# Circulating Level of Blood Iron and Copper Associated with Inflammation and Disease Activity of Rheumatoid Arthritis

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## Abstract

This study aims to compare the concentrations of circulating levels of iron, zinc, and copper in blood samples of rheumatoid arthritis (RA) patients which determine the correlations with inflammation and disease activity. A total of 102 RA patients and 66 healthy controls were enrolled. Circulation of iron, zinc, and copper levels in whole blood were assessed. Hemo-globin, C-reactive protein (CRP), erythrocyte sedimentation rate (ESR), anticyclic citrullinated peptide antibody (anti-CCP) levels were collected. A meta-analysis was performed to validate our findings. Single and multiple variate generalized linear regression were applied to identify the correlation between trace elements and clinical characteristics. Blood copper level was significantly higher in RA patients (P < 0.001), while iron and zinc levels were decreased (P < 0.001 and P = 0.02, respectively). Meta-analysis confirmed our findings for zinc (SMD = -1.17, P < 0.001) and copper (SMD = 1.24, P < 0.001). Copper level was positively correlated with DAS28-CRP (r = 0.35, P < 0.01), CRP (r = -0.46, P < 0.01) and ESR (r = -0.55, P < 0.01). Circulating blood copper was significantly higher and positively correlated with DAS28-CRP and inflammatory markers, while circulating blood iron was decreased and negatively correlated with DAS28-CRP and inflammatory markers in RA patients.

Keywords Rheumatoid arthritis · Copper · Iron · Zinc · Disease activity · DAS28-CRP

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# Introduction

Rheumatoid arthritis (RA) is a chronic, autoimmune-based disease characterized by articular inflammation and progressive joint destruction [1]. Iron, zinc, and copper are important trace elements that are essential for the regulation of metabolism and immunological functions of the human body [2]. Zinc plays an important role in the immune system by affecting both innate and adaptive immune cells [3]. Many studies have found that endogenous zinc levels can affect both the number and the function of immune cells, including macrophages, T cells, and B cells. Activation of the immune cells mentioned above can release pro-inflammatory cytokines, such as interleukin 1 (IL-1), interleukin 6 (IL-6), and tumor necrosis factor (TNF), leading to joint damage due to RA inflammation. Patients with autoimmune diseases, such as type 1 diabetes and RA, may benefit from zinc supplementation [4, 5]. Iron is an important trace element in the synthesis of hemoglobin, and iron deficiency can lead to anemia. Anemia is a common complication in RA patients and is an important extra-articular manifestation



that correlates with physical disability accordingly increased mortality. The pathogenesis of anemia in RA is complex and multifactorial, perhaps due to inflammation, iron deficiency, or both [6]. Copper is also an important trace element. Most of the copper found in the human body is in a protein-bound form, such as ceruloplasmin and superoxide dismutase, which are important for antioxidant activity and maintenance of the immune system [7]. It has been reported that inflammation can increase the concentration of copper, because copper production in hepatocytes is stimulated by pro-inflammatory interleukins [8]. This study aimed to compare the concentrations of iron, zinc, and copper in RA patients with healthy individuals in order to determine their correlation with disease activity.

# **Materials and Methods**

## **Participants**

The subjects were recruited from Guanghua Hospital Precision Medicine Research Cohort (PMRC) [9] which is a hospital-based longitudinal cohort to investigate risk factors, genetic susceptibility, and pharmacogenetics for rheumatology diseases such as RA, osteoarthritis, and ankylosing spondylitis. Healthy individuals are derived from those with an annual physical exam without rheumatological disease. Currently, PMRC has enrolled > 30,000 disease patients and 10,000 healthy individuals as controls. A total of 102 patients diagnosed with RA based on the 2010 American College of Rheumatology (ACR)/European League Against Rheumatism classification criteria for RA [10] were enrolled, and 66 age- and sex-matched controls were included for comparison from PMRC [9]. Participants with other rheumatologic diseases, acute inflammatory, renal, hepatic, gastrointestinal, metabolic, or endocrine conditions were excluded.

