REVIEW



Epidemiology of Mutations in the 65-kDa Retinal Pigment Epithelium (*RPE65*) Gene-Mediated Inherited Retinal Dystrophies: A Systematic Literature Review

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ABSTRACT

Introduction: Inherited retinal dystrophies (IRDs) represent a genetically diverse group of progressive, visually debilitating diseases. Adult and paediatric patients with vision loss due to IRD caused by biallelic mutations in the 65-kDa retinal pigment epithelium (*RPE65*) gene are often clinically diagnosed as retinitis pigmentosa (RP), and Leber congenital amaurosis

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J. M. F. Sallum Instituto de Genética Ocular, São Paulo, Brazil (LCA). This study aimed to understand the epidemiological landscape of *RPE65* gene-mediated IRD through a systematic review of the literature, as the current evidence base for its epidemiology is very limited.

Methods: Medline, Embase, and other databases were searched for articles on the epidemiology of *RPE65* gene-mediated IRDs from inception until June 2021. Studies were included if they were original research articles reporting the epidemiology of RP and LCA and/or proportion of *RPE65* gene mutations in these clinically diagnosed or molecularly confirmed IRDs patients.

Results: A total of 100 studies with relevant data were included in this systematic review. The range for prevalence of LCA and RP in the literature was 1.20-2.37 and 11.09-26.43 per 100,000, respectively. The proportion of *RPE65* mutations in clinically diagnosed patients with LCA was found to be between $\sim 2-16\%$ within the US and major European countries (France, Germany, Italy, Spain, and the UK). This range was also comparable to our findings in the Asian region for RPE65-LCA (1.26–16.67%). Similarly, for these European countries, RPE65-RP was estimated between 0.23 and 1.94%, and RPE65-IRD range was 1.2–14%. Further, in the Americas region, mutations in RPE65 were reported to cause 1-3% of RP and 0.8-3.7% of IRD cases. Lastly, the RPE65-IRD range was 4.81–8% in the Middle East region.

Conclusions: There are significant variations in reporting of *RPE65* proportions within

countries as well as regions. Generating robust epidemiological evidence on *RPE65* gene-mediated IRDs would be fundamental to support rare disease awareness, timely therapeutic intervention, and public health decision-making.

Keywords: Epidemiology; Inherited retinal dystrophies (IRD); Leber congenital amaurosis (LCA); Prevalence; Retinitis pigmentosa (RP); *RPE65* gene; Systematic review

Key Summary Points

Robust epidemiology data for *RPE65*mediated inherited retinal dystrophies (IRD) is limited and therefore accurate assessments of prevalence and incidence are challenging.

The prevalence of Leber congenital amaurosis (LCA) was estimated to be 1.20–2.37 per 100,000.

The proportion of *RPE65* mutations in clinically diagnosed cases of LCA:

European region: ranged between 1.79 and 22.22%. Americas region: ranged between 1.69 and 15.55%. The US and major European countries (France, Germany, Italy, Spain, and the UK): ranged between \sim 2 and 16%. Asian region: ranged between 1.26 and 16.67% (comparable to US and major European countries findings).

The prevalence of retinitis pigmentosa (RP) ranged between 11.09 and 26.43 per 100,000.

The proportion of *RPE65* gene mutation in clinically diagnosed cases with RP:

European region: ranged between 0.23 and 4.27%. Americas region: ranged between 0.81 and 3.28%. Major European countries (France, Germany, Italy, Spain, and the UK): ranged between 0.23 and 1.94%. The US: ranged between 0.81 and 1.85%.

INTRODUCTION

Inherited retinal dystrophies (IRDs) comprise a wide range of phenotypically and genetically heterogeneous group of rare genetic diseases that are generally characterised by progressive loss of vision [1, 2]. Mutations in more than 270 different genes have been identified as the cause of IRDs [3–5]. Among these, biallelic mutations in the RPE65 gene, i.e., the gene that encodes the 65-KDa retinal pigment epithelium (RPE) mutations affect the visual cycle in the retinal epithelium, resulting in a progressive loss of photoreceptors. Rods and cones are two main types of photoreceptor cells. Rods are mainly found in the peripheral regions of the retina and are responsible for peripheral and night vision. Cone density is higher at the macular area and is responsible for colour vision and perception of fine details.

The gene RPE65 retinoid isomerohydrolase is associated with three Online Mendelian Inheritance in Man (OMIM #180069) phenotypes, (1) autosomal recessive Retinitis pigmentosa 20 (RP), (2) autosomal recessive Leber's congenital amaurosis 2 (LCA), and (3) autosomal dominant Retinitis pigmentosa 87 with choroidal involvement [6]. While signs and symptoms are heterogeneous, RP is diagnosed in patients with gradual rod photoreceptor degeneration and good central vision within the first decade of life. RP is characterised by visual field (VF) loss and nyctalopia (poor night vision), and may progress to blindness [7]. In contrast, LCA is diagnosed in patients with rod-cone dystrophy who were either born blind or lost their low vision within the first year of life, and is characterised by a severe dystrophy of the retina [8–11]. LCA is mostly inherited as an autosomal recessive form, whereas RP can have any of the commonly recognized Mendelian inheritance patterns or maternal (mitochondrial) or digenic inheritance [6]. However, when RP is due to RPE65 biallelic variants, it has an autosomal recessive pattern of inheritance.

RP does not show any ethnic predilection; however, the most frequent pathogenic variants for RP-associated genes may vary for certain populations with a high rate of consanguinity

[12]. Further, it seems that there is no universally accepted diagnostic term for patients with retinal degeneration who lose vision during the first few years of life; various diagnostic terms were used in the literature with merely any genotypic differences, such as LCA type 2, earlyonset severe retinal dystrophy (EOSRD), autosomal recessive childhood-onset severe retinal dystrophy (arCSR), autosomal recessive retinal dystrophy, severe early childhood-onset retinal dystrophy (SECORD), and/or early severe RP. EOSRD/SECORD is defined as a severe retinal dystrophy presenting after the first year of life and usually before the age of 5 years, whereas LCA is congenital or presents within the first few months of life, and both are characterised clinically by severe congenital/early infancy low vision, nystagmus, amaurotic pupils and markedly reduced/absent full-field electroretinograms [6, 13–15].

Although various terms are being used for clinical classification of the disease, the key for specifying a disease should be based on genetic testing, genotyping or molecular diagnosis [16]. It is also important to evaluate and classify the identified variants as pathogenic, non-pathogenic or likely pathogenic; or classify based on the American College of Medical Genetics and Genomics and the Association for Molecular Pathology guidelines for genetic variants [17, 18]. The pathogenicity classification of the variants and novel variants in the RPE65 gene are described in some of the online databases for variants in human genetic mutations, which can support the molecular diagnosis of the disease, genetic counselling and medical management of the patient's condition [17, 19-21]. A natural history study highlighted the need for genetic testing, as it reported that there was limited information on phenotype correlations related to biallelic RPE65 mutations, and that a number of clinical diagnostic terms were used for the same genotype with wider variation across all the different types of LCA [13].

The current evidence base for the epidemiology of *RPE65* gene-mediated IRDs is very limited. Such epidemiological evidence will be crucial for evaluating the impact of the disease in the population in terms of the burden of disease and identifying unmet clinical needs. Thus, this study aims to understand the epidemiology landscape of *RPE65* gene-mediated IRD, and to identify key knowledge gaps/unmet needs through a systematic review of the literature, focussing on mutations in the *RPE65* gene that are often clinically diagnosed as LCA and/or as RP [22, 23].

METHODS

Literature Search Strategy and Selection Criteria

A review of the medical literature was conducted as per the guidance of the Cochrane handbook for systematic reviews [24] and was reported in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) [25]. The electronic databases (Embase, Medline, Medline in-process and Cochrane Library) were searched from inception until the 25 June 2021 to retrieve studies reporting the prevalence and incidence of LCA and RP and the proportion of RPE65 gene mutations in these IRDs patients. As RPE65 gene-mediated IRDs are often clinically diagnosed as LCA or RP, the literature search was not designed to investigate other types of IRDs due to insufficient information in the publications. The Orphanet rare diseases platform [26, 27] and the bibliography of relevant literature reviews was also screened for including potential studies. Additionally, conference abstracts were hand-searched from the publication years of 2015 to June 2021 to retrieve the latest studies that have not yet been published in journals as full text articles or to supplement results of previously published studies. This article is based on previously conducted studies and does not contain any new studies with human participants or animals performed by any of the authors.

The databases were searched for terms related to *RPE65*, *RPE65*-IRD, *RPE65*-RP, *RPE65*-LCA, incidence, prevalence and/or epidemiology. The search results were limited to English language. The detailed literature search strategy for the various databases is presented in supplementary file 1. Studies related to *RPE65*

clinical trials, animal or in-vitro studies, case reports, review papers, studies describing pathogenicity/ molecular genetic features caused by RPE65 mutations in a family or studies with mutations other than RPE65, etc. were excluded. Titles and abstracts of all the unique citations obtained were screened by two independent reviewers, and any discrepancies between the reviewers were reconciled by a third independent reviewer. The citations that did not match the eligibility criteria were excluded at the 'first pass', while unclear citations were included. Duplicates of citations (due to overlap in the coverage of the databases) were also excluded at the first pass stage. The full-text copies of all references that could potentially meet the eligibility criteria were downloaded.

