



Copy Number Variations and Schizophrenia

Kamila Szczówka¹ · Błażej Misiak² · Izabela Łaczmńska¹  · Dorota Frydecka² · Ahmed A. Moustafa^{3,4}

Received: 15 June 2022 / Accepted: 21 December 2022 / Published online: 29 December 2022
© The Author(s) 2022

Abstract

Schizophrenia is a neurodevelopmental disorder with genetic and environmental factors involved in its aetiology. Genetic liability contributing to the development of schizophrenia is a subject of extensive research activity, as reliable data regarding its aetiology would enable the improvement of its therapy and the development of new methods of treatment. A multitude of studies in this field focus on genetic variants, such as copy number variations (CNVs) or single-nucleotide variants (SNVs). Certain genetic disorders caused by CNVs including 22q11.2 microdeletion syndrome, Burnside-Butler syndrome (15q11.2 BP1-BP2 microdeletion) or 1q21.1 microduplication/microdeletion syndrome are associated with a higher risk of developing schizophrenia. In this article, we provide a unifying framework linking these CNVs and their associated genetic disorders with schizophrenia and its various neural and behavioural abnormalities.

Keywords Schizophrenia · Genetics · Copy number variations (CNVs) · Neural studies

Introduction

Schizophrenia is a chronic and complex mental disorder that affects about 0.5% of the population [1]. Patients usually present first symptoms of schizophrenia around the age of 16–30 years [2, 3]. Schizophrenia is mainly characterized by relapsing episodes of psychosis; however, occurring symptoms can be divided into two main categories: positive and negative symptoms. Positive symptoms include various types of hallucinations and delusions. In turn, negative symptoms include social withdrawal, diminished emotional range, avolition, anhedonia and alogia. There is accumulating evidence that specific dimensions of psychopathology in schizophrenia might be characterized by distinct

neurobiological mechanisms. For instance, Carpenter et al. [4] differentiated the deficit subtype of schizophrenia, which is characterized by primary and enduring negative symptoms, from non-deficit schizophrenia, which is characterized by mild negative symptoms. Further studies have demonstrated that patients showing deficit and non-deficit schizophrenia differ in clinical characteristics, neurobiological features, risk factors and family history [5–8]. Unlike non-deficit schizophrenia, deficit schizophrenia is associated with alterations of the brain structure, brain activation, worse sensory integration and impaired motor coordination. The existence of a significant relationship between deficit schizophrenia and more severe cognitive impairment has also been suggested [9–14].

Numerous mental disorders, including schizophrenia, have multifactorial backgrounds based on complex interactions between genetic and environmental factors [5]. Among known genetic alterations reported in patients with schizophrenia, copy number variants (CNVs) are one of the most often genetic alterations that might be causative. There are some reports on the presence of the highest burden of larger (> 500 kb) exonic CNVs in schizophrenia and the possibility that carrying these alterations may modify the phenotype in patients at risk of schizophrenia [15–18]. The phenotype of patients carrying CNVs might include schizophrenia as the only clinical manifestation. However, in other cases, schizophrenia develops as a part of known genetic

✉ Izabela Łaczmńska
izabela.laczmanska@umw.edu.pl

¹ Department of Genetics, Wrocław Medical University, Marcinkowskiego 1, 50-368 Wrocław, Poland

² Department of Psychiatry, Wrocław Medical University, Pasteura 10, 50-367 Wrocław, Poland

³ Department of Human Anatomy and Physiology, The Faculty of Health Sciences, University of Johannesburg, Johannesburg, South Africa

⁴ School of Psychology, Faculty of Society and Design, Centre of Data Analytics, Bond University, Gold Coast, Queensland, Australia

syndromes attributable to CNVs, such as 1q21.1 microdeletion/microduplication syndrome, 15q11.2 BP1-BP2 microdeletion syndrome or 22q11.2 deletion syndrome [19–22]. The development of the array comparative genomic hybridization (aCGH) and the next-generation sequencing (NGS) methods together with an array of bioinformatic approaches has largely improved our understanding of the association between CNVs and various neurodevelopmental disorders, including schizophrenia [23].

CNVs can be defined as alterations of the number of copies in specific regions of the genome (chromosomal duplications and deletions), which vary between individuals. Notably, CNVs represent 4.8–9.5% of the human genome and have been observed to exert an effect on the expression, as well as the function of genes [24]. It has been demonstrated that approximately 70% of all individuals carry at least one rare CNV. Among them, deletions are less common than duplications [25–28].

Existing methods for detecting CNVs in the genome are typically based on DNA probes, such as the multiplex ligation-dependent probe amplification (MLPA), aCGH or NGS. Results for alterations above 40 kbp can also be verified using the fluorescence in situ hybridization (FISH). In recent years, there has been a rapid technological development of molecular biology methods, which is why an increasing use of aCGH and NGS is observed both in the diagnostics of SNVs and CNVs. However, in many countries, this is still limited by the relatively high cost of the test and the high skill requirements of personnel [29].

CNVs in Schizophrenia

However, CNVs associated with schizophrenia lack diagnostic specificity, that is, the presence of certain CNVs in the genome can increase a risk of several other neurodevelopmental disorders, including autism spectrum disorder, attention-deficit hyperactivity disorder and intellectual disability [30–32]. According to recent studies, schizophrenia has been associated with several recurrent CNVs, which are major risk factors of the disorder, occurring in 3.5–5% of cases (see Table 1 and Fig. 1) [20, 33, 34]. However, due to the fact that those CNVs are not completely penetrant (reduced penetrance) and expressed on different levels (variable expressivity), they are not sufficient to cause schizophrenia by themselves. The percentage of individuals with CNVs who develop symptoms may vary depending on the type of CNVs and other factors (i.e., those related to interactions with other genes and the impact of environment) [35].

Interestingly, CNVs associated with schizophrenia have also been observed in healthy individuals, and thus, it may suggest that either environmental factors or additional genetic abnormalities contribute to the development of

Table 1 Overview of selected genetic syndromes associated with schizophrenia. p. 6, 7

Disorder	Region/genes	Symptoms	Breakpoint location
1q21.1 microdeletion/microduplication syndrome	Class I (1.35 Mb) CNV involves distal area between BP3 and BP4 Class II (3 Mb) CNV involves both proximal area (TAR region) and distal area between BP2 and BP4	<ul style="list-style-type: none"> - Macrocephaly (microduplication), microcephaly (microdeletion) [38] - Cognitive impairment [38] - Reduced synaptic plasticity [38] - Psychosis [38] - Agenesis (failure to develop during embryonic growth) of the corpus callosum [39] - Hypoplasia of the cerebellar vermis [39] - Psychosis [40] - Developmental and language delay [40] - Reduced volume in fusiform gyrus [41] - Learning difficulties [42] - Whole-brain volume reduction [43, 44] - Increased cortical thickness [45] - Psychosis [46] - Deficit in cognitive skills such as emotion recognition, visual learning or executive function [47, 48] 	Class I deletion: 1q21.1 BP3-BP4 Class II deletion: 1q21.1 BP2-BP4
15q11.2 BP1-BP2 microdeletion syndrome	This type of 15q11.2 microdeletion includes four highly conservative genes: <i>NIPAI</i> , <i>NIPAZ</i> , <i>CYFIP1</i> and <i>TUBGCP5</i>		15q11-q13
22q11.2 deletion syndrome	Typical deletion occurs in ~90% cases, and it includes around 90 genes (~3 Mb) Atypical deletion occurs in ~10% cases, and its size varies 1.5–2 Mb		Typical deletion: 22q11.2 (LCR22A–LCR22D) Atypical deletion: 22q11.2 (LCR22A–LCR22B)

