



Guess who? Taxonomic problems in the genus *Eiseniella* revisited by integrated approach

Irene de Sosa¹ · Daniel F. Marchán² · Marta Novo¹ · Tímea Szederjesi³ · Misel Jelic⁴ · Aleksandra Jabłońska⁵ · Raúl Navarro¹ · Ana Almodóvar¹ · Darío J. Díaz Cosín¹

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Abstract

Eiseniella neapolitana is a semi-aquatic, diploid earthworm that for many years was related to the cosmopolitan species *Eiseniella tetraedra* and even considered a subspecies of it. *Norealidys andaluciana* was described in Spain and is usually synonymized with *E. neapolitana*. We collected 69 specimens from Italy, Spain, and Cyprus and studied five molecular markers (COI, 16S, 28S, 12S, and ND1) and their morphology to solve this taxonomic problem. Phylogenetic analyses reveal the possible existence of two separate genera confounded under the name *Eiseniella*, but the study of more molecular markers and species of the genus would be necessary to confirm this. Therefore, the synonymy between *Eiseniella* and *Norealidys* is maintained. Various genetic analyses, including species delimitation, confirm the separation between *E. neapolitana* and *E. andaluciana* (= *N. andaluciana*) and excluded that *E. neapolitana* is a subspecies of *E. tetraedra*. The resemblance in external appearance despite clear genetic differences of the three species could be explained by convergent adaptation to the aquatic habitat. Despite the expected low haplotype diversity based on the 28S gene, we found a surprisingly high variability in the *E. andaluciana* (= *N. andaluciana*) population in Spain. However, its stable predicted secondary structure and its high content of G + C reject the presence of a pseudogene.

Keywords Earthworm · Genetic diversity · Morphology · Phylogeny · Semi-aquatic habitat

Introduction

The genus *Eiseniella* Michaelsen, 1900 includes two distinct groups of species, all of which belong to the riparian earthworms (Omodeo & Rota, 1991). The first group includes *Eiseniella tetraedra* (Savigny, 1826), a parthenogenetic and

cosmopolitan earthworm with morphological variability, but in general: quadrangular cross-section, less than 95 segments, male pore usually in XIII, tumid porophores, and unique position of the female pores below the line of *setae a* (Omodeo & Rota, 1991). This species showed a high genetic diversity in the Iberian Peninsula (de Sosa et al., 2017). Perel (1967) and Zicsi (1972) revised the other group. It includes about ten species, and they are also characterized by a quadrangular cross-section, the number of segments mainly ranging from 95 to 160, male pores without porophores, and female pores open dorsally to *setae b* or behind (Omodeo & Rota, 1991). The distribution of the individual species of this second group is geographically restricted (Omodeo & Rota, 1991). Currently, there are no data on the genetic diversity of these species.

Eiseniella neapolitana (Örley, 1885) belongs to the latter group. It is a semi-aquatic, circum Mediterranean and diploid earthworm with biparental reproduction (Omodeo, 1952). It was long considered a subspecies of *E. tetraedra* (Michaelsen, 1900; Stephenson, 1924; Bodenheimer, 1937; Cernovitov, 1938, 1940; Pop, 1952; Mršić, 1991; Pavlicek

✉ Irene de Sosa
iscarrasco@ucm.es

¹ Biodiversity, Ecology and Evolution Department, Faculty of Biology, Complutense University of Madrid, C/José Antonio Nováis 12, 28040 Madrid, Spain

² Centre d'Ecologie Fonctionnelle et Evolutive UMR 5175 CNRS 1919 Route de Mende, Cedex 5 34293 Montpellier, France

³ Medipredict Health Ltd, 4B Irinyi József Str, 1117 Budapest, Hungary

⁴ Department of Natural Sciences, Varaždin City Museum, Strossmayerovo šetalište 3, 42000 Varaždin, Croatia

⁵ Department of Invertebrate Zoology and Hydrobiology, University of Lodz, Banacha 12/16, 90-237, Łódź, Poland

et al., 2003). But already, Omodeo (1952, 1956), Omodeo and Rota (1991), and more recently Csuzdi and Pavlicek (2005) considered it a valid species.

Qiu and Bouché (1998) described *Reynoldsia andaluciana* on the basis of a few specimens from the locality Salobreña (Granada, Spain). The new genus, *Reynoldsia*, was characterized by rudimentary or absent of calciferous glands (Morren's glands) and typhlosole and by practically nonexistent nephridial vesicles (Qiu & Bouché, 1998). Blakemore (2008) did not accept *Reynoldsia* due to homonymy with a genus of Diptera and renamed it *Norealidys*. To date, *N. andaluciana* is considered a synonym of *E. neapolitana* in various databases such as DriloBASE (Drilobase Project, 2020), Earthworm species, a searchable database (<http://earthworm.uw.hu>; Csuzdi, 2012), or FADA database (<http://www.fada.biodiversity.be>; Balian et al., 2007). However, in Fauna Europaea (Rota & de Jong, 2015), they are treated as separate species.

In recent years, molecular tools are helping with taxonomic problems of earthworms (e.g., Huang et al., 2007; Marchán et al., 2018; de Sosa et al., 2019; Jiménez-Pinadero et al., 2021). Thus, a solution for the systematics of this group of species could arise from the study of molecular markers.

In their large-scale molecular phylogenetic analysis of Lumbricidae, Domínguez et al. (2015) found that the sole member of the genus in the study, *E. tetraedra*, appeared closely related to Iberian representatives of the genus *Eiseniona* Omodeo, 1956 (allocated by other authors in *Iberoscolex* Qiu & Bouché, 1998). The inclusion of new specimens and species of the *Eiseniella* and supposedly related genera in the phylogenetic analysis could help to prove or reject this hypothesis.

The aim of the present study is (1) to verify whether *E. neapolitana* is a valid species, distinct from *E. tetraedra*; (2) to investigate the genetic diversity of *E. neapolitana* for the first time; (3) to prove whether *N. andaluciana* is actually a synonym of *E. neapolitana*; (4) to place the genus

Eiseniella in the phylogenetic tree of lumbricids; and (5) to verify whether *Norealidys* should fall into synonymy with *Eiseniella* or not.

Material and methods

Earthworm sampling and morphological studies

We collected 55 specimens of *Eiseniella neapolitana* from nine localities in Italy (including the type locality) and one specimen from Cyprus. Also, we gathered thirteen specimens of *Norealidys andaluciana* from Salobreña (Granada, Spain) (type locality of the species) (Fig. 1, Supplementary Table 1). All individuals were collected by manual sorting in semi-aquatic areas, usually at the edge of bodies of water, washed in distilled water, fixed in 96% ethanol, and stored at -20°C in the earthworm collection of the Department of Biodiversity, Ecology and Evolution, Complutense University of Madrid (UCM-LT) with the exception of the specimen from Cyprus, which is housed in the Hungarian Natural History Museum. All newly collected material (except the Cypriot) and specimens of *E. tetraedra* from de Sosa et al. (2017) were examined morphologically with emphasis on length, weight after fixation, number of segments, position of the clitellum and the tubercula pubertatis, position of male pores, number and position of seminal vesicles and spermatheca, and form and position of the Morren's glands, typhlosole, and nephridial vesicles. In the case of *E. tetraedra*, only two specimens found in Naples, Italy, from the de UCM-LT collection (voucher numbers: 30851 and 30854) were examined for the last three characteristics. A portion of the posterior body section was collected and carefully cleaned under a stereomicroscope to remove gut and soil particles. Samples were then stored in ethanol and preserved at -20°C until DNA extraction.



