**REPORTS OF ORIGINAL INVESTIGATIONS** 



# A performance comparison of the most commonly used minimally invasive monitors of cardiac output

# Comparaison des performances des moniteurs minimalement invasifs du débit cardiaque les plus couramment utilisés

Jonathan J. Gamble, MD, FRCPC D · William P. McKay, MD, FRCPC · Barbara Ambros, DVM, MSc · Grant G. Miller, MD, FRCSC · Andrea Vasquez Camargo, MD MSc · Jonathan Norton, PhD · Jayden Cowan, MD · Jean du Rand, MBChB, FRCSC · Erick D. McNair, PhD · Kris Milbrandt, MD, FRCSC, FACS · Martin Gérard, MD, FRCPC · Maria Valentina Carrozzo, DVM

Received: 25 June 2020/Revised: 25 June 2021/Accepted: 28 June 2021/Published online: 9 August 2021 © Canadian Anesthesiologists' Society 2021

#### Abstract

**Background** Shock is common in critically ill and injured patients. Survival during shock is highly dependent on rapid restoration of tissue oxygenation with therapeutic goals based on cardiac output (CO) optimization. Despite the clinical availability of numerous minimally invasive monitors of CO, limited supporting performance data are available.

**Supplementary Information** The online version contains supplementary material available at https://doi.org/10.1007/s12630-021-02085-0.

J. J. Gamble, MD, FRCPC (⊠) · W. P. McKay, MD, FRCPC · J. Cowan, MD · M. Gérard, MD, FRCPC Department of Anesthesia, University of Saskatchewan, Saskatoon, SK, Canada e-mail: jonathan.gamble@saskhealthauthority.ca

B. Ambros, DVM, MSc · M. V. Carrozzo, DVM Western College of Veterinary Medicine, University of Saskatchewan, Saskatoon, SK, Canada

G. G. Miller, MD, FRCSC  $\cdot$  J. Norton, PhD  $\cdot$  J. du Rand , MBChB, FRCSC  $\cdot$  K. Milbrandt, MD, FRCSC, FACS Department of Surgery, University of Saskatchewan, Saskatoon, SK, Canada

A. V. Camargo, MD MSc Department of Academic Family Medicine, University of Saskatchewan, Saskatoon, SK, Canada

E. D. McNair, PhD

Department of Pathology and Laboratory Medicine, University of Saskatchewan, Saskatoon, SK, Canada

**Methods** Following approval of the University of Saskatchewan Animal Research Ethics Board, we assessed the performance and trending ability of PiCCOplus<sup>TM</sup>, FloTrac<sup>TM</sup>, and CardioQ-ODM<sup>TM</sup> across a range of CO states in pigs. In addition, we assessed the ability of invasive mean arterial blood pressure (iMAP) to follow changes in CO using a periaortic transit-time flow probe as the reference method. Statistical analysis was performed with function-fail, bias and precision, percent error, and linear regression at all flow, low-flow (> 1 standard deviation [SD] below the mean), and high-flow (> 1 SD above the mean) CO conditions.

Results We made a total of 116,957 paired CO measurements. The non-invasive CO monitors often failed to provide a CO value (CardioQ-ODM: 40.6% failed measurements; 99% confidence interval [CI], 38.5 to 42.6; FloTrac: 9.6% failed measurements; 99% CI, 8.7 to 10.5; PiCCOplus: 4.7% failed measurements; 99% CI, 4.5 to 4.9; all comparisons, P < 0.001). The invasive mean arterial pressure provided zero failures, failing less often than any of the tested CO monitors (all comparisons, P <0.001). The PiCCOplus was most interchangeable with the flow probe at all flow states: PiCCOplus (20% error; 99%) CI, 19 to 22), CardioQ-ODM (25% error; 99% CI, 23 to 27), FloTrac (34% error; 99% CI, 32 to 38) (all comparisons, P < 0.001). At low-flow states, CardioQ-ODM (43% error; 99% CI, 32 to 63) and Flotrac (45% error; 99% CI, 33 to 70) had similar interchangeability (P = 0.07), both superior to PiCCOplus (48% error; 99%) CI, 42 to 60) (P < 0.001). Regarding CO trending, the CardioQ-ODM (correlation coefficient, 0.82; 99% CI, 0.81 to 0.83) was statistically superior to other monitors including iMAP, but at low flows iMAP (correlation coefficient, 0.58; 99% CI, 0.58 to 0.60) was superior to all minimally invasive CO monitors (all comparisons P < 0.001).

**Conclusions** None of the minimally invasive monitors of CO performed well at all tested flows. Invasive mean arterial blood pressure most closely tracked CO change at critical flow states.

# Résumé

**Contexte** L'état de choc est fréquent chez les patients blessés et en urgence absolue. La survie pendant le choc dépend fortement de la restauration rapide de l'oxygénation tissulaire avec des objectifs thérapeutiques basés sur l'optimisation du débit cardiaque (DC). Malgré la disponibilité clinique de nombreux moniteurs minimalement invasifs du DC, il n'existe que des données limitées sur leur performance pour appuyer leur utilisation.

**Méthode** À la suite de l'approbation du comité d'éthique de la recherche animale de l'Université de la Saskatchewan, nous avons évalué la performance et la capacité de suivi des tendances des appareils PiCCOplus<sup>TM</sup>, FloTrac<sup>TM</sup> et CardioQ-ODM<sup>TM</sup> sur une vaste gamme d'état de DC chez des cochons. Nous avons également évalué la capacité de la tension artérielle moyenne invasive (iMAP) à suivre les changements de DC en utilisant une sonde périaortique de débit basée sur le temps de transit comme méthode de référence. L'analyse statistique a été réalisée avec fonction-échec, biais et précision, pourcentage d'erreur et régression linéaire à des conditions de DC de tous les débits, de faible débit (> 1 écart-type [ET] au-dessous de la moyenne).