Publications were included in the full-text review if they reported on prevalence/incidence of RP or LCA (irrespective of any mutation) or epidemiology of IRDs caused by RPE65 gene mutations or proportion of RPE65 gene mutation in RP or LCA. Additionally, some papers had presented results for LCA, EOSRD, severe early-onset retinal dystrophy (SEORD) and other similar terms separately, although there is no clear distinction between these diseases. In such cases, the data were included for patients identified with only LCA, RP, RPE65-IRD and/or related molecular diagnosis from populations to avoid discrepancies with such study methods. The percent proportions of the RPE65 mutation within RP or LCA or IRD patients were calculated (if not explicitly mentioned in the paper) as the number of patients with RPE65 mutations in a disease divided by the total number of patients with that disease.

The data from the included studies were extracted to data extraction grids by one reviewer, and checked by a second independent reviewer, with reconciliation of any differences by a third independent reviewer.

RESULTS

The literature search from the databases yielded 1066 citations from which 98 were removed as duplicates. Following the first pass of the citations, 118 potentially relevant references were

identified for the second pass. Following detailed examination of these full-text articles, 65 were identified for inclusion after the second pass. Additionally, 35 were obtained by screening of bibliography citations and from hand-searching of relevant conference abstracts including two Orphanet site links [26, 27]. Thus, the final number of included citations for this review were 100 (Fig. 1).

While extracting the data from included papers, it was noted that some papers were reporting the study population for LCA, RP or IRD based on a diagnostic method, i.e., by either clinical and/or molecular/genetically confirmed diagnosis of the disease. This resulted in differences in the calculated proportions of *RPE65*-affected patients out of the study population. Thus, where applicable, we report both the clinical and/or the molecular data for a country in Tables 1 and 2 by individual patients as well as by families affected by the disease.

While reporting the percentage proportions, we tried to distinguish the populations with the relatively high prevalence of consanguineous marriages [28–31]. It was found that patients with high rate of consanguinity and those affected with recessive disease will be homozygous for their mutant allele [32].

Few papers only reported the proportion of *RPE65* gene mutation in patients with combined cohorts of RP and LCA patients (Supplementary Table A) and the proportion of *RPE65* gene mutation in an IRD cohort (Supplementary Table B).

Prevalence of LCA and the Proportion of *RPE65* Gene Mutation in LCA

The prevalence of LCA has been reported in three studies conducted in Denmark, Norway and the United States of America (US), and was estimated to be 1.20–2.37 per 100,000 [33–35].

The worldwide proportion of *RPE65* gene mutations in LCA families was estimated at 6.10% [36]. Within the clinically diagnosed cases of LCA, the proportions of *RPE65*-LCA across the global regions varied from 1.26% in China to 22.22% in the Netherlands [37, 38]. The proportion of *RPE65* mutations in LCA

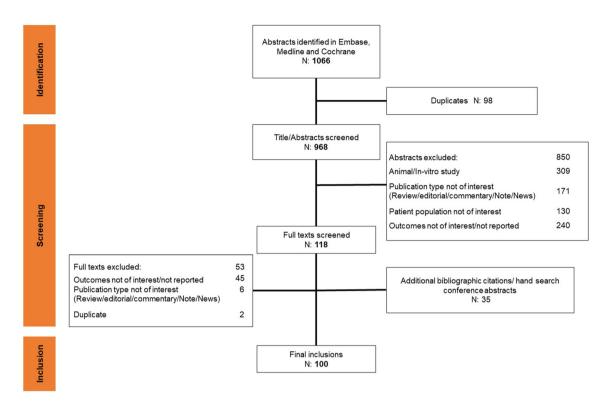


Fig. 1 PRISMA flow diagram of included and excluded publications

ranged between 1.79 and 16% in the five major European (EU-5) countries of France, Germany, Italy, Spain, and the United Kingdom (UK) [5, 39-44] and between 3.0 and 15.55% in the US [32, 45–49] (Fig. 2). A recent Costa Rican study [50] reported a very high prevalence of RPE65-LCA mutations at 95% which although outlier from our defined an ranges (1.26–22.22%) has nevertheless been included in Fig. 2. This phenomenon could be a genetic drift that may be due to a founder effect in the analysed samples from only the affected children and their immediate family members. Further, a Canadian hospital study reported 35.82% of RPE65-LCA mutations based on its sample analysing only the patients diagnosed with LCA [51]. The proportion of *RPE65* mutations in molecularly diagnosed LCA patients was based on a few studies varying from 3.95% in China to 20.51% in Brazil and up to 40% in Tunisia [37, 52–59] (Table 1).

Prevalence of RP and the Proportion of *RPE65* Gene Mutation in RP

Prevalence of RP was reported to range from 11.09 to 26.43 per 100,000 in the literature [34, 60–72] (excluding 47.62 per 100,000 for Israel's population due to high consanguinity [73]) (Fig. 3).

In the US, the prevalence of RP was 21.03 per 100,000 [65]; in the UK, it was between 20.54 and 25.03 per 100,000 [62, 66], while in the European region it ranged from 11.97 to 25.36 per 100,000 [61, 70]; for the Asian countries of China and Korea, it was the same as the global prevalence, i.e., 11.09–26.43 per 100,000 [60, 72]. For the Middle East country of Israel, it was high (47.62 per 100,000 [73].

It was also noted that populations with high rates of consanguinity and a high number of siblings per family report a much higher

Country	Proportion of <i>RPE65</i> mutations in LCA	Author, year of publication	Remarks
Ex-US and	Ex-EU range for molec	ularly diagnosed patien	ts was 3.95-20.51%
Chinaª	3.95 ^a -15.0% ^a	Xu et al. [37] and Li et al. [52]	<i>RPE65</i> in 3/76 probands with LCA and the frequency of <i>RPE65</i> was at 15% in LCA/EOSRD/LCA + EOSRD patients
Oman ^ª	7.41% ^a	Bruwer [53]	2 siblings out of 27 patients with LCA in consanguineous population genetically identified with disease-causing genes
Saudi Arabia	8.70% ^a	Khan et al. [54]	<i>RPE65</i> gene mutations identified in 2/23 consanguineous LCA children
India ^a	11.11% ^a	Viswarubhiny et al. [55]	RPE65 gene mutations identified in 1/9 patients with LCA
Mexico ^a	13.33% ^a	Zenteno et al. [56]	<i>RPE65</i> gene variants were identified in 2/15 patients with LCA
Brazil ^a	19.08 ^a -20.51% ^a	Motta et al. [57] and Sallum et al. [58]	<i>RPE65</i> gene mutations were identified in 16/78 and 29/152 patients with LCA
Tunisiaª	40.0% ^a	El Matri et al. [59]	RPE65 gene mutations were identified in 6/15 patients with LCA
Data as rep	ported for molecularly di	agnosed families	
Pakistan ^a	7.14% ^a	McKibbin et al. [127]	RPE65 in 1/14 families diagnosed molecularly with LCA
UK ^a	12.18% ^a	Hull et al. [128]	<i>RPE65</i> gene mutations were identified in 24/197 EOSRD families genetically identified with disease-causing genes
Poland ^a	13.64% ^a	Skorczyk-Werner et al. [129]	<i>RPE65</i> gene variants were identified in 3/22 families diagnosed molecularly with LCA

Table 1 Proportion of RPE65 gene mutations in molecularly diagnosed cases with LCA

EOSRD early-onset severe retinal dystrophy, EU European Union, LCA Leber congenital amaurosis

^aThe % proportion is based on molecularly diagnosed patients

Key messages from the table on the proportions of RPE65 gene mutations in molecularly diagnosed LCA cases:

The proportions of *RPE65*-LCA mutations varied across the world. It was 3.95–15% in China; 8.7% in Saudi Arabia; 13.33% in Mexico; 20.51% in Brazil; and up to 40% in Tunisia

The frequency of *RPE65* mutations in a Chinese cohort of patients with LCA and EOSRD was 15% not only in the cohort total patients but also in LCA and/or EOSRD patient groups

