# **EDITORIAL**



# CMR quantitative measurements of myocardial blood flow: Not ready for routine clinical application

Henry Gewirtz, MDa

<sup>a</sup> Cardiac Unit, Department of Medicine (Cardiology Division), Massachusetts General Hospital, Harvard Medical School, Boston, MA

Received Jun 8, 2019; accepted Jun 10, 2019 doi:10.1007/s12350-019-01812-x

# See related article, pp. 1252-1266

Editorial (Invited):

In this issue of the Journal, Kero et al. report results of quantitative measurements of myocardial blood flow (MBF) obtained with 3 T, PET/MR (CMR; with Gd-DOTA) compared with simultaneously acquired <sup>15</sup>Owater-quantitative MBF. The study was performed in a small (n = 12) group of subjects with known or suspected CAD. CMR MBF [global, regional and myocardial perfusion reserve) was compared to PET measurements, which were considered the gold standard. The authors note that prior studies comparing CMR MBF with PET measurements typically were done on separate instruments at separate times<sup>2</sup> and thus susceptible to physiological variation, which was nullified in the current study by simultaneous data acquisitions with the hybrid PET/MR scanner. What was to all intents and purposes an identical study, save with <sup>13</sup>N-ammonia as the PET tracer, and larger sample size (n = 29), has been previously reported.<sup>3</sup> The CMR pulse sequences employed as well as other specifics (e.g. native T1 signal and hematocrit corrections, signal deconvolution algorithms) have varied between studies in efforts to optimize transformation of CMR signal into true blood and tissue Gd concentrations required for purposes of tracer kinetic modeling. Thus, CMR estimates of MBF are heavily dependent on the accuracy of

Reprint requests: Henry Gewirtz, MD, Cardiac Unit, Department of Medicine (Cardiology Division), Massachusetts General Hospital, Harvard Medical School, Boston, MA 02114; hgewirtz@partners.org

J Nucl Cardiol 2021;28:1267-70.

1071-3581/\$34.00

Copyright © 2019 American Society of Nuclear Cardiology.

these transforms and as noted by others may produce very different estimate of MBF depending on which one is chosen.<sup>3</sup> Further, as noted by the authors of the present study,<sup>1</sup> CMR methodology typically permits sampling of only 4 LV slices in contrast with PET which samples the entire LV. Accordingly, the issue of image registration as well as incomplete LV sampling may further confound efforts to compared CMR and PET estimates of absolute MBF, both global and regional. That said, a particular strength of the present study is the nullification of physiological variation as a source of differences between methods and so offers a more straightforward comparison of the CMR methodology with that of the <sup>15</sup>O-water PET gold standard.

Given that CMR methodology will affect the comparison of MBF measurements with that of PET, what can be learned from results of the present study? As noted above small sample size limits any conclusion(s) drawn. Absence of invasive or CCTA anatomical, and more importantly, invasive, coronary physiological data also is another limitation since such information could shed light on the issue of when the measurements are most likely to agree and when they may diverge with respect to an independent physiological gold standard (e.g. FFR, 4 IFR, 5 IMR<sup>6</sup>). The authors data indicate CMR and PET differences are most pronounced at MBF levels  $\geq \sim 1.8 \text{ mL} \cdot \text{min}^{-1} \cdot \text{g}^{-1}$  (Figures 3A, 4A); an observation which has important implications both for use of the CMR stress MBF measurement for assessment of hemodynamic severity of focal (Ref. # 1, 48-51) and diffuse CAD, as well as MFR (see below).