Fig. 1 Localities sampled. GPS coordinates and number of specimens collected can be found in Supplementary Table 1. Specimens from Italy (3, 4, 5, 6, 7, 8, and 9) and Cyprus (2) belong to *E. neapolitana*. Specimens from Spain (1) belong to *N. andaluciana*

DNA extraction, amplification, and sequencing

Total genomic DNA was extracted using the SpeedTools Tissue DNA Kit (Biotools). Four mitochondrial markers, a fragment of cytochrome *c* oxidase subunit I (COI), 16S rRNA + tRNAs Leu, Ala and Ser (16S), 12S rRNA (12S) and NADH dehydrogenase (ND1), and one nuclear marker (a fragment of 28S rRNA) were amplified. For COI (632 bp), primer sequences, and polymerase chain reactions (PCR) followed (Pop et al., 2003). For 16S-tRNAs (774 bp) and 28S (805 bp), primer sequences and PCR conditions followed (Fernández et al., 2015). For 12S (974 bp) and ND1 (947 bp), primer sequences and PCR conditions followed (Pérez-Losada et al., 2015). Due to the unusual high variability found for the 28S gene (see the “Results” section), four primer pairs were designed to verify the sequences and exclude a possible pseudogene. However, none of the attempts worked.

All PCRs were specific and resolved via 1% agarose gel electrophoresis; they were visualized with GelRed stain (Biotium). All products were purified using ExoSAP-IT reagent (Thermo Fisher Scientific). PCR products were sequenced by Macrogen Spain Inc.

Data analyses

Sequences were aligned in MAFFT v.7 (Kato & Standley, 2013) using default settings and concatenated with BioEdit v7.0.9 (Hall, 1999). Two phylogenetic trees were constructed. The first contains sequences of the studied populations only (COI-16S + tRNAs + 28S), and the second considers the above populations and other lumbricid species (COI-16S + tRNAs + 28S + 12S + ND1) in order to place them in a wider phylogenetic context. Phylogenetic trees based on the concatenated sequences of COI-16S + tRNAs + 28S (2,211 bp) and COI-16S + tRNAs + 28S + 12S + ND1 (3296 bp) were constructed by the Bayesian Inference (BI) with MrBayes v3.1.2 (Ronquist & Huelsenbeck, 2003) and maximum likelihood (ML) using RaxML v7.03 software (Stamatakis, 2006) both implemented in CIPRES Science Gateway v.3.3 (Miller et al., 2010). The best-fitting substitution model selected by jModelTest2 (Darriba et al., 2012) for all markers in the short dataset was GTR + Γ + I, while for the longer dataset, HKY + Γ + I was chosen for 16S, and GTR + Γ was chosen for 12S. Maximum likelihood analysis with rapid bootstrapping was performed with 1000 replicates. Parameters in MrBayes were set to ten million generations, and 10,000 trees were sampled every 1000th generation, initiating the analysis with a random tree. Two independent analyses were performed, and 20% of the trees were discarded as burn-in. Sequences of each molecular marker for a representative of *E. tetraedra* (two sequences belonged to Clade I and three to Clade II (de Sosa et al., 2022) were

retrieved from Domínguez et al. (2015), Pérez-Losada et al. (2015), and de Sosa et al. (2017) and used as outgroup for the studied populations. In the tree containing other lumbricids, *Criodrilus lacuum* Hoffmeister, 1845 was used as an outgroup (Supplementary Table 2). Obtained phylogenetic trees were visualized using FigTree v1.3.1 (Morariu et al., 2008).

We calculated haplotype and nucleotide diversity for populations and genes using DNAsp v.6 (Rozas et al., 2017). We also estimated uncorrected pairwise distances for COI and 16S-tRNAs within and between our populations (*N. andaluciana*/*E. neapolitana*), as well as with *E. tetraedra* and two more lumbricids: *Dendrobaena byblica* Rosa, 1893 and *Iberoscolex oliveirae* (Rosa, 1894) (Supplementary Table 2). Haplotype networks based on each gene were constructed in PopART 1.7 (Leigh & Bryant, 2015) using the TCS inference method.

We employed two different methods for species delimitation with single locus (COI) analyses. Firstly, the Poisson Tree Processes model (PTP) (Zhang et al., 2013) was used to infer molecular clades based on our inferred molecular phylogeny. Secondly, we used bPTP, which is an updated version of the original maximum likelihood PTP. It adds Bayesian support values to delimited species on the input tree. Both analyses were conducted on the web server for PTP (available at <https://species.h-its.org/ptp>) using the MrBayes topology as advocated for this method (Zhang et al., 2013; Tang et al., 2014).

The RNAfold web server (Gruber et al., 2008) was used in order to estimate the secondary structure of the 28S rRNA for Spanish and Italian specimens and six other different species: *Aporrectodea trapezoides* (Dugès, 1828), *Allolobophora dubiosa* (Örley, 1881), *Dendrobaena octaedra* (Savigny, 1826), *D. byblica*, *I. oliveirae*, and *E. tetraedra* (Supplementary Table 2). Both MFE (minimum free energy) and centroid secondary structures were estimated. We calculated the G + C content of Spanish specimens by DNAsp v.6 (Rozas et al., 2017). Statistical analyses of morphological data were conducted in Statgraphics Centurion 19 (StatPoint Technologies Inc., USA). We used length, dry weight (after letting it drip on filter paper for 30 s), and number of segments of mature specimens to investigate differences between Spanish (*N. andaluciana*) and Italian (*E. neapolitana*) populations and *E. tetraedra* (de Sosa et al., 2017) through non-parametric analyses (Kruskal–Wallis) followed by Fisher LSD post hoc test.

Results

Phylogenetic analyses

Bayesian and Maximum Likelihood approaches reveal trees with congruent topology for the studied populations

(Fig. 2). Sequences are nested in three well-supported clades corresponding to each country studied.

Bayesian inference for the studied lumbricid species shows a polytomy for the genus *Iberoscolex*, *E. tetraedra*, and the studied populations of *N. andaluciana*/*E. neapolitana* (Fig. 3). The Spanish population (*N. andaluciana*) appears to be more related to the Italian and Cypriot populations (*E. neapolitana*) than to *E. tetraedra*, supporting the idea that they may all belong to a different genus, *Norealidys*, other than *Eiseniella*.

Genetic diversity and genetic divergence

Within *N. andaluciana*/*E. neapolitana*, a total of 17 haplotypes have been identified among 42 sequences for the COI gene, 22 haplotypes within 60 sequences for 16S-tRNAs, and 21 haplotypes within 68 sequences for 28S. Haplotype diversity (H) and nucleotide diversity (π) based on COI including all the specimens within the study are 0.91 and 0.11, respectively. H and π based on 16S-tRNAs are 0.90 and 0.04. Genetic diversity parameters based on 28S are 0.58 (H) and 0.01 (π). Values of haplotype and nucleotide diversity for each population are shown in Table 1. A remarkably high haplotype diversity based on 28S has been found for the Spanish population.