Country	Proportion of <i>RPE65</i> mutations in RP	Author, year of publication	Remarks
European coun	tries range for clinically	diagnosed was 0.23-4.27%	
Germany	0.23%	Weisschuh et al. [42]	One <i>RPE65</i> pathogenic variant in 434 patients with sporadic RP
Spain	1.02%	González-del Pozo et al. [74]	Potentially pathogenic 1 <i>RPE</i> 65 variant detected out of 98 ARRP patients
France	1.11%	Bocquet et al. [75]	<i>RPE</i> 65 was found in 1/90 ARRP individuals with consanguineous parents
Netherlands	1.55-4.27%	Haer-Wigman et al. [76] and Pierrache et al. [78]	<i>RPE</i> 65 was found in 2/129 and 44/1031 individuals with RP
Italy	1.94%	Colombo et al. [77]	RPE65 variant in 2/103 patients with ARRP
Ireland	7.41%	Whelan et al. [5]	7.41% <i>RPE65</i> mutation was in patients with dominant RP
United States 1	range for clinically diagno	osed was 0.81–1.85%	
US	0.81-1.85%	Wang et al. [79] and Morimura et al. [49]	RPE65 in 1/123 and 3/162 cases with RP
			Morimura 1998: study is of 147 individuals with ARRP and 15 with isolate RP
Mexico	2.7-10.34%	Zenteno et al. [56]	<i>RPE65</i> reported, in cases with 1/37 ARRP and 3/29 Simplex RP
Mexico	3.28%	Zenteno et al. [56]	Assumption calculation: <i>RPE65</i> in 4/122 RP patients
Range for mole	ecularly diagnosed patien	ts was 3–9.98% in EU and .	Americas
Spain ^a	3.00%*	Perea-Romero et al. [80]	<i>RPE65</i> was at 3% in the genetically solved 666 ARRP cases
US ^a	3.23%*	Wang et al. [79]	<i>RPE65</i> in 1/31 cases genetically identified with disease-causing genes
Mexico ^a	3.45*-21.43%*	Zenteno et al. [56]	<i>RPE65</i> in cases with 1/29 ARRP and 3/14 Simplex RP
Mexico ^a	5.00%*	Zenteno et al. [56]	Assumption calculation: RPE65 in 4/80 RP patients

 Table 2 Proportion of RPE65 gene mutation in RP

Country	Proportion of <i>RPE65</i> mutations in RP	Author, year of publication	Remarks
Israel	0.43%	Kimchi et al. [124]	<i>RPE</i> 65 mutation in 1/230 Ashkenazi Jewish descent clinically diagnosed families with RP
			While <i>RPE</i> 65-RP was 1.16% (1/86) within the families genetically identified with disease-causing genes
India	2.94%	Singh et al. [125]	<i>RPE</i> 65 variants in 1/34 clinically diagnosed ARRP consanguineous families
China	3.95%	Dan et al. [126]	<i>RPE</i> 65 variants in 3/76 clinically diagnosed RP families
			While <i>RPE65-RP</i> was 7% (3/43) within the families genetically identified with disease-causing genes

Table 2 continued

ARRP autosomal recessive retinitis pigmentosa, EU European Union, RP retinitis pigmentosa

^aThe % proportion is based on molecularly diagnosed patients

Key messages from table on proportions of RPE65 gene mutations in clinically diagnosed RP cases:

• The proportions of RPE65-RP in the EU-5 counties ranged between 0.23% in Germany and 1.94% in Italy

• The proportions of *RPE65*-RP across the European region ranged between 0.23% in Germany to 4.27% in the Netherlands (excluding Ireland's rare *RPE65* cases in patients with dominant RP)

• The proportions of *RPE65*-RP in the Americas region ranged between $\sim 1\%$ in the US and 3.28% in the Mexico (excluding Mexico's calculation for subgroup population with simplex RP)

Key messages from table on proportions of RPE65 gene mutations in molecularly diagnosed RP cases:

• The proportions of *RPE65*-RP across the European and Americas region ranged between 3% in the US and Spain to 9.98% in the Netherlands (excluding Mexico's calculation for subgroup population with simplex RP)

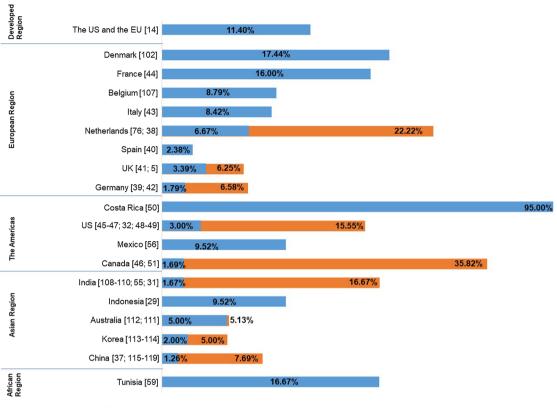
prevalence. One such example was the study in Israel in which, due to high rates of consanguinity and a high number of siblings per family in some of the ethnic groups, a much higher prevalence of 1 in 2100 was found for nonsyndromic RP (both in the Jewish and the Arab Muslim populations) in the Jerusalem area [73]. The prevalence was also found to be highest in the age group of 65 years and older [68, 69].

The proportion of the *RPE65* mutation in clinically diagnosed RP patients was reported to be between 0.23% (Germany) and 4.27% (the Netherlands) for the European region [42, 74–78]. For the EU-5, the proportion ranged from 0.23–1.94% [42, 77], while in the US, it was between 0.81 and 1.85% [49, 79] (Table 2). A Mexican study reported that *RPE65* in RP occurred in 3.28% of clinically diagnosed and

5% of the molecularly diagnosed probands [56]. The study by Wang et al. in the US [79] reported the proportion of *RPE65* mutation in molecularly diagnosed RP patients as 3.23% (vs. 0.81% in clinically diagnosed RP patients). The proportion of *RPE65* mutation in molecularly diagnosed RP patients across the European and Americas region was between 3% in the US and Spain to 9.98% in the Netherlands [56, 78–80] (Table 2).

Proportion of *RPE65*-Mediated RP and LCA Combined

The following studies reported the proportion of the *RPE65* mutation out of the combined population of RP and LCA patients



Max range
Proportion of RPE65 in LCA

Fig. 2 Proportion of RPE65 gene mutation in clinically diagnosed LCA patients. Source: Whelan et al. [5], Thompson et al. [14], Sitorus et al. [29], Verma et al. [31], Lotery et al. [32], Xu et al. [37], Booij et al. [38], Eisenberger et al. [39], Vallespin et al. [40], Henderson et al. [41], Weisschuh et al. [42], Simonelli et al. [43], Bocquet et al. [44], Dharmaraj et al. [45], Zernant et al. [46], Galvin et al. [47], Simovich et al. [48], Morimura et al. [49], Glen et al. [50], Heon et al. [51], Viswarubhiny et al. [55], Zenteno et al. [56], El Matri et al. [59], Haer-Wigman et al. [76], Astuti et al. [102], Coppieters et al. [107], Mamatha et al. [108], Sundaresan et al. [109], Srikrupa et al. [110], Thompson et al. [111], Lamey et al. [112], Surl et al. [113], Seong et al. [114], Liu, Bu [115], Li et al. [116], Chen et al. [117], Zhong et al. [118], Xu et al. [119]. The high RPE65-LCA 16.67% in South India [31] is based on 27/30 probands born through consanguineous marriage. The high RPE65-LCA 35.82% in Canada [51] is probably related to the paper's methodology to analyse only LCA patients with clinically and molecularly confirmed diagnosis identified at one of the hospitals with an ethnically diverse population. The very high prevalence of *RPE65* mutations (95%) in Costa Rica [50] is due to four founder mutations in RPE65 which have been maintained in this genetically isolated population. The paper's methodology was to analyse samples from affected children and their immediate family members only. Additional data by families (not shown in Fig. 2): The Chinese paper [120] had reported RPE65 mutations in 1% (1/100) families with LCA. The Chinese paper [82] had reported biallelic RPE65 mutations in 2.97% (8/269) families with LCA. The Saudi Arabia paper [121] had reported RPE65 mutations in 5.41% (2/37) consanguineous families with LCA. The Spanish paper [122] had reported *RPE65* mutations in 16.51% (18/109) families with LCA. The Indian paper [123] had reported RPE65 mutations in 18.18% (2/11) families with LCA. Worldwide paper [36] had reported RPE65 mutations in 6.15% (11/179) families with LCA. Key messages from Fig. 2 on proportions of RPE65 gene mutations in clinically diagnosed cases of LCA: The proportions of RPE65-LCA across the world ranged between 1.26% in China to 22.22% in the Netherlands (excluding outliers from Costa Rica and Canada). The proportions of RPE65-LCA in the EU-5 counties ranged between 1.79% in Germany to 16% in France. The proportions of RPE65-LCA across the European region ranged between 1.79% in Germany to 22.22% in the Netherlands. The proportions of RPE65-LCA in the Americas region ranged between 1.69% in Canada to 15.55% in the US (excluding outliers from Costa Rica and Canada). The proportions of RPE65-LCA in the Asian region ranged between 1.26% in China to 16.67% in India. The proportion of RPE65-LCA across the US and Europe was at 11.4%

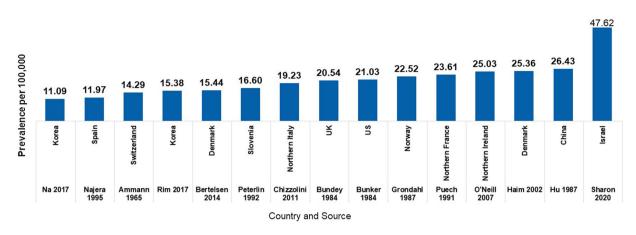


Fig. 3 Country-wise RP prevalence. Source: Bertelsen et al. [34], Hu [60], Haim [61], O'Neill et al. [62], Puech et al. [63], Grondahl [64], Bunker et al. [65], Bundey, Crews [66], Chizzolini et al. [67], Peterlin et al. [68], Rim

et al. [69], Ammann et al. [70], Najera et al. [71], Na et al. [72], Sharon et al. [73]. The high prevalence (47.62) of RP in Israel [73] is based on consanguineous populations

(Supplementary Table A). In a Norwegian study with 513 isolated RP and LCA patients, 0.6% of the cases of *RPE65* mutation were observed [35]. A German study reported that the proportion of the RPE65 mutation was 0.79% for the 126 patients with RP and LCA [39]. In a Dutch study, it was reported that the proportion of the RPE65 mutation was 6% for the 35 patients diagnosed with juvenile autosomal recessive RP (n = 17), juvenile isolated RP (n = 9) and LCA (n = 9) [38]. Further, a study in Mexico reported that the proportion of the RPE65 mutation was 4.20% for the 143 clinically diagnosed patients with RP and LCA. This proportion increased to 6.32% for the molecularly diagnosed 95 patients with RP and LCA [56].