## Data Collection

Medical history, clinical data, and physical examination findings of patients with RA, including disease course, hemoglobin, platelet count, C-reactive protein (CRP), rheumatoid factor (RF), anticyclic citrullinated peptide antibody (anti-CCP) levels, general health scores, joint swelling, and tenderness, were collected.

Fasting peripheral blood samples were collected from patients with RA and healthy controls. Iron, zinc, and copper levels in whole blood were measured by atomic absorption spectrometry using an acetylene flame. Laboratory tests, such as platelet count, hemoglobin, CRP, ESR, RF immunoglobulin G, RF immunoglobulin A, RF immunoglobulin M, and anti-CCP, were performed using standard laboratory methods.

Disease Activity Score with 28 joints using CRP (DAS28-CRP) was calculated to estimate disease activity. The 102 patients with RA were classified into two groups (57 patients with active disease vs 45 patients in remission) based on DAS28-CRP. RA patients with DAS28-CRP < 3.2 were divided into the RA remission group, while patients with DAS28-CRP  $\geq$  3.2 were divided into the active RA group.

## **Meta-analysis**

The standard mean difference (SMD) and 95% confidence interval (CI), based on the random effects model, were applied to evaluate the difference in serum levels of zinc and copper between patients with RA and healthy controls. The significance of pooled SMD between groups was assessed using the Z-test. Heterogeneity between studies was calculated according to Cochran's Q statistic, and the I<sup>2</sup> test was used to quantify the degree of inconsistency by calculating the percentage of total between-studies variation due to heterogeneity rather than chance [11]. Statistical significance was set at P < 0.05. Meta-analysis was conducted using the R software (R Foundation for Statistical Computing, Vienna, Austria).

## **Statistical Analysis**

SPSS version 21 (SPSS Inc., Chicago, IL, USA) was used for all statistical analyses, and data are expressed as the mean  $\pm$  standard error of mean. A one-way analysis of variance test was used when data were normally distributed, and the variance between groups was not significantly different. Nonparametric tests were used for non-normal distributions or unequal variance data. Stepwise multiple linear regression analysis was used to analyze the relationship between disease activity and trace element concentrations and different clinical, laboratory, and other parameters. The correlation coefficient (r) was calculated to evaluate the linear association between the level of trace elements and other parameters and the disease activity of RA. Statistical significance was set at p < 0.05.

## Results

## **Comparison of RA and Control Groups**

In the RA group, 15 patients were men and 87 were women, while in the control group, there were 12 men and 54 women. The mean blood copper level in in the RA group was significantly higher than that of the control group  $(18.11 \pm 4.24 \ \mu mol/L \ versus 14.96 \pm 2.45 \ \mu mol/L)$ ,

P < 0.001). The mean blood iron level in the RA group was  $6.74 \pm 1.06$  mmol/L, which was significantly lower than that of the control group  $(8.03 \pm 1.44 \text{ mmol/L}, P < 0.001)$ . The mean blood zinc level in the patients with RA was  $81.68 \pm 14.14 \mu mol/L$ , which was also significantly lower than that of the controls  $(87.06 \pm 14.76 \text{ }\mu\text{mol/L}, P = 0.02)$ . (Table 1 and Fig. 1).

#### Meta-analysis of Common Trace Metals in RA

Meta-analysis of the previously published results of RA trace element levels with our results was performed. Significantly decreased zinc levels were found in patients with RA (SMD = -1.17, 95% CI = -1.68, -0.66, Z = -4.51, P < 0.001). Copper was significantly higher in patients with RA than in controls (SMD = 1.24, 95% CI = 0.64 to - 1.85, Z = 4.04, P < 0.001) (Fig. 2).

## Comparison Between RA Remission, Active RA, and Control Group.

Patients in the RA remission group had higher hemoglobin and iron levels, and lower platelet counts, RF-IgG, RF-IgA, and copper levels compared to those in the active RA group. There were no statistically significant differences between the two groups with respect to age, sex, disease duration, RF-IgM, anti-CCP, and zinc levels (Table 2). The mean iron level in the RA group was significantly lower than that in the control group (P < 0.001). However, the mean copper and zinc levels were similar, with no significant difference between the two groups (P = 0.058 and 0.168, respectively). A comparison of the levels of the three trace elements in the active RA group and the control group revealed that copper levels were significantly higher in patients with active RA than in controls (P < 0.001). The levels of iron and zinc in the active RA group were both lower than those in the control group (P < 0.001 and P = 0.014, respectively) (Table 2, Fig. 3).