Genetic divergence among populations based on COI ranges from 1.57 to 17.81% (Table 2). Values above 9–15% fall into the ambiguous gap between intraspecific and interspecific divergence in earthworms suggested by Chang and James (2011) (which, on the other hand, was based on the slightly higher Kimura-2-parameters distances). Thus, if *E. neapolitana* from Italy seems to represent by itself a cryptic species complex, according to genetic divergence, the samples from the Spanish population, Cyprus, and Italy seem to belong to three different species. Values obtained for the populations based on 16S are in the range of 0.38 to 10.22% and also show a high variability (Table 2). Genetic distances between the studied populations of *N. andaluciana*/*E. neapolitana* and *E. tetraedra*, *D. byblica*, and *I. oliveirae* are also shown in Table 2. COI gene-based distances among *E. tetraedra* and the studied populations of *N. andaluciana*/*E. neapolitana* are higher than the threshold of 9–15% proposed by Chang and James (2011) for interspecific distances. Moreover, distances based on 16S are even higher, reaching about 30%.

The haplotype network for COI shows each population to be highly distant from other neighboring populations, with number of mutational steps from 14 (Italy 4–Italy 6) to 73 (Italy 5–Italy 6) (Fig. 4). Samples from different countries are separated from one another by a high number of mutational steps. Italy 5 is the most heterogeneous population. In contrast, the haplotype network based on 28S shows high

homogeneity for Italian haplotypes, whereas Spanish haplotypes show remarkably high differences, as evidenced in their haplotype diversity (Fig. 5). Here as well, populations from different countries appear well separated from one another.

To reject the amplification of a pseudogene in the sequences of 28S for the Spanish population (*N. andaluciana*), we examined the secondary structure of this gene in the studied populations and other Lumbricidae species (Fig. 6). The Italian specimens (*E. neapolitana*) show the same structure as other species such as *A. trapezoides*, *D. octaedra*, and *I. albolineatus*. However, the Spanish specimens, *A. dubiosa* and *D. byblica*, show different forms. The minimum free energy prediction of all sequences was low, ranged from –397 to –420 kcal/mol. The G + C content for the Spanish population is 66.50%.

Species delimitation

The two methods used to delineate species produced the same results (Fig. 7). They recognized seven different species. These analyses perfectly separate *E. tetraedra* and *E. neapolitana*, as well as the populations of the latter species (species 1, 2–5, and 6). For *E. neapolitana*, as many as four different species are delineated in Italy, coinciding with populations in the south (species 2), populations in the north (species 3), and populations in the center of the peninsula (species 4 and 5). Support values are presented in Table 3.

Morphological studies

Morphological characters of the specimens are shown in Table 4. Only mature individuals were used for the morphological analyses. The Cypriot specimen is not included in the morphological analysis because we only have the postclitellar region of the body. Spermatheca and male funnels are in 10 and 11 and are iridescent in all specimens. The clitellum position of all individuals from the Spanish population is 21–25, while the Italian populations show some variability in this trait. There is also some variability in the position of the *tubercula pubertatis* in the Italian populations, but in all Spanish specimens, they are 22–23–24. No variability has been found for the male pore, which is always located at segment 15.

The Italian specimens show the Morren's glands in 10–14 with diverticula in 10, lamellae in 11–13, and little distinct in 14, while none of these structures are observed in the individuals from the Spanish population (except for a very doubtful structure in 11, which should be confirmed histologically) (Fig. 8A; compare with Fig. 8B: *E. tetraedra*). In the nephridia, the glandular part is broadened both in the Italian and Spanish specimens, and a well-differentiated nephridial vesicle does not appear, but the nephridial tube gradually tapers to open into the nephridial pore (Fig. 8C;

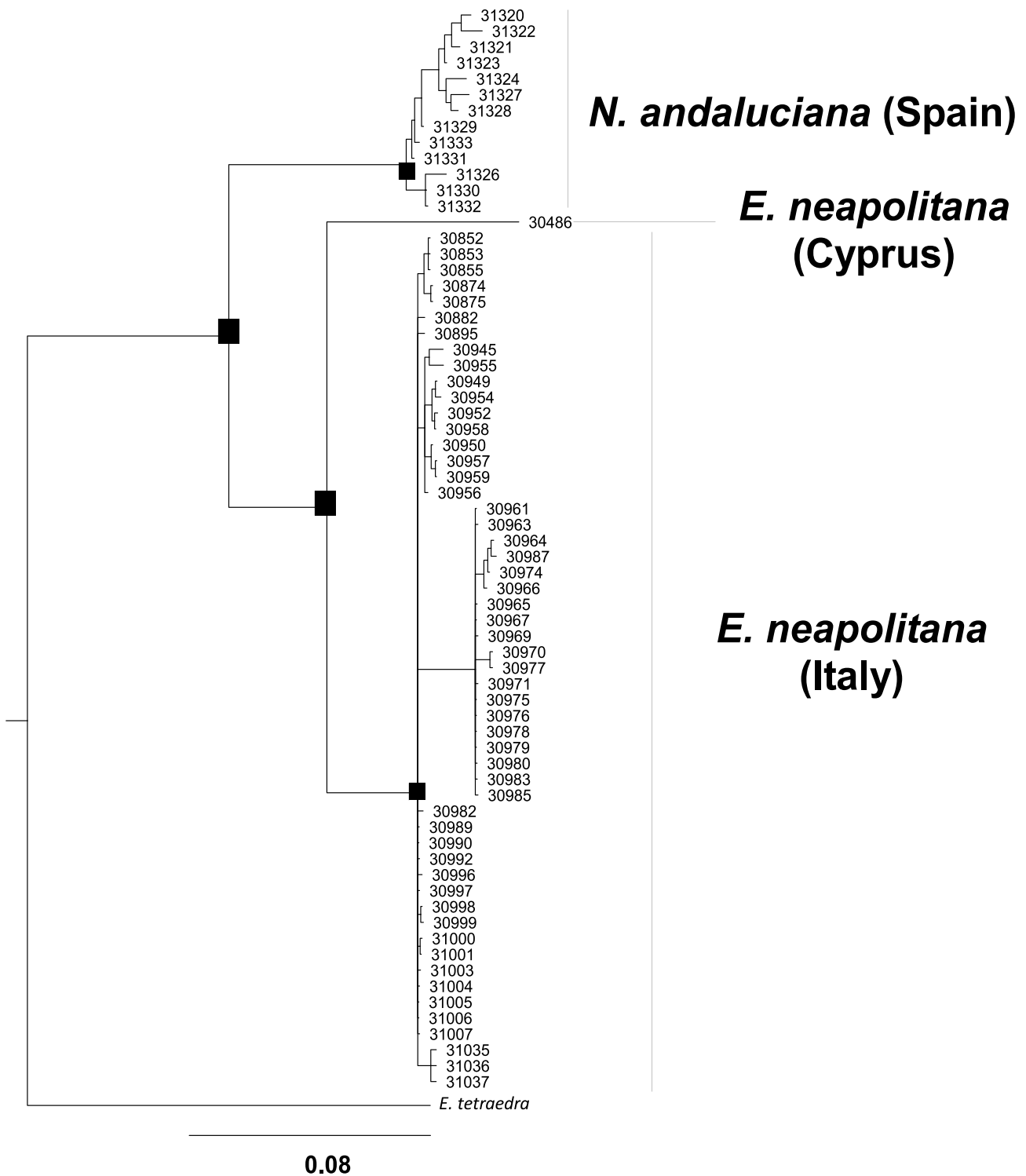


Fig. 2 Bayesian inference (BI) of the phylogenetic tree based on conj-Posterior probability/bootstrap support values (of maximum likelihood analysis, ML) are shown as black squares when higher than 0.9/0.7