Proportion of *RPE65* Gene Mutation in IRD

Few studies have reported the proportions of *RPE65* gene mutations in clinically diagnosed cases with IRD, varying from ~ 1% in China, US, UK and Israel to 14% in Germany [73, 76, 81–91], including the proportion of 8% in Iran and Tunisia's high-consanguineous population [89, 90]. One study reported the proportion of *RPE65*-IRD across the US and Europe as 2.06% [14]. The proportions of *RPE65*-IRD in the EU-5 counties and the

European region ranged between 1.2% in the UK and 14% in Germany [83, 91]. Lastly, RPE65 mutations were estimated at $\sim 1\%$ in the US, while for the Middle East region, it varied between 4.81% in Saudi Arabia and 8% in Iran's diagnosed clinically cases with IRD [81, 85, 87, 89]. Also, based on the available data, the proportions of RPE65 in molecularly diagnosed cases with IRD was reported at 1.78% in China, 2.82% in the United Arab Emirates (UAE) and 4% in Brazil [57, 84, 92] (Supplementary Table B).

Another study analysed and predicted worldwide genetic prevalence of autosomal recessive IRD for biallelic *RPE65* gene in a total of 15,620 biallelic *RPE65* patients. This study expected that over 60% (9484 individuals) of these affected individuals would be from the African population, while only 9% would be Europeans [93].

Incidence of IRDs, RP and LCA

The genetic incidence of *RPE65*-IRD was 680 per 100,000 for the US population in a genetic testing study with 5879 cases submitted to the 'My Retina Tracker Genetic Testing Program' [94].

A population-based study conducted in Maine, US, between 1976 and 1980 evaluated

the incidence of RP. The study used medical and social service sources as the data source and estimated the birth incidence of persons who will become affected with non-syndromic RP as 28 per 100,000. The incidence of newly diagnosed RP cases per year was estimated to be 0.6 per 100,000 [65]. An epidemiological study of RP in Denmark reported that the average incidence between 1990 and 1997 was 0.79 persons per 100,000 population per year [61]. A nationwide population-based study from South Korea reported the incidence of RP over 4 years. This study found the average incidence of RP to be 1.64 cases per 100,000 person-years and that it was similar in men and women [69]. Further, a national registry study spanning over 15 years in Kuwait reported that RP was the leading cause for registered legal blindness driven by a high prevalence of consanguineous marriages. This study also observed fluctuations in RP incidence rates by age groups due to delay in visual impairment certification of many patients years after the onset of their disability or diagnosis [28].

The incidence of LCA was computed to be between 1 in 50,000 and 1 in 100,000 persons (i.e., approximately 1/75,000) in a multi-centre study analysing LCA patient cohorts from Europe and the US [46]. Additionally, LCA incidence also seems to be high in the consanguineous population of South India [29–31].

DISCUSSION

The clinical diagnosis of most *RPE65* patients is based on signs/symptoms, ophthalmologic features, and age of onset, while the molecular diagnosis of patients with *RPE65*-mediated IRD is based on pathogenic classification of variants identified on genetic tests with the use of various methods like next-generation sequencing, Sanger sequencing, whole exome-analysis or arrayed primer extension. Molecular diagnosis is important for the ophthalmologist to be able to analyse if the affected patient's condition can benefit from potential therapeutic interventions, like the new gene therapy treatments or IRDs clinical trials. Early diagnosis is important to deliver the potential gene therapy treatment to improve and preserve the visual function and therefore diminish the disease burden in affected patients [95]. Findings from the literature indicate that several disorders due to biallelic mutations in RPE65 are named differently, and that the clinical aspects are overlapping, poorly defined and unreliable. They are all a clinical spectrum of the same disease generated by the RPE65 enzymatic dysfunction, such as the phenotype of LCA 2 which is a little more severe and therefore clinically evident in a young population versus RP [13, 96-101]. Further, RPE65-mediated IRD may have different names based on the physician's assessment but have a similar set of symptoms. Therefore, the epidemiology of RPE65-mediated IRD should include all these diagnoses. However, in the literature, there are limited data for the epidemiology of RPE65-mediated IRDs as a group of diseases and instead the data for individual aetiologies were reported. Based on our review results, LCA and RP are ultra-rare IRDs with a prevalence ranging from 1.20 to 2.37 per 100,000 and from 11.09 to 26.43 per 100,000 persons, respectively. Also, the incidence of newly diagnosed RP cases per year was estimated to be about 0.6-1.64 per 100,000 population [61, 65, 69].

Epidemiological studies based on RPE65 mutations in molecularly confirmed RP or LCA or IRD were also found to be scarce in the literature. Although the data on prevalence and incidence of RPE65-mediated IRD was limited, they indicated significant variation between countries and regions. Based on the clinical diagnosis of the disease in the UK, Ireland, Western Europe (France, Germany, Switzerland, Netherlands and Belgium), and Southern European countries (Italy and Spain), the frequency for RPE65-LCA cases was between 1.79 and 22.2%, RPE65-RP was 0.23-4.27%, and RPE65-IRD was around 1.2-14% [38, 39, 42, 76, 78, 83, 91]. RPE65 variants have been reported to be most prevalent cause of LCA in the Nordic country of Denmark (17.44%) [102]. In North America, mutations in RPE65 were reported to cause 3.0-15.55% of clinically diagnosed LCA cases in the US, 0.81-1.85% of RP in the US, 3.28% for RPE65-RP in Mexico and ~ 1% for

clinically diagnosed IRD cases in the US [45, 49, 56, 79, 81, 85]. In South America, the proportion were high due to the identification in the molecularly diagnosed Brazilian patients with only RPE65-LCA at 20.51% and RPE65-IRD at 4% [57, 58]. This could be because there were negative and inconclusive cases or could be attributable to the likelihood of the physician asking only suspected patients for a genetic test. This bias on asking for genetic tests would be much higher if the physician thinks that it could be a treatable IRD. For the Middle East region (Israel, Saudi Arabia and Iran), the proportion for RPE65 mutations in IRD ranged between 1 and 8% [73, 87, 89]. In the African region (Tunisia), the frequency for RPE65-LCA was 16.67% [59]. Lastly, for the Asian region (Australia, China, India, Indonesia and Korea), the RPE65-LCA was 1.26-16.67% [31, 37] and the frequency for RPE65-IRD in China was 0.84-1.32% [82, 84].

While interpreting the results from this review, it is important to acknowledge several limitations. The main ones are related to factors like study design, population demographics (age, consanguineous marriages), enrolment criteria for patients, diagnostic criteria (clinical or molecular/genetic testing), ascertainment bias, classification, biological assaying techniques, etc., which made it difficult to compare the data between studies. It was also found that terms like probands, families, or individuals were used interchangeably in some of the studies, for which we had to rely on the term which was first described in the methods section or was mentioned on the figure/table from where the data were extracted. The literature search was mainly focussed on papers reporting RPE65 in RP and/or LCA, so some other phenotypes with RPE65 may have been missed due to a change in nomenclature. There might be intrinsic bias in the studies while distinguishing RPE65 in RP or LCA due to a lack of clarity on the age of onset. The evaluation for the quality of included studies was not relevant as these were mainly genotyping studies and do not fit into conventional study types defined in the the quality assessment scale [103]. Further, the applicability of quality assessment checklist was limited as this literature review's scope was limited to the collation of epidemiological data without any quantitative meta-analysis.

CONCLUSIONS

Patients with RPE65 gene-mediated IRD are reported from across the world, including in Europe, North America, South America, Middle East, and Asian countries. The proportion of RPE65 gene mutation in IRD cohorts was around 1% in the countries of UK, Netherlands, US, Israel and China. It was noted that the range of ~ 2- ~ 16% for the proportions of RPE65 mutations in patients with LCA was comparable between the EU-5 countries, the US and the Asian countries. Also, the proportions of RPE65 mutations in patients with autosomal recessive RP was found to be between 1.02 and 2.7%. These ranges for RPE65 in LCA and ARRP are comparable to previously reported reviews [104. 105].

Robust epidemiological data for *RPE65*-mediated IRDs are limited, and therefore accurate assessments of prevalence and incidence are challenging. Insufficient data on prevalence have contributed to the insufficient funding and resources available to conduct genetic testing for IRDs, and to provide genetic counselling for IRD patients and families [106].

