CASE REPORT



Spur cell anemia related to alcoholic liver cirrhosis managed without liver transplantation: a case report and literature review

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Abstract

Spur cell anemia is an acquired hemolytic anemia associated with liver cirrhosis and is characterized by the presence of increased large red blood cells, which are covered with spike-like projections that vary in width, length, and distribution. A 26-year-old man was referred to our hospital presenting with jaundice, lower limb edema, and dyspnea. The patient was subsequently diagnosed with spur cell anemia related to alcoholic liver cirrhosis. Spur cell anemia is an independent predictor of mortality in liver cirrhosis and has been associated with extremely poor prognosis. The most effective treatment for spur cell anemia is liver transplantation. As seen in the literature, the treatment of spur cell anemia without liver transplantation is quite challenging. This report highlights the importance of management and treatment strategies, including control of fluid retention, blood transfusion, plasma diafiltration, and administration of diuretics. Our treatment strategies might be useful in patients who are not candidate of liver transplantation or patients waiting for liver transplantation.

Keywords Alcoholic liver cirrhosis · Case report · Liver cirrhosis · Plasma diafiltration · Spur cell anemia

Introduction

Anemia of diverse etiology is commonly seen in patients with liver cirrhosis. Causes of anemia related to liver cirrhosis are complex and multifactorial. Gastrointestinal hemorrhage secondary to the formation of esophageal and gastric varices, thrombocytopenia, and coagulation disorders are some of the major and life-threatening causes of anemia in liver cirrhosis [1, 2]. In addition, in patients with cirrhosis, portal hypertension gastropathy and gastric vascular ectasia are common causes of chronic anemia [3]. Gastrointestinal hemorrhage may also lead to iron-deficiency anemia in cirrhosis [4]. Hemolysis secondary to splenomegaly, caused by

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portal hypertension, may lead to increased plasma volume, macrocytosis, and megaloblastic anemia [5]. In addition, autoimmune hemolytic anemia associated with autoimmune hepatitis [6], Wilson's disease with increased non-cerulo-plasmin-bound copper released from impaired hepatocytes [7], Zieve's syndrome [8], and spur cell anemia (SCA) [9] are all causes of hemolytic anemia associated with liver disease. Aplastic anemia, characterized by the development of pancytopenia and hypocellular bone marrow, is sometimes associated with hepatitis [10]. Alcoholic liver disease is one of the major causes of liver cirrhosis. Alcohol is known to be toxic to the bone marrow, and up to 60% of alcoholics with cirrhosis also have some degree of malnutrition, which may lead to anemia (e.g., folic acid deficiency, vitamin B12 deficiency) [11] (Table 1).

SCA is an acquired hemolytic anemia associated with liver cirrhosis and characterized by the presence of increased large red blood cells which are covered with spike-like projections that vary in width, length, and distribution. SCA was first reported in 1964 [9, 12]. The prognosis of SCA is extremely poor and liver transplantation is the only curative treatment [13–17]. The treatment of SCA without liver transplantation is quite challenging. We herein report a case

 Table 1
 Causes of anemia

 related to liver disease

Cause of anemia related to liver disease	References	
1. Gastrointestinal bleeding		
a. Esophageal varices, gastric varices, gastric vascular ectasia, etc	[1, 3]	
b. Thrombocytopenia, coagulation disorder	[2]	
2. Hemolysis		
a. Hypersplenism secondary to portal hypertension	[5]	
b. Autoimmune hemolytic anemia associated with autoimmune hepatitis	[<mark>6</mark>]	
c. Wilson's disease with increased non-ceruloplasmin-bound copper	[7]	
d. Zieve's syndrome, spur cell anemia	[<mark>8, 9</mark>]	
3. Aplastic anemia associated with hepatitis	[10]	
4. Alcohol		
a. Bone marrow toxicity	[11]	
b. Malnutrition (e.g. folic acid deficiency, vitamin B12 deficiency)	[11]	

of SCA related to alcoholic liver disease, which was successfully controlled by intensive care management.