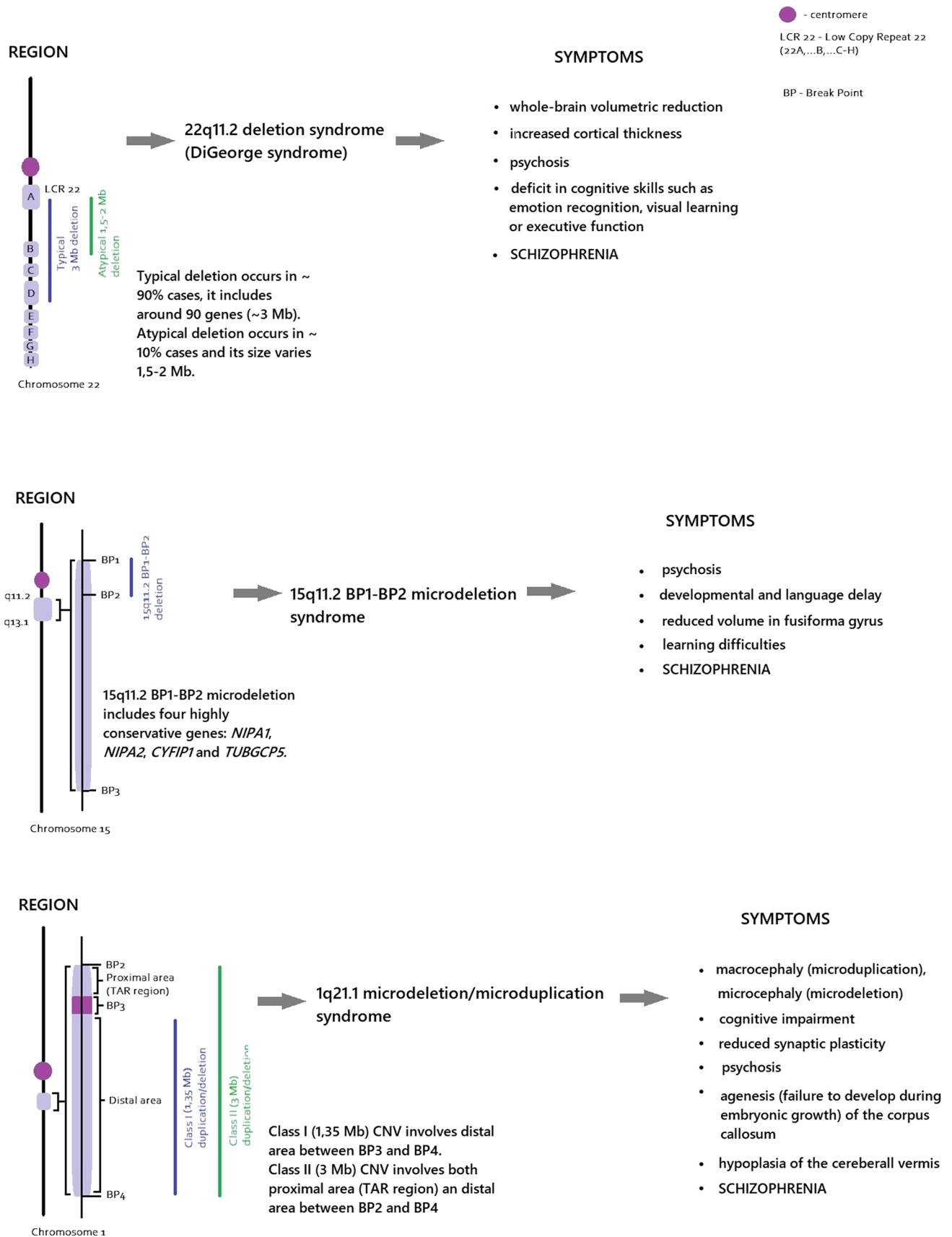


Fig. 1 Simplified overview of the phenotype and specific genes associated with CNVs located at 22q11.2, 15q11.2 and 1q21.1. p. 12

schizophrenia. Moreover, some genetic syndromes such as 22q11.2 microdeletion syndrome, Burnside-Butler syndrome (15q11.2 microdeletion) or 1q21.1 microduplication/microdeletion are strongly associated with a risk of schizophrenia among other CNVs [34, 36, 37]. Table 1 and Fig. 1 show an overview of clinical characteristics of these genetic syndromes.

In the next subsections, we review genetic syndromes commonly associated with a risk of schizophrenia. Recent studies report that other CNVs can also be associated with a higher risk of schizophrenia. Those CNVs (listed in Table 2) include deletions and duplications on chromosomes 2, 3, 7, 8, 9, 13, 16 and sex chromosome X [20, 49].

Table 2 The comparison of schizophrenia-associated CNV loci according to “Schizophrenia-associated genomic copy number variants and subcortical brain volumes in the UK Biobank” [20] and “Contribution of copy number variants to schizophrenia from a genome-wide study of 41,321 subjects” [65]. Only two publications were used as sources to create the said table, as they contained the most information about multiple CNVs compared to others. p. 14, 15

Schizophrenia-associated CNV loci according to Warland et al., 2019	Schizophrenia-associated CNV loci according to Marshall et al., 2017
Deletions	Deletions
1q21.1	1q21.1
2p16.3 (<i>NRXN1</i>)	2p16.3
3q29	3q29
-	7q11.21 (<i>ZNF92</i>)
-	7p36.3 (<i>VIPR2; WDR60</i>)
-	8q22.2 (<i>VPS13B</i>)
-	9p24.3 (<i>DMRT1</i>)
15q11.2	15q11.2
15q13.3	15q13.3
16p12.1	-
-	16p11.2 (<i>distal</i>)
22q11.2	22q11.2(1)
Duplications	Duplications
1q21.1	1q21.1
-	7q11.21 (<i>ZNF92</i>)
7q11.23	7q11.23
-	7p36.3 (<i>VIPR2; WDR60</i>)
-	9p24.3 (<i>DMRT1</i>)
-	13q12.11 (<i>ZMYM5</i>)
15q11-q13	-
16p11.2	16p11.2 (<i>proximal</i>)
16p13.11	-
-	22q11.21
-	Xq28 (<i>distal</i>)
-	Xq28 (<i>MAGEA11</i>)

15q11.2 Microdeletion Syndrome

Burnside-Butler syndrome is a neurodevelopmental disorder with a genetic basis, i.e., the occurrence of this syndrome is correlated with the presence of pathogenic CNV. Symptoms of Burnside-Butler syndrome include altered brain morphology, cognitive impairment and behavioural alterations. This disease is caused by the 15q11.2 BP1-BP2 deletion, which is a rare CNV spanning about 500 kbp. This region includes four highly conservative genes, including *TUBGCP5*, *CYFIP1*, *NIPA1* and *NIPA2* that are expressed in the brain. The dysfunction of their protein products is also associated with Prader-Willi syndrome (PWS) and Angelman syndrome (AS). The 15q11.2 BP1-BP2 deletion is present in 0.57–1.27% of the world population [50]. Inter-individual variability in clinical manifestation might be the consequence of incomplete penetrance and variable expression levels of genes located in the 15q11.2 BP1-BP2 region [40, 50].