Résultats Nous avons effectué un total de 116 957 mesures de DC appariées. Les moniteurs non invasifs de la DC n'ont souvent pas réussi à fournir une valeur de DC (CardioQ-ODM : 40,6% de mesures échouées; intervalle de confiance [IC] de 99 %, 38,5 à 42,6; FloTrac : 9,6 % de mesures échouées; IC 99 %, 8,7 à 10,5; PiCCOplus : 4,7 % de mesures échouées; IC 99 %, 4,5 à 4,9; toutes les comparaisons, P < 0,001). La tension artérielle moyenne invasive n'a fourni aucun échec plus souvent que n'importe lequel des moniteurs de DC testés (toutes les comparaisons, P < 0,001). Le PiCCOplus était le plus interchangeable avec la sonde de débit à tous les états de débit : PiCCOplus (erreur de 20 %; IC 99 %, 19 à 22), CardioQ-ODM (erreur de 25 %; IC 99 %, 23 à 27), FloTrac (erreur de 34 %; IC 99 %, 32 à 38) (toutes les comparaisons, P < 0.001). Aux états de débit faible, les moniteurs CardioQ-ODM (erreur de 43 %; IC 99 %, 32 à 63) et FloTrac (erreur de 45 %; IC 99 %, 33 à 70) présentaient une interchangeabilité similaire (P = 0.07), tous deux supérieurs au PiCCOplus (erreur de 48 %; IC 99 %, 42 à 60) (P < 0,001). En ce qui concerne le suivi des tendances de DC, le CardioQ-ODM (coefficient de correlation, 0,82; IC 99 %, 0,81 à 0,83) était statistiquement supérieur aux autres moniteurs, y compris au iMAP, mais à faibles débits, l'iMAP (coefficient de correlation, 0,58; IC 99 %, 0,58 à 0,60) était supérieure à tous les moniteurs de DC minimalement invasifs (toutes les comparaisons, P < 0,001).

**Conclusion** Aucun des moniteurs de DC minimalement invasif n'a donné de bons résultats à tous les débits testés. La tension artérielle moyenne invasive était le moniteur qui a suivi de plus près les changements de DC dans des états critiques de débit

**Keywords** Cardiac output · Monitoring · physiologic · Arterial pressure · Anesthesia and analgesia · Hemodynamics

# Introduction

Shock is common in critically ill and injured patients.<sup>1,2</sup> Patient survival in shock states is dependent on rapid restoration of tissue oxygenation.<sup>3,4</sup> Therapeutic restoration of tissue oxygenation involves complex hemodynamic strategies in which cardiac output (CO) optimization plays an important role.<sup>5–7</sup> Low CO is associated with significant adverse events including death, and therapeutic intervention can improve these outcomes.<sup>8,9</sup> Traditionally, clinical CO measurements used pulmonary artery catheter (PAC)-based thermodilution,<sup>10</sup> but PAC use has decreased because of risk of complications.<sup>10</sup>

The development of minimally invasive monitors of CO arose from beliefs in the benefits of CO monitoring and manipulation while reducing the harms of PAC.<sup>11</sup> Numerous minimally invasive monitors of CO are commercially available, and uncertainty exists as to which is "best."<sup>12</sup> Three systems with the greatest market share are FloTrac<sup>TM</sup> (Edwards Lifesciences Corp, Irvine, CA, USA), PiCCOplus<sup>TM</sup> (Pulsion Medical Systems, Munich, Germany), and CardioQ-ODM<sup>TM</sup> (Deltex Medical Limited, Chichester, UK), each employing a different technology.<sup>13</sup> These devices have been validated against the PAC under limited hemodynamic conditions.<sup>14,15</sup> The PAC is a poor standard, reference making it а suboptimal comparator.<sup>16,17</sup> Further, most studies do not report results at various flow states (i.e., all, high, or low). This is potentially important as CO monitoring is likely most important at low-flow states, and an aggregate all flow analysis, even if reassuring, may fail to accurately inform clinicians of device performance at critical CO states. Moreover, few studies have investigated CO trending using a gold standard reference device.

Despite the enthusiasm for CO-guided hemodynamic management, the literature does not robustly support its clinical benefit over standard invasive arterial blood pressure-guided strategies.<sup>18–20</sup> Many practitioners do not routinely monitor CO and continue to use arterial blood pressure guided hemodynamic optimization in high-risk patients.<sup>11</sup> This later strategy may also be due to the optimistic assumption that arterial blood pressure and CO are closely related.<sup>21–23</sup> Despite the limited correlation of blood pressure and CO, guiding therapy by invasive mean arterial pressure (iMAP) may be as efficacious as minimally invasive monitors of CO.

Given the interest in CO-guided hemodynamic management, we sought to evaluate 1) the reliability, bias, precision, interchangeability, and trending (correlation) of the three mentioned minimally invasive monitors, and 2) the reliability and trending of mean iMAP. Both objectives used a gold standard comparator (aortic flow probe), were measured over a wide range of hemodynamic conditions in a pig model, and were analyzed at all flow, low flow (> 1 standard deviation [SD] below the mean), and high flow (> 1 SD above the mean) states.

# Methods

This prospective interventional study was approved by the University of Saskatchewan Animal Research Ethics Board (AUP 20130131 in February 2014 and 20170023 in May 2017) and the study protocol adhered to the Canadian Council on Animal Care guidelines for humane animal use. This report adheres to the ARRIVE guidelines for animal research.<sup>24</sup>

Bias considerations of the study design

To minimize bias, all measurements were objective and concurrent. A convenience sample of 12 pigs was chosen as this was affordable and feasible within our institution. The data analysis plan was modified post hoc. We planned to use polar plots, but published recommendations suggest simple regression is satisfactory for comparison with a gold standard.<sup>14</sup>

# Animals

Healthy male pigs (Camborough/PIC Boar 327) median (interquartile range) weight, 30(26–32) kg were used. We used a porcine model because of its similarities with human cardiovascular anatomy and physiology.<sup>25</sup> All animals

were group housed in a climate-controlled room with 12-hr light-dark cycles with free access to standard food and water. They were fasted eight hours prior to the experimentation but were not denied water.

