology. *Proteomics.* 2001;1:377–96.
- Zhou G, Li H, DeCamp D, Chen S, Shu H, Gong Y, et al. 2D differential in-gel electrophoresis for the identification of esophageal scans cell cancer-specific protein markers. *Mol Cell Proteomics.* 2002;1:117–24.
- Wolters DA, Washburn MP, Yates III JR. An automated multidimensional protein identification technology for shotgun proteomics. *Anal Chem.* 2001;73:5683–90.
- Aebersold R, Mann M. Mass spectrometry-based proteomics. *Nature.* 2003;422:198–207.
- MacCoss MJ, McDonald WH, Saraf A, Sadygov R, Clark JM, Tasto JJ, et al. Shotgun identification of protein modifications from protein complexes and lens tissue. *Proc Natl Acad Sci USA.* 2002;99:7900–5.
- Regnier FE, Rigs L, Zhang R, Xiong L, Liu P, Chakraborty A, et al. Comparative proteomics based on stable isotope labeling and affinity selection. *J Mass Spectrom.* 2002;37:133–45.
- Smolka MB, Zhou H, Purkayastha S, Aebersold R. Optimization of the isotope-coded affinity tag-labeling procedure for quantitative proteome analysis. *Anal Biochem.* 2001;297:25–31.
- Zhu H, Pan S, Gu S, Bradbury EM, Chen X. Amino acid residue specific stable isotope labeling for quantitative proteomics. *Rapid Commun Mass Spectrom.* 2002;16:2115–23.
- Gu S, Pan S, Bradbury EM, Chen X. Precise peptide sequencing and protein identification in the human proteome through *in vivo* lysine-specific mass tagging. *J Am Soc Mass Spectrom.* 2003;14:1–7.
- Casado-Vela J, Martinez-Esteso MJ, Rodriguez E, Borrás E, Elortza F, Bru-Martinez R. iTRAQ-based quantitative analysis of protein mixtures with large fold change and dynamic range. *Proteomics.* 2010;10:343–7.
- Griffin TJ, Xie H, Bandhakavi S, Popko J, Mohan A, Carlis JV, et al. iTRAQ reagent-based quantitative proteomic analysis on a linear ion trap mass spectrometer. *J Proteome Res.* 2007;6:4200–9.
- Pierce A, Unwin RD, Evans CA, Griffiths S, Carney L, Zhang L, et al. Eight-channel iTRAQ enables comparison of the activity of six leukemogenic tyrosine kinases. *Mol Cell Proteomics.* 2008;7:853–63.
- Zhan X, Desiderio DM. Signaling pathway networks mined from human pituitary adenoma proteomics data. *BMC Med Genomics.* 2010;3:13.
- Evans CO, Moreno CS, Zhan X, McCabe MT, Vertino PM, Desiderio DM, et al. Molecular pathogenesis of human prolactinomas identified by gene expression profiling, RT-qPCR, and proteomic analyses. *Pituitary.* 2008;11:231–45.
- Zhan X, Giorgianni F, Desiderio DM. The proteomics analysis of growth hormone isoforms in human pituitary. *Proteomics.* 2005;5:1228–41.
- Boguszewski CL, Johannsson G, Bengtsson BA, Johannsson A, Carlsson B, Carlsson LM. Circulating non-22-kilodalton growth hormone isoforms in acromegalic men before and after transphenoidal surgery. *J Clin Endocr Metab.* 1997;82:1516–21.
- Boguszewski CL, Jansson C, Boguszewski MC, Rosberg S, Carlsson B, Albertsson-Wikland K, et al. Increased proportion of circulating non-22-kilodalton growth hormone isoforms in short children: a possible mechanism for growth failure. *J Clin Endocr Metab.* 1997;82:2944–9.
- Giorgianni F, Beranova-Giorgianni S, Desiderio DM. Identification and characterization of phosphorylated proteins in human pituitary. *Proteomics.* 2004;4:587–98.
- Gambino GM, Beck-Peccoz P, Borgato S, Faglia G, Spada A, Persani L. Bioactivity and glycosylation of circulating prolactin in various physiological and pathological conditions. *Pituitary.* 1999;2:225–31.
- Hoffmann T, Penel C, Ronin C. Glycosylation of human prolactin regulates hormone bioactivity and metabolic clearance. *J Endocrinol Invest.* 1993;16:807–16.

30. Hoffmann T, Gunz G, Brue T, Jaquet P, Ronin C. Prolactin isoforms secreted by human prolactinomas. *Horm Res.* 1992;38:164–70.