Case report

A 26-year-old man was referred to our hospital presenting jaundice, lower limb edema, and dyspnea. He had a drinking history of 200-300 g alcohol per day since 20 years of age. Laboratory test and computed tomography (CT) a year prior to hospitalization showed alcoholic liver disease without decompensated liver cirrhosis. The patient had no familial predisposition to the disease and no past surgical history, history of pancreatitis, or history of blood transfusion. He had jaundice, lower limb edema, and dyspnea since 2 months prior to hospital referral. Despite the progressive increase in symptoms, he did not stop drinking until 1 week prior to referral, by which time he had difficulty in walking due to the lower limb edema and dyspnea. He then visited another hospital before being referred to our hospital. At the time of arrival, the patient was jaundiced, had conjunctival anemia and systemic edema which was especially severe in the lower limbs. The patient was also hypoxemic, and was administered nasal oxygen at 2 L/min. Laboratory test (Table 2) results revealed severe liver dysfunction [Child-Pugh score 11 points, Model for End-stage Liver Disease (MELD) score 24 points, MELD-Na score 28 points] with a total bilirubin of 12.7 mg/dL, direct bilirubin of 5.4 mg/dL, prothrombin time of 30%, and Mac-2 binding protein glycan isomer of 12.16 COI. Brain natriuretic peptide levels were elevated. In addition, the patient had a hemoglobin level of 2.1 g/dL with macrocytic anemia, and a peripheral blood smear revealed the presence of a large number of spur cells (Fig. 1). Haptoglobin was decreased, despite direct and indirect Coombs tests results being normal. No abnormality was detected in upper gastrointestinal endoscopy and bone marrow examination. Chest CT revealed pleural effusion and ground-glass opacity with partial consolidation, suggesting pulmonary edema (Fig. 2a), whilst contrast CT of the abdomen and pelvis revealed chronic liver disease and ascites estimated to be less than 1 L without portal vein thrombosis or hepatocellular carcinoma (Fig. 2b). Pathological findings of liver biopsy were consistent with advanced alcoholic liver cirrhosis (Fig. 3a–c).

With presence of spur cells in the peripheral blood smear, laboratory test, liver biopsy, and exclusion of other causes of anemia and liver disease, the patient was diagnosed with SCA related to alcoholic liver cirrhosis. He was administered a blood transfusion as part of the treatment regimen. In addition, he was supplemented with branched-chain amino acid, levocarnitine, rifaximin, zinc, folic acid, vitamin B1, vitamin B6, vitamin B12, vitamin C, and vitamin B12. Whilst treatment, PaO₂/FiO₂ ratio decreased to 100 and the diagnosis of acute pulmonary respiratory distress was established, so a decision was made to intubate the patient. Hemoglobin level modification was necessary to improve oxygenation. Large amounts of blood transfusions were thought to worsen pulmonary edema, so plasma diafiltration (PDF) was performed during blood transfusion. The PaO_2/FiO_2 ratio improved to > 200, and the furosemide dose was increased to decrease pulmonary edema. The patient experienced a rapid decrease in body weight and was consequently weaned off from ventilatory support. Although MELD score did not improve, there was improvement in anemia and respiratory failure (Fig. 4). The patient was discharged and advised to abstain from alcohol. Laboratory test at the time of discharge showed improvement of hemoglobin level (Hb 8.7 g/dL) and slight improvement of bilirubin level (Total-bilirubin 9.5 mg/ dL, Direct-bilirubin 3.7 mg/dL). Liver function was also improved compared to that on admission (Child-Pugh score 9 points, MELD score 19 points, MELD-Na score 23 points). CT at the time of discharge revealed recovery of pleural effusion and consolidation (Fig. 5a). Subcutaneous