Apart from CNVs covering the above-mentioned genes, the presence of pathogenic variants, commonly referred to as single nucleotide variants (SNVs), is reported as the mechanism underlying the development of Burnside-Butler syndrome which has been previously reported [40, 50].

22q11.2 Microdeletion Syndrome

Around 30 years ago, it was demonstrated that schizophrenia tends to co-occur with velocardiofacial syndrome [51]. In the same year, both velocardiofacial syndrome and 22q11.2 microdeletion syndrome (formerly known as DiGeorge syndrome), two clinical syndromes that were later classified into the spectrum of one syndrome of congenital abnormalities, were associated with deletions of the 22q11.2 region [52, 53]. However, the first genome-wide analysis of CNVs in patients with schizophrenia was published in 2007 [54].

The 22q.11.2 deletion occurs approximately in 1 of every 4000 live births and is equally distributed between males and females. The 22q11.2 microdeletion syndrome is associated with clinical manifestations with considerable inter-individual variability; however, most frequently, they include immunodeficiency, congenital cardiac anomalies and palatal abnormalities (each one being present in ~75% of individuals diagnosed with 22q11.2 microdeletion syndrome) [55]. The prevalence of schizophrenia in subjects with 22q11.2 microdeletion syndrome has been estimated at 25%, and this syndrome is known to be the strongest genetic factor of schizophrenia [56].

Major microdeletions, causative for the syndrome, are classified as CNVs and are usually larger than 1 kb. The inheritance of 22q11.2 microdeletion syndrome is autosomal dominant, as deletion of one copy chromosome 22 is sufficient to cause the disorder. However, most frequently,

the 22q11.2 microdeletion occurs spontaneously during gametogenesis. Parental studies show no association of the deletion with age of the parents [57–60]. This hemizygous deletion includes typically around 90 genes (~3 Mb). Parental studies show that the deletion inheritance occurs approximately in 10% of cases, while ~90% are caused by random deletion during gametogenesis or early development of the foetus [61]. Most of 22q11.2 microdeletions (~90%) are approximately 3 Mb in size; the remaining 10% include a nested deletion (~1.5–2 Mb) or atypical, non-nested distal microdeletions. Recent studies on CNVs in schizophrenia demonstrated that 5% of cases showed associations with rare CNVs rather than common CNVs. Furthermore, individuals with previously mentioned rare CNVs are about twenty times more likely to develop schizophrenia, indicating that the 22q11.2 microdeletion should be considered when diagnosing schizophrenia [26, 62, 63] (Table 1). Notably, microdeletions of various sizes in the 22q11.2 region also predispose to the development of other neurodevelopmental disorders, including attention-deficit/hyperactivity disorder (ADHD) and autism spectrum disorder [57, 64].

1q21.1 Microdeletion/Microduplication Syndrome

The 1q21.1 region contains many low-copy repeat sequences, and thus, it is susceptible to deletions and duplications occurring during meiotic division. The 1q21.1 locus is divided into four segment breakpoints (BP): BP1, BP2, BP3 and BP4. Therefore, two classes of the 1q21.1 microdeletion/microduplication-related CNVs have been described and include smaller 1.35 Mb class I CNVs and larger ~3 Mb class II CNVs. Class I CNVs involve the segment between BP3 and BP4, which is localised in the distal part of the 1q21.1 region. Class II microdeletion/microduplications are placed from the distal 1q21.1 to the proximal 1q21.1 regions, between BP2 and BP4 [17, 65]. Both microdeletions and microduplications can occur *de novo* or can be inherited in an autosomal dominant manner. Importantly, CNVs in the 1q21.1 region are characterized by incomplete penetration and variable expression [66].

The proximal part of the 1q21.1 region includes the *RBM8A* gene that is causally associated with the thrombocytopenia-absent radius (TAR) syndrome. However, microdeletions covering the distal part of the 1q21.1 region may manifest in developmental disorders, microcephaly and schizophrenia [67]. Many patients with the 1q21.1 microduplication show other signs of psychopathology, including anxiety, depression and ADHD. Neurological manifestations that include epilepsy and hypotonia might also occur leading to developmental delay. In paediatric patients, microduplication in this region might be associated with macrocephaly, developmental delay and autism spectrum disorder. In turn,

the 1q21.1 microdeletions are predominant in patients diagnosed with schizophrenia and microcephaly [67].

Association Between CNVs and Clinical Features of Schizophrenia

There are many possibilities of how the different CNVs contribute to the development of schizophrenia and its numerous clinical features, including deficits in social skills, learning processes, emotional recognition and cognitive flexibility [47, 63, 68–70]. Moreover, it is reported that patients with schizophrenia and pathogenic CNVs are more likely to present treatment resistance [71]. The mechanism of how CNVs influence symptoms of schizophrenia remains unclear. However, mouse models and induced pluripotent stem cells have successfully been used by studies addressing the neurobiology of CNVs associated with schizophrenia [72–77]. Abnormalities of basic associative learning processes have long been correlated with this particular disorder. These observations have been followed by many researchers, including studies conducted in 2017, results of which show that CNVs impact inhibitory learning in schizophrenia, which potentially contributes to the development of core symptoms in this disorder [63]. Genes associated with processes involved in synaptic plasticity (such as genes encoding components of the NMDA receptor complex, connected with glutamatergic signalling, but also genes involved in inhibitory GABAergic modulation of neuronal signalling) have been shown to be affected by CNVs [78, 79]. Products of these genes are synaptic proteins regulating the molecular processes directly related to associative learning and memory. Deficits in associative learning in individuals with schizophrenia have been assumed to contribute to the development and persistence of psychotic and cognitive symptoms. Patients with schizophrenia typically show the persistence of delusional beliefs, and thus, impairment of extinction learning could be an explanation of the core symptoms of schizophrenia [19, 33, 80, 81].

The study conducted in young patients diagnosed with 22q11.2 deletion syndrome has demonstrated that both negative and positive symptoms of schizophrenia were escalated in individuals with worst social skills. In the study from 2016, symptoms typical for schizophrenia, such as social anxiety and lack of close friends, were found to be increased in individuals with 22q11.2 DS [68–70].

22q11.2 deletion and other high-risk CNVs also contribute to a large extent to the intellectual disability, which was proved in a study using genome-wide microarrays. Moreover, scores of attention tests correlate with particular symptoms of schizophrenia. Lower scores are associated with negative symptoms, but not positive symptoms in this disorder. Also, low IQ test scores achieved by children can predict the likelihood of developing schizophrenia during

adulthood. Lower scores for social and cognitive tests are frequent in young individuals at risk mental states, compared with controls. All mentioned low scores provide probabilistic, not categorical, predictors for schizophrenia development [34, 61, 62, 82, 83].

Single Nucleotide Variants (SNVs) and Gene Expression in Schizophrenia

Studies on single nucleotide variants in schizophrenia are crucial for understanding the pathological mechanism of this condition. This subject has been discussed previously in details by several authors [22, 84–91]. Two different mechanisms (deletion and point mutations) for loss of function observed for genes critical to any condition with genetic background confirm their clinical significance and support the hypothesis about double-hits while loss of heterozygosity (LOH) co-occurring with a functional variants may be relevant [85].