#### Anesthesia

The pigs were initially sedated with intramuscular ketamine (5 mg·kg<sup>-1</sup>) and midazolam (0.5 mg·kg<sup>-1</sup>). Intravenous access was established, and alfaxalone (mean [SD], 2.5 [0.8]  $mg \cdot kg^{-1}$ ) was administered intravenously to effect and the pigs' tracheas were intubated with appropriately sized cuffed endotracheal tubes. All animals were positive pressure ventilated by a Merlin Small Animal Ventilator in 100% oxygen (Vetronic Services Ltd, Torquay, England). Anesthesia was maintained with intravenous remifentanil (20) $\mu g \cdot k g^{-1} \cdot h r^{-1}$ ) in combination with either isoflurane (end tidal concentration, 0.8-1.2%) in oxygen or propofol  $mg \cdot kg^{-1} \cdot hr^{-1}$ ). Initial ventilation settings (12 - 15)included a tidal volume of 15 mL $\cdot$ kg<sup>-1</sup> and a rate of 12 breaths  $min^{-1}$  then titrated to achieve an arterial oxygen saturation > 94% and a partial pressure of carbon dioxide of 40-50 mm Hg. A multichannel physiologic monitor (Datex-Ohmeda CardiocapTM/5, GE Healthcare, Helsinki, Finland) was used to monitor electrocardiography, arterial oxygenation, heart rate, blood pressure (systolic, mean, and diastolic), respiratory rate, tidal volume, minute volume, end-tidal carbon dioxide level, and expired isoflurane concentration. A femoral artery was percutaneously cannulated with a 5F 20-cm thermistor-tipped aortic catheter, connected to the PiCCOplus monitoring system for measurement of invasive arterial pressure, and slaveconnected to the physiologic monitor. An internal jugular vein was surgically exposed and anterograde cannulated with a 7F 16-cm three-lumen catheter (Arrow; Teleflex, Wayne, PA, USA), to allow for continuous recording of central venous pressure and central venous blood gas sampling. Temperature was continuously monitored via the PiCCOplus monitoring system. Hypothermia was avoided by covering exposed skin and by active forced air warming.

#### Cardiac output monitoring

#### Aortic root flow (gold standard)

After appropriate positioning, a left-sided thoracotomy was performed. The aortic root was dissected free and separated from the pulmonary artery. A pre-calibrated appropriately sized ultrasonic transit-time flow probe (PAU-SERIES COnfidence Flowprobes® with Ultrafit Liners 20 or 24 mm, Transonic Systems Inc., Ithaca, NY, USA; accuracy,  $\pm$  2%) was positioned without constriction on the aortic

root for recording of instantaneous flow and connected to its flow meter (TS420 Perivascular Flow Module). The flow meter's analogue display and signal quality indicator were used to assess and ensure appropriate CO measurement.

# Less invasive CO monitors

All the minimally invasive CO monitors require input of demographic data. The minimal age, weight, and height varies according to monitor type and associated disposables. The DP12 probe with a CardioQ-ODM monitor had the most restrictive low input settings: age, 16 yr; height, 149 cm; and weight, 30 kg. For consistency, we inputted these settings for all animals in all devices.

PiCCOplus (CO measured by arterial pulse contour analysis calibrated to transpulmonary thermodilution)

The previously cannulated femoral artery catheter was connected to the PiCCOplus monitoring system calibrated by the average of three consecutive measurements of thermodilution by injecting 10 mL of iced saline randomly throughout the respiratory cycle into the central venous catheter. This calibration was done immediately prior to experimentation, and six hours later if required.

FloTrac (CO measured by uncalibrated arterial pulse contour analysis)

A brachial artery was cannulated with a 20-G catheter. This catheter was connected to the FloTrac sensor kit and coupled to a Vigileo<sup>TM</sup> monitor (Edwards Lifesciences, Irvine, CA, USA) to evaluate CO.

CardioQ-ODM (CO measured by esophageal doppler ultrasound)

The probe connected to the monitoring system was advanced into the esophagus until blood flow signals were detected. Before and after each experiment, the probe was repositioned until an ideal sharp velocity outline with narrow spectral dispersion was obtained. Device output was visually monitored throughout experimentation.

Invasive mean arterial blood pressure

Invasive mean arterial blood pressure was measured and recorded via the PiCCOplus monitoring system.

#### Measurements

Continuous measurements of invasive blood pressure (systolic, mean, and diastolic) as well as CO from the aortic flow probe, FloTrac, PiCCOplus, and CardioQ-ODM were recorded throughout the experiment. After baseline measurements were made and between each of the following experiments, the animal was allowed to recover for 30 min.

# Experiments

# Effect of increasing CO and afterload

A continuous infusion of epinephrine was administered. The initial dose was 0.1  $\mu$ g·kg<sup>-1</sup>·min<sup>-1</sup>, increased to 0.2  $\mu$ g·kg<sup>-1</sup>·min<sup>-1</sup>, increased again to 0.3  $\mu$ g·kg<sup>-1</sup>·min<sup>-1</sup>, then stopped. Each infusion dose was continued until stability in CO and blood pressure was achieved and continued for an additional five minutes.

# Fluid loading

A rapid bolus of 20 mL·kg<sup>-1</sup> Ringer's Lactate solution was administered intravenously. Cardiac output data were continuously collected until stability in CO and blood pressure was achieved.

# Effect of increasing CO

A continuous infusion of sodium isoproterenol was administered. The initial dose was 0.01  $\mu$ g·kg<sup>-1</sup>·min<sup>-1</sup>, increased to 0.03  $\mu$ g·kg<sup>-1</sup>·min<sup>-1</sup>, increased again to 0.1  $\mu$ g·kg<sup>-1</sup>·min<sup>-1</sup>, and then stopped.

# Effect of decreasing cardiac afterload

A continuous infusion of sodium nitroprusside was administered. The initial dose was 0.3  $\mu$ g·kg<sup>-1</sup>·min<sup>-1</sup>, increased to 0.6  $\mu$ g·kg<sup>-1</sup>·min<sup>-1</sup>, increased again to 0.9  $\mu$ g·kg<sup>-1</sup>·min<sup>-1</sup>, and then stopped.

# Effect of decreasing CO

Intravenous boluses of esmolol (2 and then 4 mg·kg<sup>-1</sup>) were administered to decrease CO. The 2 mg·kg<sup>-1</sup> dose was administered first, followed 15 min later with the 4 mg·kg<sup>-1</sup> dose.