## **Multiple Linear Regression Analysis** of the Relationship Between DAS28-CRP and Trace **Elements, and Other Clinical Parameters**

In the multiple linear regression analysis of the relationship between disease activity (DAS28-CRP), trace elements, and other clinical parameters (sex, age, disease duration, hemoglobin, platelet count, RF-IgG, RF-IgA, RF-IgM, and anti-CCP), four items, including age, hemoglobin, copper, and RF-IgG levels, were found to be correlated with DAS28-CRP (R-square = 0.305) (Table 3).

Table 1 Comparison between the RA and control groups

	RA group $(n=102)$	Control group $(n=66)$	MD	SED	Р
Sex (women/men)	87/15	54/12	NA	NA	0.668
Age (years)	$57.19 \pm 12.06$	$57.95 \pm 13.69$	-0.77	2.01	0.703
Copper (µmol/L)	$18.11 \pm 4.24$	$14.96 \pm 2.45$	3.15	0.52	< 0.001
Iron (mmol/L)	$6.74 \pm 1.06$	$8.03 \pm 1.44$	-1.29	0.19	< 0.001
Zinc (µmol/L)	$81.68 \pm 14.14$	$87.06 \pm 14.76$	-5.38	2.27	0.02

MD, mean difference; SED, std. error difference



Fig. 1 Comparison of mean blood copper, iron, and zinc between RA and control group. A Copper level; B Iron level; C Zinc level

#### A Zinc level

	6	Experimenta	al		Control				
Study	Total	Mean	SD	Total	Mean	SD		Weight	SMD [95% CI]
Aaseth J 1978	23	10	3	30	13	3	Her	4.17%	-0.99 [ -1.56, -0.41]
Sahebari M 2015	110	902.56	158.69	100	748.87	162.11		4.34%	0.96 [ 0.67, 1.24]
Afridi HI 2015	53	5.97	0.55	52	9.37	1.41	H#H	4.17%	-3.17 [ -3.74, -2.59]
Yazar M 2005	25	66.3	23.51	25	65.78	15.1	H	4.18%	0.03 [ -0.53, 0.58]
Li J 2014	60	86.52	16.36	60	141.66	20	HH	4.21%	-3.00 [ -3.52, -2.48]
Bacon MC 1900	34	130.53	33.18	9	100	13		4.00%	0.99 [ 0.22, 1.75]
Niedermeier W 1971	105	72.8	19.5	105	115.4	39.4	-	4.34%	-1.37 [ -1.67, -1.07]
Banford JC 1982	85	12.1	2.1	49	12.1	1.6	in the second se	4.31%	0.00 [ -0.35, 0.35]
Morgenstern H 1983	30	58.5	10.3	30	74.6	14.3	H <b>H</b> H	4.18%	-1.28 [ -1.83, -0.72]
Zoli A 1998	57	85.6	21.9	20	108.1	19.2	HH	4.20%	-1.05 [ -1.58, -0.51]
Tuncer S 1999	39	108.68	0.32	22	125.3	6.38	H#H	3.84%	-4.30 [ -5.233.38]
Wanchu A 2002	39	824	386	22	1024	428	H	4.20%	-0.49 [ -1.02, 0.04]
Silverio Amancio OM 2003	41	93.7	14.6	23	92.6	11.3	101	4.21%	0.08 [ -0.43, 0.59]
Ala S 2009	40	0.71	0.19	40	1.04	0.36	нн	4.24%	-1.14 [ -1.61, -0.66]
Ullah Z 2016	61	85.6	32.3	61	95.9	16.4	<b>.</b>	4.31%	-0.40 [ -0.76, -0.04]
Afridi HI 2013	202	6.98	1.15	203	9.97	1.33		4.35%	-2.40 [ -2.66, -2.14]
Mierzecki A 2011	74	80.1	12.5	30	72	8	101	4.26%	0.71 [ 0.27, 1.14]
Dijkmans BA 1987	36	10.29	0.4	18	14.4	0.6		2.94%	-8.53 [-10.24, -6.83]
Cimmino MA 1986	33	14	3.2	85	13.9	3	-	4.28%	0.03 [ -0.37, 0.43]
Pandey SP 1985	28	72.53	7.75	31	105.13	11.39	H#H	3.99%	-3.27 [ -4.05, -2.49]
Kennedy AC 1975	113	85.7	19	100	99	11		4.34%	-0.84 [ -1.12, -0.56]
Hansson 1975	78	105.87	13.34	132	103.08	14.54		4.34%	0.20 [ -0.08, 0.48]
Onal S 2011	32	0.44	0.182	52	0.74	0.297	HH	4.24%	-1.15 [ -1.620.67]
Wang HJ 2021	102	81.68	14.14	66	87.06	14.76	×	4.33%	-0.37 [ -0.68, -0.06]
Random effects model							•	100.00%	-1.17 [ -1.680.66]

