

The effect of *trans*-10,*cis*-12 conjugated linoleic acid on lipogenesis is tissue dependent in hamsters

I. Churruca · A. Fernández-Quintela · A. Zabala ·
M. T. Macarulla · V. Navarro · V. M. Rodríguez ·
E. Simón · F. Milagro · M. P. Portillo

Published online: 21 September 2007
© Springer-Verlag and NuGO 2007

Keywords Conjugated linoleic acid · Lipogenesis ·
Adipose tissue · Liver · Hamster

Introduction

Conjugated linoleic acid (CLA) has been reported as a potent modulator of body composition, especially by reducing the accumulation of body fat into adipose tissue [1]. Moreover, CLA can also modify triacylglycerol metabolism in other organs and tissues that play an important role in lipid metabolism, such as the liver [1]. There is now strong evidence showing that the *trans*-10,*cis*-12 CLA isomer is mostly responsible for these effects [2]. The mechanisms by which *trans*-10,*cis*-12 CLA alters triacylglycerol metabolism have not been completely clarified.

The aim of the present work was to investigate the effects of *trans*-10,*cis*-12 CLA intake on the expression of two lipogenic enzymes, acetyl-CoA carboxylase (ACC) and fatty acid synthase (FAS) and the transcriptional factors that regulate their activity, sterol regulatory element binding proteins SREBP-1a and SREBP-1c in adipose tissue and liver from hamsters fed an atherogenic diet.

Materials and methods

The experiment was conducted with twenty 9-week-old (105 ± 1), male Syrian Golden hamsters, individually housed in metabolic cages. Hamsters were randomly divided into two dietary groups and fed a semi-purified atherogenic diet consisting of 200 g/kg of casein, 4 g/kg of L-methionine, 200 g/kg of wheat starch, 405 g/kg of sucrose, 100 g/kg of palm oil, 30 g/kg of cellulose, 4 g/kg of choline-HCl, 1 g/kg of cholesterol, vitamin mix 11 g/kg and mineral mix 40 g/kg, supplemented with 0.5% linoleic acid (control group) or 0.5% *trans*-10,*cis*-12 CLA, respectively. Animals had free access to food and water. Food intake and body weight were measured daily. At the end of the experimental period (6 weeks) animals were sacrificed under anaesthesia. Liver and adipose tissues from different anatomical locations (epididymal, perirenal and subcutaneous) were dissected, weighed, sliced and immediately frozen. Liver triacylglycerol content was determined spectrophotometrically by using a commercial kit. ACC, FAS, SREBP-1a and SREBP-1c mRNA levels in epididymal adipose tissue and liver were assessed by RT-PCR. Results are presented as means \pm SE of the means. Statistical analysis was performed using SPSS 11.0. Data were analysed by one-way ANOVA followed by Newman-Keuls post hoc test. Statistical significance was set-up at the $P < 0.05$ level.

Results

Differences neither in food intake nor in final body weight were found between both experimental groups. Hamsters fed the *trans*-10,*cis*-12 CLA isomer showed significantly reduced adipose tissue sizes and increased liver weight, but

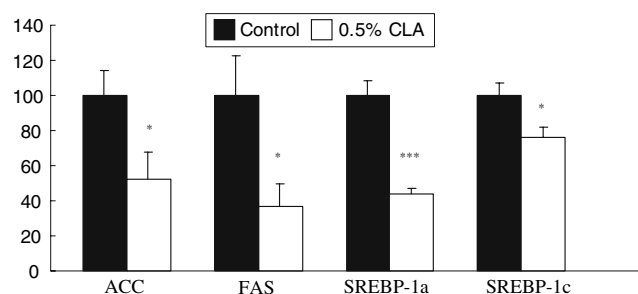
I. Churruca · A. Fernández-Quintela · A. Zabala ·
M. T. Macarulla · V. Navarro · V. M. Rodríguez ·
E. Simón · M. P. Portillo (✉)
Department of Nutrition and Food Science,
Faculty of Pharmacy,
University of the Basque Country, Vitoria, Spain
e-mail: mariapuy.portillo@ehu.es

F. Milagro
Department of Physiology and Nutrition,
University of Navarra, Pamplona, Spain

Table 1 Final body weight, food intake, adipose tissue and liver weights, and hepatic triacylglycerol content of hamsters fed on the experimental diets for 6 weeks