#### Hypovolemia

Each animal was phlebotomized with two 20 mL $\cdot$ kg<sup>-1</sup> of blood draws. Data were continuously collected until

stability in CO and blood pressure were achieved and continued for an additional five minutes; then the second blood draw commenced. At the conclusion of this experiment, the animal was euthanized with a lethal dose of pentobarbital and the experiment terminated.

#### Data acquisition

Analogue flow signals from the aortic flow meter were digitized at 100 Hz with an analog-to-digital converter (National Instruments USB-6259 multifunction DAQ, National Instruments, Austin, TX, USA), and sampled by customized data logging software (Biobench, National Instruments) operating on a personal computer for online display and storage of data. The CO values from the FloTrac, PiCCOplus, and CardioQ-ODM were copied to computer files by proprietary programs. The iMAP was measured with the PiCCOplus monitor, recording the average mean blood pressure for sequential five second intervals, and copied to computer files by its proprietary program.

#### Data analysis

The commercial CO monitors produced printouts of their measurements from proprietary programs at different rates, averaging the CO from epochs of different durations: PiCCOplus (1-sec epochs); Flotrac (20-sec epochs); and CardioQ-ODM (30-sec epochs). These data were used at the original output rate for regression against the flow probe. Flow probe data were averaged for epoch lengths corresponding to each device.

# Statistical analysis

To simplify results for the reader, data from all pigs was analyzed together as if it were a single end-to-end experiment. We justified this on the basis of the pigs being of the same age, sex, and strain, and having similar weights. To adjust for the differences in CO attributable to different pig sizes or physiology, we adjusted the data by normalizing the mean flow for each pig to the overall mean flow and used the normalization factor for each pig to adjust the point-by-point CO output of each device for each pig before aligning the device outputs end to end. calculations Statistical were performed with SIGMAPLOT version 13.0 (Jandel Scientific, Div. of Jandel Corp., San Rafeal, CA, USA). Because of the large number of comparisons, the statistical significance level was set at P < 0.01. Primary data were tested for normality by the Kolmogorov-Smirnov test and compared with the use of non-parametric Kruskal–Wallis analysis of variance on ranks. We conducted the analyses at all flows > 1  $L \cdot min^{-1}$ , low-flow ( > 1 SD below the mean) and highflow ( > 1 SD above the mean) states. Categorical variables were analyzed with Chi square, except when comparing data containing a category with zero incidence (iMAP failure), in which case McNemar's test was used.

Function: failure analysis (reliability of data output)

We compared the frequency of failure of the devices (epochs when no CO data were produced) using Chi square and McNemar's test. Analysis was done for all outputs of each device that occurred during flow outputs > 0.1 L·min<sup>-1</sup> as well as at low, medium, and high CO states as measured by the flow probe. Ninety-nine percent confidence intervals (CIs) are reported.

Bias (accuracy) and limits of agreement (precision)

Because the CO devices all incorporate demographics (height, weight, and age) into the CO measurement using confidential proprietary algorithms, and because porcine anatomic proportions are different from those in humans, we considered that we could not assume comparable bias among devices. Thus, we calculated overall limits of agreement from the normalized data. Cardiac output data were not normally distributed, so non-parametric analysis was done to estimate bias and precision with Olofsen's method and calculator.<sup>26,27</sup> Ninety-nine percent CIs are reported.

Percent error (device interchangeability with flow probe)

Percent error was calculated from normalized absolute differences of each CO datum from the corresponding flow reading as follows:

$$100\left(\sum_{0}^{n}\left(\frac{|diff_{n}|}{Flow_{n}}\right) \div n\right)$$

Ninety-nine percent CIs are reported.

#### Regression (trending)

Correlation coefficients were calculated with linear regression of each device upon flow probe CO using data output when the device was functioning and flow was > 0.1 L·min<sup>-1</sup>. Correlation coefficients were compared by the method of Steiger.<sup>28</sup> Ninety-nine percent CIs are reported.

#### Results

Cardiac output from all experimental conditions was collected for ten of 12 animals. Data for animal 1 were not analyzed because of an unrecognized flow probe data acquisition storage failure. FloTrac data for animal 7 were not collected because of an unplanned data storage failure. Table 1 shows the number of paired measurements for the various monitoring modalities for the data in aggregate and at high and low flow states. A representative plot from animal 5 shows the sequence of experimental interventions and measures from all three studied minimally invasive CO monitoring devices (Fig. 1). Time plots for each animal are presented in the Electronic Supplementary Material (ESM) (eFig 1a–i). The results of all animals are summarized in ESM eFig. 1k.

None of the CO measures were normally distributed. The flow meter measured flows ranged from 0 to 9.1  $L\cdot min^{-1}$ ; the median [1<sup>st</sup> to 99<sup>th</sup> percentile] CO over all measurements was 3.2 [1.0 to 5.9]  $L\cdot min^{-1}$ . Cardiac outputs less than 2.2  $L\cdot min^{-1}$  were considered low flow while those greater than 4.4  $L\cdot min^{-1}$  were considered high flow, representing flows > 1 SD below and above the mean,

respectively. None of the minimally invasive monitors of CO consistently provided output at all times (percent failed measurements), although the aortic root flow probe did (Table 1). The PiCCOplus had the fewest failures at all flows and low flows (4.7%; 99% CI, 4.5 to 4.9) and (6.9%: 99% CI, 6.5 to 7.3) (all comparisons P < 0.001). At all flow and low-flow states, iMAP always (with zero failures) provided data (P < 0.001) (Table 1).