Neurostructural Abnormalities Associated with Schizophrenia Risk CNVs

Schizophrenia risk CNVs have been noticed to have correlation with certain morphological brain abnormalities. Especially early-onset schizophrenia patients have been noticed to show dysmorphic features. Early detection of individuals with high risk of schizophrenia development is crucial for creating suitable prevention and/or therapy [92].

Windows of vulnerability occurring multiple times during the brain development are when abnormalities might progress which can lead to the development of the symptoms of mental disorders. One of the most popular neurodevelopmental theories in schizophrenia pathogenesis is the two-hit model, which states that there are two points of aberrant development (early brain development and adolescence) that significantly increase the risk for schizophrenia-like symptoms in the individual [93].

The study conducted by Warland et al. (2019) focused on the association of schizophrenia CNVs with subcortical brain volumes [20]. The authors used samples of participants from the UK Biobank. The analysis of MRI data focused on 15 metrics of subcortical volumes, out of which 5 brain regions (right thalamus, left putamen, right pallidum, right hippocampus and right accumbens) presented the significant association with schizophrenia-related CNVs. All of these subcortical structures showed a reduction in volume in patients with schizophrenia and CNVs [20]. Reduced volume of subcortical structures in patients carrying schizophrenia-related CNVs compared to unaffected CNVs carriers were also reported by previous studies [94, 95]. Reduction in

volumes of subcortical structures, such as the hippocampus and thalamus, was observed in individuals with high risk of developing schizophrenia, compared to healthy controls [96]. Changes in brain cortical anatomy associated with the presence of rare CNVs have been observed by Caseras et al. in 2021 [97]. The 1q21.1 deletion and the 15q11.2 deletion CNVs were associated with reduced gyrus surface area in carriers. Also, it appeared that the 15q11.2 deletion correlates with thicker cortex in carriers [97]. Finally, the 22q11.2 microdeletion has been associated with thicker cortex and reductions in the cortical surface area [98].

Discussion

The correlation of selected CNVs at specific *loci* with risk factors for several neuropsychiatric disorders, such as schizophrenia, autism spectrum disorder, intellectual disability and depression, has been reported previously [21, 31, 99]. Further research is needed to determine which genetic abnormalities or genetic variants are related to the phenotypic expression of schizophrenia symptoms, including negative, positive and cognitive abnormalities. For example, the *BCL9* gene polymorphisms are thought to be associated with negative symptoms in schizophrenia, as mentioned gene product is involved in the Wnt signalling pathway, a conserved pathway regulating crucial processes of cell fate determination in metazoan animals, including humans, which is significant in neuroplasticity, neurogenesis and cell survival [46, 100–103].

Genetic background for schizophrenia should not be considered without other crucial aspects of the expressed phenotype—such as differences in synaptic signalling and plasticity, abnormalities in micro and macrostructures of the brain or even environmental factors, which are significant in many psychiatric disorders. Determining which genes are involved in the development of schizophrenia is the first step of determining the role of mentioned genes' products in molecular pathways leading to the expression of schizophrenia symptoms. The phenotype may also depend on the type of particular CNVs—microdeletion and microduplication phenotypes often appear to be on the two ends of the spectrum. This situation is especially apparent in patients with 1q21.1 microdeletion/microduplication syndrome. Microdeletion in this region is correlated with schizophrenia and microcephaly; microduplication, on the other hand, is prevalent in individuals with autism spectrum disorder and macrocephaly [38].

There are also some reports on 3q29 deletion syndrome which is connected with > 40-fold higher risk for schizophrenia and the presence of treatment-resistant psychotic symptoms, multiple medical comorbidities and early-onset

dementia [104, 105], 16p11.2 duplication and [106] and 17q12 deletion [107].

In some cases, like in Burnside-Butler syndrome, the clinical phenotype of the child depends on the origin of parental deletion—if deletion is inherited from the father, there is a higher risk of congenital heart defects in the offspring; however, if deletion is maternal then the risk for intellectual disability and autism is increased [108]. In this situation, awareness of the genetic profiles of the parents can determine abnormalities which their future offspring is more prone to, and it can also increase the quality of genetic counselling.

Conclusions and Futures Directions

Numerous disorders with a genetic background such as previously mentioned 22q11.2 microdeletion syndrome, Burnside-Butler Syndrome (15q11.2 BP1-BP2 microdeletion) or 1q21.1 microduplication/microdeletion syndrome have been noticed to contribute to a higher risk of schizophrenia development. Patients with said disorders often display psychiatric (i.e., cognitive impairment, psychosis) and neurostructural symptoms (i.e., brain volumetric reduction, increased cortical thickness) that are observed in schizophrenic patients which further highlights the importance of CNVs in the understanding of the genetic background of schizophrenia [38, 40, 43, 109].

It is worth acknowledging that many CNVs occur among the human population and show no pathogenic impact, while others can be associated with a predisposition to various psychiatric disorders. Only 3.5–5% of individuals with schizophrenia are carriers of major-risk CNVs. Moreover, mentioned CNVs have also been observed in healthy individuals, which may suggest that other factors such as epigenetics or environmental conditions could be involved in schizophrenia development [34, 36].

Further research of CNVs that are associated with the development of schizophrenia could contribute to new prospects of therapies and prevention in people with a higher genetic risk of the disorder. More GWAS analyses could also reveal novel copy number variations associated with schizophrenia which, combined with current knowledge about single-gene copy number variants and gene expression, could be vital in the understanding of the aetiology of this psychiatric disorder.

The application of genome-wide experiments not only allows us to better understand the genetic background of schizophrenia but also supports the thesis about its polygenic inheritance and genetic overlap with other mental disorders including autism spectrum disorder or bipolar disorder. A thorough understanding of the genetic basis of this disorder could enable predictive testing, early

diagnosis and more effective therapy. Due to the complexity of schizophrenia and a variety of underlying genetic mechanisms, including CNVs, this appears to be a difficult task, but a promising next step may be to apply machine learning techniques together with high-throughput gene research technologies and clinical data analysis.

It is also important to note the complexity of schizophrenia in terms of exposure to environmental factors known to affect a risk of psychosis, clinical manifestation, course of the disorder and clinical and functional outcomes. Moreover, certain aspects of psychopathological symptoms and behavioural abnormalities, which are known to be present in schizophrenia, cross traditional diagnostic boundaries established by ICD and DSM classification systems [110]. These include positive, negative, mood and disorganization symptoms and cognitive impairment [8, 111]. In this regard, future studies that aim to provide new insights into the role of CNVs need to deconstruct the psychosis spectrum by dissecting specific dimensions of psychopathology or symptom clusters. Studies in this field may also use existing approaches related to the Research Domain Criteria (RDoC) and the Hierarchical Taxonomy of Psychopathology (HiTOP) [111, 112]. Also, investigating CNVs with respect to specific endophenotypes that capture phenotype constructs characterized by the association with illness in the population, heritability, state-independent manifestation (expression of the phenotype independent of the illness activity) and the association with illness in families might be helpful in addressing the heterogeneity of the diagnostic construct called schizophrenia [113, 114]. Nevertheless, large samples of individuals with psychosis spectrum disorders will be needed due to relatively low effect size estimates of specific CNVs. However, these research efforts might uncover on how CNVs build up specific phenotypes at the continuum of psychosis.

