



Optical biopsy in gastroenterology: Focus on confocal laser endomicroscopy

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Introduction

The quality of endoscopy images has improved remarkably over the last several decades. Current generation endoscopes provide high-definition images with superior resolution and are capable of magnification as well. This means that subtle lesions are less likely to be missed on routine endoscopy. However, high definition alone may not be superior to standard white light endoscopy for in vivo characterization of polyps [1]. Differentiation of the lesions as neoplastic vs. non-neoplastic is essential as it bears implication on subsequent management and surveillance strategies. For example, hyperplastic polyps bear no malignant potential. On the other hand, adenomatous polyps harbor a definite risk of malignant transformation and need to be resected. The integration of image enhancement techniques like narrow band imaging (NBI) in the endoscopes have largely overcome the issue of discriminating neoplastic and non-neoplastic polyps in the gastrointestinal (GI) tract. Novel image enhancement techniques allow better characterization of the lesions as compared with high-definition endoscopy alone [2].

Image enhanced and optical endoscopic techniques

Conventional white light endoscopy is largely inaccurate (accuracy 59% to 84%) in differentiating neoplastic from non-neoplastic polyps. The incorporation of optical techniques in the currently available endoscopes has largely overcome the unmet need generated by standard or high-definition white light endoscopy. Current endoscopic image enhancement techniques

include NBI (Olympus Medical Co., Tokyo, Japan), i-scan (Pentax Hoya, Tokyo, Japan), flexible spectral imaging color enhancement (FICE, Fujifilm Medical Co., Tokyo, Japan), and blue laser imaging (BLI, Fujifilm Medical Co., Tokyo, Japan). Other techniques in various phases of development include autofluorescence imaging (AFI, Olympus Medical Co, Tokyo, Japan), endocytoscopy, confocal laser endomicroscopy (CLE, Cellvizio System Mauna Kea, Paris, France), optical coherence tomography, multiphoton microscopy, and Raman spectroscopy.

In this issue of the *Journal*, one such technique, i.e. probe-based CLE (pCLE), was evaluated for the characterization of GI lesions in the upper and lower GI tracts [3]. In the following section, we will discuss the current status and the future directions with respect to the applications of CLE in digestive tract.

Confocal laser endomicroscopy: principle and technique

In CLE, a low-power laser is used to illuminate the tissue of interest. The fluorescence of the light subsequently deflected from the tissue is refocused by the same lens onto the confocal detection system. The presence of a pinhole ensures that only the light reflected from a specific plane gets detected and processed further, whereas the light deflected from other planes and not in line with the pinhole is rejected (Fig. 1). This characteristic feature of CLE enables high spatial resolution and evaluation of tissue architecture at cellular level [4].

There are two basic systems of CLE: endoscope-integrated system (eCLE) and probe-based system (pCLE). Dedicated endoscopes (EC3870K, Pentax, Tokyo, Japan) have integrated CLE system at the tip of the endoscope. On the other hand, pCLE system (Cellvizio Endomicroscopy System; Mauna Kea Technologies, Paris, France) utilizes miniature probes (0.9–2.5-mm diameter), which can be passed through the accessory channel of most of the commercially available endoscopes. These probes are essentially composed of a fiberoptic bundle

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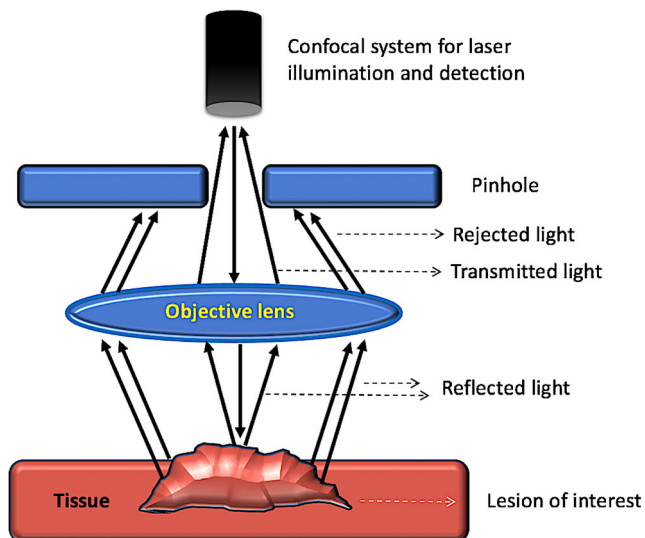


Fig. 1 Principle of confocal laser endomicroscopy

with integrated distal lens and can be used approximately 20 times [4]. Various types of confocal miniprobes are available for use in the GI tract including GastroFlex ultra high definition (UHD) (upper GI tract), ColoFlex UHD (colon), CholangoFlex (ERCP), and AQ-Flex 19 (needle-based CLE).