The measures of accuracy and precision are presented in Table 2 and Fig 2a-c (Bland–Altman plots). Interchangeability (percent error), and trending ability (correlation coefficients) for each monitoring device at all flow, low flow, and high flow are presented in Tables 3 and 4. The PiCCOplus was most interchangeable with the flow probe at all flow states (20%; 99% CI, 19 to 22) (P <0.001); at low flows, the CardioQ-ODM (43%; 99% CI, 32 to 63) and Flotrac (45%; 99% CI, 33 to 70) interchangeability were similar (P = 0.07) and both superior to PiCCOplus (48%; 99% CI, 42 to 60) (all comparisons, P < 0.001). Regarding CO trending, the CardioQ-ODM (0.82; 99% CI, 0.81 to 0.83) was statistically superior to the other devices at all flows, but at low flows, iMAP (0.58; 99% CI, 0.58 to 0.60) was best

Table 1 Number of paired measurements for the various monitoring modalities and failure rates

Device	Total samples	% failure 99% CI		P value <sup>†</sup>	
All flows ( $> 1$ to 9.1 L·m	iin <sup>-1</sup> )				
iMAP	116,957	0			
PiCCOplus	116,957	4.7	4.5 to 4.9	< 0.001	
FloTrac	4,549	9.6	8.7 to 10.5	< 0.001	
CardioQ-ODM	3,792	40.6	38.5 to 42.6	< 0.001	
Low flows ( $< 2.2 \text{ L} \cdot \text{min}^-$	-1)				
iMPAP	19,312	0			
PiCCOplus	19,312	6.9	6.5 to 7.3	< 0.001	
FloTrac	424	13.8	10.9 to 16.7	< 0.001	
CardioQ-ODM	376	40.5	35.0 to 45.9	< 0.001	
Medium flow (2.2 to 4.4 L	$2 \cdot \min^{-1}$ )				
iMAP	76,396	0			
PiCCOplus	76,396	5.2	4.8 to 5.6	< 0.001	
FloTrac	3,534	6.0	5.2 to 6.9	0.02	
CardioQ-ODM	2,179	42.5	39.8 to 45.2	< 0.001	
High flow ( $> 4.4 \text{ L} \cdot \text{min}^{-1}$	<sup>1</sup> )				
iMAP	21,249	0			
PiCCOplus	21,249	1.0	0.8 to 1.2	< 0.001	
FloTrac	591	23.5	20.1 to 26.9	< 0.001	
CardioQ-ODM	1,237	59.0	53.4 to 64.5	< 0.001	

Failure was determined when no cardiac output value was registered by the device.

Note that devices are ranked by failure rate in each flow state reported and compared with the device above<sup>†</sup>

CI = confidence interval; iMAP = invasive mean arterial pressure

**Fig. 1** Representative time plot from animal 5. Flow probe measured cardiac output under labelled experimental conditions. X-axis: time in minutes; Y-axis: normalized cardiac output in L·min<sup>-1</sup>.



Table 2 Bias and precision of non-invasive cardiac output devices

Device	Bias	99% CI* of bias	Precision lower LOA	Precision upper LOA
CardioQ-ODM	-0.1	-1.5 to 0.5	-3.1	4.3
PiCCOplus	-0.1	-0.6 to 0.2	-2.4	3.6
FloTrac	0.1	-0.2 to 1.1	-3.6	3.9

Values are L·min<sup>-1</sup>. Reference flow was determined by a periaortic transit-time flow probe.

\*1st to 99th percentiles.

CI = confidence interval; LOA = limit of agreement.

(all comparisons, P < 0.001). Scatterplots for low-flow states for each device and iMAP are shown in Fig 3 a–d while plots for all flow states can be found in the supplement in ESM eFig 2a–d. Four quadrant plots of all devices at all- and low-flow states can be found in ESM eFig 3a–h.

#### Discussion

This study compared the most commonly used minimally invasive CO monitors to a gold standard ultrasonic aortic root flow probe over a large range of COs (0 to 9.1  $L \cdot min^{-1}$ ). None of the studied devices nor iMAP gave useful information under all study conditions. At all flows and when functioning, percent error was lowest with PiCCOplus, then CardioQ-ODM, and greatest with FloTrac. PiCCOplus also failed least often, while FloTrac failed less frequently than CardioQ-ODM. Crucially, at low flow, none of the CO measurement devices performed well, and iMAP trended CO change better than the assessed devices did.

The CardioQ-ODM appeared to provide good graphical signals at low CO when judging the output by the device's manual, yet the monitor reported failure. This suggests the CardioQ-ODM was measuring CO signal, but the computer algorithm should be updated to provide reliable data at low CO.

The primary objective of this study was to assess the accuracy, precision, interchangeability, and trending ability of the most commonly used minimally invasive monitors of CO over a wide range of hemodynamic conditions; the strengths of our study reflect this goal. The aortic flow probe is described as the gold standard comparator as it provides direct continuous flow measurement, independent of rotation, hemoglobin concentration, or electrically



Fig 2 a-c Bland-Altman plots for each studied monitor of cardiac output: PiCCOplus (2a), FloTrac (2b), CardioQ-ODM (2c).



Fig 2 continued

charged molecules.<sup>29</sup> We followed the recommendations of a widely quoted paper on methodology for assessing trending ability.<sup>14</sup> In pigs, the anatomical and physiologic characteristics of the cardiovascular system are similar to those in humans.<sup>25</sup>Furthermore, the experimental conditions created grossly reflect the hemodynamics in a wide range of clinically encountered disease states. The extremely invasive nature of the study precludes similar study in humans.

The level of performance needed from a cardiac monitor to improve clinical outcomes is unknown. Despite this,  $\pm$  30% percent error (PE) is a widely quoted threshold for acceptable agreement to consider CO devices interchangeable.<sup>30</sup> At first consideration, both the CardioQ-ODM and PiCCOplus devices would seem to meet this threshold; however, this initial analysis is flawed for two major reasons. First, the 30% threshold is based on PAC-based CO measurements as the comparator device and incorporates the PAC error into overall error. The flow probe error is negligible; therefore, errors can be attributed to the devices studied alone.<sup>29,31</sup> In this situation, the magnitude of acceptable error is less clear, but some authors have suggested that an accuracy of 10% is desirable as this is the error of non-invasive arterial blood pressure measurements;<sup>32</sup> others have suggested 15% as this is similar to successive PAC CO measurements;<sup>33,34</sup> and still others arbitrarily suggested 20%.<sup>35</sup> Using even the most liberal of these thresholds, all the devices fall short. The second problem of accepting 30% PE is the PiCCOplus and CardioQ-ODM met this only at all flow states, whereas clinicians are more likely concerned about CO assessment at low-flow states. Although our definition of low flow is arbitrary (CO < 1SD below the mean), the best performing device at low flow (CardioQ-ODM) had a PE of 43%, not meeting any described definition of acceptable. Two of the monitoring systems tested rely solely on examination of peripheral pulse data (FloTrac and PiCCOplus) while one (CardioQ-ODM) relies solely on central flow as measured indirectly with ultrasound. Early work found that the accuracy of minimally invasive CO measurement is greatly enhanced by combining central and peripheral information.<sup>36</sup>

