SYSTEMATIC REVIEW



Gastrointestinal viral shedding in children with SARS-CoV-2: a systematic review and meta-analysis

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Abstract

Background Coronavirus disease 2019 (COVID-19) has different manifestations in pediatric cases. It is assumed that they might present more gastrointestinal symptoms with a different viral shedding pattern in gastrointestinal samples. In this systematic review and meta-analysis, we aimed to evaluate the viral shedding pattern in gastrointestinal specimens of children with COVID-19.

Methods We searched all published studies in English language in PubMed, Web of Science, and Scopus, up to date as of October 2021. Our search included the term "severe acute respiratory syndrome coronavirus 2, COVID-19, SARS-CoV-2, novel coronavirus, or coronavirus; and shed, excrete, secret, or carriage; and stool or rectal; and children or pediatrics". We included studies evaluating SARS-CoV-2 shedding in gastrointestinal specimens, including rectal swabs and stool samples of children with COVID-19 infection. We excluded duplicated data, case reports, and studies without original data.

Results Twelve studies met the eligibility criteria for the qualitative synthesis, 10 of which were included in the meta-analysis. The pooled prevalence of gastrointestinal severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) RNA in children with COVID-19 was 86% (95% confidence interval 73%–96%, I^2 =62.28%). After respiratory specimen had become negative, 72% (43/60) had persistent shedding in gastrointestinal specimens. The gastrointestinal RNA had a positive test result for more than 70 days after symptoms onset.

Conclusions Gastrointestinal shedding of SARS-CoV-2 might occur in a substantial portion of children and might persist long after negative respiratory testing. Further research is recommended to find the role of SARS-CoV-2 gastrointestinal shedding in transmission in children.

Keywords Coronavirus disease 2019 (COVID-19) · Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) · Viral shedding

Introduction

The outbreak of the novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) led to coronavirus disease 2019 (COVID-19). The disease spread quickly to the

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world and led to a major crisis, the COVID-19 pandemic [1, 2]. The most common clinical presentation of patients with COVID-19 are fever and respiratory symptoms; however, an increasing number of studies report that a variety of gastrointestinal (GI) symptoms such as diarrhea, vomiting, abdominal pain, as well as some atypical findings can also be found in adults and also pediatric patients [3-6]. COVID-19 mostly occurs in adults and children only represent 1%-8% of the total laboratory-confirmed cases [7]. SARS-CoV-2 is primarily transmitted through respiratory droplets and person-to-person close contact [8]. Based on this transmission route, COVID-19 infection is diagnosed by the presence of SARS-CoV-2 RNA in upper and lower respiratory specimens by real-time reverse transcription polymerase chain reaction (RT-PCR). However, viral RNA can be found in other samples [9]. SARS-CoV-2 RNA can also be detected

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in anal/rectal swabs and stool specimens from COVID-19 patients, due to the pathogenicity of the virus in the GI tract, raising concerns about the fecal–oral route of transmission. Interestingly, viral shedding through feces could continue even after respiratory tests are, or have become, negative [10–12].

A few studies have reviewed the possibility of the fecal–oral transmission in patients with COVID-19 [13–16], and a comprehensive result of fecal viral shedding in children is still lacking. On the other hand, children show different clinical characteristics, less severe infection, and different viral load and shedding pattern compared to the adults [17–20]. Therefore, this study aims to assess viral shedding through GI samples in children infected with COVID-19 and provide an overview of the available literature regarding the fecal–oral transmission of the SARS-CoV-2.

Methods

Search strategies

This study was performed according to the PRISMA guidelines (preferred reporting items for systematic reviews and meta-analysis) [21]. We searched electronic databases, including PubMed, Scopus, and Web of Science, up to date as of October 2021. Search studies were limited to those written in English. Our search included the terms: severe acute respiratory syndrome coronavirus 2, COVID-19, SARS-CoV-2, novel coronavirus, or coronavirus; and shed, excrete, secret, or carriage; and stool or rectal; and children or pediatrics. References cited by the identified studies were also screened for additional references.

Study selection

We included studies that met the following criteria: (1) laboratory-confirmed children (defined as being < 18 years) with respiratory COVID-19 infection based on positive RT-PCR results and (2) evaluation of GI specimen including stool collection or rectal swabs for SARS-CoV-2 RNA. We excluded (1) duplicated data; (2) case reports (< 2 patients); and (3) studies without original data (e.g., review articles).