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Heterogeneity:df = 23,t\*2= 1.53,p = 0.00, I<sup>2</sup> = 97.15%

-10 -5 0 5

Standardized Mean Difference

#### B Copper level

	E	xperiment	tal		Control									
Study	Total	Mean	SD	Total	Mean	SD							Weight	SMD [95% CI]
Aaseth J 1978	23	28	5	30	14	3			÷	HH			3.48%	3.46 [ 2.61, 4.31]
Sahebari M 2015	110	990.75	239.54	100	1112.21	315.78			ĸ.				3.71%	-0.43 [-0.71, -0.16]
Afridi HI 2015	53	0.78	0.16	52	1.68	0.26	н	H	:				3.57%	-4.15 [-4.833.47]
Yazar M 2005	25	110.19	27.76	25	81.14	32.07			HH				3.61%	0.95 [ 0.37. 1.54]
Li J 2014	60	14.36	3.22	60	26.86	4.12		HHH	:				3.62%	-3.36 [-3.91, -2.80]
Bacon MC 1900	34	103.24	14.6	9	109	11		۲	•				3.54%	-0.40 [-1.14, 0.33]
Niedermeier W 1971	105	110.4	29.1	105	97	32.1			<u>ج</u>				3.71%	0.44 [ 0.16, 0.71]
Banford JC 1982	85	0.69	0.26	49	0.74	0.13			÷.				3.69%	-0.22 [-0.58, 0.13]
Morgenstern H 1983	30	106.3	37.6	30	102	31.8			нн				3.64%	0.12 [-0.38, 0.63]
Zoli A 1998	57	151.8	33.2	20	117.8	22.5			-				3.63%	1.09 [ 0.55, 1.63]
Tuncer S 1999	39	154.31	6.37	22	108.25	4.01			:				3.02%	8.06 [ 6.53, 9.58]
Wanchu A 2002	39	1426	324	22	946	446			1	1			3.62%	1.27 [ 0.70, 1.84]
Silverio Amancio OM 2003	41	143.5	35.7	23	126	25.9			HH				3.64%	0.53 [ 0.01, 1.05]
Ala S 2009	40	1.08	0.3	40	1	0.2			(m)				3.67%	0.31 [-0.13, 0.75]
Ullah Z 2016	61	130.3	42.8	61	102.7	22.5			1				3.69%	0.80 [ 0.43, 1.17]
Afridi HI 2013	202	1.07	0.18	203	1.44	0.31			1				3.72%	-1.46 [-1.68, -1.24]
Chakraborty M 2015	50	175.8	22.19	50	111.16	24.95			1	HH			3.63%	2.72 [ 2.17, 3.26]
Strecker D 2013	74	122.3	28.2	30	91.2	16.9			1				3.66%	1.21 [ 0.76, 1.67]
Louro MO 2000	40	153.9	5	95	104	1.8			1			<b></b>	2.69% 1	6.00 [14.05, 17.94]
Marrella M 1990	77	143.8	23.8	25	108.6	21.8			н	н			3.65%	1.50 [ 1.00, 1.99]
Youssef AA 1983	60	26.9	6.7	14	21.4	3.7			-				3.61%	0.87 [ 0.27, 1.47]
Scudder PR 1978	100	25.6	6.6	100	18.5	3.4			: <b>H</b>				3.70%	1.35 [ 1.04, 1.65]
Aiginger P 1978	12	193.3	22	7	130.3	12.8			: 1				3.14%	3.12 [ 1.76, 4.49]
McMurray W 1975	146	175.8	14.07	45	130.88	10.97			1	104			3.65%	3.34 [ 2.86, 3.81]
Scudder P 1976	50	13.66	2.38	50	14.42	1.35		1	-				3.68%	-0.39 [-0.79, 0.01]
Hansson L 1975	78	150.69	42.12	132	137.12	40.33			ж.				3.71%	0.33 [ 0.05, 0.61]
Onal S 2011	32	1.66	0.285	52	1.13	0.194			: '				3.62%	2.26 [ 1.70, 2.81]
Wang HJ 2021	102	18.11	4.24	66	14.96	2.45							3.70%	0.86 [ 0.54. 1.18]
Random effects model Heterogeneity:df = 27.t*2= 2	.53.p =	0.00; I <sup>2</sup> =	98.2%						٠				100.00%	1.24 [ 0.64, 1.85]
							Г		+		1	1		
							-5		0	5	10	15	20	