31. Zhan X, Desiderio DM. Nitroproteins from a human pituitary adenoma tissue discovered with a nitrotyrosine affinity column and tandem mass spectrometry. *Anal Biochem.* 2006;354:279–89.
32. Zhan X, Desiderio DM. The human pituitary nitroproteome: detection of nitrotyrosyl-proteins with two-dimensional Western blotting, and amino acid sequence determination with mass spectrometry. *Biochem Biophys Res Commun.* 2004;325:1180–6.
33. Zhan X, Desiderio DM. Linear ion-trap mass spectrometric characterization of human pituitary nitrotyrosine-containing proteins. *Int J Mass Spectrom.* 2007;259:96–104.
34. Maceykaa M, Payneb SG, Milstienb S, Spiegel S. Sphingosine kinase, sphingosine-1-phosphate, and apoptosis. *Biochim Biophys Acta.* 2002;1585:193–201.
35. Schwab SR, Pereira JP, Matloubian M, Xu Y, Huang Y, Cyster JG. Lymphocyte sequestration through S1P lyase inhibition and disruption of S1P gradients. *Science.* 2005;309:1735–9.
36. Hla T. Dietary factors and immunological consequences. *Science.* 2005;309:1682–3.
37. Tamura T, Lee DH, Osaka F, Fujiwara T, Shin S, Chung CH, et al. Molecular cloning and sequence analysis of cDNAs for Wve major subunits of human proteasomes (multi-catalytic proteinase complexes). *Biochem Biophys Acta.* 1991;1089:95–102.
38. Kristensen P, Johnsen AH, Uerkvitz W, Tanaka K, Hendil KB. Human proteasome subunits from 2-dimensional gels identified by partial sequencing. *Biochem Biophys Res Commun.* 1994;205:1785–9.
39. Pagliarini DJ, Calvo SE, Chang B, Sheth SA, Vafai SB, Ong SE, et al. A mitochondrial protein compendium elucidates complex I disease biology. *Cell.* 2008;134:112–23.
40. Koene S, Smeitink J. Mitochondrial medicine: entering the era of treatment. *J Intern Med.* 2009;265:193–209.
41. Modica-Napolitano JS, Singh KK. Mitochondria as targets for detection and treatment of cancer. *Expert Rev Mol Med.* 2002;4:1–19.
42. Modica-Napolitano JS, Singh KK. Mitochondrial dysfunction in cancer. *Mitochondrion.* 2004;4:755–62.
43. Schapira AH. Mitochondrial dysfunction in neurodegenerative diseases. *Neurochem Res.* 2008;33:2502–9.
44. Ballinger SW. Mitochondrial dysfunction in cardiovascular disease. *Free Radic Biol Med.* 2005;38:1278–95.
45. Anderson CM. Mitochondrial dysfunction in diabetes mellitus. *Drug Dev Res.* 1999;46:67–79.
46. Rabinovich RA, Bastos R, Ardite E, Llinas L, Orozco-Levi M, Gea J, et al. Mitochondrial dysfunction in COPD patients with low body mass index. *Eur Respir J.* 2007;29:643–50.
47. Mabaliraja U, Dinda AK, Kumar S, Roshan R, Gupta P, Sharma SK, et al. Mitochondrial structural changes and dysfunction are associated with experimental allergic asthma. *J Immunol.* 2008;181:3540–8.
48. Halabe Bucay A. The biological significance of cancer: Mitochondria as a cause of cancer and the inhibition of glycolysis with citrate as a cancer treatment. *Med Hypotheses.* 2007;69:826–8.
49. Goebel HH, Schulz F, Rama B. Ultrastructurally abnormal mitochondria in the pituitary oncocyoma. *Acta Neurochir.* 1980;51:195–201.
50. Horoupian DS. Large mitochondria in a pituitary adenoma with hyperprolactinemia. *Cancer.* 1980;46:537–42.
51. Niroi N, Chrousos GP, Kohn B, Lafferty A, Abu-Asab M, Bonat S, et al. Adrenocortical-pituitary hybrid tumor causing Cushing's syndrome. *J Clin Endocrinol Metab.* 2001;86:2631–7.
52. Saeger W, Ludecke DK, Buchfelder M, Fahlbusch R, Quabbe HJ, Petersenn S. Pathohistological classification of pituitary tumors: 10 years of experience with the German pituitary tumor registry. *Eur J Endocrinol.* 2007;156:203–16.
53. Garber K. Targeting mitochondria emerges as therapeutic strategy. *J Natl Cancer Inst.* 2005;97:1800–1.
54. Galluzzi L, Larochette N, Zamzami N, Kroemer G. Mitochondria as therapeutic targets for cancer chemotherapy. *Oncogene.* 2006;25:4812–30.