(BI/ML). The scale bar represents 0.08 substitutions per position. Sample numbers correspond to references in the UCM-LT collection and can be found at Supplementary Table 1

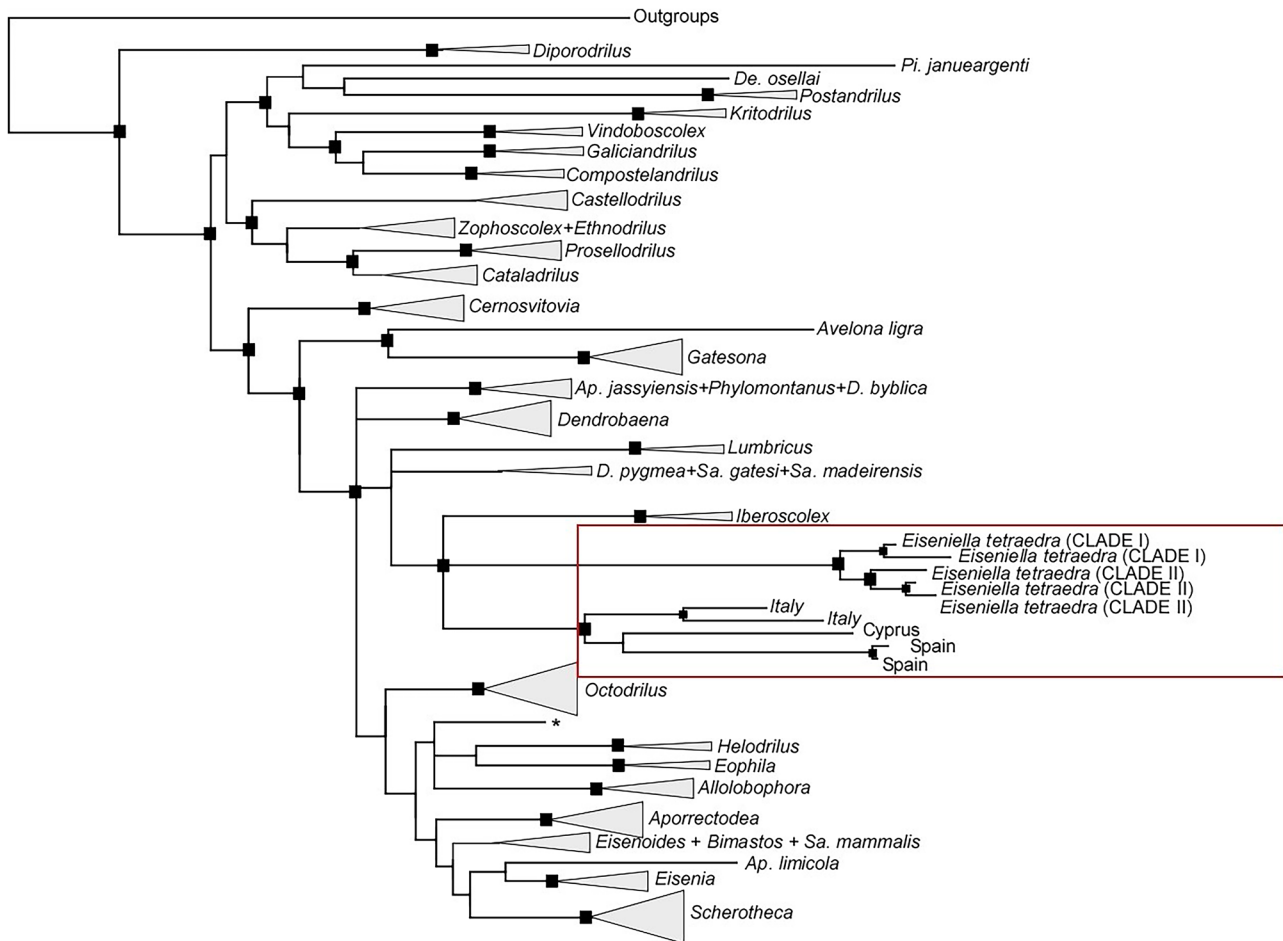


Fig. 3 Bayesian inference (BI) of the phylogenetic tree of lumbricids based on concatenated sequences of COI, 16S, 28S, 12S, and ND1 (long dataset). Posterior probability/bootstrap support values (of maximum likelihood analysis, ML) are shown as black squares when higher than 0.9/0.7 (BI/ML). Gray triangles represent two or more species of a genus-level clade collapsed to facilitate interpretation of

the figure. Asterisk represents several singleton species in an unresolved polytomy: *Aporrectodea rosea*, *Aporrectodea georgii*, *Panoniona leoni*, *Allolobophora bartolii*, *Helodrilus patriarchalis*, and *Proctodrilus antipai*. Specimens from Italy and Cyprus belong to *E. neapolitana*. Specimens from Spain belong to *N. andaluciana*. Complete names of species are shown in Supplementary Table 2

compare with Fig. 8D: *E. tetraedra*). The typhlosole is present in all specimens. It is small and simple at first, and then

Table 1 Haplotype and nucleotide diversity values of each population studied. NA indicates populations with only one specimen and sequence. Hyphens indicate no sequences for a gene

Populations	COI (H/ π)	16S (H/ π)	28S (H/ π)
Spain	0.69/0.005	0.67/0.004	0.98/0.01
Cyprus	NA	NA	NA
Italy 1	-	0	0
Italy 2	-	0.66/0.004	0.66/0.0008
Italy 3	NA	NA	NA
Italy 4	0.90/0.01	-	0.2/0.0002
Italy 5	0.92/0.01	0.52/0.005	0
Italy 6	0.53/0.0008	0	0.27/0.0003
Italy 7	-	-	0

a slightly pronounced central furrow appears, giving it a certain bilobed appearance (Fig. 8E; compare with Fig. 8F: *E. tetraedra*).

No statistically significant differences ($p > 0.05$) are found between the Spanish and Italian populations and *E. tetraedra* for length and weight. In contrast, a number of segments show statistically significant differences ($p = 0.002$) between two homogenous groups: Italian and Spanish populations vs. *E. tetraedra* (Fig. 9).