Indeed, the data of the present study are best understood with reference to Figures 3A, B; 4A, B and 6A, B, which, respectively, demonstrate correlations and Bland–Altman (BA) plots for global (Figure 3) and regional (Figure 4) rest and stress MBF taken together and global MFR (Figure 6). It should be noted dynamic data for both CMR and PET were fit to a one tissue

compartment model with standard corrections applied for PET <sup>15</sup>O-water (Ref. #36). CMR data, however, were fit to a model which incorporated an empirically derived permeability surface area product (PS) based on Renkin-Crone model<sup>8</sup> to correct K1<sup>9</sup> but did not use the complete extraction fraction (EF) since MBF values obtained "...correlated poorly with PET MBF at high values." The EF of Gd-DOTA rolls off markedly beginning at MBF  $\sim 1.25 \text{ mL} \cdot \text{min}^{-1} \cdot \text{g}^{-1}$  (Figure 2A). Although the correlations for global  $(R^2 = 0.74)$  and regional  $(R^2 = 0.56)$  MBF both are at least moderate, the data exhibit considerable scatter around the regression line at higher flow. The BA plots and the authors' own analysis bear this out in recognizing the "wide limits of agreement" between CMR and PET rest and stress MBF measurements. The limits of agreement (95%) for global MBF ranged from -1.24 to +1.25 and for regional values -2.17 to +2.17. Noteworthy, too, is the fact that CMR global rest MBF failed to correlate with PET (Figure 5A) though stress (global, Figure 5C) did  $(R^2 = 0.48)$ . Thus, there was no correlation between MFR determined by CMR and PET (Figures 6A, B). Similar data concerning poor agreement between 3T PET/CMR and PET <sup>13</sup>N-ammonia determined global MFR has been reported by others.<sup>3</sup> In that study, systematically higher CMR rest MBF (vs PET) resulted in consistently lower MFR,<sup>3</sup> whereas in the present study, the differences were random in nature (Figures 6A, B). In light of these findings, it is clear currently employed PET-based stress MBF as well as MFR (commonly referred to as coronary flow reserve CFR, even though strictly speaking it is not) cut points both for diagnosis and prognosis in a variety of cardiovascular disease states<sup>7,10</sup> do not necessarily apply to CMR based measurements of these parameters.

A word about statistical bias in BA plots is in order here. In brief, the mean of differences for each measurement of methods being compared (i.e., CMR vs PET) is equal to bias while ± 1.96 \* SD of the mean defines the limits of agreement, also known as 95% coefficient of reproducibility. 11,12 The authors note bias for CMR vs PET global and regional MBF measurements is "negligible" (i.e. near or equal to zero). However, it is readily apparent CMR may provide an unbiased estimate of PET MBF either because relatively large negative and positive differences cancel and so the average approaches zero, the case in the present study or because the one provides almost identical measurements as the other and so individual differences are near or at zero, hence the mean of difference also approaches zero, certainly not the case in the study under consideration. Accordingly, as the authors correctly note it is the limits of agreement which matter. Characterization of bias as "negligible", while mathematically correct,

uninformative without calling attention to the associated limits of agreement (quite wide), as the authors, to their credit, do. Thus, "negligible" bias in the present study should not be construed as supportive of CMR vs PET measurements of MBF.

Since PET/MR scanners are limited in availability, it is worth considering what results may be obtained with current standalone 1.5 T MR and PET/CT instruments. In a recent study of 21 patients, with known or suspected CAD, rest and stress (adenosine) MBF measurements were made on the same day; once with the 1.5 T MR instrument (Gd-DOTA contrast) and then again with the PET/CT scanner (13N-ammonia, reference method).<sup>2</sup> The correlation between global MBF by CMR and PET was strong  $(R^2 = 0.85, \text{ with})$ slope = 0.94 and intercept = 0.14). Bias was small  $\sim -0.1$  and 95% CR  $\sim \pm 0.8$ . However, there was greater scatter at global MBF  $\geq 2 \text{ mL} \cdot \text{min}^{-1} \cdot \text{g}^{-1}$ . Regional CMR MBF demonstrated even wider scatter. Thus,  $R^2 = 0.69$ , with small bias ( $\sim -0.1$ ) but larger CR ( $\sim \pm 1.1$ ) and many MBF  $\geq 2$  at or beyond the 95% CR limits. Finally, CMR MFR on a global and regional basis correlated only modestly with PET  $(R^2 = 0.48 \text{ and } 0.32, \text{ respectively})$ . Thus, while the overall results of the study indicate closer agreement between CMR and PET measurements of MBF on both global and regional basis than that of current report, both demonstrate (1) closer agreement for global in comparison with that of regional MBF measurements (2) considerably greater divergence of CMR vs PET regional stress MBF at levels  $\geq 2 \text{ mL} \cdot \text{min}^{-1} \cdot \text{g}^{-1}$  and (3) weakest correlations between CMR and PET assessment of both global and regional MFR (nil in the present study and 0.48 and 0.32, respectively, in that of the other<sup>2</sup>).