There was a high heterogeneity in the reporting of the data, while the lack of sufficient high-quality studies highlights the need to conduct better quality studies on rare genetic diseases. We found that there were variations within the countries as well as between the regions in the reporting of the proportions of RPE65 gene mutations, resulting in broad ranges for estimating the affected patient populations. Consanguineous populations were found to have outliers, as inheritance strongly influences the rate of prevalence in such rare genetic disorders. Therefore, due to limited evidence, further research is needed to generate robust evidence for better understanding of the epidemiology of RPE65-mediated IRDs and to diagnose the disease at an early stage. This would determine if the potential therapeutic intervention can provide the best chance to prevent progression to severe visual impairment or complete blindness in affected patients. With the improvement in patient access and the quality of genetic tests, there is a possibility that, in coming years, ophthalmologists will be able to diagnose more cases with *RPE65*-mediated IRDs.

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Data Availability. Data sharing is not applicable to this article as no datasets were generated or analyzed during the current study.

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REFERENCES

- 1. AAO: Clinical statement: recommendations on clinical assessment of patients with inherited retinal degenerations. https://www.aao.org/clinical-statement/recommendations-on-clinical-assessment-of-patients. (2016). Accessed 11 May 2018.
- 2. Stone EM, Andorf JL, Whitmore SS, DeLuca AP, Giacalone JC, Streb LM, et al. Clinically focused molecular investigation of 1000 consecutive families with inherited retinal disease. Ophthalmology.

2017;124(9):1314–31. https://doi.org/10.1016/j. ophtha.2017.04.008.

- 3. RetNet: Summaries of genes and loci causing retinal diseases. https://sph.uth.edu/retnet/ (2020). Accessed 30 Sep 2020.
- 4. Ziccardi L, Cordeddu V, Gaddini L, Matteucci A, Parravano M, Malchiodi-Albedi F, et al. Gene therapy in retinal dystrophies. Int J Mol Sci. 2019;20(22):5722.
- 5. Whelan L, Dockery A, Wynne N, Zhu J, Stephenson K, Silvestri G, et al. Findings from a genotyping study of over 1000 people with inherited retinal disorders in Ireland. Genes (Basel). 2020;11(1):105. https://doi.org/10.3390/genes11010105.
- 6. OMIM.Database: 180069. Retinoid Isomerohydrolase RPE65. https://www.omim.org/entry/ 180069?search=RPE65&highlight=rpe65 (2021). Accessed 25 May 2021.
- Hartong DT, Berson EL, Dryja TP. Retinitis pigmentosa. Lancet (Lond, Engl). 2006;368(9549): 1795–809. https://doi.org/10.1016/S0140-6736(06)69740-7.
- 8. Pennesi M, Weleber R, Yang P. BMJ best practice retinitis pigmentosa. 2018.
- 9. RNIB: Understanding—retinitis pigmentosa and other inherited retinal dystrophies. https://www.rnib.org.uk/eye-health/eye-conditions/retinitis-pigmentosa (2020). Accessed 7 Jan 2021.
- 10. Allikmets R. Leber congenital amaurosis: a genetic paradigm. Ophthalmic Genet. 2004;25(2):67–79. https://doi.org/10.1080/13816810490514261.
- 11. den Hollander AI, Roepman R, Koenekoop RK, Cremers FPM. Leber congenital amaurosis: genes, proteins and disease mechanisms. Prog Retin Eye Res. 2008;27(4):391–419. https://doi.org/10.1016/j. preteyeres.2008.05.003.
- 12. Fahim A, Daiger S, Weleber R, et al. Nonsyndromic retinitis pigmentosa overview. In: Pagon R, Adam M, Ardinger H, et al., editors. GeneReviews. Seattle: University of Washington; 2017.
- 13. Chung DC, Bertelsen M, Lorenz B, Pennesi ME, Leroy BP, Hamel CP, et al. The natural history of inherited retinal dystrophy due to biallelic mutations in the RPE65 gene. Am J Ophthalmol. 2019;199:58–70. https://doi.org/10.1016/j.ajo. 2018.09.024.
- 14. Thompson DA, Gyurus P, Fleischer LL, Bingham EL, McHenry CL, Apfelstedt-Sylla E, et al. Genetics and phenotypes of RPE65 mutations in inherited retinal

degeneration. Invest Ophthalmol Vis Sci. 2000;41(13):4293–9.

- 15. Cideciyan AV. Leber congenital amaurosis due to RPE65 mutations and its treatment with gene therapy. Prog Retin Eye Res. 2010;29(5):398–427. https://doi.org/10.1016/j.preteyeres.2010.04.002.
- 16. Sodi A, Banfi S, Testa F, Della Corte M, Passerini I, Pelo E, et al. RPE65-associated inherited retinal diseases: consensus recommendations for eligibility to gene therapy. Orphanet J Rare Dis. 2021;16(1): 257. https://doi.org/10.1186/s13023-021-01868-4.
- 17. Motta FL, Martin RP, Porto FBO, Wohler ES, Resende RG, Gomes CP, et al. Pathogenicity reclassification of RPE65 missense variants related to leber congenital amaurosis and early-onset retinal dystrophy. Genes (Basel). 2020;11(1):24.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17(5):405–24. https://doi.org/10.1038/gim. 2015.30.
- Consortium TU. UniProt: a worldwide hub of protein knowledge. Nucleic Acids Res. 2018;47(D1): D506–15. https://doi.org/10.1093/nar/gky1049.
- 20. Landrum MJ, Lee JM, Benson M, Brown GR, Chao C, Chitipiralla S, et al. ClinVar: improving access to variant interpretations and supporting evidence. Nucleic Acids Res. 2017;46(D1):D1062–7. https://doi.org/10.1093/nar/gkx1153.
- 21. Stenson PD, Mort M, Ball EV, Evans K, Hayden M, Heywood S, et al. The Human Gene Mutation Database: towards a comprehensive repository of inherited mutation data for medical research, genetic diagnosis and next-generation sequencing studies. Hum Genet. 2017;136(6):665–77. https:// doi.org/10.1007/s00439-017-1779-6.
- 22. Gu SM, Thompson DA, Srikumari CR, Lorenz B, Finckh U, Nicoletti A, et al. Mutations in RPE65 cause autosomal recessive childhood-onset severe retinal dystrophy. Nat Genet. 1997;17(2):194–7.
- 23. Chao DL, Burr A, Pennesi M. RPE65-related leber congenital amaurosis/early-onset severe retinal dystrophy. 2019 Nov 14. In: Adam MP, Ardinger HH, Pagon RA et al. (eds) GeneReviews® [Internet]. University of Washington, Seattle; 1993–2020. Available from: https://www.ncbi.nlm.nih.gov/ books/NBK549574/ Accessed 13 Oct 2020.
- 24. Higgins. JP, Green. S. Cochrane handbook for systematic reviews of interventions. 2011.

- 25. Moher D, Liberati A, Tetzlaff J, Altman DG, Group TP. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. J Clin Epidemiol. 2009;62(10):1006–12.
- 26. Orphanet: retinitis pigmentosa. . https://www. orpha.net/consor/cgi-bin/OC_Exp.php?Expert=791 (2014). Accessed 13 Oct 2020.
- 27. Orphanet: Leber congenital amaurosis. https:// www.orpha.net/consor/cgi-bin/OC_Exp.php?Lng= GB&Expert=65 (2015). Accessed 13 Oct 2020.
- Pandova MG, Al-Merjan JI, Sadeq NA. Registered blindness in Kuwait—15 years of dynamic changes. Ophthalmic Epidemiol. 2019;26(2):75–83. https:// doi.org/10.1080/09286586.2018.1521981.
- Sitorus RS, Lorenz B, Preising MN. Analysis of three genes in Leber congenital amaurosis in Indonesian patients. Vision Res. 2003;43(28):3087–93. https:// doi.org/10.1016/j.visres.2003.08.008.
- 30. Kumaramanickavel G, Joseph B, Vidhya A, Arokiasamy T, Shridhara SN. Consanguinity and ocular genetic diseases in south India: analysis of a fiveyear study. Community Genet. 2002;5(3):182–5. https://doi.org/10.1159/000066334.
- 31. Verma A, Perumalsamy V, Shetty S, Kulm M, Sundaresan P. Mutational screening of LCA genes emphasizing RPE65 in South Indian cohort of patients. PLoS ONE. 2013. https://doi.org/10.1371/ journal.pone.0073172.
- 32. Lotery AJ, Namperumalsamy P, Jacobson SG, Weleber RG, Fishman GA, Musarella MA, et al. Mutation analysis of 3 genes in patients with Leber congenital amaurosis. Arch Ophthalmol. 2000;118(4):538–43.
- 33. Stone EM. Leber congenital amaurosis—a model for efficient genetic testing of heterogeneous disorders: LXIV Edward Jackson Memorial Lecture. Am J Ophthalmol. 2007;144(6):791–811. https://doi.org/ 10.1016/j.ajo.2007.08.022.
- Bertelsen M, Jensen H, Bregnhoj JF, Rosenberg T. Prevalence of generalized retinal dystrophy in Denmark. Ophthalmic Epidemiol. 2014;21(4): 217–23. https://doi.org/10.3109/09286586.2014. 929710.
- 35. Holtan JP, Selmer KK, Heimdal KR, Bragadottir R. Inherited retinal disease in Norway—a characterization of current clinical and genetic knowledge. Acta Ophthalmol. 2019. https://doi.org/10.1111/ aos.14218.
- 36. Hanein S, Perrault I, Gerber S, Tanguy G, Barbet F, Ducroq D, et al. Leber congenital amaurosis: comprehensive survey of the genetic heterogeneity,

refinement of the clinical definition, and genotypephenotype correlations as a strategy for molecular diagnosis. Hum Mutat. 2004;23(4):306–17. https:// doi.org/10.1002/humu.20010.