Discussion

Eiseniella neapolitana was described by Örley (1885) as *Allurus neapolitanus* and considered by many authors (Michaelsen, 1900; Stephenson, 1924; Bodenheimer, 1937; Cernosvitov, 1938, 1940; Pop, 1952; Mršić, 1991; Pavlicek

Table 2 Percentage of uncorrected pairwise distances based on COI (above diagonal) and 16S (below diagonal) between the studied populations and three different species: *Eiseniella tetraedra*, *Dendrobaena*

byblica, and *Iberoscolex oliveirae*. Hyphens indicate no sequences for a gene in one or two of the compared populations

16S/COI	Spain	Cyprus	Italy 1	Italy 2	Italy 4	Italy 5	Italy 6	<i>E. tetraedra</i>	<i>I. oliveirae</i>	<i>D. byblica</i>
Spain	0.57/0.43	16.04	-	-	12.17	17.81	15.14	20.51	18.26	19.44
Cyprus	10.22	0/0	-	-	10.09	16.33	13.13	20.61	17.87	18.67
Italy 1	7.33	7.49	-/0	-	-	-	-	-	-	-
Italy 2	7.24	7.62	0.38	-/0.43	-	-	-	-	-	-
Italy 4	7.62	7.5	1.01	0.89	0.49/0.9	9.11	1.57	16.27	13.38	14.14
Italy 5	7.89	8.62	5.09	5.17	5.28	1.52/0.55	11.85	21.38	18.17	19.24
Italy 6	7.46	7.49	0.51	0.47	0.70	5.22	0.83/0	20.34	17.29	16.98
<i>E. tetraedra</i>	29.75	29.47	29.51	29.52	29.36	29.33	29.46	7.10/0.93	19.20	19.50
<i>I. oliveirae</i>	31.07	31.26	30.74	29.58	30.90	30.70	31.00	5.98	0/0	12.65
<i>D. byblica</i>	29.78	29.97	29.45	29.58	29.64	29.28	29.71	5.22	3.22	0/0

et al., 2003) as a subspecies of *Eiseniella tetraedra*, while Omodeo (1952, 1956), Omodeo and Rota (1991), and, more recently, Csuzdi and Pavlicek (2005) claimed it a valid

species. The holotype of *E. neapolitana* was collected by Örley (1885) in Sebeto River (Napoli, Italy). Our specimens from Italy 1 (i.e., locality 3 in Fig. 1) originate from the type

Fig. 4 Haplotype network based on COI. Each black circle represents a different haplotype; their size is proportional to the number of samples belonging to that haplotype. Each perpendicular line indicates a mutational step; if there were more than four, it is shown with the number of mutational steps. White circles are hypothetical intermediate haplotypes. Branch length does not contain any information

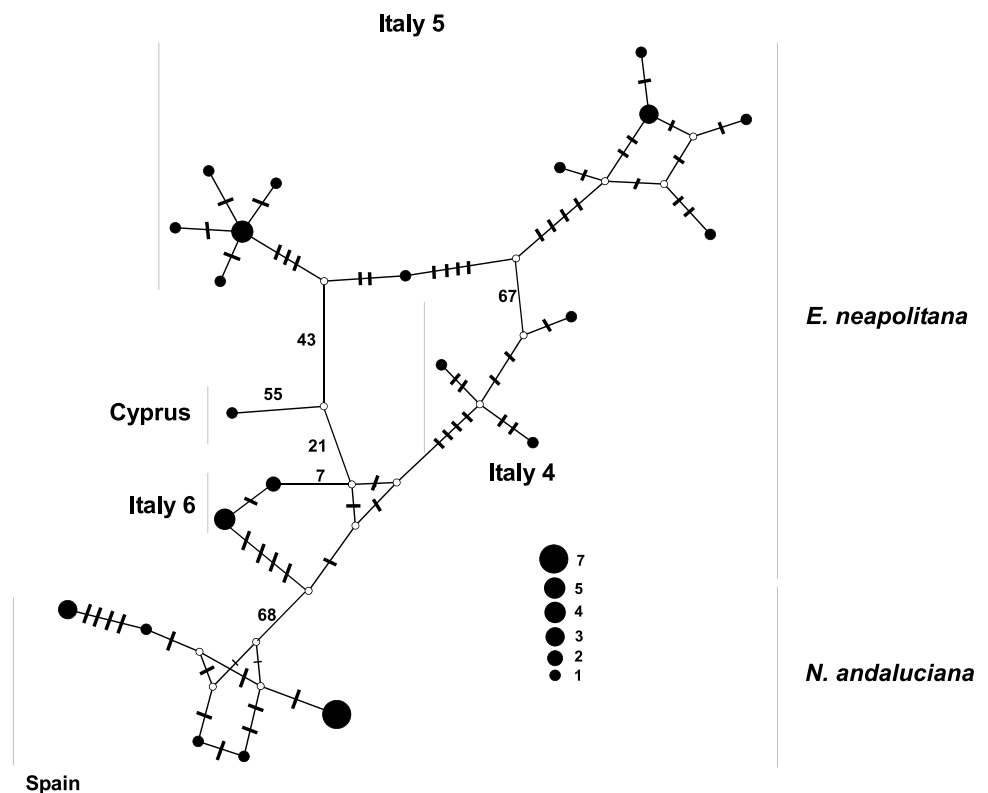
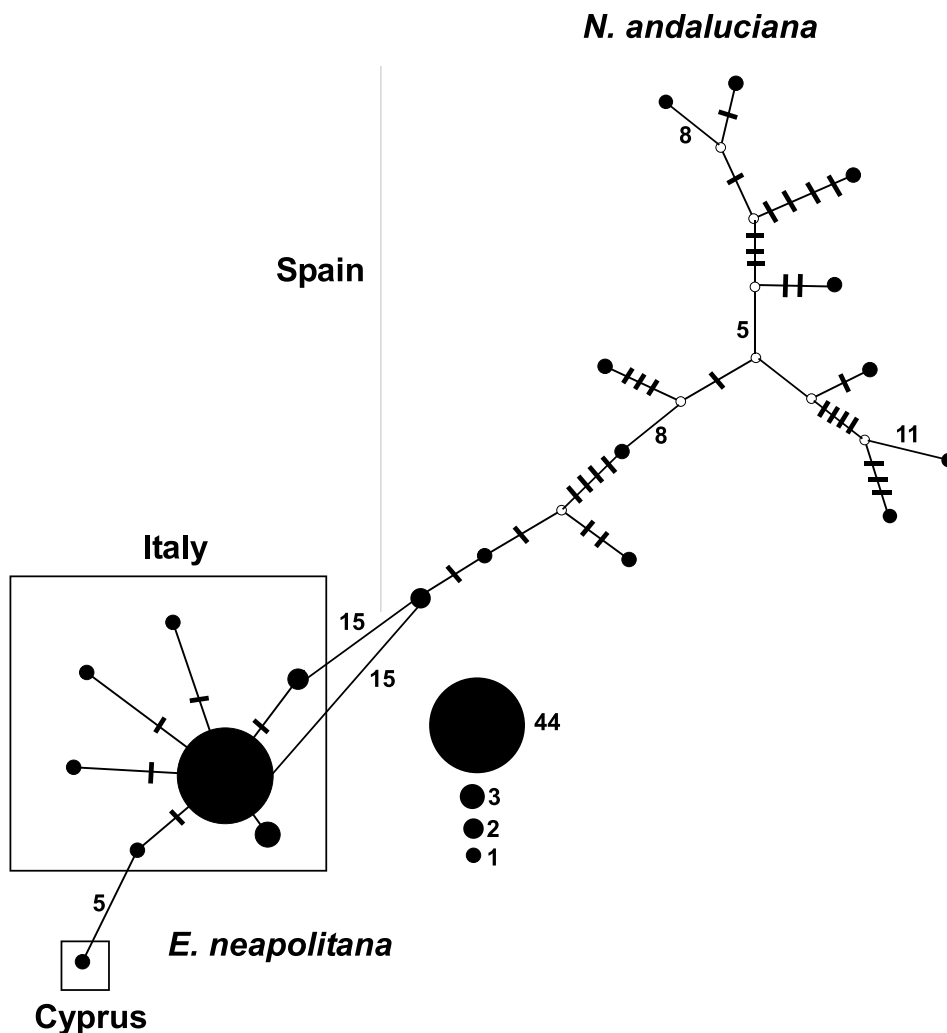


Fig. 5 Haplotype network based on 28S. Each black circle represents a different haplotype; their size is proportional to the number of samples belonging to that haplotype. Each perpendicular line indicates a mutational step, if there were more than four it is shown with the number of mutational steps. White circles are hypothetical intermediate haplotypes. Branch length does not contain any information



locality. However, all Italian specimens of *E. neapolitana* collected in this study correspond to the morphological characters indicated by Örley (1885) as diagnostic for this species.