Experimental groups	Control CLA		0.5% <i>trans</i> -10, <i>cis</i> -12 statistical significance	
	Mean	SEM	Mean	SEM
Final body weight (g)	121	3	119	1
Food intake (g/day)	6.0	0.14	5.7	0.08
Adipose tissue weight (g)				
Epididymal	2.48	0.18	1.96*	0.10
Perirenal	1.68	0.10	1.19**	0.09
Subcutaneous	3.78	0.25	2.67**	0.14
Liver				
Weight (g)	4.62	0.16	5.10*	0.11
Triacylglycerols (mg/g)	7.21	0.6	4.79**	0.48

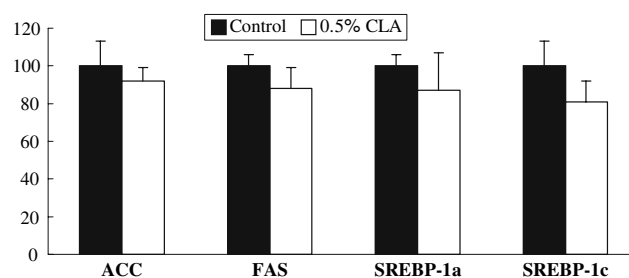
* $P < 0.05$, ** $P < 0.01$

**Fig. 1** Acetyl-CoA carboxylase, fatty acid synthase (FAS), SREBP-1a and SREBP-1c mRNA levels in epididymal adipose tissue. * $P < 0.05$, *** $P < 0.001$

reduced hepatic triacylglycerol content (Table 1). ACC, FAS, SREBP-1a, SREBP-1c expression were significantly reduced in epididymal adipose tissue (Fig. 1). In contrast, no significant differences were observed in liver (Fig. 2).

Discussion

Conflicting results have been reported when the effects of CLA on lipogenic enzyme expression were assessed in adipose tissue. Kang et al. [3] found an increase in FAS mRNA induced by 0.2% *trans*-10,*cis*-12 CLA feeding for 4 weeks in mice. In contrast, Azain et al. [4] did not find changes in FAS activity induced by CLA feeding in rats. Other authors found reduced expression of FAS and ACC in mice [5, 6]. In the present work, *trans*-10,*cis*-12 CLA feeding significantly reduced the mRNA expression of the two specific lipogenic enzymes measured (ACC, FAS). SREBP-1a and SREBP-1c are potent activators of enzymes involved in fatty acid synthesis, and have been shown to be

**Fig. 2** Acetyl-CoA carboxylase, FAS, SREBP-1a and SREBP-1c mRNA levels in liver

affected by nutritional factors [7]. The reduction that we found in the mRNA expression of these transcriptional factors, induced by *trans*-10,*cis*-12, paralleled changes observed in ACC and FAS. Concerning the effects of CLA on liver, several studies in mice have shown that *trans*-10,*cis*-12 increased liver weight [8–13]. In some of these trials, hepatic triacylglycerol content was measured and it was concluded that hepatomegaly was due to lipid accumulation [5, 14–16]. In contrast, the studies performed on rats reported no effects on liver weight induced by CLA [4, 10, 17–20].

The present results show that feeding the *trans*-10,*cis*-12 CLA led to greater liver weight. This effect was similar to that found in mice, but in hamsters the effect was not due to increased fat accumulation; on the contrary, the triacylglycerol content in liver from animals fed this CLA isomer was significantly lower than that in the control group. With regard to the expression of lipogenic enzymes and their transcriptional factors, no significant changes were induced by *trans*-10,*cis*-12 CLA in liver.

All these results suggest that the effects of the *trans*-10,*cis*-12 CLA isomer on the expression of ACC and FAS, and the transcriptional factors that regulates these enzymes (SREBPs) are tissue-specific in hamsters. Decreased lipogenic enzyme expression in adipose tissue could contribute to reduced adipose tissue size. However, reduced liver triacylglycerol content was not mediated by decreased lipogenesis.

Acknowledgments This study was supported by grants from the Ministerio de Ciencia y Tecnología (BFI2002-00273), the Government of País Vasco (Biogune, Programa Etorrek) and University of País Vasco (00101.125-E-14788/2002 and 9/00101.125-15340/2003). V. Navarro is a recipient of a doctoral fellowship from the Ministerio de Educación y Ciencia.