- 37. Xu Y, Xiao X, Li S, Jia X, Xin W, Wang P, et al. Molecular genetics of Leber congenital amaurosis in Chinese: new data from 66 probands and mutation overview of 159 probands. Exp Eye Res. 2016;149: 93–9. https://doi.org/10.1016/j.exer.2016.06.019.
- 38. Booij JC, Florijn RJ, ten Brink JB, Loves W, Meire F, van Schooneveld MJ, et al. Identification of mutations in the AIPL1, CRB1, GUCY2D, RPE65, and RPGRIP1 genes in patients with juvenile retinitis pigmentosa. J Med Genet. 2005;42(11):e67.
- 39. Eisenberger T, Neuhaus C, Khan AO, Decker C, Preising MN, Friedburg C, et al. Increasing the yield in targeted next-generation sequencing by implicating CNV analysis, non-coding exons and the overall variant load: the example of retinal dystrophies. PLoS ONE. 2013;8(11): e78496. https://doi. org/10.1371/journal.pone.0078496.
- 40. Vallespin E, Cantalapiedra D, Riveiro-Alvarez R, Wilke R, Aguirre-Lamban J, Avila-Fernandez A, et al. Mutation screening of 299 Spanish families with retinal dystrophies by leber congenital amaurosis genotyping microarray. Invest Ophthalmol Vis Sci. 2007;48(12):5653–61. https://doi.org/10.1167/iovs. 07-0007.
- 41. Henderson RH, Waseem N, Searle R, Van Der Spuy J, Russell-Eggitt I, Bhattacharya SS, et al. An assessment of the Apex microarray technology in genotyping patients with leber congenital amaurosis and early-onset severe retinal dystrophy. Invest Ophthalmol Vis Sci. 2007;48(12):5684–9. https://doi. org/10.1167/iovs.07-0207.
- 42. Weisschuh N, Obermaier CD, Battke F, Bernd A, Kuehlewein L, Nasser F, et al. Genetic architecture of inherited retinal degeneration in Germany: a large cohort study from a single diagnostic center over a 9-year period. Hum Mutat. 2020;41(9): 1514–27. https://doi.org/10.1002/humu.24064.
- Simonelli F, Ziviello C, Testa F, Rossi S, Fazzi E, Bianchi PE, et al. Clinical and molecular genetics of Leber's congenital amaurosis: a multicenter study of Italian patients. Invest Ophthalmol Vis Sci. 2007;48(9):4284–90. https://doi.org/10.1167/iovs. 07-0068.
- 44. Bocquet B, Lacroux A, Surget MO, Baudoin C, Marquette V, Manes G, et al. Relative frequencies of inherited retinal dystrophies and optic neuropathies in Southern France: assessment of 21-year data management. Ophthalmic Epidemiol. 2013;20(1):13–25. https://doi.org/10.3109/ 09286586.2012.737890.

- 45. Dharmaraj S, Silva E, Pina AL, Li YY, Yang JM, Carter RC, et al. Mutational analysis and clinical correlation in Leber congenital amaurosis. Ophthalmic Genet. 2000;21(3):135–50.
- 46. Zernant J, Külm M, Dharmaraj S, Den Hollander AI, Perrault I, Preising MN, et al. Genotyping microarray (disease chip) for leber congenital amaurosis: detection of modifier alleles. Invest Ophthalmol Vis Sci. 2005;46(9):3052–9. https://doi.org/10.1167/ iovs.05-0111.
- 47. Galvin JA, Fishman GA, Stone EM, Koenekoop RK. Evaluation of genotype-phenotype associations in Leber congenital amaurosis. Retina. 2005;25(7): 919–29. https://doi.org/10.1097/00006982-200510000-00016.
- 48. Simovich MJ, Miller B, Ezzeldin H, Kirkland BT, McLeod G, Fulmer C, et al. Four novel mutations in the RPE65 gene in patients with Leber congenital amaurosis. Hum Mutat. 2001;18(2):164.
- 49. Morimura H, Fishman GA, Grover SA, Fulton AB, Berson EL, Dryja TP. Mutations in the RPE65 gene in patients with autosomal recessive retinitis pigmentosa or Leber congenital amaurosis. Proc Natl Acad Sci USA. 1998;95(6):3088–93. https://doi.org/ 10.1073/pnas.95.6.3088.
- 50. Glen WB, Peterseim MMW, Badilla R, Znoyko I, Bourg A, Wilson R, et al. A high prevalence of biallelic RPE65 mutations in Costa Rican children with Leber congenital amaurosis and early-onset retinal dystrophy. Ophthalmic Genet. 2019;40(2): 110–7. https://doi.org/10.1080/13816810.2019. 1582069.
- 51. Heon E, Perez-Araya M, Trang H, Roadhouse C, Vincent A. RPE65 is the predominant LCA gene in a Canadian ethnically diverse LCA patient population. Investig Ophthalmol Vis Sci. 2015;56(7):3847.
- 52. Li Y, Xu KE, Zhang X, Xie Y, Jiang F, Liu L. Comprehensive molecular screening in a cohort of Chinese patients with Leber congenital amaurosis or severe early childhood onset retinal dystrophy. Investig Ophthalmol Vis Sci. 2016;57(12):666.
- 53. Bruwer AASAAGFAZ. Phenotypic and genotypic characterization of leber congenital amaurosis in Omani families: a Sultan Qaboos University Hospital experience. Oman Medical Specialty Board Career and Research Forum 2018: Abstracts: Oman Medical Journal 2018.
- Khan AO, Al-Mesfer S, Al-Turkmani S, Bergmann C, Bolz HJ. Genetic analysis of strictly defined leber congenital amaurosis with (and without) neurodevelopmental delay. Br J Ophthalmol. 2014;98(12): 1724–8. https://doi.org/10.1136/bjophthalmol-2014-305122.

- 55. Viswarubhiny S, Anjanamurthy R, Vanniarajan A, Bharanidharan D, Perumalsamy V, Sundaresan P. Clinical exome sequencing facilitates the understanding of genetic heterogeneity in Leber congenital amaurosis patients with variable phenotype in southern India. Eye Vis. 2021;8(1):20. https://doi. org/10.1186/s40662-021-00243-5.
- 56. Zenteno JC, García-Montaño LA, Cruz-Aguilar M, Ronquillo J, Rodas-Serrano A, Aguilar-Castul L, et al. Extensive genic and allelic heterogeneity underlying inherited retinal dystrophies in Mexican patients molecularly analyzed by next-generation sequencing. Mol Genet Genomic Med. 2019. https://doi.org/10.1002/mgg3.1044.
- 57. Motta FL, Martin RP, Filippelli-Silva R, Salles MV, Sallum JMF. Relative frequency of inherited retinal dystrophies in Brazil. Sci Rep. 2018;8(1):15939. https://doi.org/10.1038/s41598-018-34380-0.
- 58. Sallum JMF, Motta FL, Arno G, Porto FBO, Resende RG, Belfort R Jr. Clinical and molecular findings in a cohort of 152 Brazilian severe early onset inherited retinal dystrophy patients. Am J Med Genet C. 2020;184(3):728–52. https://doi.org/10.1002/ajmg. c.31828.
- 59. El Matri K, Falfoul Y, Habibi I, Turki A, Hassairi A, Chebil A, et al. Clinical and genetic characteristics of leber congenital amaurosis in the Tunisian population: experience of the oculogenetic laboratory LR14SP01. Acta Ophthalmol. 2018;96:35. https:// doi.org/10.1111/aos.13972.
- 60. Hu DN. Prevalence and mode of inheritance of major genetic eye diseases in China. J Med Genet. 1987;24(10):584–8.
- 61. Haim M. Epidemiology of retinitis pigmentosa in Denmark. Acta Ophthalmol Scand Suppl. 2002;233: 1–34.
- 62. O'Neill JJ, McKay GJ, Simpson DA, Silvestri G. The epidemiology of retinitis pigmentosa in Northern Ireland. Investig Ophthalmol Vis Sci. 2007;48(13): 3724.
- 63. Puech B, Kostrubiec B, Hache JC, Francois P. Epidemiology and prevalence of hereditary retinal dystrophies in the Northern France. J Fr Ophtalmol. 1991;14(3):153–64.
- 64. Grondahl J. Estimation of prognosis and prevalence of retinitis pigmentosa and Usher syndrome in Norway. Clin Genet. 1987;31(4):255–64.
- 65. Bunker CH, Berson EL, Bromley WC, Hayes RP, Roderick TH. Prevalence of retinitis pigmentosa in Maine. Am J Ophthalmol. 1984;97(3):357–65.