Author Contribution All authors contributed to the study's conception and design. The first draft of the manuscript was written by Kamila Szećowka and Izabela Laczmańska, and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

Availability of Data and Materials Data sharing is not applicable to this article.

Declarations

Ethics Approval Not applicable.

Consent to Participate Not applicable.

Consent for Publication Not applicable.

Research Involving Human Participants and/or Animals Not applicable.

Informed Consent Not applicable.

Competing Interests The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Saha S, Chant D, Welham J, McGrath J (2005) A systematic review of the prevalence of schizophrenia. *PLoS Med* 2(5):e141. <https://doi.org/10.1371/journal.pmed.0020141>
- Jaeschke K, Hanna F, Ali S et al (2021) Global estimates of service coverage for severe mental disorders: findings from the WHO Mental Health Atlas 2017. *Glob Ment Heal (Cambridge, England)* 8. <https://doi.org/10.1017/GMH.2021.19>
- McGrath J, Saha S, Chant D, Welham J (2008) Schizophrenia: a concise overview of incidence, prevalence, and mortality. *Epidemiol Rev* 30:67–76. <https://doi.org/10.1093/EPIREV/MXN001>
- Carpenter WT, Heinrichs DW, Wagman AMI (1988) Deficit and nondeficit forms of schizophrenia: the concept. *Am J Psychiatry*. 145(5):578–583. <https://doi.org/10.1176/ajp.145.5.578>
- Moustafa AA (2021) Cognitive and behavioral dysfunction in schizophrenia. Academic Press, London
- Ganguly P, Soliman A, Moustafa AA (2018) Holistic management of schizophrenia symptoms using pharmacological and non-pharmacological treatment. *Front Public Heal* 6:166. <https://doi.org/10.3389/FPUBH.2018.00166>
- Moustafa AA, Garami JK, Mahlberg J et al (2016) Cognitive function in schizophrenia: conflicting findings and future directions. *Rev Neurosci* 27:435–448. <https://doi.org/10.1515/REVNEURO-2015-0060>
- Frydecka D, Eissa AM, Hewedi DH et al (2014) Impairments of working memory in schizophrenia and bipolar disorder: the effect of history of psychotic symptoms and different aspects of cognitive task demands. *Front Behav Neurosci* 8:416. <https://doi.org/10.3389/FNBEH.2014.00416>
- Dickerson F, Kirkpatrick B, Boronow J et al (2006) Deficit schizophrenia: association with serum antibodies to cytomegalovirus. *Schizophr Bull* 32:396–400. <https://doi.org/10.1093/schbul/sbi054>
- Kirkpatrick B, Galderisi S (2008) Deficit schizophrenia: an update. *World Psychiatry* 7:143–147. <https://doi.org/10.1002/j.2051-5545.2008.tb00181.x>
- Galderisi S, Maj M (2009) Deficit schizophrenia: an overview of clinical, biological and treatment aspects. *Eur Psychiatry* 24:493–500. <https://doi.org/10.1016/j.eurpsy.2009.03.001>
- Bora E, Binnur Akdede B, Alptekin K (2017) Neurocognitive impairment in deficit and non-deficit schizophrenia: a meta-analysis. *Psychol Med* 47:2401–2413. <https://doi.org/10.1017/S0033291717000952>
- Arango C, Kirkpatrick B, Buchanan RW (2000) Neurological signs and the heterogeneity of schizophrenia. *Am J Psychiatry* 157:560–565. <https://doi.org/10.1176/appi.ajp.157.4.560>
- Galderisi S, Maj M, Mucci A et al (2002) Historical, psychopathological, neurological, and neuropsychological aspects of deficit schizophrenia: a multicenter study. *Am J Psychiatry* 159:983–990. <https://doi.org/10.1176/appi.ajp.159.6.983>
- Bray NJ, O'Donovan MC (2018) The genetics of neuropsychiatric disorders. *Brain Neurosci Adv* 2:239821281879927. <https://doi.org/10.1177/2398212818799271>
- Grozeva D, Kirov G, Ivanov D et al (2010) Rare copy number variants: a point of rarity in genetic risk for bipolar disorder and schizophrenia. *Arch Gen Psychiatry* 67:318–327. <https://doi.org/10.1001/ARCHGENPSYCHIATRY.2010.25>
- Rees E, Walters JTR, Georgieva L et al (2013) Analysis of copy number variations at 15 schizophrenia-associated loci. *Br J Psychiatry* 204:108–114. <https://doi.org/10.1192/BJP.BP.113.131052>
- Szatkiewicz JP, O'Dushlaine C, Chen G et al (2014) Copy number variation in schizophrenia in Sweden. *Mol Psychiatry* 19:762–773. <https://doi.org/10.1038/MP.2014.40>
- Hall J, Trent S, Thomas KL et al (2015) Genetic risk for schizophrenia: convergence on synaptic pathways involved in plasticity. *Biol Psychiatry* 77:52–58. <https://doi.org/10.1016/j.biopsych.2014.07.011>
- Warland A, Kendall KM, Rees E et al (2020) Schizophrenia-associated genomic copy number variants and subcortical brain volumes in the UK Biobank. *Mol Psychiatry* 25:854–862. <https://doi.org/10.1038/s41380-019-0355-y>
- Rees E, Kirov G (2021) Copy number variation and neuropsychiatric illness. *Curr Opin Genet Dev* 68:57–63. <https://doi.org/10.1016/J.GDE.2021.02.014>
- Shnyder NA, Novitsky MA, Neznanov NG et al (2022) Genetic predisposition to schizophrenia and depressive disorder comorbidity. *Genes (Basel)* 13(3):457. <https://doi.org/10.3390/GENES13030457>
- Martinez-Granero F, Blanco-Kelly F, Sanchez-Jimeno C et al (2021) Comparison of the diagnostic yield of aCGH and genome-wide sequencing across different neurodevelopmental disorders npj. *Genomic Med* 61(6):1–12. <https://doi.org/10.1038/s41525-021-00188-7>
- Pös O, Radvanszky J, Buglyó G et al (2021) DNA copy number variation: main characteristics, evolutionary significance, and pathological aspects. *Biomed J* 44:548–559. <https://doi.org/10.1016/J.BJ.2021.02.003>
- Zarrei M, MacDonald JR, Merico D (2015) Scherer SW (2015) A copy number variation map of the human genome. *Nat Rev Genet* 16(16):172–183. <https://doi.org/10.1038/nrg3871>
- Girirajan S, Brkanac Z, Coe BP et al (2011) Relative burden of large CNVs on a range of neurodevelopmental phenotypes. *PLoS Genet* 7:e1002334. <https://doi.org/10.1371/JOURNAL.PGEN.1002334>
- Thapar A, Cooper M (2013) Copy number variation: what is it and what has it told us about child psychiatric disorders? *J Am Acad Child Adolesc Psychiatry* 52:772–774. <https://doi.org/10.1016/j.jaac.2013.05.013>
- Ruderfer DM, Hamamsy T, Lek M et al (2016) Patterns of genic intolerance of rare copy number variation in 59,898 human exomes. *Nat Genet* 48:1107–1111. <https://doi.org/10.1038/ng.3638>
- Singh AK, Olsen MF, Lavik LAS et al (2021) Detecting copy number variation in next generation sequencing data from diagnostic gene panels. *BMC Med Genomics* 14:1–12. <https://doi.org/10.1186/S12920-021-01059-X/TABLES/2>