55. Modica-Napolitano JS, Kulawiec M, Singh KK. Mitochondria and human cancer. *Cur Mol Med.* 2007;7:121–31.
56. Bell EL, Klimova T, Chandel NS. Targeting the mitochondria for cancer therapy: regulation of hypoxia-inducible factor by mitochondria. *Antioxid Redox Signal.* 2008;10:635–40.
57. Tandon VR, Sharma S, Mahajan A, Bardi GH. Oxidative stress: a novel strategy in cancer treatment. *J K Science.* 2005;7:1–3.
58. Toyokuni S. Molecular mechanisms of oxidative stress-induced carcinogenesis: from epidemiology to oxygenomics. *IUBMB Life.* 2008;60:441–7.
59. Pinilla L, Tena-Sempere M, Aguilar E. Nitric oxide stimulates growth hormone secretion in vitro through a calcium—and cyclic guanosine monophosphate-independent mechanism. *Horm Res.* 1999;51:242–7.
60. Cuttica CM, Giusti M, Bocca L, Sessarego P, De Martini D, Valenti S, et al. Nitric oxide modulates in vivo and in vitro growth hormone release in acromegaly. *Neuroendocrinology.* 1997;66:426–31.
61. Bocca L, Valenti S, Cuttica CM, Spaziante R, Giordano G, Giusti M. Nitric oxide biphasically modulates GH secretion in cultured cells of GH-secreting human pituitary adenomas. *Minerva Endocrinol.* 2000;25:55–9.
62. Valko M, Rhodes CJ, Moncol J, Izakovic M, Mazur M. Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem Biol Interact.* 2006;160:1–40.
63. Yates MS, Kensler TW. Chemopreventive promise of targeting the Nrf2 pathway. *Drug News Perspect.* 2007;20:109–17.
64. Lim JH, Kim KM, Kim SW, Hwang O, Choi HJ. Bromocriptine activates NQO1 via Nrf2-PI3K/Akt signaling: novel cytoprotective mechanism against oxidative damage. *Pharmacol Res.* 2008;57:325–31.
65. Turner HE, Nagy ZS, Nullivan N, Esiri MM, Wass JAH. Expression analysis of cyclins in pituitary adenomas and the normal pituitary gland. *Clin Endocrinol.* 2000;53:337–44.
66. Musat M, Vax VV, Borboli N, Georguev M, Bonner S, Korbonits M, et al. Cell cycle dysregulation in pituitary oncogenesis. *Front Horm Res.* 2004;32:34–62.
67. Saeger W. Proliferation markers and cell cycle inhibitors in pituitary adenomas. *Front Horm Res.* 2004;32:110–26.
68. Heaney AP. Pituitary tumour pathogenesis. *Br Med Bull.* 2006;75–76:81–97.
69. Farrell WE, Clayton RN. Molecular pathogenesis of pituitary tumors. *Front Neuroendocrinol.* 2000;21:174–98.
70. Yu R, Ren SG, Melmed S. Proteasome inhibitors induce apoptosis in growth hormone—and prolactin-secreting rat pituitary tumor cells. *J Endocrinol.* 2002;174:379–86.
71. Dhillon AS, Hagan S, Rath O, Kolch W. MAP kinase signaling pathways in cancer. *Oncogene.* 2007;26:3279–90.
72. Wada T, Penninger JM. Mitogen-activated protein kinases in apoptosis regulation. *Oncogene.* 2004;23:2838–49.
73. Johnson GL, Lpadat R. Mitogen-activated protein kinase pathways mediated by ERK, JNK, and p38 protein kinases. *Science.* 2002;298:1911–2.
74. English JM, Cobb MH. Pharmacological inhibitors of MAPK pathways. *Trends Pharmacol Sci.* 2002;23:40–5.
75. Olson JM, Hallahan AR. p38 MAP kinase: a convergence point in cancer therapy. *Trends Mol Med.* 2004;10:125–9.
76. Ewing I, Pedder-Smith S, Franchi G, Ruscica M, Emery M, Vax V, et al. A mutation and expression analysis of the oncogene BRAF in pituitary adenomas. *Clin Endocrinol (Oxf).* 2007;66:348–52.
77. Fougner SL, Bollerslev J, Latif F, Hald JK, Lund T, Ramm-Petersen J, et al. Low levels of raf kinase inhibitory protein in

- growth hormone-secreting pituitary adenomas correlate with poor response to octreotide treatment. *J Clin Endocrinol Metab.* 2008; 93:1211–6.