- Bundey S, Crews SJ. A study of retinitis pigmentosa in the City of Birmingham. I Prevalence J Med Genet. 1984;21(6):417–20.
- 67. Chizzolini M, Galan A, Milan E, Sebastiani A, Costagliola C, Parmeggiani F. Good epidemiologic practice in retinitis pigmentosa: from phenotyping to biobanking. Curr Genomics. 2011;12(4):260–6. https://doi.org/10.2174/138920211795860071.
- Peterlin B, Canki-Klain N, Morela V, Stirn B, Rainer S, Cerar V. Prevalence of retinitis pigmentosa in Slovenia. Clin Genet. 1992;42(3):122–3.
- 69. Rim TH, Park HW, Kim DW, Chung EJ. Four-year nationwide incidence of retinitis pigmentosa in South Korea: a population-based retrospective study from 2011 to 2014. BMJ Open. 2017. https://doi.org/10.1136/bmjopen-2016-015531.
- 70. Ammann F, Klein D, Franceschetti A. Genetic and epidemiological investigations on pigmentary degeneration of the retina and allied disorders in Switzerland. J Neurol Sci. 1965;2(2):183–96.
- Najera C, Millan JM, Beneyto M, Prieto F. Epidemiology of retinitis pigmentosa in the Valencian community (Spain). Genet Epidemiol. 1995;12(1): 37–46. https://doi.org/10.1002/gepi.1370120105.
- 72. Na KH, Kim HJ, Kim KH, Han S, Kim P, Hann HJ, et al. Prevalence, age at diagnosis, mortality, and cause of death in retinitis pigmentosa in Korea—a nationwide population-based study. Am J Ophthalmol. 2017;176:157–65. https://doi.org/10.1016/ j.ajo.2017.01.014.
- 73. Sharon D, Ben-Yosef T, Goldenberg-Cohen N, Pras E, Gradstein L, Soudry S, et al. A nationwide genetic analysis of inherited retinal diseases in Israel as assessed by the Israeli inherited retinal disease consortium (IIRDC). Hum Mutat. 2020;41(1):140–9. https://doi.org/10.1002/humu.23903.
- 74. González-del Pozo M, Borrego S, Barragán I, Pieras JI, Santoyo J, Matamala N, et al. Mutation screening of multiple genes in Spanish patients with autosomal recessive retinitis pigmentosa by targeted resequencing. PLoS ONE. 2011. https://doi.org/10.1371/journal.pone.0027894.
- 75. Bocquet B, Marzouka NA, Hebrard M, Manes G, Senechal A, Meunier I, et al. Homozygosity mapping in autosomal recessive retinitis pigmentosa families detects novel mutations. Mol Vis. 2013;19: 2487–500.
- 76. Haer-Wigman L, van Zelst-Stams WA, Pfundt R, van den Born LI, Klaver CC, Verheij JB, et al. Diagnostic exome sequencing in 266 Dutch patients with visual impairment. Eur J Hum Genet EJHG.

2017;25(5):591–9. https://doi.org/10.1038/ejhg. 2017.9.

- 77. Colombo L, Maltese PE, Castori M, El Shamieh S, Zeitz C, Audo I, et al. Molecular epidemiology in 591 Italian probands with nonsyndromic retinitis pigmentosa and usher syndrome. Investig Oph-thalmol Vis Sci. 2021;62(2):13. https://doi.org/10. 1167/iovs.62.2.13.
- Pierrache LHM, Thiadens AAHJ, Van Den Born LI, Klaver CCW. Prevalence of retinitis pigmentosa subtypes in two Rotterdam-based tertiary care centers. Acta Ophthalmol. 2018;96:26–7. https://doi. org/10.1111/aos.13736.
- 79. Wang F, Wang H, Tuan HF, Nguyen DH, Sun V, Keser V, et al. Next generation sequencing-based molecular diagnosis of retinitis pigmentosa: identification of a novel genotype-phenotype correlation and clinical refinements. Hum Genet. 2014;133(3): 331–45. https://doi.org/10.1007/s00439-013-1381-5.
- Perea-Romero I, Gordo G, Iancu IF, Del Pozo-Valero M, Almoguera B, Blanco-Kelly F, et al. Author Correction: Genetic landscape of 6089 inherited retinal dystrophies affected cases in Spain and their therapeutic and extended epidemiological implications. Sci Rep. 2021;11(1):10340. https://doi.org/10.1038/ s41598-021-89275-4.
- Alastalo T-P, Kämpjärvi K, Guidugli L, Känsäkoski J, Wells K, Västinsalo H, et al. Prevalence and genetic characteristics of RPE65-associated retinal disease. Investig Ophthalmol Vis Sci. 2019;60(9):400.
- 82. Li S, Xiao X, Yi Z, Sun W, Wang P, Zhang Q. RPE65 mutation frequency and phenotypic variation according to exome sequencing in a tertiary centre for genetic eye diseases in China. Acta Ophthalmol. 2019. https://doi.org/10.1111/aos.14181.
- 83. Pontikos N, Arno G, Jurkute N, Schiff E, Ba-Abbad R, Malka S, et al. Genetic basis of inherited retinal disease in a molecularly characterized cohort of more than 3000 families from the United Kingdom. Ophthalmology. 2020;127(10):1384–94. https:// doi.org/10.1016/j.ophtha.2020.04.008.
- 84. Gao F-J, Wang D-D, Li J-K, Hu F-Y, Xu P, Chen F, et al. Frequency and phenotypic characteristics of RPE65 mutations in the Chinese population. Orphanet J Rare Dis. 2021;16(1):174. https://doi. org/10.1186/s13023-021-01807-3.
- 85. Ramkumar HL, Gudiseva HV, Kishaba KT, Suk JJ, Verma R, Tadimeti K, et al. A report on molecular diagnostic testing for inherited retinal dystrophies by targeted genetic analyses. Genet Test Mol Biomark. 2017;21(2):66–73. https://doi.org/10.1089/ gtmb.2016.0251.

- 86. Villanueva A, Biswas P, Kishaba K, Suk J, Tadimeti K, Raghavendra PB, et al. Identification of the genetic determinants responsible for retinal degeneration in families of Mexican descent. Ophthalmic Genet. 2018;39(1):73–9. https://doi.org/10.1080/ 13816810.2017.1373830.
- Magliyah M, Saifaldein AA, Schatz P. Late presentation of RPE65 retinopathy in three siblings. Doc Ophthalmol. 2020;140(3):289–97. https://doi.org/ 10.1007/s10633-019-09745-z.
- Tran VH, Vaclavik V, Houghton S, Tiab L, Schorderet DF, Munier FL. Genetics of retinitis pigmentosa and other hereditary retinal disorders in Western Switzerland. Invest Ophthalmol Vis Sci. 2014;55(13):4514.
- 89. Tayebi N, Akinrinade O, Khan MI, Hejazifar A, Dehghani A, Cremers FPM, et al. Targeted next generation sequencing reveals genetic defects underlying inherited retinal disease in Iranian families. Mol Vis. 2019;25:106–17.
- 90. Habibi I, Falfoul Y, Turki A, Hassairi A, El Matri K, Chebil A, et al. Genetic spectrum of retinal dystrophies in Tunisia. Sci Rep. 2020;10(1):11199. https:// doi.org/10.1038/s41598-020-67792-y.
- 91. Feldhaus B, Kohl S, Weisschuh N, Nasser F, Zrenner E, Zobor D. Leber congenital amaurosis (LCA): prevalence of mutations in a large German cohort and clinical characterization of the associated phenotype. Investig Ophthalmol Vis Sci. 2018;59(9): 1832.
- 92. Khan AO. Phenotype-guided genetic testing of pediatric inherited retinal disease in the United Arab Emirates. Retina. 2020;40(9):1829–37. https://doi.org/10.1097/iae.00000000002675.
- 93. Hanany M, Rivolta C, Sharon D. Worldwide carrier frequency and genetic prevalence of autosomal recessive inherited retinal diseases. Proc Natl Acad Sci. 2020;117(5):2710–6. https://doi.org/10.1073/ pnas.1913179117.
- 94. Mansfield BC, Yerxa BR, Branham KH. Implementation of a registry and open access genetic testing program for inherited retinal diseases within a nonprofit foundation. Am J Med Genet C . 2020;184(3): 838–45. https://doi.org/10.1002/ajmg.c.31825.
- 95. Lorenz B, Tavares J, van den Born LI, Marques JP, Scholl HPN. Current management of patients with RPE65 mutation-associated inherited retinal degenerations in Europe: results of a multinational survey by the European Vision Institute Clinical Research Network. Ophthalmic Res. 2021. https:// doi.org/10.1159/000515688.