Two reviewers performed the initial screening of titles and abstracts independently to identify potentially relevant studies. The full texts of the selected studies were screened by two independent authors. There was a complete agreement regarding the excluded studies. Two authors independently performed data extraction from each study included. If necessary, we tried to contact the author of the original study by e-mail. Disagreements and uncertainties were resolved by consensus. EndNote (version X8) was used to manage the articles and citations. Information including the first author, study period, country of origin, age, infection status (symptomatic infection or asymptomatic infection), symptoms, number of confirmed patients with COVID-19, number of patients with positive SARS-CoV-2 RNA in GI samples before and after the loss of respiratory RNA, and maximum duration of positive GI samples from symptoms onset or hospitalization were extracted.

Quality assessment

The scale known as quality assessment of diagnostic accuracy studies (QUADAS) was used to assess the quality of the included studies [22]. The scale consists of 14 items, and 1 point is given to each item if the conditions are met.

Statistical analysis

We calculated a weighted pooled estimate of the proportion testing positive from the GI samples using random-effects modeling, which in case of heterogeneity provides more conservative estimates than fixed effects modeling. Heterogeneity was assessed visually using a forest plot, which represents the estimated prevalence and their relevant 95% confidence interval (CI). The heterogeneity of studies was examined using Cochran's heterogeneity statistic (Q test) and summarized as I^2 . The I^2 values of 25%, 50% and 75% were considered as low, medium, and high heterogeneity, respectively. Publication bias was assessed with funnel plots and Egger's regression test [23]. All the analyses were undertaken using STATA software (Release 12. statistical software. College Station, Texas: STATA Corp LP).

Results

Search strategies

The search strategies resulted in 169 studies, which were reduced to 156 citations after removing the duplicates. The titles and abstracts of these records were screened, and then 142 studies were excluded, and 14 records were selected for full-text review. After a full-text review, two studies that did not have the required information, the number of positive results for SARS-CoV-2 RNA in GI specimens of children, were excluded [24, 25]. Finally, 12 studies met the eligibility criteria and were included in the qualitative synthesis. Two studies did not have the required information to be included in the meta-analysis [17, 18]. Figure 1 shows the study selection process based on the PRISMA guidelines.

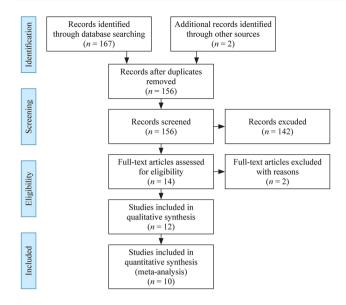


Fig. 1 Summary of the literature search and study selection

Study characteristics

Most studies were performed in China (83%, 10/12), except for two studies that were conducted in Germany and Italy (Table 1). Of the included studies, all had case report/case series designs. All studies were conducted between January and April 2020. In total, 171 children were included in this study for qualitative analysis. All children had respiratory COVID-19 infection and were tested for SARS-CoV-2 RNA in GI specimens. The age of the patients ranged from 8 days to 210 months.

None of the children had severe disease. Of 171 patients, 38 (22%) were asymptomatic, and 41 had no information on symptoms available. The main symptoms were fever (52%, 64/124) and cough (33%, 41/124), and GI manifestations including vomiting, diarrhea, abdominal pain, and constipation were reported totally in 12% (15/124) of children. Nine (75%) studies tested SARS-CoV-2 presence in stool samples, 1 (8%) in rectal swabs, and 2 (17%) in rectal swabs or stool samples. All studies performed real-time RT-PCR to detect SARS-CoV-2 in GI specimens. Further details on study characteristics, along with the QUADAS score, are depicted in Table 1.

Prevalence of fecal SARS-CoV-2 RNA

The pooled prevalence of fecal SARS-CoV-2 RNA in patients with COVID-19 confirmed by respiratory samples was 86% (95% CI 73%–96%) (Fig. 2). The highest prevalence was 100% in three studies, and the lowest was 33% (3/10) in a study by Tan et al. [28]. Eight studies had available data to compare fecal SARS-CoV-2 positivity rate in symptomatic and asymptomatic patients. From symptomatic

and asymptomatic children, 90% (45/50) and 69% (9/13) were positive for fecal RNA, respectively. There was no significant difference in the positivity rate of fecal RNA between symptomatic and asymptomatic patients (P > 0.05).

Persistent shedding of fecal SARS-CoV-2 RNA

All studies had serial SARS-CoV-2 test results for both respiratory and GI specimens, and all reported persistently positive results for SARS-CoV-2 RNA in GI specimens after the virus converted negative in respiratory samples. Nine studies had available data on the prevalence of SARS-CoV-2 RNA in GI samples after respiratory specimen had become negative. From children who had a positive GI specimen test collectively, 72% (43/60) had persistent positive GI specimen test despite negative respiratory test.