Table 2 Laboratory data on admission

<peripheral blood=""></peripheral>			γ-GTP	208	U/L	Haptoglobin	<10	mg/dL
WBC	4900	/µL	ChE	102	U/L	ANA	<40	×
Neutro	59.2	%	T-chol	215	mg/dL	AMA (M2)	(-)	
Eosi	0.2	%	TG	83	mg/dL	Intrinsic factor Ab	(-)	
Baso	0.2	%	HDL-chol	39	mg/dL	Gastric Parietal Cell Ab	(-)	
Lym	30.7	%	LDL-chol	56	mg/dL	<tumor markers=""></tumor>		
Mono	9.7	%	UA	7.6	mg/dL	AFP	5.9	ng/mL
RBC	41×10^{4}	/µL	BUN	7.1	mg/dL	PIVKA-II	6033	ng/mL
Hb	2.1	g/dL	Cr	0.41	mg/dL	< Viral markers >		
Ht	6.2	%	NH ₃	70	µg/dL	IgM-HA Ab	(-)	
MCV	151.2	fl	Na	131	mEq/L	HBs-Ag	(-)	
MCH	51.2	pg	Κ	2.8	mEq/L	HBs-Ab	(-)	
MCHC	33.9	%	Cl	92	mEq/L	HBc-Hb	(-)	
Plt	10.7×10^{4}	/µL	Fe	146	µg/dL	HBV-DNA (PCR)	(-)	
Ret	5	%	UIBC	12	µg/dL	HCV-Ab	(-)	
<coagulation></coagulation>			Ferritin	1020	ng/mL	HCV-RNA (PCR)	(-)	
PT	30	%	Vit. B ₁₂	1500	pg/mL	IgA-HEV	(-)	
PT-INR	2.11		Folic acid	22	pg/mL	IgM-CMV	(-)	
APTT	37	S	Cu	90	µg/dL	EBV VCA-IgM	< 10	×
FIB	123	mg/dL	Zn	41	µg/dL	EBV VCA-IgG	80	×
FDP	37.1	µg/mL	CRP	1.09	µg/dL	EBNA	< 10	×
D-dimer	13	µg/mL	HGF	0.75	ng/mL	<coombs test=""></coombs>		
ATIII	47	%	M2BPGi	12.16	COI	Direct	(-)	
<biochemistry></biochemistry>			Hyaluronic acid	4050	ng/mL	Indirect	(-)	
ТР	5.9	g/dL	Type IV collagen	19	ng/mL	< Urinalysis >		
Alb	3.0	g/dL	BTR	3.75		Protein	(+++)	
T-Bill	12.7	mg/dL	BNP	874.8	pg/mL	Sugar	(-)	
D-Bill	5.4	mg/dL	KL-6	279	U/mL	Occult blood	(+)	
AST	58	U/L	<immunology></immunology>			Urobilinogen	(++)	
ALT	18	U/L	IgG	1569	mg/dL	Bilirubin	(+)	
LDH	449	U/L	IgA	254	mg/dL	WBC	(-)	
ALP	175	U/L	IgM	87	mg/dL			

WBC white blood cell, *Neutro* neutrophil, *Eosi* eosinophil, *Baso* basophil, *Lyn* lymphocyte, *Mono* monocyte, *Ret* reticulocyte, *RBC* red blood cell, *Ht* hematocrit, *MCV* mean corpuscular volume, *MCHC* mean corpuscular hemoglobin concentration, *Plt* platelet, *PT* prothrombin time, *INR* international normalized ratio, *APTT* active partial thromboplastin time, *FIB* fibrinogen, *FDP* fibrin and fibrinogen degradation products, *ATIII* antithrombinIII, *TP* total protein, *Alb* albumin, *T-Bill* total bilirubin, *D-bill* direct bilirubin, *AST* aspartate aminotransferase, *ALT* alanine aminotransferase, *LDH* lactate dehydrogenase, *ALP* alkaline phosphatase, *GTP* glutamyl transferase, *ChE* cholinesterase, *T-chol* total cholesterol, *TG* triglyceride, *HDL-chol* high-density lipoprotein cholesterol, *LDL-chol* low-density lipoprotein cholesterol, *UA* uric acid, *BUN* blood urea nitrogen, *Cr* creatine, *NH*₃ ammonia, *Na* natrium, *K* kalium, *Cl* cholride, *Fe* ferrum, *UIBC* unsaturated iron-binding capacity, *Vit* vitamin, *Cu* copper, *Zu* zinc, *CRP* C-reactive protein, *HGF* hepatocyte growth factor, *M2BPGi* mac-2-binding protein glycan isomer, *BTR* ratio of branched-chain amino acid to tyrosine, *BNP* brain natriuretic peptide, *KL-6* Krabs von den Lungen-6, *ANA* anti-nuclear antibody, *AMA* anti-mitochondrial antibody, *AFP* alfa-fetoprotein, *PIVKA-II* protein-induced by vitamin K absence or antagonist-II, *HA Ab* hepatitis A antibody, *HBs-Ag* hepatitis B surface antibody, *HBc-Ab* hepatitis B core antibody, *HBV-DNA* hepatitis B virus deoxyribonucleic acid, *PCR* polymerase chain reaction, *HCV-Ab* hepatitis C virus antibody, *HCV-RNA* hepatitis C virus ribonucleic acid, *HEV* hepatitis E virus, *CMV* cytomegalovirus, *EBV* VCA-*IgG* Ab epstein–barr virus-viral capsid antigen immunoglobulin M antibody, *EBV* VCA-*IgG* Ab epstein–barr virus virus nuclear antigen