The genus *Reynoldsia* was established by Qiu and Bouché (1998). This name entered into homonymy with the Diptera *Reynoldsia* Malloch, 1934, so Blakemore (2008) replaced it with the name *Norealidys*, with the only species *N. andaluciana*. This genus was discovered in Salobreña, in the south of Spain, on the sea coast, in a sugar cane cultivation, on a sandy soil rich in organic matter, and very moist (Qiu & Bouché, 1998). Our specimens of the Spanish population were collected in the same locality. Currently, the sugar cane fields have disappeared, so the specimens were searched for in the area where this culture used to be found, on a riverbank with sandy and moist soil that was not flooded. As for the morphological details, the position of the clitellum and the *tubercula pubertatis* of the studied individuals are consistent with those diagnostic for *N. andaluciana*. The external appearance is very similar to that of the genus

Eiseniella. The anatomical traits of *N. andaluciana*, namely the absence or reduction of the Morren's glands, typhlosole, and nephridial vesicles, probably indicating an adaptation to a humid environment (Qiu & Bouché, 1998) are all found in the individuals collected by us at the Salobreña site, so we can conclude that they do indeed belong to this species. Qiu and Bouché (1998) described the species as earthworms without skin pigmentation and of small size (between 47 and 72 segments). Our studied individuals were pigmented and had a larger number of segments. This contradiction could be due to the fact that only two individuals were used to describe the species, and they were not sufficient to capture the intraspecific variability of *N. andaluciana*.

Specimens from Italy (*E. neapolitana*) and Spain (*N. andaluciana*) were nested in the same clade in our phylogenetic tree, but with long and well-separated branches. The genetic distances among them, as well as the distances from the other species studied (*E. tetraedra*, *D. byblica*, and *I. oliveirae*), are high (from 16.27 to 21.38%). However, Szederjesi et al. (2018) proved that within *Eisenia lucens*

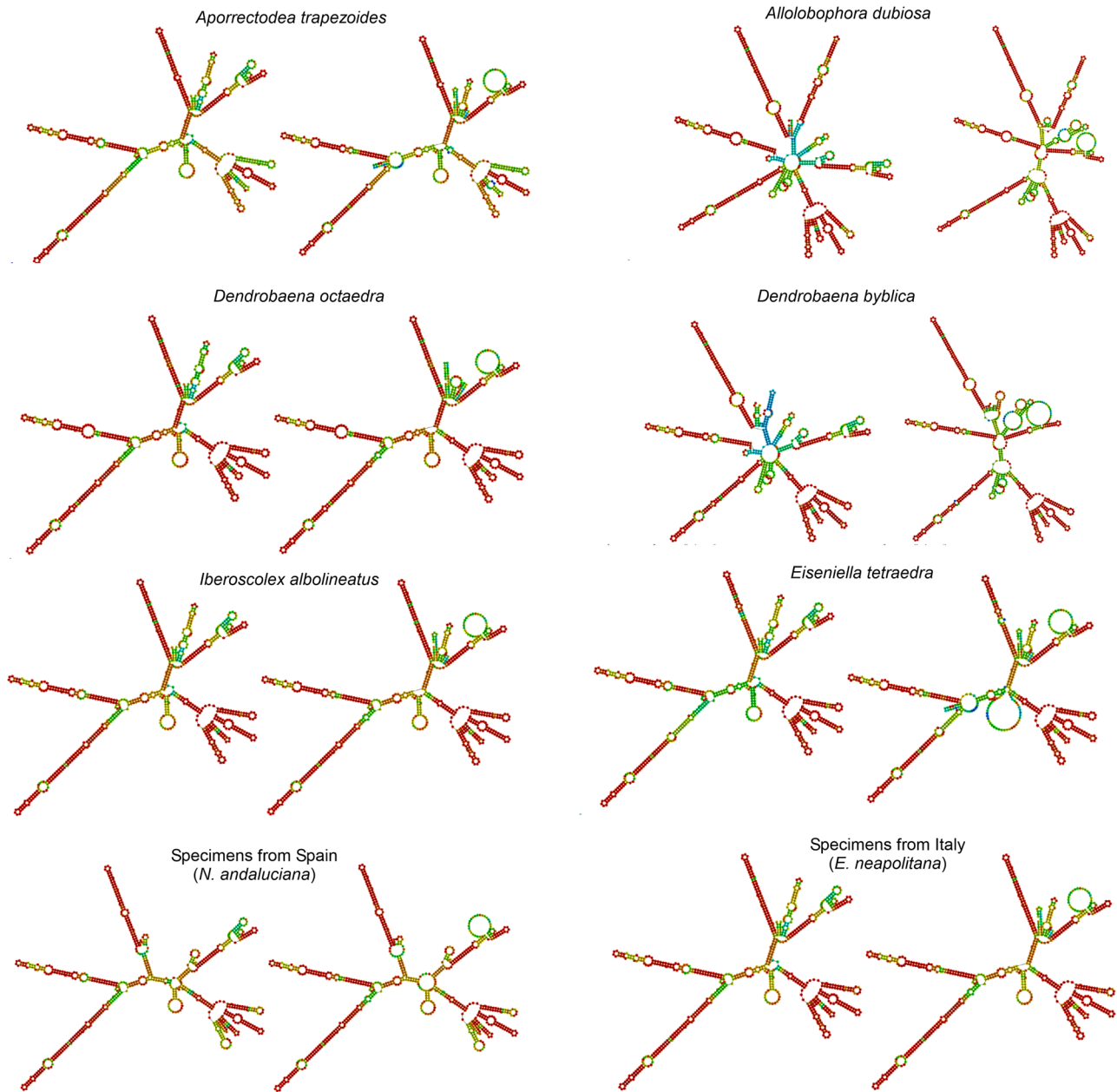


Fig. 6 Predicted secondary structures of 28S gene for Spanish and Italian specimens from the studied populations and of six different species: *A. trapezoides*, *A. dubiosa*, *D. octaedra*, *D. byblica*, *I. albo-*

lineatus, and *E. tetraedra*. The structure is colored according to the probability of base pairing: high probability in red, medium probability in green and yellow, and low probability in blue

(Waga, 1857) genetic distances can be higher than 19%. These authors also highlighted that genetic distances are rather clade-specific; therefore, setting a general threshold is not possible. Nevertheless, species delimitation analyses confirmed that *E. tetraedra* is a separate species from *E. neapolitana*. On the other hand, they also showed that *N. andaluciana* is a different species from *E. neapolitana*, which invalidates the proposal of synonymy between these two species. Moreover, the specimen from Cyprus seems to represent a species distinct from *E. neapolitana*. However,

since it is a single specimen and we only had the posterior section of its body, we cannot describe or name this species until we have examined more material from this population. Finally, these analyses and the haplotype networks demonstrate the status of species complex of *E. neapolitana* in Italy, which needs to be further investigated.