The advantages of eCLE include provision of a wider field of view and an adjustable depth of scanning (0–250 μm) as compared with pCLE, in which the depth of scanning ($\sim 50 \mu\text{m}$) is fixed depending on the type of probe being used. On the other hand, image acquisition rate is faster with pCLE (12 frames/second) as compared with eCLE (0.8–1.6 frames/second). eCLE system is no longer available for commercial use, and therefore, most studies have evaluated the pCLE system in the GI and hepatobiliary tract.

Contrast agents for CLE

CLE requires the administration of topical (acriflavine hydrochloride and cresyl violet) and/or intravenous (fluorescein sodium and indocyanine green) fluorescent contrast agents immediately before the procedure. Acriflavine hydrochloride strongly labels the nuclei and superficial epithelial cells, whereas fluorescein sodium does not stain nuclei but enables deeper imaging by highlighting the vasculature, the lamina propria, and the intercellular spaces [5]. Besides the less favorable staining characteristics, there are concerns over the mutagenic potential of acriflavine. Therefore, intravenous fluorescein is generally preferred over the topical contrast agents for CLE.

Technique of pCLE

In pCLE, the confocal miniprobe is passed through the working channel of the standard endoscope and placed gently in

contact with the tissue of interest after the administration of the contrast agent (2.5–5.0 mL of 10% fluorescein sodium). The probe should be perpendicular to the tissue for better image acquisition. Since blood and mucus can interfere with the quality of imaging, the area of interest should be washed and care should be taken while placing the probe on friable tissues. For the same reason, biopsies should be avoided before CLE examination. Optimal images can be obtained from 30 s to 8–10 min (up to 60 min) after the administration of contrast agents. The images can also be saved and stored for subsequent offline analysis and interpretation.

Current and potential applications of CLE in the GI tract

In the GI tract, CLE has been evaluated for the following indications: identification of dysplasia in Barrett's esophagus [6], classification of gastric and colorectal polyps (Table 1), assessment of disease activity and development of dysplasia in inflammatory bowel disease [18, 19], evaluation of indeterminate biliary strictures [20], and characterization of solid and cystic pancreatic masses [21, 22]. In addition, limited data suggests that CLE may be potentially useful for the *in vivo* identification of celiac disease (villous atrophy and increased intraepithelial lymphocytes) [23], ampullary lesions [24], and follow up after endoscopic mucosal resection (EMR) of colorectal polyps [25].

The subsequent discussion will focus on the application and utility of CLE for polypoidal lesions in the GI tract.

CLE for polypoidal lesions in the GI tract

CLE provides real-time histology of the polyps in both the upper and lower GI tracts. It enables the differentiation of hyperplastic or non-neoplastic polyps from neoplastic polyps with a reasonable degree of accuracy. There are two main classification systems, which are utilized to characterize the dysplastic and non-dysplastic GI epithelium, i.e. Mainz and Miami classification systems.

CLE for gastric polyps

CLE has been found useful and accurate in characterizing gastric lesions like intestinal metaplasia, gastric intraepithelial neoplasia, adenomas, and cancers. However, there is no well-defined classification system for gastric polyps or lesions, and experts have utilized the cellular morphology and the architecture of glands and vessels to differentiate between different gastric lesions [10]. In the normal gastric mucosa, the glands and epithelial cells are regularly arranged, homogenous with good polarity, and the

Table 1 Outcomes of confocal laser endomicroscopy in the gastrointestinal tract (gastric and colonic)

Study	Number of lesions (number of patients)	GI tract	Size (mean or median)	Sensitivity/specificity	Accuracy
Hurlstone et al. (2008)* [7]	162 (39)	Colon	1–34 mm	97.4%/99.3%	99.1%
Gomez et al. (2010) [8]	75 (53)	Colonic	NA	76%/72%	75%
De Palma et al. (2010) [9]	32 (20)	Colorectal	13 mm	100%/84.6%	92.3%
Li et al. (2011) [10]	182 (phase I)	Gastric lesions	21 mm	84%/92.1%	88.5%
	1786 (phase II)		14 mm	88.9%/99.3%	98.8%
Jeon et al. (2011) [11]	35 (31)	Gastric lesions	NA	NA	94.2%
Shahid et al. (2012) [12]	130 polyps (65)	Small colorectal polyps	4.6 mm	86%/78%	82%
Kuiper et al. (2012) [13]	154 (64)	Colorectal lesions	5.0 mm	59.5%/77.4%	71.9%
Buchner et al. (2010) [14]	119 (75)	Colonic polyps	10 mm	91%/76%	91%
Gong et al. (2015) [15]	86 (82)	Gastric lesions	16.7 mm	91.9%/90%	93.5%
Belderbos et al. (2017) [16]	113 (52)	Colonic polyps	8 mm	88%/NA	76%
Chen et al. (2018) [17]	322	Gastric lesions	NA	72.4%/93.1%	88.9%
Current study [3]	50 (50)	Upper and lower GI polypoidal lesions	13.7 mm	87.5%/79.1%	