SARS-CoV-2 RNA was detected in GI specimens for more than 70 days after onset of illness and more than 5 weeks after hospital admission. In most studies, researchers did not continue follow-up until the point of negative results. One study compared the mean duration of viral shedding in the GI tract between symptomatic and asymptomatic cases, and no significant difference was reported (28.1 \pm 13.3 days vs. 30.8 \pm 18.6 days, P > 0.05).

Results revealed a medium heterogeneity among the selected studies ($I^2 = 62.28\%$; P < 0.01). The funnel plot shows the presence of a possible publication bias. Based on Egger's regression test, the publication bias among included studies could not be ignored (P < 0.001; Fig. 3). The pooled estimates of prevalence might be reduced because of concerns regarding risk of bias (selection bias, detection bias, and attrition bias), and inconsistency of the tested patient populations; which may have contributed to the heterogeneity of findings across studies.

Discussion

We performed a systematic review and meta-analysis to clarify the viral shedding of SARS-CoV-2 through GI specimens in children with positive COVID-19 respiratory tests. In the previous study conducted by Wang et al. [35], GI symptoms and fecal nucleic acid detection in pediatric COVID-19 patients were analyzed. In this study, we performed a meta-analysis to find the prevalence of SARS-CoV-2 RNA in GI specimens. Our research did not identify any studies on children with severe disease. Based on the reports published, most infected children showed mild disease [36], however, approximately 4.4% of infected children have severe disease with a mortality rate of 0.2% [37]. In a study by Mamishi et al. [38], 71% (n=24) of pediatric cases showed severe SARS-CoV-2 infection, and the mortality of 12.5% (n=3) was reported. Based on our results regarding

Authors	Study period	Country	Number of patients with COVID-19	Age	Symptoms	Type of GI specimens	Number of positive patients (GI)	Positive GI but negative respiratory	Max duration positive GI	QUADAS score
Cai et al. [26]	Jan-Apr 2020	China	49	Mean±SD: 11.5±5.12 y	21 asym Cough, fever, sore throat, stuffy nose, rhinorrhea, pneumo- nia, loss of taste and smell	Stool or rectal	45	NS	70 d (admission)	13
Hua et al. [27]	Feb 2020	China	35	NS	NS	Stool or rectal	32	18/35	> 70 d (illness onset)	12
Ma et al. [17]	Jan 2020	China	NS	NS	4 asym Fever	Stool	9	6/6	4 wk (illness onset)	11
Tan et al. [28]	Jan-Mar 2020	China	10	Mean: 7 y	3 asym Fever, cough, abdomi- nal pain, vomiting, constipation, convul- sion	Stool	ε	2/3	>9 d (test)	12
Xu et al. [29]	Feb 2020	China	10	NS	Fever, cough, sore throat, rhinorrhea, diarrhea	Rectal	×	8/8	>27 d (test)	13
Liu et al. [30]	Jan-Feb 2020	China	6	Median (range): 85 (7–139) mon	Cough, fever, sore throat	Stool	8	8/8	66 d (illness onset)	13
Ma et al. [31]	Feb-Mar 2020	China	10	Median (range): 4.7 y (11 mon–14 y and 9 mon)	6 asym Fever, cough	Stool	10	7/10	5 wk (admission)	13
Wolf et al. [32]	Jan 2020	Germany	5	NS	Gastroenteritis, soft stool, fever, vomiting	Stool	5	2/2	4 wk (admission)	12
Xing et al. [20]	Jan-Feb 2020	China	ß	NS	Fever	Stool	Э	3/3	>4 wk (admis- sion)	12
Cai et al. [33]	Jan-Feb 2020	China	9	NS	Fever, cough, sore throat, stuffy nose, sneezing, rhinorrhea	Stool	5	5/5	> 30 d (illness onset)	13
Su et al. [18]	Jan-Feb 2020	China	6	Mean: 4.5 y, Median: 3.5 y	6 NS 1 wild cough 2 mild fever	Stool	SN	5/9	NS	12
De Ioris et al. [34]	Mar–Apr 2020	Italy	22	Median (range): 84 mon (8 d–210 mon)	4 asym Fever, respiratory symptoms, diarrhea, vomiting, seizure	Stool	15	2/15	15 d (illness onset)	13
COVID-19 coron	avirus disease 201	9, QUADAS	quality assessm	lent of diagnostic accurat	COVID-19 coronavirus disease 2019, QUADAS quality assessment of diagnostic accuracy studies, NS not specified, asym asymptomatic infection, GI gastrointestinal, SD standard deviation	d, <i>asym</i> asympto	matic infection,	GI gastrointesti	nal, <i>SD</i> standard de	viation