Earlier work in which CMR estimates of endocardial (endo) and epicardial (epi) MBF was compared to that of fluorescent microspheres understandably has been more encouraging and demonstrated what is possible in human studies as well. Thus, pixel wise (32  $\mu$ L/voxel resolution) CMR images (2-3 slices, 7-mm thick) were obtained which permitted detailed transmural resolution of MBF distribution. After correction for misregistration, CMR MBF correlated quite closely with that of microspheres for transmural, epi and endo layers (all  $R^2 = 0.94$ ) with little bias ( $\sim 0$  to -0.1) though CR was more substantial ( $\sim \pm 1.0$  mL·min<sup>-1</sup>·g<sup>-1</sup> for transmural and endo but ( $\sim \pm 1.5$  for epi with bias  $\sim -0.12$ ). The substantial ( $\sim \pm 1.5$  for epi with bias  $\sim -0.12$ ).

There were, however, important disparities between CMR and microspheres measurements of endo and epi MBF at baseline and with adenosine induced hyperemia. Thus, per Table 1, "remote" CMR rest endo/epi was  $0.99 \pm 0.19$  vs expected  $> 1^{14-16}$  and observed

PET MBF ratio  $1.16 \pm 0.17$ . However, with local hyperemia (ado) CMR endo/epi ratio was essentially unchanged  $(1.05 \pm 0.15)$  compared with baseline, whereas microspheres demonstrated expected greater epi vs endo dilator capacity such that the ratio inverted  $(0.88 \pm 0.14)$ . Both layers it should be noted exhibiting substantial increases in MBF in comparison with that of the unstimulated remote region. Nonetheless, CMR examples provided of patients with known IHD (n = 5)demonstrated excellent correlations with invasive coronary angiography. Expected stress-induced transmural gradients in endo epi MBF were demonstrated as well as transmural MBF deficits relative to baseline. Comparison with an external gold standard (possibly CCTA) was not available. Nevertheless, the methodology clearly holds promise particularly if quantitative MBF data are employed.

Indeed, current practice guidelines consider CMR a class I/II indication for evaluation of stable IHD in patients unable to exercise. 17,18 Validation studies typically have been against invasive coronary angiography (anatomical stenosis generally at 50% diameter reduction cut point<sup>19</sup>) though at least one study used FFR at 0.75 cut point.<sup>20</sup> CMR images were evaluated qualitatively or semi-quantitatively for comparison of stress vs rest defect(s) and presence or absence of scar (late Gd enhancement). 21,22 Qualitative assessment of rest, stress endo/epi contrast distribution also has been reported to enhance CAD detection particularly with multi-vessel CAD.<sup>20,21</sup> Quantitative MBF data, however, generally has not been employed for clinical use. This is unfortunate since the practice considerably degrades the capabilities of the methodology and subjects it to many of the limitations of SPECT MPI in terms of relative imaging comparisons. Though much higher spatial resolution and technically superior images are available with CMR, they are proportionately more difficult and time consuming to obtain and analyze; a problem, along with other administrative and financial issues which together have worked to preclude high capacity, high throughput clinical service in comparison with that of PET, which the current study and others<sup>23</sup> recognizes as the gold standard for quantitative MBF determination in patients.

In conclusion, the data obtained in the current small study<sup>1</sup> suggest CMR methods for quantitative MBF measurements require considerably more development before their full potential is realized and they can offer accurate, reproducible data suitable for high throughput, routine, clinical use. Until then, quantitative PET measurements of MBF will remain the gold standard for clinical practice.