*Confocal laser endoscope

GI gastrointestinal, NA not available

vessels are normal in caliber (honeycomb-like in the gastric body or coil-shaped in the antrum). The presence of villous architecture and goblets cells defines intestinal metaplasia. In contrast, the glandular and cellular polarity are impaired or lost in intraepithelial neoplasia and gastric cancer, respectively. In addition, the vessels are dilated and irregular in caliber in neoplasia.

Several studies have assessed the utility of CLE in the real-time evaluation of gastric pre-neoplastic and neoplastic lesions. In a large study including 182 (phase I) and 1786 (phase II) patients, real-time eCLE had a higher sensitivity (88.9% vs. 72.2%), specificity (99.3% vs. 95.1%), and accuracy (98.8% vs. 94.1%) for gastric superficial cancer than white light endoscopy [10]. In another study, CLE was compared with magnifying endoscopy NBI (ME-NBI) in 82 patients with suspected superficial gastric cancers. CLE was highly accurate, but not superior to ME-NBI for differentiating cancerous from non-cancerous gastric lesions (accuracy 93.5% vs. 93.7%) [15]. A recent systematic review and meta-analysis including twenty-three studies analyzed the diagnostic value of CLE for gastric cancer and precancerous lesions among Asian population [26]. The pooled sensitivity and specificity of CLE were 91% and 99% in gastric cancer and 81% and 98% in gastric intraepithelial neoplasia, respectively [26]. In one study, CLE (accuracy 94.2%) outperformed endoscopic biopsy (accuracy 85.7%) for the diagnosis of gastric adenomas and adenocarcinomas [11]. Therefore, CLE has the potential to replace endoscopic biopsy or at least provide a guidance to obtain targeted biopsy specimens. Unlike colonic polyps, the data is more limited on the application of CLE in gastric lesions. In addition, a standardized classification system is warranted for uniform reporting of CLE images in different gastric lesions.

CLE for colonic polyps

CLE has been more widely assessed for its role in the evaluation of colonic polyps as compared with gastric polyps. In the initial studies by Kiesslich et al. (2004) and Hurlstone et al. (2008), CLE was highly accurate (99%) in the prediction of neoplastic changes in the colonic polyps [5, 7]. Of note, integrated confocal endoscope (no longer available) was used in both these studies as compared with majority of the later studies in which the pCLE system was used. Such a high degree of accuracy could not be replicated in the subsequent studies, which revealed a diagnostic yield ranging from 72% to 99% (Table 1). Several studies have compared pCLE with the currently established narrow spectrum imaging techniques or virtual chromoendoscopy. Buchner et al. compared pCLE with virtual chromoendoscopy (NBI or FICE) in 75 patients with 119 colonic polyps. The sensitivity of pCLE was better than virtual chromoendoscopy for the classification of polyps (91% vs. 77%) [14]. In contrast, the post-hoc accuracy of pCLE was found to be inferior to NBI and chromoendoscopy in a study by Kuiper and colleagues [13]. Poor quality of videos was presumably responsible for the inferior results with pCLE in this study. Similar concerns have been raised in the other studies regarding the acquisition of good-quality pCLE images for offline evaluation [16, 27]. Another factor that may influence the accuracy of in vivo histology is the experience of the operator and inter-observer agreement. There is limited literature regarding the learning curve of pCLE [28, 29]. In a study by Buchner et al., the accuracy improved from 63% for the first twenty lesions to 86% after 60 lesions [28]. A few other studies also concluded that accurate post-hoc interpretation of

eCLE confocal images can be learned quickly [29, 30]. In addition, the inter-observer agreement for the classification of neoplasia has been found to be moderate to good in another recent study [8]. These studies suggest that interpretation and acquisition of pCLE images are not difficult to learn by novices.