Further, all the devices frequently failed to provide any data output, especially at low-flow states. Guidance on how to interpret these failure rates is not available, but the range of 20–60% failure seems excessive. The issue of monitor

	Table 3	Percent	error for	non-inv	vasive	cardiac	output	devices
--	---------	---------	-----------	---------	--------	---------	--------	---------

Device	Percent error*	99% CI	P value <sup>†</sup>
All flows			
PiCCOplus	20%	19% to 22%	
CardioQ-ODM	25%	23% to 27%	< 0.001
FloTrac	34%	32% to 38%	< 0.001
Low flow			
CardioQ-ODM	43%	32% to 63%	
FloTrac	45%	33% to 70%	0.07
PiCCO plus	48%	42% to 60%	< 0.001

\*Percent error was calculated using available measurements (i.e., excludes failures).

Devices ranked in table according to percent error within each flow state.

<sup>†</sup> To the device above

CI = confidence interval

 Table 4
 Tracking: a comparison of coefficients of correlation among non-invasive cardiac output monitoring devices and invasive mean arterial pressure

	R*	99% CI	P value <sup>†</sup>
All flows			
CardioQ-ODM	0.82	0.81 to 0.83	
PiCCOplus	0.48	0.45 to 0.51	< 0.001
FloTrac	0.33	0.32 to 0.34	< 0.001
iMAP	0.31	0.30 to 0.32	< 0.001
Low flows			
iMAP	0.58	0.58 to 0.60	
CardioQ-ODM	0.46	0.43 to 0.49	< 0.001
PiCCOplus	0.43	0.41 to 0.45	0.035
FloTrac	0.25	0.13 to 0.36	< 0.001

R = correlation coefficient

<sup>†</sup> Compared with the device above

CI = confidence interval

output failure is likely more problematic than simply a lack of data output. These devices fail more at low-flow states, but also fail at medium and high flows. Thus, failure may or may not indicate a low-flow state. Hence, device failure increases cognitive load for the care provider, uncertainty of appropriate response, and distraction from other tasks.

Trending ability is another important measurement for CO assessment.<sup>14,37</sup> The ability to accurately detect change in CO as an indication of deterioration or response to intervention may be more important than absolute accuracy. It is unclear how to clinically interpret regression analysis of CO monitoring. Although both the CardioQ-ODM and PiCCOplus are highly correlated at all

flow states, none of the monitors are well correlated at low-flow states.<sup>38</sup>

Although the study of the relationship between blood pressure and CO is not new, it remains interesting.<sup>22,39</sup> Our rigorous results (116,957 paired measurements) confirm previous data that iMAP has a low positive correlation with CO at all flow states.<sup>22</sup>Our results show iMAP to be more correlated to CO than any of the studied devices at low-flow states.

A limitation of the study is that young animals were used ( $\sim$  ten weeks old). These animals presumably did not suffer from adult human diseases including cardiovascular disease, potentially limiting the generalization of our results. Additionally, the animals weighed much less than



Fig 3 a-d Scatter plots for each studied monitor in low-flow states: PiCCOplus (3a), Flotrac (3b), CardioQ-ODM (3c), and invasive mean arterial pressure (3d).



Normalzied Aortic Root Flow Probe Measured Cardiac Output (L/min)

Fig 3 continued

an average adult. We chose a young porcine model for several reasons: 1) the known similarities of human and porcine cardiovascular anatomy and physiology,<sup>25</sup> 2) the expense of using a primate animal model, and 3) the danger to investigators of using adult pigs, which can weight more than 300 kg.

Another limitation of our study relates to the various experimental hemodynamic conditions created to mimic physiologic stress and disease states. It is likely the hemodynamic effects of most disease states, especially septic shock, are far more complex than those we created, limiting the generalizability of our results to sick humans. Additionally, the relevance of our analysis at various flow states could be questioned as the clinician would not know the true flow state and therefore could not apply our flow state findings to observed device measurements. This criticism is valid if these devices were used in isolation. We contend that in clinical practice, CO monitoring would rarely be used as a sole monitor, rather would be used in conjunction with numerous other monitors in addition to the context of the clinical scenario. The clinicians' integration of the totality of the available data would likely influence the assessment of patient flow state and therefore application of our results. Further, should a device not perform well at critical low-flow states, we contend that its utility is limited regardless of its performance at other or aggregate flow states. Finally, we did not evaluate all commercially available minimally invasive monitors of CO.

Numerous studies have assessed the performance of minimally invasive CO devices using the PAC as a comparator. For studies investigating CO compared with a gold standard under a wide range of hemodynamic conditions, the body of literature is much smaller.<sup>35,40–44</sup>

Acute care physicians often work on the premise that achieving adequate CO is an essential management component for care of critically ill patients especially considering that signals of instability, need for intervention, and resuscitation end points are often clinically challenging. The logical extension of this management framework is that the use of CO monitors should improve patient outcomes. A systematic review suggests that CO-guided hemodynamic management does not improve mortality, but may reduce important morbidities and hospital length of stay (the underlying data are of limited quality).<sup>20</sup> The knowledge that survivors of major surgical procedures have higher CO<sup>45</sup> and, likely more importantly, the belief that monitoring and manipulating CO should improve patient outcomes, continues to drive intensive research and commercial interest. The reasons for weak improvements in clinical outcomes are probably numerous, but poor monitor performance may be an important factor; our results suggest that there is significant room for improvement in this domain. Further, the studies investigating CO-guided hemodynamic management compared it with either goal directed or usual care, both primarily based on arterial blood pressure measurement. This general lack of outcome improvement could be explained knowing that the minimally invasive CO devices perform similar to iMAP monitoring under low-flow conditions.