edema was remarkably improved, without obvious change in liver shape (Fig. 5b). A year on from initial hospitalization, MELD score and spur cell rate in peripheral blood smear did not improve. Though, the patient succeeded in abstaining from alcohol and is doing well without exacerbation of hemoglobin level.

Discussion

Several abnormal red blood cells such as acanthocytes, echinocytes, target cells, stomatocytes, and spur cells are seen in patients with liver cirrhosis [17]. Although the precise mechanism is unknown, the central role of abnormal lipid

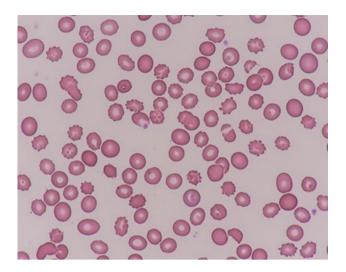


Fig. 1 Peripheral blood smear (May-Giemsa stain, $\times 1000$) revealed approximately 25% of spur cells with multiple spicules irregularly distributed over the red blood cell

metabolism in the formation of abnormal red blood cells (including spur cells and target cells) has been demonstrated by in vitro observation [18]. SCA was originally reported as a severe form of hemolytic anemia, with the presence of spur cells on a peripheral blood smear [9]. Recently, various etiologies of liver cirrhosis were reported as causes of SCA, which is shown to occur in any etiology of liver cirrhosis [19–21]. The precise mechanism of SCA in liver cirrhosis is still unknown, although abnormal ratio of cholesterol and phospholipids in red blood cell membrane is thought to be the main cause of SCA [9, 15]. According to previous reports, the diagnosis of SCA differ [9, 12, 15, 17, 22]. In a recent study, patients presenting with > 5% spur cells in peripheral blood were diagnosed with SCA [22]. In addition, the exclusion of other causes of anemia is an essential

Fig. 3 Pathological findings of liver biopsy. **a** (hematoxylin and eosin stain, scale bar: 50 μ m): hematoxylin and eosin stain shows hepatocellular ballooning and Mallory bodies (blue arrowhead). **b** (periodic acid-Schiff stain, scale bar: 250 μ m), **c** (Azan stain, scale: same as **b**): periodic acid-Schiff stain and Azan stain show lobular distortion with scattered small hepatic cell nests. Note that fatty change of hepatocyte is minimal

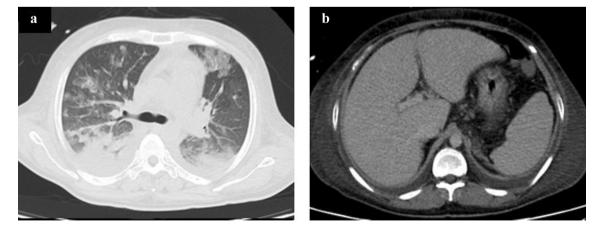
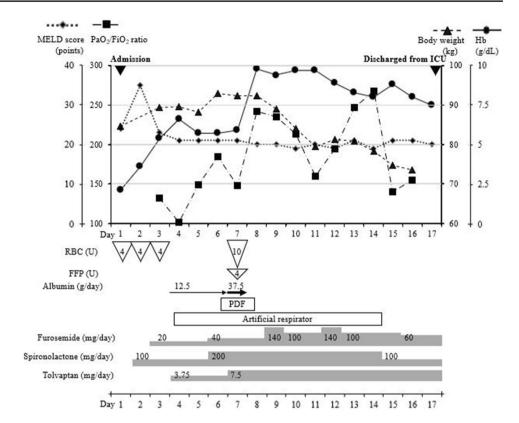