The phylogenetic tree distinguishes two different clades comprising, on one hand, *E. tetraedra* and on the other hand all the other populations studied in this work. Based on the depth of the branches, one can assume the existence of two

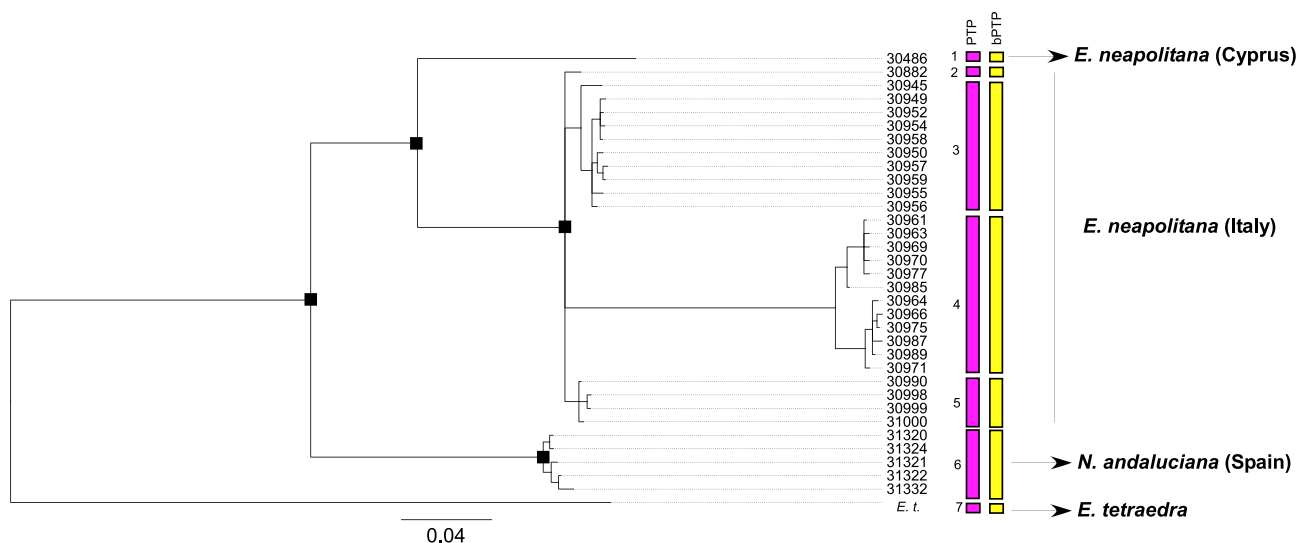


Fig. 7 Bayesian inference (BI) of the phylogenetic tree based on sequences of COI of the studied populations and *E. tetraedra*. Posterior probability is shown as black squares when higher than 0.9. The scale bar represents 0.04 substitutions per position. Sample numbers correspond to references in the

UCM-LT collection and can be found at Supplementary Table 1. Results of the PTP model are shown in pink vertical lines. Results of the bPTP model are shown in yellow vertical lines

different genera. However, other molecular markers that could resolve the polytomy obtained by our phylogenetic analyses need to be investigated in addition to the other species within the genus *Eiseniella*. Thus, the most conservative stance is to maintain the synonymy between the genera *Eiseniella* and *Norealidys* in the meantime.

The genus *Eiseniella* appears in the phylogenetic tree in a polytomy with *Iberoscolex* (= *Eiseniona*), a genus whose relationships with the other lumbricids and internal phylogeny will be the subject of future studies. Although the general appearance of *Eiseniella* species is very similar, such as the quadrangular posterior section with dorsal groove and the anterior position of the clitellum, there are differences in the position of clitellum and tubercula pubertatis. We also found significant differences in the number of segments between *E. neapolitana* and *E. andaluciana* (= *N. andaluciana*) with *E. tetraedra*. Their similar external appearance (small size, anterior clitellum, and posterior dorsal sulcus) could be due to convergence to aquatic or

semi-aquatic habitat. Earthworms from the semi-aquatic family Almidae also present quadrangular body at the posterior end and round body at the anterior and an anterior clitellum (Chanabun et al., 2020). Other semi-aquatic species, such as those of the family Criodrilidae, also share some of these morphological characteristics (Blakemore, 2008; Omodeo, 1984). However, the most closely related genus in the phylogenetic tree, *Iberoscolex* (= *Eiseniona*), does not present these characters. Therefore, these traits do not appear to have a phylogenetic signal, being shared by species from different families.

E. neapolitana showed well-developed Morren’s glands, which would place it close to *E. tetraedra*. However, molecular markers placed it together with *E. andaluciana* (= *N. andaluciana*), which tends to lack or underdevelop these structures (Qiu & Bouché, 1998). Future studies could resolve these inconsistencies and show whether structures such as the Morren’s glands could correspond to analogous organs and what evolutionary pressures intervene in their development, similar to those indicated by Marchán et al. (2016) for the typhlosole of the Hormogastridae.

As a result of concerted evolution, all copies of rDNA families are generally rapidly homogenized within individuals and species, but interspecific divergence can be high (Hillis & Dixon, 1991). Intraspecific divergence has been found to be high in animals such as corals (Márquez et al., 2003), grasshoppers (Keller et al., 2006), and nematodes (Hugall et al., 1999; Pereira & Baldwin, 2016). At protein-coding loci, pseudogenes can be detected by the presence of stop codons and frame shifts, the absence of substitution bias at third position, or the changes in otherwise invariant amino

Table 3 Support values of species delimitation analyzes (PTP and bPTP). The delimited species numbers are the same as those used in Fig. 7

Delimited species	PTP	bPTP
1	1	1
2	1	1
3	0.88	0.76
4	0.86	0.55
5	0.94	0.94
6	0.94	0.99
7	1	1

Table 4 Morphological traits for mature specimens from Italy and Spain

ID UCM-LT	Locality	Length (mm)	Dry weight (mg)	N° of segments	Clitellum	<i>Tubercula pubertatis</i>	Male pore	Seminal vesicles	Spermatheca	Male funnels
31,320	Spain	40	180	127	21–25	22–23–24	15	9–10–11–12	Iridescent	Iridescent
31,321	Spain	37	150	118	21–25	22–23–24	15	9–10–11–12	Iridescent	Iridescent
31,324	Spain	25	80	101	21–25	22–23–24	15	9–10–11–12	Iridescent	Iridescent
31,329	Spain	30	110	102	21–25	22–23–24	15	9–10–11–12	Iridescent	Iridescent
31,332	Spain	31	100	103	21–25	22–23–24	15	9–10–11–12	Iridescent	Iridescent
31,333	Spain	29	80	100	21–25	22–23–24	15	9–10–11–12	Iridescent	Iridescent
30,853	Italy 1	52	226.8	168	21–25	21–22–23–24	15	9–10–11–12	Iridescent	Iridescent
30,964	Italy 5	21	45.2	97	21–26	22–23–24	15	9–10–11–12	Iridescent	Iridescent
30,966	Italy 5	30	45.8	132	21–26	22–23–24–25	15	9–10–11–12	Iridescent	Iridescent
30,976	Italy 5	35	75.7	135	21–27	22–23–24–IN25	15	9–10–11–12	Iridescent	Iridescent
30,997	Italy 6	33	114	114	21–25	22–23–24–25	15	9–10–11–12	Iridescent	Iridescent