- 96. Weleber RG, Michaelides M, Trzupek KM, Stover NB, Stone EM. The phenotype of severe early childhood onset retinal dystrophy (SECORD) from mutation of RPE65 and differentiation from Leber congenital amaurosis. Invest Ophthalmol Vis Sci. 2011;52(1):292–302. https://doi.org/10.1167/iovs. 10-6106.
- 97. Leroy BP, Dharmaraj, S.: Leber Congenital Amaurosis. Orphanet Encyclopedia. https://www.orpha.net/data/patho/GB/uk-LCA.pdf (2003). Accessed 5 Jan 2019.
- 98. Berson EL. Retinitis pigmentosa and allied retinal diseases. In: Duane TD, Tasman W, Jaeger EA, editors. Duane's clinical ophthalmology. Philadelphia: Lippincott Williams & Wilkins; 2013.
- 99. Traboulsi E. Genetic diseases of the eye. New York: Oxford University Press; 1998.
- 100. NHx. Data on File. CSR RPE65 NHx:3-6,53–57. January 11, 2017a. Philadelphia, PA. Spark Therapeutics, Inc. 2017.
- 101. Russell S, Bennett J, Wellman JA, Chung DC, Yu ZF, Tillman A, et al. Efficacy and safety of voretigene neparvovec (AAV2-hRPE65v2) in patients with RPE65-mediated inherited retinal dystrophy: a randomised, controlled, open-label, phase 3 trial. The Lancet. 2017;390(10097):849–60. https://doi.org/ 10.1016/S0140-6736(17)31868-8.
- 102. Astuti GDN, Bertelsen M, Preising MN, Ajmal M, Lorenz B, Faradz SMH, et al. Comprehensive genotyping reveals RPE65 as the most frequently mutated gene in Leber congenital amaurosis in Denmark. Eur J Hum Genet. 2016;24(7):1071–9. https://doi. org/10.1038/ejhg.2015.241.
- 103. NHLBI-NIH: Quality assessment tool for observational cohort and cross-sectional studies. https:// www.nhlbi.nih.gov/health-topics/study-qualityassessment-tools (2017). Accessed 28 May 2018.
- 104. Tsang SH, Sharma T. Leber congenital amaurosis. In: Tsang SH, Sharma T, editors. Atlas of inherited retinal diseases. Cham: Springer ; 2018. p. 131–7.
- 105. Tsang SH, Sharma T. Retinitis pigmentosa (nonsyndromic). In: Tsang SH, Sharma T, editors. Atlas of inherited retinal diseases. Cham: Springer ; 2018. p. 125–30.
- 106. Galvin O, Chi G, Brady L, Hippert C, Del Valle RM, Daly A, et al. The impact of inherited retinal diseases in the Republic of Ireland (ROI) and the United Kingdom (UK) from a cost-of-illness perspective. Clin Ophthalmol. 2020;14:707–19. https://doi.org/ 10.2147/OPTH.S241928.

- 107. Coppieters F, Casteels I, Meire F, De Jaegere S, Hooghe S, van Regemorter N, et al. Genetic screening of LCA in Belgium: predominance of CEP290 and identification of potential modifier alleles in AHI1 of CEP290-related phenotypes. Hum Mutat. 2010;31(10):E1709–66. https://doi.org/10. 1002/humu.21336.
- 108. Mamatha G, Srilekha S, Meenakshi S, Kumaramanickavel G. Screening of the RPE65 gene in the Asian Indian patients with Leber congenital amaurosis. Ophthalmic Genet. 2008;29(2):73–8. https://doi. org/10.1080/13816810802008259.
- 109. Sundaresan P, Vijayalakshmi P, Thompson S, Ko AC, Fingert JH, Stone EM. Mutations that are a common cause of Leber congenital amaurosis in northern America are rare in Southern India. Mol Vis. 2009;15:1781–7.
- 110. Srikrupa NN, Srilekha S, Sen P, Arokiasamy T, Meenakshi S, Bhende M, et al. Genetic profile and mutation spectrum of Leber congenital amaurosis in a larger Indian cohort using high throughput targeted re-sequencing. Clin Genet. 2018;93(2): 329–39. https://doi.org/10.1111/cge.13159.
- 111. Thompson JA, De Roach JN, McLaren TL, Montgomery HE, Hoffmann LH, Campbell IR, et al. The genetic profile of Leber congenital amaurosis in an Australian cohort. Mol Genet Genomic Med. 2017;5(6):652–67. https://doi.org/10.1002/mgg3. 321.
- 112. Lamey T, McLaren T, Montgomery H, Hoffmann L, Kap C, De Roach J. Genetic analysis for australians clinically diagnosed with Leber congenital amaurosis. Clin Exp Ophthalmol. 2013;41:119. https:// doi.org/10.1111/ceo.12231.
- 113. Surl D, Shin S, Lee ST, Choi JR, Lee J, Byeon SH, et al. Copy number variations and multiallelic variants in Korean patients with Leber congenital amaurosis. Mol Vis. 2020;26:26–35.
- Seong MW, Kim SY, Yu YS, Hwang JM, Kim JY, Park SS. Molecular characterization of leber congenital amaurosis in Koreans. Mol Vis. 2008;14:1429–36.
- 115. Liu J, Bu J. A gene scan study of RPE65 in Chinese patients with leber congenital amaurosis. Chin Med J. 2017;130(22):2709–12. https://doi.org/10.4103/ 0366-6999.218007.
- 116. Li L, Xiao X, Li S, Jia X, Wang P, Guo X, et al. Detection of variants in 15 genes in 87 unrelated Chinese patients with Leber congenital amaurosis. PLoS ONE. 2011. https://doi.org/10.1371/journal. pone.0019458.
- 117. Chen Y, Zhang Q, Shen T, Xiao X, Li S, Guan L, et al. Comprehensive mutation analysis by whole-

exome sequencing in 41 Chinese families with leber congenital amaurosis. Invest Ophthalmol Vis Sci. 2013;54(6):4351–7. https://doi.org/10.1167/iovs. 13-11606.

- 118. Zhong Z, Rong F, Dai Y, Yibulayin A, Zeng L, Liao J, et al. Seven novel variants expand the spectrum of RPE65-related leber congenital amaurosis in the Chinese population. Mol Vis. 2019;25:204–14.
- 119. Xu K, Xie Y, Sun T, Zhang X, Chen C, Li Y. Genetic and clinical findings in a Chinese cohort with Leber congenital amaurosis and early onset severe retinal dystrophy. Br J Ophthalmol. 2019. https://doi.org/ 10.1136/bjophthalmol-2019-314281.
- 120. Xu F, Dong Q, Liu L, Li H, Liang X, Jiang R, et al. Novel RPE65 mutations associated with leber congenital amaurosis in Chinese patients. Mol Vis. 2012;18:744–50.
- 121. Li Y, Wang H, Peng J, Gibbs RA, Lewis RA, Lupski JR, et al. Mutation survey of known LCA genes and loci in the Saudi Arabian population. Invest Ophthalmol Vis Sci. 2009;50(3):1336–43. https://doi.org/10. 1167/iovs.08-2589.
- 122. Lopez-Rodriguez R, Lantero E, Blanco-Kelly F, Avila-Fernandez A, Martin Merida I, del Pozo-Valero M, et al. RPE65-related retinal dystrophy: mutational and phenotypic spectrum in 45 affected patients. medRxiv. 2021. https://doi.org/10.1101/2021.01. 19.21249492.
- 123. Srilekha S, Arokiasamy T, Srikrupa NN, Umashankar V, Meenakshi S, Sen P, et al. Homozygosity mapping in leber congenital amaurosis and autosomal recessive retinitis pigmentosa in south indian families. PLoS ONE. 2015. https://doi.org/10.1371/journal.pone.0131679.
- 124. Kimchi A, Khateb S, Wen R, Guan Z, Obolensky A, Beryozkin A, et al. Nonsyndromic retinitis pigmentosa in the Ashkenazi Jewish population: genetic and clinical aspects. Ophthalmology. 2018;125(5):725–34. https://doi.org/10.1016/j. ophtha.2017.11.014.
- 125. Singh HP, Jalali S, Narayanan R, Kannabiran C. Genetic analysis of Indian families with autosomal recessive retinitis pigmentosa by homozygosity screening. Invest Ophthalmol Vis Sci. 2009;50(9): 4065–71. https://doi.org/10.1167/iovs.09-3479.
- 126. Dan H, Huang X, Xing Y, Shen Y. Application of targeted panel sequencing and whole exome sequencing for 76 Chinese families with retinitis pigmentosa. Mol Genet Genomic Med. 2020;8(3): e1131. https://doi.org/10.1002/mgg3.1131.
- 127. McKibbin M, Ali M, Mohamed MD, Booth AP, Bishop F, Pal B, et al. Genotype-phenotype

correlation for leber congenital amaurosis in northern Pakistan. Arch Ophthalmol. 2010;128(1): 107–13. https://doi.org/10.1001/archophthalmol. 2010.309.

128. Hull S, Henderson R, Webster A, Michaelides M, Holder GE, Moradi P, et al. Molecular investigation of a large UK cohort of early onset retinal dystrophy. Invest Ophthalmol Vis Sci. 2015;56(7):2867.

129. Skorczyk-Werner A, Niedziela Z, Stopa M, Krawczyński MR. Novel gene variants in Polish patients with Leber congenital amaurosis (LCA). Orphanet J Rare Dis. 2020;15(1):345. https://doi.org/10.1186/ s13023-020-01634-y.