Fig.2 a Chest computed tomography (CT) revealed pleural effusion and ground-glass opacity with partial consolidation suspecting pulmonary edema. b Contrast CT of the abdomen and pelvis revealed

chronic liver disease and ascites estimated to be less than 1 L without portal vein thrombosis or hepatocellular carcinoma

Fig. 4 Blood transfusion and fluid retention aggravated PaO₂/ FiO₂ ratio. Intubation, plasma diafiltration (PDF) during blood transfusion, high-dose diuretics improved anemia, oxygenation, and body weight. Without liver transplantation, Model for End-stage Liver Disease (MELD) score and presence of spur cell in peripheral blood smear did not improve, although symptoms related to anemia and liver cirrhosis were successfully managed. FFP fresh frozen plasma, MELD Model for End-stage Liver Disease, PDF plasma diafiltration, RBC red blood cell, U Unit



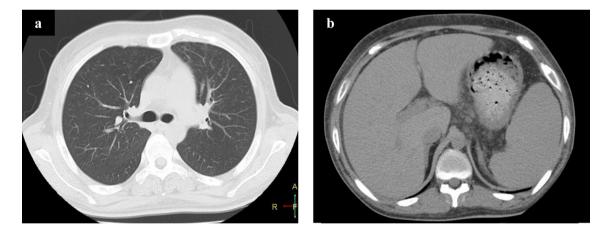


Fig. 5 a Chest computed tomography (CT) at the time of discharge revealed improvement of pleural effusion and consolidation. b Abdominal CT at the time of discharge showed improved subcutaneous edema. Though, there was no obvious change in liver shape

step in the diagnosis of SCA. The prevalence of SCA among patients with liver cirrhosis is still unclear. Vassiliadis et al. reported that 16.7% (9 out of 54) of patients with liver cirrhosis with a Child–Pugh score > 7, had SCA [12]. Alexopoulou et al. reported that 31% (36 out of 116) of patients with decompensated liver cirrhosis, had SCA [22]. When compared with patients without SCA, patients with SCA had worse liver function (higher MELD score) [12, 22], lower hemoglobin levels [12], and worse survival [12, 22]. Alexopoulou et al. also reported that in multivariate analysis, SCA was an independent predictor of mortality, and survival rate of patients with SCA at the first, second, and third month of follow-up was 77%, 45%, and 33%, respectively, with a median reported survival of 1.9 months [22]. Vassiliadis et al. reported a 25-day median survival rate in the 9 patients with SCA, and only one patient, who had undergone liver transplantation, had survived after 1 year [12]. To our knowledge, there have been 24 case reports on 26 patients

with SCA since 1983 including our case (Table 3) [13, 14, 16, 19, 20, 23–40]. On reviewing these cases, the average age was 42.6 years old, and 14 (53.8%) of the 26 patients were men. Most of the cases lack the data of severity of liver disease, such as Child–Pugh score and Model for End-stage Liver Disease (MELD) score. Though, assessable cases had severe liver disease with average Child–Pugh score 11 points, and average MELD score 26.3 points. The average hemoglobin level was 7.5 g/dL and our case had the lowest hemoglobin level. Patients underwent liver transplantation experienced recovery of SCA and had good prognosis. However, patients without liver transplantation had poor survival time (from 2 weeks to 5 months), except for our case (doing

well for a year without aggravation of liver disease and hemoglobin level). As seen in the literature, the prognosis of SCA is extremely poor and management of SCA is essential for better survival in end stage liver cirrhosis.

As for treatment, abstinence from alcohol is required to avoid the progression of alcoholic liver cirrhosis. Liver transplantation remains the only curative treatment for reversing SCA [13–17], and very few attempts have been made for treatment without liver transplantation. Flunarizine is reported to impair the morphological abnormality of red blood cells, and two cases of SCA treated by flunarizine are reported. In addition, there has been a case of SCA that was treated by combination therapy comprising