30. Malhotra D, Sebat J (2012) CNVs: harbingers of a rare variant revolution in psychiatric genetics. *Cell* 148:1223–1241. <https://doi.org/10.1016/J.CELL.2012.02.039>
31. Nakatochi M, Kushima I, Ozaki N (2021) Implications of germline copy-number variations in psychiatric disorders: review of large-scale genetic studies. *J Hum Genet* 66:25–37. <https://doi.org/10.1038/S10038-020-00838-1>
32. Lee PH, Feng YCA, Smoller JW (2021) Pleiotropy and cross-disorder genetics among psychiatric disorders. *Biol Psychiatry* 89:20–31. <https://doi.org/10.1016/J.BIOPSYCH.2020.09.026>
33. Clifton NE, Pocklington AJ, Scholz B et al (2017) Schizophrenia copy number variants and associative learning. *Mol Psychiatry* 22:178–182. <https://doi.org/10.1038/mp.2016.227>
34. Rees E, Kendall K, Pardiñas AF et al (2016) Analysis of intellectual disability copy number variants for association with schizophrenia. *JAMA Psychiat* 73:963–969. <https://doi.org/10.1001/jamapsychiatry.2016.1831>
35. Henriksen MG, Nordgaard J, Jansson LB (2017) Genetics of schizophrenia: overview of methods, findings and limitations. *Front Hum Neurosci* 11:322. <https://doi.org/10.3389/FNHUM.2017.00322/BIBTEX>
36. Costain G, Lionel AC, Merico D et al (2013) Pathogenic rare copy number variants in community-based schizophrenia suggest a potential role for clinical microarrays. *Hum Mol Genet* 22:4485–4501. <https://doi.org/10.1093/hmg/ddt297>
37. Tansey KE, Rees E, Linden DE et al (2016) Common alleles contribute to schizophrenia in CNV carriers. *Mol Psychiatry* 21:1085–1089. <https://doi.org/10.1038/mp.2015.143>
38. Yoon J, Mao Y (2021) Dissecting molecular genetic mechanisms of 1q21.1 CNV in neuropsychiatric disorders. *Int J Mol Sci* 22(11):5811. <https://doi.org/10.3390/ijms22115811>
39. Skórka A, Bielicka-Cymermann J, Gieruszczak-Białek D, Korniszewski L (2005) Thrombocytopenia - Absent radius (TAR) syndrome: a case with agenesis of corpus callosum, hypoplasia of cerebellar vermis and horseshoe kidney. *Genet Couns* 16(4):377–382
40. Rafi SK, Butler MG (2020) The 15q11.2 bp1-bp2 microdeletion (burnside-butler) syndrome: In silico analyses of the four coding genes reveal functional associations with neurodevelopmental phenotypes. *Int J Mol Sci* 21(9):3296. <https://doi.org/10.3390/ijms21093296>
41. Ulfarsson MO, Walters GB, Gustafsson O et al (2017) 15q11.2 CNV affects cognitive, structural and functional correlates of dyslexia and dyscalculia. *Transl Psychiatry* 7(4):e1109. <https://doi.org/10.1038/TP.2017.77>
42. Butler MG (2019) Magnesium supplement and the 15q11.2 BP1–BP2 microdeletion (Burnside–Butler) syndrome: a potential treatment? *Int J Mol Sci* 20(12):2914. <https://doi.org/10.3390/IJMS20122914>
43. Bearden CE, Van Erp TGM, Dutton RA et al (2007) Mapping cortical thickness in children with 22q11.2 deletions. *Cereb Cortex* 17:1889. <https://doi.org/10.1093/CERCOR/BHL097>
44. Kates WR, Burnette CP, Jabs EW et al (2001) Regional cortical white matter reductions in velocardiofacial syndrome: a volumetric MRI analysis. *Biol Psychiatry* 49:677–684. [https://doi.org/10.1016/S0006-3223\(00\)01002-7](https://doi.org/10.1016/S0006-3223(00)01002-7)
45. Jalbrzikowski M, Jonas R, Senturk D et al (2013) Structural abnormalities in cortical volume, thickness, and surface area in 22q11.2 microdeletion syndrome: relationship with psychotic symptoms. *Neuroimage (Amst)* 3:405. <https://doi.org/10.1016/J.NICL.2013.09.013>
46. Kimura H, Tanunaka S, Kushima I et al (2015) Association study of BCL9 gene polymorphism rs583583 with schizophrenia and negative symptoms in Japanese population. *Sci Rep* 5:15705. <https://doi.org/10.1038/SREP15705>
47. Antshel KM, Fremont W, Ramanathan S, Kates WR (2017) Predicting cognition and psychosis in young adults with 22q11.2 deletion syndrome. *Schizophr Bull* 43:833–842. <https://doi.org/10.1093/schbul/sbw135>
48. Kates WR, Russo N, Wood WM et al (2015) Neurocognitive and familial moderators of psychiatric risk in velocardiofacial (22q11.2 deletion) syndrome: a longitudinal study. *Psychol Med* 45:1629–1639. <https://doi.org/10.1017/S0033291714002724>
49. Marshall CR, Howrigan DP, Merico D et al (2016) Contribution of copy number variants to schizophrenia from a genome-wide study of 41,321 subjects. *Nat Genet* 49(49):27–35. <https://doi.org/10.1038/ng.3725>
50. Cox DM, Butler MG (2015) The 15q11.2 BP1–BP2 microdeletion syndrome: a review. *Int J Mol Sci* 16:4068–4082. <https://doi.org/10.3390/ijms16024068>
51. Shprintzen RJ, Goldberg R, Golding-Kushner KJ, Marion RW (1992) Late-onset psychosis in the velo-cardio-facial syndrome. *Am J Med Genet* 42:141–142
52. Driscoll DA, Budarf ML, Emanuel BS (1992) A genetic etiology for DiGeorge syndrome: consistent deletions and microdeletions of 22q11. *Am J Hum Genet* 50:924–933
53. Driscoll DA, Spinner NB, Budarf ML et al (1992) Deletions and microdeletions of 22q11.2 in velo-cardio-facial syndrome. *Am J Med Genet* 44:261–268. <https://doi.org/10.1002/ajmg.1320440237>
54. Szatmari P, Paterson AD, Zwaigenbaum L et al (2007) Mapping autism risk loci using genetic linkage and chromosomal rearrangements. *Nat Genet* 39:319–328. <https://doi.org/10.1038/ng1985>
55. McDonald-McGinn DM, Sullivan KE, Marino B et al (2015) 22q11.2 deletion syndrome. *Nat Rev Dis Prim* 1:15071. <https://doi.org/10.1038/NRDP.2015.71>
56. Owen MJ, Doherty JL (2016) What can we learn from the high rates of schizophrenia in people with 22q11.2 deletion syndrome? *World Psychiatry* 15(1):23–25. <https://doi.org/10.1002/wps.20274>
57. Murphy KC, Jones LA, Owen MJ (1999) High rates of schizophrenia in adults with velo-cardio-facial syndrome. *Arch Gen Psychiatry* 56:940–945. <https://doi.org/10.1001/archpsyc.56.10.940>
58. Kraus C, Vanicek T, Weidenauer A et al (2018) (2018) DiGeorge syndrome. *Wiener Klin Wochenschrift* 1307(130):283–287. <https://doi.org/10.1007/S00508-018-1335-Y>
59. Scambler PJ, Kelly D, Lindsay E et al (1992) Velo-cardio-facial syndrome associated with chromosome 22 deletions encompassing the DiGeorge locus. *Lancet* 339:1138–1139. [https://doi.org/10.1016/0140-6736\(92\)90734-K](https://doi.org/10.1016/0140-6736(92)90734-K)
60. Van L, Boot E, Bassett AS (2017) Update on the 22q11.2 deletion syndrome and its relevance to schizophrenia. *Curr Opin Psychiatry* 30:191–196. <https://doi.org/10.1097/ycp.000000000324>
61. Hiroi N, Takahashi T, Hishimoto A et al (2013) Copy number variation at 22q11.2: from rare variants to common mechanisms of developmental neuropsychiatric disorders. *Mol Psychiatry* 18:1153–1165. <https://doi.org/10.1038/mp.2013.92>
62. Palmer BW, Heaton RK, Kuck J et al (1997) Is it possible to be schizophrenic yet neuropsychologically normal? *Neuropsychology* 11:437–446. <https://doi.org/10.1037/0894-4105.11.3.437>
63. Lowther C, Costain G, Baribeau DA, Bassett AS (2017) Genomic disorders in psychiatry—what does the clinician need to know? *Curr Psychiatry Rep* 19(11):82. <https://doi.org/10.1007/s11920-017-0831-5>
64. Kraus C, Vanicek T, Weidenauer A et al (2018) DiGeorge syndrome: relevance of psychiatric symptoms in undiagnosed adult