#### Summary

The PiCCOplus performed better than other devices when considering all CO states, but none of the studied minimally invasive monitors of CO performed well, especially at critical low-flow states in a young pig model. This evidence points to possible poor performance in humans. None of the studied monitors tracked change in CO better than mean arterial blood pressure at low flow states.

Author contributions The manuscript is the collaborative work of 12 authors. William P. McKay and Jonathan J. Gamble conceptualized the study. Jonathan J. Gamble, William P. McKay, Jonathan Norton, and Erick D. McNair formalized the study protocol. Jonathan J. Gamble, William P. McKay, Barbara Ambros, Maria Valentina Carrozzo, Grant G. Miller, Andrea Vasquez Camargo, Jayden Cowan, Jean du Rand, Martin Gerrard, and Kris Milbrandt acquired the data. William P. McKay conducted the data analysis. Jonathan J. Gamble wrote the initial draft and all 12 authors revised the manuscript.

Acknowledgements This work was presented at Anesthesiology 2015; Oct 24 2015; San Diego. We gratefully acknowledge the assistance of Department of Anesthesiology, University of Saskatchewan and the Department of Anesthesiology, Perioperative Medicine and Pain Management, University of Saskatchewan. We also acknowledge the assistance of the Western College of Veterinary Medicine, University of Saskatchewan for their support of this project.

**Conflicts of interest** Commercial or non-commercial affiliations that are or may be perceived to be a conflict of interest with the work for each author and any other associations, such as consultancies: No author has any commercial or other affiliations that are, or may be perceived to be, a conflict of interest.

**Funding statement** Department of Anesthesiology, Perioperative Medicine and Pain Management University of Saskatchewan; College of Medicine, University of Saskatchewan; Western College of Veterinary Medicine, University of Saskatchewan.

**Disclosures** Study was approved by the University of Saskatchewan Animal Research Ethics Board (AUP 20130131 in February 2014 and 20170023 in May 2017).

**Editorial responsibility** This submission was handled by Dr. Gregory L. Bryson, former Deputy Editor-in-Chief, *Canadian Journal of Anesthesia/Journal canadien d'anesthésie.* 

#### References

- 1. Varpula M, Tallgren M, Saukkonen K, Voipio-Pulkki LM, Pettila V. Hemodynamic variables related to outcome in septic shock. Intensive Care Med 2005; 31: 1066-71.
- Kauvar DS, Lefering R, Wade CE. Impact of hemorrhage on trauma outcome: an overview of epidemiology, clinical presentations, and therapeutic considerations. J Trauma 2006; 60(6 Suppl): S3-11.
- 3. Dellinger RP, Levy MM, Rhodes A, et al.; Surviving Sepsis Campaign Guidelines Committee including The Pediatric Subgroup: Surviving Sepsis Campaign. Surviving Sepsis Campaign: international guidelines for management of severe sepsis and septic shock, 2012. Intensive Care Med 2013; 39: 165-228.
- Rossaint R, Bouillon B, Cerny V, et al.; Task Force for Advanced Bleeding Care in Trauma. Management of bleeding following major trauma: an updated European guideline. Critl Care 2010; DOI: https://doi.org/10.1186/cc8943.
- Rampal T, Jhanji S, Pearse RM. Using oxygen delivery targets to optimize resuscitation in critically ill patients. Curr Opin Crit Care 2010; 16: 244-9.
- Schumann J, Henrich EC, Strobl H, et al. Inotropic agents and vasodilator strategies for the treatment of cardiogenic shock or low cardiac output syndrome. Cochrane Database Syst Rev 2018; DOI: https://doi.org/10.1002/14651858.CD009669.pub3.
- 7. Fleming A, Bishop M, Shoemaker W, et al. Prospective trial of supranormal values as goals of resuscitation in severe trauma. Arch Surg 1992; 127: 1175-81.
- van Diepen S, Katz JN, Albert NM, et al. Contemporary management of cardiogenic shock: a scientific statement from the American Heart Association. Circulation 2017; 136: e232-68.
- 9. *Hamilton MA, Cecconi M, Rhodes A.* A systematic review and meta-analysis on the use of preemptive hemodynamic intervention to improve postoperative outcomes in moderate and high-risk surgical patients. Anesth Analg 2011; 112: 1392-402.
- Marik PE. Obituary: pulmonary artery catheter 1970 to 2013. Ann Intensive Care 2013; DOI: https://doi.org/10.1186/2110-5820-3-38.
- Cannesson M, Pestel G, Ricks C, Hoeft A, Perel A. Hemodynamic monitoring and management in patients undergoing high risk surgery: a survey among North American and European anesthesiologists. Crit Care 2011; DOI: https://doi.org/10.1186/ cc10364.
- Thiele RH, Bartels K, Gan TJ. Cardiac output monitoring: a contemporary assessment and review. Crit Care Med 2015; 43: 177-85.
- 13. Davidson RS. Deltex Medical. Edison Investment Research; 2010.
- Critchley LA, Lee A, Ho AM. A critical review of the ability of continuous cardiac output monitors to measure trends in cardiac output. Anesth Analg 2010; 111: 1180-92.
- Peyton PJ, Chong SW. Minimally invasive measurement of cardiac output during surgery and critical care: a meta-analysis of accuracy and precision. Anesthesiology 2010; 113: 1220-35.