## **Disclosures**

None.

### References

- Kero T, Johannson E, Engdtrom M, Eggers K, Johannson L, Ahlstrom H, Lubberlink M. Evaluation of quantitative CMR perfusion imaging by comparison with simultaneous <sup>15</sup>O-water-PET. J Nucl Cardiol 2019.
- Engblom H, Xue H, Akil S, Carlsson M, Hindorf C, Oddstig J, Hedeer F, Hansen MS, Aletras AH, Kellman P, Arheden H. Fully quantitative cardiovascular magnetic resonance myocardial perfusion ready for clinical use: A comparison between cardiovascular magnetic resonance imaging and positron emission tomography. J Cardiovasc Magn Reson 2017;19:78.
- Kunze KP, Nekolla SG, Rischpler C, Zhang SH, Hayes C, Langwieser N, Ibrahim T, Laugwitz KL, Schwaiger M. Myocardial perfusion quantification using simultaneously acquired (13)NH<sub>3</sub>-ammonia PET and dynamic contrast-enhanced MRI in patients at rest and stress. Magn Reson Med 2018;80:2641-54.
- Pijls NH. Optimum guidance of complex PCI by coronary pressure measurement. Heart 2004;90:1085-93.
- Sen S, Nijjer S, Petraco R, Malik IS, Francis DP, Davies J. Instantaneous wave-free ratio: numerically different, but diagnostically superior to FFR? Is lower always better? J Am Coll Cardiol 2013;62:566.
- Berry C. Fractional flow reserve, coronary flow reserve and the index of microvascular resistance in clinical practice. Radcliffe Cardiology.com. 2014:1-6.
- Gould KL, Johnson NP. Coronary physiology beyond coronary flow reserve in microvascular angina: JACC state-of-the-art review. J Am Coll Cardiol 2018;72:2642-62.
- Renkin EM. Transport of potassium-42 from blood to tissue in isolated mammalian skeletal muscles. Am J Physiol 1959;197:1205-10.
- Gewirtz H, Fischman AJ, Abraham S, Gilson M, Strauss HW, Alpert NM. Positron emission tomographic measurements of absolute regional myocardial blood flow permits identification of nonviable myocardium in patients with chronic myocardial infarction. J Am Coll Cardiol 1994;23:851-9.
- Taqueti VR, Di Carli MF. Coronary microvascular disease pathogenic mechanisms and therapeutic options: JACC state-ofthe-art review. J Am Coll Cardiol 2018;72:2625-41.
- Bland–Altman. Bland Altman Plot. https://www.enwikipediaorg/ wiki/Bland%E2%80%93Altman\_plot. Accessed 6 Mar 2019.
- Bland JM, Altman DG. Statistical methods for assessing agreement between two methods of clinical measurement. Lancet 1986;1:307-10.
- Hsu LY, Groves DW, Aletras AH, Kellman P, Arai AE. A quantitative pixel-wise measurement of myocardial blood flow by contrast-enhanced first-pass CMR perfusion imaging: Microsphere validation in dogs and feasibility study in humans. JACC Cardiovasc Imaging 2012;5:154-66.
- Gewirtz H, Dilsizian V. Integration of quantitative positron emission tomography absolute myocardial blood flow measurements in the clinical management of coronary artery disease. Circulation 2016;133:2180-96.
- Gewirtz H, Gross SL, Williams DO, Most AS. Contrasting effects of nifedipine and adenosine on regional flow distribution and metabolism distal to a severe coronary arterial stenosis: Observations in sedated, closed-chest domestic swine. Circulation 1984;69:1048-57.