Table 3 Characteristics of patients with spur cell anemia

No	Age	Sex	Etiology	Child– Pugh score	MELD score	Hemo- globin (g/ dL)	Treatment	Prognosis	References
1	26	М	Alcohol	11	24	2.1	Blood transfusion, albumin infusion, plasma diafiltra- tion, diuretics, intubation	Doing well for a year	Our case
2	31	М	Alcohol	NA	NA	9.0	Blood transfusion	Succumbed in 2 weeks	[23]
3	52	М	Alcohol	12	29	7.9	Blood transfusion, plasma- pheresis, albumin infusion	Discharged	[24]
4	64	Μ	NAFLD	10	NA	6.6	High-dose steroids, dialysis	Discharged	[25]
5	9	F	Wilson's disease	NA	NA	8.9	NA	Succumbed within 10 days	[19]
6	44	М	Alcohol	11	27	7.4	NA	Waiting for LT	[26]
7	47	F	Alcohol	NA	NA	NA	Blood transfusion	NA	[27]
8	44	F	PBC/AIH overlap	NA	30	9.0	Blood transfusion, plasma- pheresis	Waiting for LT	[28]
9	60	М	Alcohol	NA	38	6.1	Blood transfusion	Succumbed in 5-months	[29]
10	32	F	Alcohol, HCV	NA	NA	6.2	Blood transfusion	Waiting for LT	[<mark>30</mark>]
11	56	F	NA	NA	NA	6.8	Blood transfusion	Succumbed	[31]
12	61	F	PBC	NA	NA	6.2	NA	NA	[20]
13	34	М	Alcohol	NA	NA	NA	LT	Doing well	[13]
14	17	F	Biliary atresia	NA	NA	5.7	LT	Doing well	[14]
15	47	М	Alcohol	NA	NA	7.3	Blood transfusion	Waiting for LT	[32]
16	43	F	Alcohol	NA	40	11.2	NA	Succumbed in 2-months	[33]
17	48	М	NA	NA	NA	7.7	NA	Succumbed before LT	[34]
18	30	М	Alcohol	NA	NA	7.7	Flunarizine, pentoxifylline, cholestyramine	Recovery of anemia	[35]
19	41	F	Alcohol	NA	NA	8.4	Diuretics	Lost of follow-up	[36]
20	31	М	Alcohol	NA	NA	NA	LT	Doing well a year later	[<mark>16</mark>]
21	53	М	Alcohol	NA	NA	8.5	LT	Recovery of anemia	[37]
22	44	F	Alcohol	NA	NA	9.0	LT	Doing well 3-months later	[37]
23	49	М	Alcohol	NA	NA	NA	NA	NA	[38]
24	52	М	Alcohol	NA	NA	7.9	NA	Succumbed	[<mark>39</mark>]
25	47	F	Alcohol	NA	NA	6.2	Flunarizine	Recovery of anemia	[40]
26	45	F	Alcohol	NA	NA	8.3	Flunarizine	ND	[40]

AIIH autoimmune hepatitis, HCV hepatitis C virus, F female, LT liver transplantation, M male, MELD Model for End-Stage Liver Disease, NA not available, NAFLD non-alcoholic fatty liver disease, PBC primary biliary cholangitis

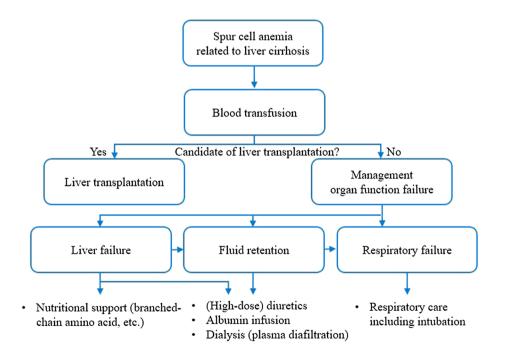
flunarizine, pentoxifylline, and cholestyramine, although flunarizine and pentoxifylline had to be discontinued 6 months later due to adverse events [35]. High-dose steroids were also reported to be effective in improving anemia in SCA [25]. Plasmapheresis has been reported to improve dyslipidemia in SCA (increase in concentrations of total cholesterol, phospholipids, and apo-AI), and as an effective treatment of SCA [24]. However, the precise mechanism of effectiveness in these cases is still unknown, and liver transplantation is the only curative treatment to reverse the morphological feature of SCA.