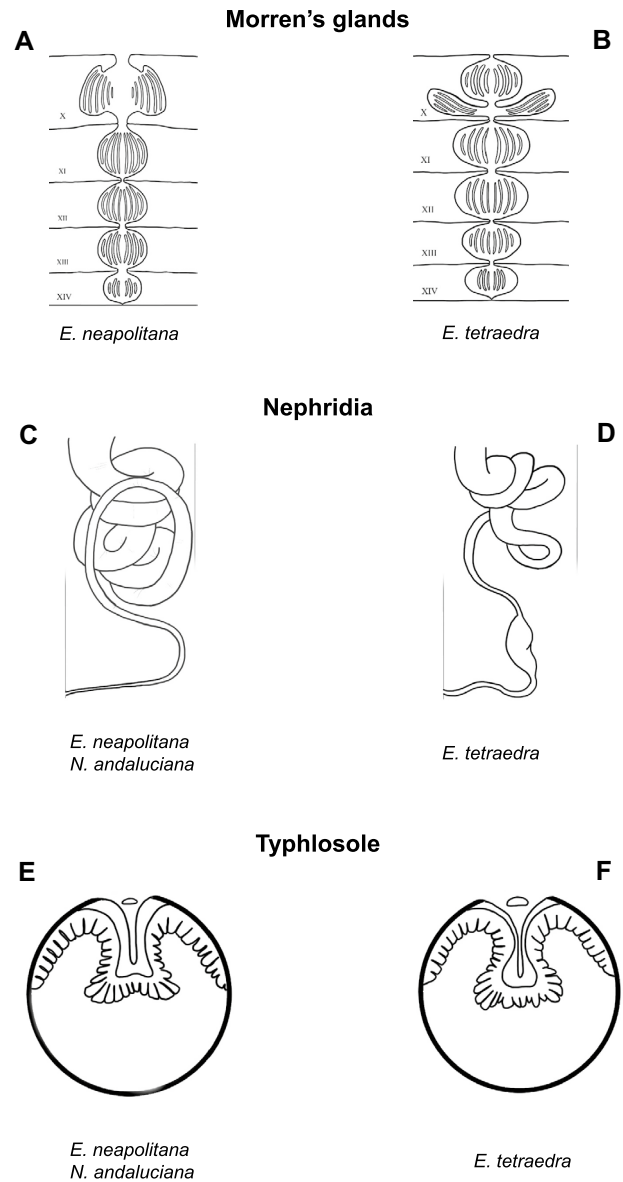


Fig. 8 Illustrations of internal anatomy of specimens studied

acid residues (e.g., Arctander, 1995). These indicators cannot be used for nonprotein-coding genes (such as 28S rRNA); however, selection acts to preserve the secondary structure of functional RNA molecules. Current methods for predicting RNA secondary structure are mainly based on the minimum free energy algorithm, which finds the optimal folding state of RNA in vivo using an iterative method to satisfy the minimum energy or other constraints (Zhang et al., 2019). In general, secondary structure stability (low free energies) and pattern of nucleotide substitution in other ribosomal genes such as 5.8S or ITS appeared to be the most powerful approaches to distinguish putative pseudogenes from presumed functional sequences (Razafimandimbison et al., 2004). Thus, the presence of a stable predicted secondary structure (Pereira &

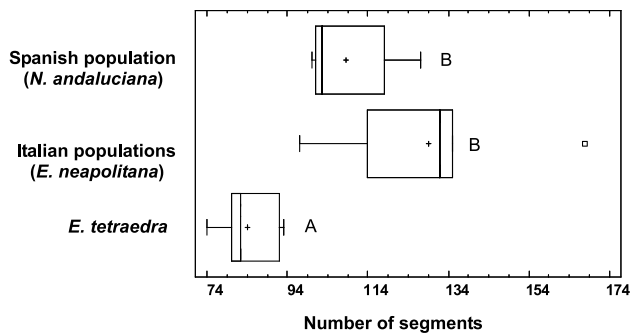


Fig. 9 Box-plot diagram of number of segments in the studied populations and *E. tetraedra*. Different letters indicate different groups in multiple range tests

Baldwin, 2016) and high G+C content (Zheng et al., 2008) do not support the presence of pseudogenes. Therefore, the high intraspecific divergence for the 28S rRNA gene in the studied Spanish populations may not be explained by the presence of pseudogenes. The mode of reproduction has also been associated with rRNA heterogeneity. Cross-fertilization may increase intraspecific variation due to recombination in the cyclic parthenogenetic *Daphnia pulex* Leydig, 1860 (Crease & Lynch, 1991) and in polyploid obligate mitotic parthenogens of the nematode genus *Meloidogyne* (Hugall et al., 1999). Although the studied Spanish population appears to be sexual based on the presence of sperm in the spermatheca, suggesting copulation with another individual, laboratory experiments are required to investigate whether it may also be parthenogenetic. Therefore, its hybrid origin could explain the hypervariability in the 28S rDNA.

Differences in predicted secondary structure of 28S rRNA between different earthworms' species may be due to hypervariability rather than pseudogenes or lack of function. They also do not appear to have a phylogenetic signal, as species of the same genus have different structures.

Conclusions

Phylogenetic analyses based on several markers, genetic divergence based on COI, and species delimitation analyses support the validity of *E. neapolitana* and *N. andaluciana* as two distinct species. They also invalidate *E. neapolitana* as a subspecies of *E. tetraedra*. *E. neapolitana* also appears as a cryptic complex in the studied populations. According to the phylogenetic analysis, we could hypothesize the existence of two different genera. However, the molecular tree shows the studied species in an unresolved polytomy position, which indicates that the markers used have no power enough to resolve their phylogenetic relationship; the dataset should be increased by adding further molecular markers and the remaining species of the genus *Eiseniella* seeking to resolve

the polytomy recovered in our analyses. Thus, the synonymy between the genera *Eiseniella* and *Norealidys* is maintained pending further confirmation. The high variability of the 28S rRNA gene for *E. andaluciana* (= *N. andaluciana*) could not be explained by the presence of pseudogenes due to its stable secondary structure and high G+C content.

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Availability of data and material We collected 55 specimens of *Eiseniella neapolitana* from nine localities in Italy (including the type locality), thirteen specimens of *Norealidys andaluciana* from Salobreña (Granada, Spain) (type locality of the species), and one specimen from Cyprus (Supplementary Table 1). Voucher specimens were deposited in the earthworm collection of the Department of Biodiversity, Ecology and Evolution, Complutense University of Madrid (UCM-LT). In addition, all nucleotide sequences generated in this study were deposited in the GenBank (Supplementary Table 2).

Declarations

Consent for publication All authors approved the final version of the manuscript for publication.

Conflict of interest The authors declare no competing interests.

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