- 16. Mills I, Fallon JT, Wrenn D, Sasken HF, Gray W, Bier J, Levine D, Berman S, Gilson M, Gewirtz H. Adaptive responses of the coronary circulation and myocardium to chronic reduction in perfusion pressure and flow. Am J Physiol (Heart Circ Physiol 35) 1994;266:H447-57.
- 17. Fihn SD, Gardin JM, Abrams J, Berra K, Blankenship JC, Dallas AP, Douglas PS, Foody JM, Gerber TC, Hinderliter AL, King SB III, Kligfield PD, Krumholz HM, Kwong RY, Lim MJ, Linderbaum JA, Mack MJ, Munger MA, Prager RL, Sabik JF, Shaw LJ, Sikkema JD, Smith CR Jr, Smith SC Jr, Spertus JA, Williams SV, American College of Cardiology Fellow, American Heart Association Task Force on Practice Guidelines, American College of Physicians, American Association for Thoracic Surgery, Preven-Cardiovascular Nurses Association, Society for Cardiovascular Angiography and Interventions and Society of Thoracic Surgeons. ACCF/AHA/ACP/AATS/PCNA/SCAI/STS Guideline for the diagnosis and management of patients with stable ischemic heart disease: A report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines, and the American College of Physicians, American Association for Thoracic Surgery, Preventive Cardiovascular Nurses Association, Society for Cardiovascular Angiography and Interventions, and Society of Thoracic Surgeons. J Am Coll Cardiol 2012;60:e44-164.
- 18. Messroghli DR, Moon JC, Ferreira VM, Grosse-Wortmann L, He T, Kellman P, Mascherbauer J, Nezafat R, Salerno M, Schelbert EB, Taylor AJ, Thompson R, Ugander M, van Heeswijk RB, Friedrich MG. Clinical recommendations for cardiovascular magnetic resonance mapping of T1, T2, T2\* and extracellular volume: A consensus statement by the Society for Cardiovascular Magnetic Resonance (SCMR) endorsed by the European Association for Cardiovascular Imaging (EACVI). J Cardiovasc Magn Reson 2017;19:75.
- 19. Giang TH, Nanz D, Coulden R, Friedrich M, Graves M, Al-Saadi N, Luscher TF, von Schulthess GK, Schwitter J. Detection of

- coronary artery disease by magnetic resonance myocardial perfusion imaging with various contrast medium doses: First European multi-centre experience. Eur Heart J 2004;25:1657-65.
- 20. Lockie T, Ishida M, Perera D, Chiribiri A, De Silva K, Kozerke S, Marber M, Nagel E, Rezavi R, Redwood S, Plein S. High-resolution magnetic resonance myocardial perfusion imaging at 3.0-Tesla to detect hemodynamically significant coronary stenoses as determined by fractional flow reserve. J Am Coll Cardiol 2011;57:70-5.
- 21. Plein S, Kozerke S, Suerder D, Luescher TF, Greenwood JP, Boesiger P, Schwitter J. High spatial resolution myocardial perfusion cardiac magnetic resonance for the detection of coronary artery disease. Eur Heart J 2008;29:2148-55.
- 22. Vincenti G, Masci PG, Monney P, Rutz T, Hugelshofer S, Gaxherri M, Muller O, Iglesias JF, Eeckhout E, Lorenzoni V, Pellaton C, Sierro C, Schwitter J. Stress perfusion CMR in patients with known and suspected CAD: Prognostic value and optimal ischemic threshold for revascularization. JACC Cardiovasc Imaging 2017;10:526-37.
- 23. Gould KL, Johnson NP, Bateman TM, Beanlands RS, Bengel FM, Bober R, Camici PG, Cerqueira MD, Chow BJ, Di Carli MF, Dorbala S, Gewirtz H, Gropler RJ, Kaufmann PA, Knaapen P, Knuuti J, Merhige ME, Rentrop KP, Ruddy TD, Schelbert HR, Schindler TH, Schwaiger M, Sdringola S, Vitarello J, Williams KA Sr, Gordon D, Dilsizian V, Narula J. Anatomic versus physiologic assessment of coronary artery disease. Role of coronary flow reserve, fractional flow reserve, and positron emission tomography imaging in revascularization decision-making. J Am Coll Cardiol 2013;62:1639-53.

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.