References

- Wang YW, Jones PJH (2004) Conjugated linoleic acid and obesity control: efficacy and mechanisms. *Int J Obes* 28:941–955
- Pariza MW, Park Y, Cook ME (2000) Mechanism of action of conjugated linoleic acid: evidence and speculation. *Proc Soc Exp Biol Med* 223:8–13

3. Kang K, Miyazaki M, Ntambi JM, Pariza MW (2004) Evidence that the anti-obesity effect of conjugated linoleic acid is independent of effects on stearoyl-CoA desaturase1 expression and enzyme activity. *Biochem Biophys Res Commun* 315:532–537
4. Azain MJ, Hausman DB, Sisk MB, Flatt WP, Jewell DE (2000) Dietary conjugated linoleic acid reduces rat adipose tissue cell size rather than cell number. *J Nutr* 130:1548–1554
5. Tsuboyama-Kasaoka N, Takahashi M, Tanemura K, Kim HJ, Tange T, Okuyama H, Kasai M, Ikemoto S, Ezaki O (2000) Conjugated linoleic acid supplementation reduces adipose tissue by apoptosis and develops lipodystrophy in mice. *Diabetes* 49:1534–1542
6. Clément L, Poirier H, Niot I, Bocher V, Guerre M-Millo, Krief S, Staels B, Besnard P (2002) Dietary *trans*-10,*cis*-12 conjugated linoleic acid induces hyperinsulinemia and fatty liver in the mouse. *J Lipid Res* 43:1400–1409
7. Horton JD, Goldstein JL, Brown MS (2002) SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver. *J Clin Invest* 109:1125–1131
8. DeLany JP, Blohm F, Truett AA, Scimeca JA, West DB (1999) Conjugated linoleic acid rapidly reduces body fat content in mice without affecting energy intake. *Am J Physiol* 276:R1172–R1179
9. Hamura M, Yamatoya H, Kudo S (2001) Glycerides rich in conjugated linoleic acid (CLA) improve blood glucose control in diabetic C57BLKS-Leprdb/leprdb mice. *J Oleo Sci* 50:889–894
10. Martin JC, Grégoire S, Siess MH, Genby M, Chardigny JM, Berdeaux O, Juanéda P, Sébédio JL (2000) Effects of conjugated linoleic acid isomers on lipid-metabolizing enzymes in male rats. *Lipids* 35:91–98
11. Miner JL, Cederberg CA, Nielsen MK, Chen X, Baile CA (2001) Conjugated linoleic acid (CLA), body fat, and apoptosis. *Obes Res* 9:129–134
12. Terpstra AH, Beynen AC, Everts H, Kocsis S, Katan MB, Zock PL (2002) The decrease in body fat in mice fed conjugated linoleic acid is due to increases in energy expenditure and energy loss in the excreta. *J Nutr* 132:940–945
13. Warren JM, Simon VA, Bartolini G, Erickson KL, Mackey BE, Kelley DS (2003) *Trans*-10,*cis*-12 CLA increases liver and decreases adipose tissue lipids in mice: possible roles of specific lipid metabolism genes. *Lipids* 38:497–504
14. Belury MA, Kempa-Steczko A (1997) Conjugated linoleic acid modulates hepatic lipid composition in mice. *Lipids* 32:199–204
15. West DB, DeLany JP, Camet P, Blohm FY, Truett AA, Scimeca J (1998) Effects of conjugated linoleic acid on body fat and energy metabolism in the mouse. *Am J Physiol* 275:R667–R672
16. Takahashi Y, Kushihiro M, Shinohara K, Ide T (2003) Activity and mRNA levels of enzymes involved in hepatic fatty acid synthesis and oxidation in mice fed conjugated linoleic acid. *Biochim Biophys Acta* 1631:265–273
17. Chin SF, Storkson JM, Albright KJ, Cook ME, Pariza MW (1994) Conjugated linoleic acid is a growth factor for rats as shown by enhanced weight gain and improved feed efficiency. *J Nutr* 124:2344–2349
18. Sakono M, Miyanaga F, Kawahara S, Yamauchi K, Fukuda N, Watanabe K, Iwata T, Sugano M (1999) Dietary conjugated linoleic acid reciprocally modifies ketogenesis and lipid secretion by the rat liver. *Lipids* 34:997–1000
19. Stangl GI (2000) High dietary levels of conjugated linoleic acid mixture alter hepatic glycerophospholipid class profile and cholesterol-carrying serum lipoproteins of rats. *J Nutr Biochem* 11:184–191
20. Wang YM, Rahman SM, Nagao K, Han SY, Buang Y, Cha JY, Yanagita T (2003) Conjugated linoleic acid reduces hepatic microsomal triacylglycerol transfer protein activity and hepatic triacylglycerol mass in obese rats. *J Oleo Sci* 52:129–134