Comparison of outcomes: current study vs. published literature

In the current issue of the *Journal*, Goenka and colleagues prospectively evaluated the histology in 50 GI polyps using pCLE [3]. Most (38, 76%) of the polyps were colorectal and gastric polyps were fewer in number (12, 24%). pCLE was performed after administration of intravenous fluorescein using the standard dosage and technique (as described above). Videos were also recorded for offline evaluation by the same endoscopist at a later date. Considering histopathology as the gold standard, the overall diagnostic accuracy of real-time pCLE and offline examination of the recorded images was 83.3% and 85.4%, respectively. The diagnostic accuracy in the current study stands in line with the published literature, i.e. 89% to 94% in gastric polyps and 72% to 99% in colonic polyps. However, a separate analysis was not performed for gastric or colonic polyps. Therefore, it is difficult to ascertain the individual diagnostic accuracy of pCLE in gastric polyps from this study. In addition, the real-time and offline analysis of the images was performed by a single endoscopist. Consequently, the inference regarding inter-observer variability cannot be drawn. Experience of the operator and quality of images for offline evaluation are the other factors that may affect the diagnostic accuracy of pCLE.

Nevertheless, this is the first study from India which evaluated the role of pCLE in GI polypoidal lesions. It is likely that with the improvement in operator's experience, the diagnostic accuracy will increase further.

Conclusions from the study

The diagnostic performance of pCLE for the in vivo prediction of histology appears reasonably good in the current study. The authors concluded that with the use of CLE, it may be possible to avoid polypectomy in some patients. However, the important question is as follows: how good is good enough? Is the accuracy sufficient to implement the strategy of “resect and discard” or “leave behind” the polyps in the GI tract? In our opinion, the diagnostic accuracy should be at least more than 90% so that there is a minimum risk of leaving potentially malignant polyps behind. The American Society of Gastrointestinal Endoscopy (ASGE) established the Preservation and Incorporation of Valuable endoscopic

Innovations (PIVI) thresholds for small colorectal polyps. As per the ASGE PIVI threshold criteria, the negative predictive value for adenomatous histology and the agreement of endoscopic technique with histopathology should be $\geq 90\%$ [31]. Therefore, there is a definite scope of improvement and the use of pCLE for avoiding polypectomy may not be ready for prime time.

CLE: current limitations and future directions

CLE is a promising imaging technique for various GI lesions. Recent studies have revealed a reasonable degree of accuracy in differentiating neoplastic and non-neoplastic lesions across the GI tract. However, the concerns potentially hampering the widespread use of CLE in clinical practice need to be addressed in future studies. First, most studies depicting excellent results of CLE have been conducted by experts at the academic centers. Consequently, these results may not be reproducible in community practice. For the same reason, there is heterogeneity in the literature in respect to the diagnostic accuracy of these techniques, which may not reach key thresholds for meaningful decisions in community settings. Second, CLE may not be as accurate for small-sized polyps (< 10 mm) as compared to larger polyps [16]. Therefore, the degree of accuracy and negative predictive value of CLE need to be improved especially in cases with smaller polyps. Third, CLE adds to the overall duration and cost of the procedure. Moreover, the field of view is limited as compared to white light endoscopy and NBI. This means that it cannot be used as a sole modality for screening purposes and has to be used in conjunction with wide-field high-definition endoscopy and the other narrow spectrum imaging techniques. Fourth, randomized comparative studies are required between CLE and the currently established narrow spectrum imaging techniques (like NBI and i-scan). Last, the quality of CLE images obtained for subsequent examination or verification is often inadequate. In the published literature, an interpretation of the lesions could not be done in a substantial proportion of patients due to the poor quality of videos obtained during the initial examination [16, 27].

In the future, novel devices like dual-axis confocal (DAC) microscopy may overcome some of the drawbacks associated with CLE. In the DAC system, the out-of-focus light is directed away from the pinhole and is more optimally rejected than the single-axis CLE (SAC), thereby improving the signal-to-background ratio [32]. Preliminary data suggest that DAC may further improve the resolution and the quality of images over the currently available SAC. The development of image interpretation methods and the use of artificial intelligence have gained a great deal of attention over the last few years. It is likely that fully automated computer-aided diagnosis will reduce the dependence on optical diagnosis obtained in real

time through CLE. In addition, it may reduce the inter-observer variability associated with qualitative CLE analysis of images [33, 34].

In summary, CLE has the potential to replace conventional histopathology in GI tract lesions provided that quality training, quality assurance, and patient acceptance are addressed adequately. The study in this issue of the *Journal* is an initial attempt to evaluate the diagnostic accuracy of CLE in Indian patients with gastric and colonic polypoidal lesions. More quality studies are required to establish the role of CLE in clinical practice.

Compliance with ethical standards

Conflict of interest ZN, and DNR declare that they have no conflict of interest.

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