- Dhingra VK, Fenwick JC, Walley KR, Chittock DR, Ronco JJ. Lack of agreement between thermodilution and fick cardiac output in critically ill patients. Chest 2002; 122: 990-7.
- Espersen K, Jensen EW, Rosenborg D, et al. Comparison of cardiac output measurement techniques: thermodilution, Doppler, CO2-rebreathing and the direct Fick method. Acta Anaesthesiol Scand 1995; 39: 245-51.
- Pearse RM, Harrison DA, MacDonald N, et al. Effect of a perioperative, cardiac output-guided hemodynamic therapy algorithm on outcomes following major gastrointestinal surgery a randomized clinical trial and systematic review. JAMA 2014; 311: 2181-90.
- Agency for Healthcare Research and Quality. Esophageal Doppler Ultrasound-Based Cardiac Output Monitoring for Real-Time Therapeutic Management of Hospitalized Patients: A Review. Rockville (MD), 2007. Available from URL: https://www. ecri.org/Resources/EPC\_Sample\_Reports/Esophageal\_Doppler\_ Ultrasound\_Based\_Cardiac\_Output\_Monitoring.pdf (accessed July 2021).
- Grocott MP, Dushianthan A, Hamilton MA, et al. Perioperative increase in global blood flow to explicit defined goals and outcomes after surgery: a Cochrane systematic review. Br J Anaesth 2013; 111: 535-48.
- 21. *Magder S.* Phenylephrine and tangible bias. Anesth Analg 2011; 113: 211-3.
- 22. Wo CC, Shoemaker WC, Appel PL, Bishop MH, Kram HB, Hardin E. Unreliability of blood pressure and heart rate to evaluate cardiac output in emergency resuscitation and critical illness. Crit Care Med 1993; 21: 218-23.
- 23. Brotmacher L. Evaluation of derivation of cardiac output from blood pressure measurements. Circ Res 1957; 5: 589-93.
- Kilkenny C, Browne WJ, Cuthill IC, Emerson M, Altman DG. Improving bioscience research reporting: the ARRIVE guidelines for reporting animal research. PLoS Biol 2010; DOI: https://doi. org/10.1371/journal.pbio.1000412
- Swindle MM, Makin A, Herron AJ, Clubb FJ Jr, Frazier KS. Swine as models in biomedical research and toxicology testing. Vet Pathol 2012; 49: 344-56.
- Olofsen E, Dahan A, Borsboom G, Drummond G. Improvements in the application and reporting of advanced Bland-Altman methods of comparison. J Clin Monit Comput 2015; 29: 127-39.
- Olofsen E. Webpage for Bland-Altman analysis. Available from URL: https://sec.lumc.nl/method\_agreement\_analysis/bae.html (accessed July 2021).
- 28. *Steiger J.* Tests for comparing elements of a correlation matrix. Psychol Bull 1980; 87: 245-51.
- 29. Dean DA, Jia CX, Cabreriza SE, et al. Validation study of a new transit time ultrasonic flow probe for continuous great vessel measurements. ASAIO J 1996; 42: M671-6.
- Critchley LA, Critchley JA. A meta-analysis of studies using bias and precision statistics to compare cardiac output measurement techniques. J Clin Monit Comput 1999; 15: 85-91.
- Bednarik JA, May CN. Evaluation of a transit-time system for the chronic measurement of blood-flow in conscious sheep. J Appl Physiol (1985) 1995; 78: 524-30.
- 32. Rutten AJ, Ilsley AH, Skowronski GA, Runciman WB. A comparative study of the measurement of mean arterial blood pressure using automatic oscillometers, arterial cannulation and auscultation. Anaesth Intensive Care 1986; 14: 58-65.
- Corley A, Barnett AG, Mullany D, Fraser JF. Nurse-determined assessment of cardiac output. Comparing a non-invasive cardiac output device and pulmonary artery catheter: a prospective observational study. Int J Nurs Stud 2009; 46: 1291-7.
- 34. Sturgess DJ, Pascoe RL, Scalia G, Venkatesh B. A comparison of transcutaneous Doppler corrected flow time, b-type natriuretic peptide and central venous pressure as predictors of fluid

responsiveness in septic shock: a preliminary evaluation. Anaesth Intensive Care 2010; 38: 336-41.

- Uemura K, Kawada T, Inagaki M, Sugimachi M. A Minimally invasive monitoring system of cardiac output using aortic flow velocity and peripheral arterial pressure profile. Anesth Analg 2013; 116: 1006-17.
- 36. *McKay WP, Gregson PH, McKay BW, Militzer J.* Sternal acceleration ballistocardiography and arterial pressure wave analysis to determine stroke volume. Clin Invest Med 1999; 22: 4-14.
- 37. Joosten A, Desebbe O, Suehiro K, et al. Accuracy and precision of non-invasive cardiac output monitoring devices in perioperative medicine: a systematic review and meta-analysis. Br J Anaesth 2017; 118: 298-310.
- Mukaka MM. Statistics corner: a guide to appropriate use of correlation coefficient in medical research. Malawi Med J 2012; 24: 69-71.
- 39. Levy MN, Brind SH, Brandlin FR, Phillips FA Jr. The relationship between pressure and flow in the systemic circulation of the dog. Circ Res 1954; 2: 372-80.
- 40. Critchley LA, Peng ZY, Fok BS, Lee A, Phillips RA. Testing the reliability of a new ultrasonic cardiac output monitor, the USCOM, by using aortic flowprobes in anesthetized dogs. Anesth Analg 2005; 100: 748-53.

- 41. *Gunn SR, Kim HK, Harrigan PW, Pinsky MR*. Ability of pulse contour and esophageal Doppler to estimate rapid changes in stroke volume. Intensive Care Med 2006; 32: 1537-46.
- 42. Lemson J, de Boode WP, Hopman JC, Singh SK, van der Hoeven JG. Validation of transpulmonary thermodilution cardiac output measurement in a pediatric animal model. Pediatr Crit Care Med 2008; 9: 313-9.
- Bajorat J, Hofmockel R, Vagts DA, et al. Comparison of invasive and less-invasive techniques of cardiac output measurement under different haemodynamic conditions in a pig model. Eur J Anaesthesiol 2006; 23: 23-30.
- 44. Phillips RA, Hood SG, Jacobson BM, West MJ, Wan L, May CN. Pulmonary artery catheter (PAC) accuracy and efficacy compared with flow probe and transcutaneous Doppler (USCOM): an ovine cardiac output validation. Crit Care Res Pract 2012; DOI: https:// doi.org/10.1155/2012/621496.
- 45. Shoemaker WC, Appel PL, Kram HB. Hemodynamic and oxygen transport responses in survivors and nonsurvivors of high-risk surgery. Crit Care Med 1993; 21: 977-90.

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.