- patients. *Wien Klin Wochenschr* 130:283–287. <https://doi.org/10.1007/s00508-018-1335-y>
65. Marshall CR, Howrigan DP, Merico D et al (2017) Contribution of copy number variants to schizophrenia from a genome-wide study of 41,321 subjects. *Nat Genet* 49:27–35. <https://doi.org/10.1038/ng.3725>
 66. Sønnderby IE, van der Meer D, Moreau C et al (2021) 1Q21.1 Distal copy number variants are associated with cerebral and cognitive alterations in humans. *Transl Psychiatry* 11. <https://doi.org/10.1038/s41398-021-01213-0>
 67. Albers CA, Newbury-Ecob R, Ouwehand WH, Ghevaert C (2013) New insights into the genetic basis of TAR (thrombocytopenia-absent radii) syndrome. *Curr Opin Genet Dev* 3(3):316–23. <https://doi.org/10.1016/j.gde.2013.02.015>
 68. Fonseca-Pedrero E, Debbané M, Schneider M et al (2016) Schizotypal traits in adolescents with 22q11.2 deletion syndrome: validity, reliability and risk for psychosis. *Psychol Med* 46:1005–1013. <https://doi.org/10.1017/S0033291715002500>
 69. Mekori-Domachevsky E, Guri Y, Yi J et al (2017) Negative subthreshold psychotic symptoms distinguish 22q11.2 deletion syndrome from other neurodevelopmental disorders: a two-site study. *Schizophr Res* 188:42–49. <https://doi.org/10.1016/j.schres.2016.12.023>
 70. Vangkilde A, Jepsen JRM, Schmock H et al (2016) Associations between social cognition, skills, and function and subclinical negative and positive symptoms in 22q11.2 deletion syndrome. *J Neurodev Disord* 8. <https://doi.org/10.1186/s11689-016-9175-4>
 71. Kushima I, Aleksic B, Nakatochi M et al (2017) High-resolution copy number variation analysis of schizophrenia in Japan. *Mol Psychiatry* 22:430–440. <https://doi.org/10.1038/MP.2016.88>
 72. Nakazawa T (2022) Modeling schizophrenia with iPSC cell technology and disease mouse models. *Neurosci Res* 175:46–52. <https://doi.org/10.1016/j.NEURES.2021.08.002>
 73. Räsänen N, Tiihonen J, Koskivi M et al (2022) The iPSC perspective on schizophrenia. *Trends Neurosci* 45:8–26. <https://doi.org/10.1016/j.TINS.2021.11.002>
 74. Forsingdal A, Jørgensen TN, Olsen L et al (2019) Can animal models of copy number variants that predispose to schizophrenia elucidate underlying biology? *Biol Psychiatry* 85:13–24. <https://doi.org/10.1016/j.BIOPSYCH.2018.07.004>
 75. Arioka Y, Shishido E, Kushima I et al (2021) Chromosome 22q11.2 deletion causes PERK-dependent vulnerability in dopaminergic neurons. *EBioMedicine* 63:103138. <https://doi.org/10.1016/j.EBIOM.2020.103138>
 76. Arioka Y, Shishido E, Kubo H et al (2018) Single-cell trajectory analysis of human homogenous neurons carrying a rare RELN variant. *Transl Psychiatry* 8(1):129. <https://doi.org/10.1038/S41398-018-0177-8>
 77. Saito R, Koebis M, Nagai T, et al (2020) Comprehensive analysis of a novel mouse model of the 22q11.2 deletion syndrome: a model with the most common 3.0-Mb deletion at the human 22q11.2 locus. *Transl Psychiatry* 10. <https://doi.org/10.1038/S41398-020-0723-Z>
 78. Kirov G, Pocklington AJ, Holmans P et al (2012) De novo CNV analysis implicates specific abnormalities of postsynaptic signaling complexes in the pathogenesis of schizophrenia. *Mol Psychiatry* 17:142–153. <https://doi.org/10.1038/mp.2011.154>
 79. Pocklington AJ, Rees E, Walters JTR et al (2015) Novel findings from CNVs implicate inhibitory and excitatory signaling complexes in schizophrenia. *Neuron* 86:1203. <https://doi.org/10.1016/j.NEURON.2015.04.022>
 80. Corlett PR, Krystal JH, Taylor JR, Fletcher PC (2009) Why do delusions persist? *Front Hum Neurosci* 3. <https://doi.org/10.3389/neuro.09.012.2009>
 81. Gruart A, Leal-Campanario R, López-Ramos JC, Delgado-García JM (2015) Functional basis of associative learning and their relationships with long-term potentiation evoked in the involved neural circuits: Lessons from studies in behaving mammals. *Neurobiol Learn Mem* 124:3–18. <https://doi.org/10.1016/j.nlm.2015.04.006>
 82. Hollis C (1995) Child and adolescent (juvenile onset) schizophrenia. A case control study of premorbid developmental impairments. *Br J Psychiatry* 166:489–495. <https://doi.org/10.1192/bjp.166.4.489>
 83. Cannon TD, Rosso IM, Bearden CE et al (1999) A prospective cohort study of neurodevelopmental processes in the genesis and epigenesis of schizophrenia. *Dev Psychopathol* 11:467–485. <https://doi.org/10.1017/S0954579499002163>
 84. Gejman P, Sanders A, Duan J (2010) The Role of Genetics in the Etiology of Schizophrenia. *Psychiatr Clin North Am* 33:35. <https://doi.org/10.1016/j.PSC.2009.12.003>
 85. Vorstman JAS, Olde Loohuis LM, Kahn RS, Ophoff RA (2018) Double hits in schizophrenia. *Hum Mol Genet* 27:2755. <https://doi.org/10.1093/HMG/DDY175>
 86. Singh T, Walters JTR, Johnstone M et al (2017) The contribution of rare variants to risk of schizophrenia in individuals with and without intellectual disability. *Nat Genet* 49:1167. <https://doi.org/10.1038/NG.3903>
 87. Lencz T, Yu J, Khan RR et al (2021) Novel ultra-rare exonic variants identified in a founder population implicate cadherins in schizophrenia. *Neuron* 109:1465–1478.e4. <https://doi.org/10.1016/j.neuron.2021.03.004>
 88. Singh SM, Castellani CA, Hill KA (2020) Postzygotic somatic mutations in the human brain expand the threshold-liability model of schizophrenia. *Front Psychiatry* 11:1093. <https://doi.org/10.3389/FPSYT.2020.587162/BIBTEX>
 89. Hathy E, Szabó E, Varga N et al (2020) Investigation of de novo mutations in a schizophrenia case-parent trio by induced pluripotent stem cell-based in vitro disease modeling: convergence of schizophrenia- and autism-related cellular phenotypes. *Stem Cell Res Ther* 11:1–15. <https://doi.org/10.1186/S13287-020-01980-5/FIGURES/5>
 90. Tromp A, Mowry B, Giacomotto J (2021) Neurexins in autism and schizophrenia—a review of patient mutations, mouse models and potential future directions. *Mol Psychiatry* 26:747–760. <https://doi.org/10.1038/S41380-020-00944-8>
 91. Sekiguchi M, Sobue A, Kushima I, et al (2020) ARHGAP10, which encodes Rho GTPase-activating protein 10, is a novel gene for schizophrenia risk. *Transl Psychiatry* 10. <https://doi.org/10.1038/S41398-020-00917-Z>
 92. Priol AC, Denis L, Boulanger G, et al (2021) Detection of morphological abnormalities in schizophrenia: an important step to identify associated genetic disorders or etiologic subtypes. *Int J Mol Sci* 22. <https://doi.org/10.3390/IJMS22179464>
 93. Debnath M, Venkatasubramanian G, Berk M (2015) Fetal programming of schizophrenia: select mechanisms. *Neurosci Biobehav Rev* 49:90. <https://doi.org/10.1016/j.NEUBIOREV.2014.12.003>
 94. Okada N, Fukunaga M, Yamashita F et al (2016) Abnormal asymmetries in subcortical brain volume in schizophrenia. *Mol Psychiatry* 21:1460. <https://doi.org/10.1038/MP.2015.209>
 95. Van Erp TGM, Hibar DP, Rasmussen JM et al (2016) Subcortical brain volume abnormalities in 2028 individuals with schizophrenia and 2540 healthy controls via the ENIGMA consortium. *Mol Psychiatry* 21:547. <https://doi.org/10.1038/MP.2015.63>
 96. Harrisberger F, Buechler R, Smieskova R et al (2016) Alterations in the hippocampus and thalamus in individuals at high risk for psychosis. *NPJ Schizophr* 2:16033. <https://doi.org/10.1038/NPJSCHZ.2016.33>
 97. Caseras X, Kirov G, Kendall KM et al (2021) Effects of genomic copy number variants penetrant for schizophrenia on cortical