In our case, acute respiratory distress syndrome occurred secondary to SCA related to alcoholic liver disease. There are several causes of acute respiratory distress syndrome considered in our case. First, there was a possibility of acuteon-chronic liver failure in our case [41]. Acute-on-chronic liver failure patients experience systematic inflammatory response syndrome because of cytokine storm in relation acute insult and/or subsequent development of sepsis [42]. In our case, blood culture remained negative, so sterile inflammation response or occult infection could be considered. However, our case was not visiting our hospital regularly and we lacked the laboratory data for the diagnosis of acuteon-chronic liver failure in Japan. Secondary, fluid retention (e.g., ascites, pleural effusion, edema) are common complications of end stage liver cirrhosis, and could cause respiratory failure. The mechanism of fluid retention in cirrhosis is complicated, although the major causes can be explained by malnutrition and increased activity in the renin-angiotensin-aldosterone system, which leads to hyperdynamic circulation [43, 44].

We successfully managed SCA without liver transplantation. Blood transfusion is necessary to improve oxygenation and increase the hemoglobin level. Liver transplantation is the only curative treatment of spur cell anemia. According to the Japan society of hepatology, the candidate of liver transplantation should be considered by liver function (Child-Pugh score 10 points or more, of MELD score), period of alcohol abstinence (more than 18 months), and hepatocellular carcinoma (within the Milan criteria [45] or within the 5-5-500 rule [46]). When the patient is not a candidate of liver transplantation, SCA, which could be accompanied with fluid retention, organ failure, such as liver failure, respiratory failure (aggravated by aggressive transfusion), must be managed by preserved treatment. During treatment, nutrition support for liver failure, respiratory care, including intubation, for respiratory failure, and fluid retention control, including diuretics, albumin infusion, plasma diafiltration (PDF), was helpful (Fig. 6). In addition, PDF, a blood purification therapy in which simple plasma exchange is performed using a selective plasma separator while the dialysate flows outside the hollow fibers [47], was useful in the removal of water-soluble and albumin-bound toxins [47]. The benefit of PDF compared to simplex plasma exchange is that PDF enabled the performance of dialysis simultaneously with plasma exchange, without worsening fluid retention [47].

Several limitations to our report exist. First, Japanese health care insurance does not cover PDF in chronic liver disease, and careful informed consent must, therefore, be obtained before initiating treatment. Secondary, our report luck the evaluation of cholesterol/phospholipid ratio in red

Fig. 6 Blood transfusion is necessary for the management of spur cell anemia. Liver transplantation is the only curative treatment of spur cell anemia, considered by liver function, period of alcohol abstinence, and hepatocellular carcinoma. When the patient is not a candidate of liver transplantation, nutrition support for liver failure, respiratory care, including intubation, for respiratory failure, and fluid retention control, including diuretics, albumin infusion, plasma diafiltration, was helpful for management during blood transfusion



bold cell membrane, serum chenodeoxycholic acid level and serum deoxycholic + cholic acid/chenodeoxycholic acid ratio, which is observed in SCA [16]. However, recent diagnostic criteria of SCA do not include these laboratory data [22]. Finally, although our treatment strategies successfully managed severe SCA with alcoholic liver cirrhosis, SCA could not be reversed without liver transplantation. Liver transplantation is essential in severe SCA accompanied by worse liver function as curative treatment, and clinicians should look for the possibility of liver transplantation in candidates of liver cirrhosis. Our managing strategies might be useful in patients with SCA who is not a candidate of liver transplantation or patients waiting for liver transplantation.

In conclusion, blood transfusion and control of fluid retention were successful in managing SCA related to alcoholic liver cirrhosis. This finding, which is dependent on the assumption of abstinence from alcohol, warrants further investigations for the management of SCA without liver transplantation.

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Author contributions TM treated the patient and wrote the paper; YH evaluated pathologically; TK, MA, KT, HG, NN, NK, and KI supervised the treatment; MS mainly reviewed and amended the manuscript; all authors issued final approval for the version to be submitted.

Compliance with ethical standards

Conflict of interest Takao Miwa, Yuichiro Hatano, Takahiro Kochi, Masashi Aiba, Katsuhisa Toda, Hideko Goto, Noriaki Nakamura, Naoki Katsumura, Kenji Imai, and Masahito Shimizu declare that they have no conflict of interest.

Human and animal rights All procedures followed have been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments.

Informed consent Informed consent was obtained from the patient for publication of this case report.

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