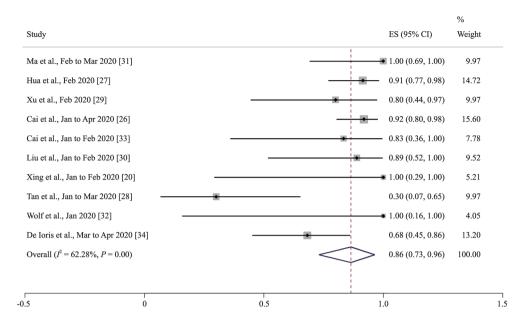


Fig. 2 Forest plot diagram for the pooled prevalence of detectable SARS-CoV-2 RNA in gastrointestinal samples of children with confirmed COVID-19 infection. *COVID-19* coronavirus disease 2019, *SARS-CoV-2* severe acute respiratory syndrome coronavirus 2, *ES* effect size, *CI* confidence interval

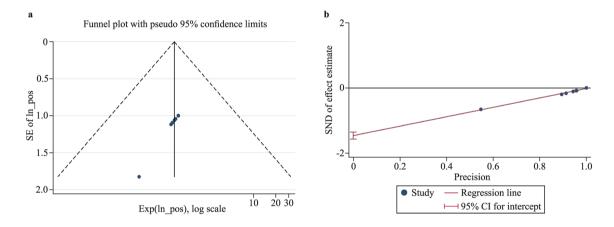


Fig. 3 a The funnel plot diagram for publication bias; b the Egger's test for publication bias ($P \le 0.001$). SE standard error, SND standardized normal distribution, CI confidence interval

the main clinical manifestations, of 124 cases with available data on symptoms, fever was reported in 64 (52%), cough in 41 (33%), and GI symptoms including vomiting, diarrhea, abdominal pain, and constipation in 15 (12%).

In our study, the pooled prevalence of fecal SARS-CoV-2 RNA through GI specimens was 86% in children. Similarly, in systematic reviews by Xu et al. [39] (n=42) and Wang et al. [35] (n=106), the nucleic acid positivity rates of feces in children were 89% and 85.7%, respectively. However, based on the study by Cheung et al. [40] on adult patients (n=4243), the pooled prevalence of fecal RNA was lower (54%). The lower prevalence might be due to the fact that children might present more often GI symptoms compared

to adults, and a higher proportion of fecal SARS-CoV-2 RNA is observed in patients with GI manifestation [16, 41].

All included studies reported persistent detectable RNA in GI specimens after the loss of RNA in the respiratory tract. The positivity rate of fecal RNA after the respiratory tract specimens turned negative was 72% (43/60). In the systematic review by Wang et al. [35], 71.2% (52/73) had persistent positive fecal RNA despite negative respiratory tests. Small studies involving both adults and pediatric cases reported a higher incidence of positive fecal RNA after negative respiratory swab in children compared to adults [18]. However, in the systematic review by Cheung et al. [40], 70.3% (87/124) of adult patients with detectable RNA in stool samples remained positive after the loss of virus from respiratory specimens, which is near to studies on children.

SARS-CoV-2 RNA can be detected through GI specimens of children for more than 70 days. This prolonged shedding observed in pediatric cases also occurs in adults but to a less extent [41]. However, the duration of viral shedding might mainly be due to the host immune status leading to longer viral shedding [14, 42]. According to a previous report, a high level of IL-10 and reduced counts of T-cell and B-cell are associated with prolonged viral RNA shedding [42]. In a study by Ma et al. [17], the possible reasons behind this phenomenon are discussed and considered to be less hand hygiene and different expression of angiotensin converting enzyme 2, a functional receptor for SARS-CoV-2, in the intestine of children. Moreover, high-dose corticosteroids might result in delayed virus clearance and prolonged viral shedding time [14].

The detection of viral RNA in GI specimens does not necessarily indicate that viable infectious virions are present or can transmit through the fecal route [43]. However, some studies demonstrate live viruses through culturing positive SARS-CoV-2 fecal samples [24, 44, 45]. Moreover, since the genetics of SARS-CoV-2 is similar to SARS-CoV that had confirmed fecal–oral transmission, the possibility of a similar transmission route remains to be determined [46]. One limitation of our study is the small number of included studies, with small sample sizes. Studies with higher quality are required to better understand the fecal viral shedding of SARS-CoV-2 in children.

In conclusion, GI shedding of SARS-CoV-2 might occur in a substantial portion of children and also persist long after negative respiratory testing and symptoms resolution. Further research is recommended to find the role of SARS-CoV-2 GI shedding in transmission in children.

Author contributions MM involved in designing and interpretations the manuscript. BS involved in gathering and grouping the articles and writing the draft of the manuscript. MS involved in statistical analysis and revised the manuscript. All authors read and approved the final manuscript.

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Data availability The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethical approval Not applicable.

Conflict of interest The authors declare that they have no competing interests.

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