- thickness and surface area in healthy individuals: analysis of the UK Biobank. *Br J Psychiatry* 218:104. <https://doi.org/10.1192/BJP.2020.139>
98. Sun D, Ching CRK, Lin A et al (2020) Large-scale mapping of cortical alterations in 22q11.2 deletion syndrome: convergence with idiopathic psychosis and effects of deletion size. *Mol Psychiatry* 25:1822. <https://doi.org/10.1038/S41380-018-0078-5>
99. Kirov G (2015) CNVs in neuropsychiatric disorders. *Hum Mol Genet* 24:R45–R49. <https://doi.org/10.1093/hmg/ddv253>
100. de la Roche M, Worm J, Bienz M (2008) The function of BCL9 in Wnt/beta-catenin signaling and colorectal cancer cells. *BMC Cancer* 8. <https://doi.org/10.1186/1471-2407-8-199>
101. Li J, Zhou G, Ji W et al (2011) Common variants in the BCL9 gene conferring risk of schizophrenia. *Arch Gen Psychiatry* 68:232–240. <https://doi.org/10.1001/ARCHGENPSYCHIATRY.2011.1>
102. Xu C, Aragam N, Li X, et al (2013) BCL9 and C9orf5 are associated with negative symptoms in schizophrenia: meta-analysis of two genome-wide association studies. <https://doi.org/10.1371/journal.pone.0051674>
103. Komiya Y, Habas R (2008) Wnt signal transduction pathways *Organogenesis* 4:68. <https://doi.org/10.4161/ORG.4.2.5851>
104. Harner MK, Lichtenstein M, Farrell M et al (2020) Treatment-resistant psychotic symptoms and early-onset dementia: a case report of the 3q29 deletion syndrome. *Schizophr Res* 224:195–197. <https://doi.org/10.1016/J.SCHRES.2020.08.012>
105. Nawa Y, Kushima I, Aleksic B et al (2022) Treatment-resistant schizophrenia in patients with 3q29 deletion: a case series of four patients. *Psychiatry Clin Neurosci* 76:338–339. <https://doi.org/10.1111/PCN.13361>
106. Hayashi Y, Kushima I, Aleksic B et al (2022) Variable psychiatric manifestations in patients with 16p11.2 duplication: a case series of 4 patients. *Psychiatry Clin Neurosci* 76:86–88. <https://doi.org/10.1111/PCN.13324>
107. Kushima I, Uematsu M, Ishizuka K et al (2022) Psychiatric patients with a de novo 17q12 deletion: two case reports. *Psychiatry Clin Neurosci* 76:345–347. <https://doi.org/10.1111/PCN.13367>
108. Davis KW, Serrano M, Loddo S et al (2019) Parent-of-origin effects in 15q11.2 BP1-BP2 microdeletion (Burnside-Butler) syndrome. *Int J Mol Sci* 20:1459. <https://doi.org/10.3390/IJMS20061459>
109. Karlsgodt KH, Sun D, Cannon TD (2010) Structural and functional brain abnormalities in schizophrenia. *Curr Dir Psychol Sci* 19:226. <https://doi.org/10.1177/0963721410377601>
110. Murray RM, Quattrone D (2022) The Kraepelinian concept of schizophrenia: dying but not yet dead. *Schizophr Res* 242:102–105. <https://doi.org/10.1016/J.SCHRES.2021.12.005>
111. Dikeos DG, Wickham H, McDonald C et al (2006) Distribution of symptom dimensions across Kraepelinian divisions. *Br J Psychiatry* 189:346–353. <https://doi.org/10.1192/BJP.BP.105.017251>
112. Kotov R, Krueger RF, Watson D (2018) A paradigm shift in psychiatric classification: the Hierarchical Taxonomy Of Psychopathology (HiTOP). *World Psychiatry* 17:24. <https://doi.org/10.1002/WPS.20478>
113. Gottesman II, Gould TD (2003) The endophenotype concept in psychiatry: etymology and strategic intentions. *Am J Psychiatry* 160:636–645. <https://doi.org/10.1176/APPI.AJP.160.4.636/ASSET/IMAGES/LARGE/L58F2.JPEG>
114. Misiak B, Frydecka D, Rybakowski JK (2016) Editorial: Endophenotypes for schizophrenia and mood disorders: Implications from genetic, biochemical, cognitive, behavioral, and neuroimaging studies. *Front Psychiatry* 7:83. <https://doi.org/10.3389/FPSYT.2016.00083/BIBTEX>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.