78. Dekker LJ, Burgers PC, Kros JM, Smitt PA, Luijckers TM. Peptide profiling of cerebrospinal fluid by mass spectrometry. *Expert Rev Proteomics.* 2006;3:297–309.
79. Weston AD, Hood L. Systems biology, proteomics, and the future of health care: toward predictive, preventative, and personalized medicine. *J Proteome Res.* 2004;3:179–96.
80. Cheng L, Zhou L, Tao L, Zhang M, Cui J, Li Y. SELDI-TOF MS profiling of serum for detection of laryngeal squamous cell carcinoma and the progression to lymph node metastasis. *J Cancer Res Clin Oncol.* 2008;134:769–76.
81. Peticoin EF, Ardekani AM, Hitt BA, Levine PJ, Fusaro VA, Steinberg SM, et al. Use of proteomic patterns in serum to identify ovarian cancer. *Lancet.* 2002;359:572–7.
82. Dekker LJ, Boogerd W, Stockhammer G, Dalebout JC, Siccama I, Zheng P, et al. MALDI-TOF mass spectrometry analysis of cerebrospinal fluid tryptic peptide profiles to diagnose leptomeningeal metastases in patients with breast cancer. *Mol Cell Proteomics.* 2005;4:1341–9.
83. Diamandis EP. Point: Proteomic patterns in biological fluids: do they represent the future of cancer diagnostics? *Clin Chem.* 2003;49:1272–5.
84. Diamandis EP. Analysis of serum proteomic patterns for early cancer diagnosis: drawing attention to potential problems. *J Natl Cancer Inst.* 2004;96:353–6.
85. Diamandis EP. Mass spectrometry as a diagnostic and a cancer biomarker discovery tool: opportunities and potential limitations. *Mol Cell Proteomics.* 2004;3:367–78.
86. Diamandis EP, van der Merwe DE. Plasma protein profiling by mass spectrometry for cancer diagnosis: opportunities and limitations. *Clin Cancer Res.* 2005;11:963–5.
87. Dekker LJ, Dalebout JC, Siccama I, Jenster G, Sillevius-Smitt PA, Luijckers TM. A new method to analyze matrix-assisted laser desorption/ionization time-of-flight peptide profiling mass spectra. *Rapid Commun Mass Spectrom.* 2005;19:865–70.
88. Villanueva J, Philip J, Entenberg D, Chaparro CA, Tanwar MK, Holland EC, et al. Serum peptide profiling by magnetic particle-assisted, automated sample processing and MALDI-TOF mass spectrometry. *Anal Chem.* 2004;76:1560–70.
89. Villanueva J, Martorella AJ, Lawlor K, Philip J, Fleisher M, Robbins RJ, et al. Serum peptidome patterns that distinguish metastatic thyroid carcinoma from cancer-free controls are unbiased by gender and age. *Mol Cell Proteomics.* 2006; 5:1840–52.
90. Rosenling T, Slim CL, Christin C, Coulier L, Shi S, Stoop MP, et al. The effect of preanalytical factors on stability of the proteome and selected metabolites in cerebrospinal fluid (CSF). *J Proteome Res.* 2009;8:5511–22.
91. Kader HA, Tchernev VT, Satyaraj E, Lejnine S, Kotler G, Kingsmore SF, et al. Protein microarray analysis of disease activity in pediatric inflammatory bowel disease demonstrates elevated serum PLGF, IL-7, TGF-beta1, and IL-12p40 levels in Crohn's disease and ulcerative colitis patients in remission versus active disease. *Am J Gastroenterol.* 2005;100:414–23.
92. Joos T, Bachmann J. Protein microarrays: potentials and limitations. *Front Biosci.* 2009;14:4376–85.
93. Andresen H, Bier FF. Peptide microarrays for serum antibody diagnostics. *Methods Mol Biol.* 2009;509:123–34.
94. Ray S, Mehta G, Srivastava S. Label-free detection techniques for protein microarrays: prospects, merits and challenges. *Proteomics.* 2010;10:731–48.
95. Zhu H, Bilgin M, Bangham R, Hall D, Casamayor A, Bertone P, et al. Global analysis of protein activities using proteome chips. *Science.* 2001;293:2101–5.
96. Liu Y, Wu J, Yan G, Hou R, Zhuang D, Chen L, et al. Proteomic analysis of prolactinoma cells by immuno-laser capture microdissection combined with online two-dimensional nano-scale liquid chromatography/mass spectrometry. *Proteome Sci.* 2010;8:2.