Standardized Mean Difference

Fig. 2 Forest plots of serum trace metal in RA patients compared with controls. A Zinc level; B Copper level

	RA		Control	Р				
	Remission $n = 45$	Active $n=57$	n=66	Remission vs active	Remission vs control	Active vs control		
Copper(mmol/L)	15.99±3.21	$19.79 \pm 4.22$	$14.96 \pm 2.5$	< 0.001	0.058	< 0.001		
Iron (µmol/L)	$7.14 \pm 0.96$	$6.42 \pm 1.03$	$8.03 \pm 1.44$	< 0.001	< 0.001	< 0.001		
Zinc (µmol/L)	$83.18 \pm 14.06$	$80.50 \pm 14.22$	$87.06 \pm 14.8$	0.346	0.168	0.014		

Table 2 Copper, iron, and zinc levels of patients with RA with different disease activities



Fig. 3 Comparison of mean blood copper, iron, and zinc between RA remission, RA activity, and control group. A Iron level; B Zinc level; C Copper level

 Table 3
 Multiple linear regression analysis for DAS28-CRP with the included parameters in patients with RA

Element	Variables	Coefficients	Р
DAS28-CRP	Hemoglobin	-0.313	0.001
DAS28-CRP	Copper	0.220	0.017
DAS28-CRP	Age	0.214	0.013
DAS28-CRP	RF-IgG	0.174	0.048

## Linear Association Between Levels of Trace Elements and Other Parameters and the Disease Activity in Patients with RA

The linear association between the level of trace elements and other parameters and the RA disease activity showed that hemoglobin levels (r = -0.41, P < 0.01) and blood iron (r = -0.37, P < 0.01) were negatively correlated with DAS28-CRP (Supplementary Table 2, Fig. 4A, B). Blood copper level was positively correlated with DAS28-CRP (r = 0.35, P < 0.01), CRP (r = 0.45, P < 0.01) and ESR (r = 0.58, P < 0.01) (Supplementary Table 2, Fig. 4C-E). Blood iron level was highly positively correlated with blood zinc (r = 0.56, P < 0.01) and hemoglobin (r = 0.70, P < 0.01) while negatively correlated with CRP (r = -0.46, P < 0.01) and ESR (r = -0.55, P < 0.01) (Supplementary Table 2 and Fig. 4F-I).

# Discussion

This study presents a comparison of trace element levels in healthy controls and patients with RA with different disease activities. We found higher copper concentrations and lower zinc concentrations in patients with RA compared to healthy controls, confirming the reports from other investigators [12, 13]. This result was also consistent with that of Xin et al., who conducted a meta-analysis suggested that increased serum levels of copper and decreased serum levels of zinc are generally present in patients with RA [14]. These changes may be related to the antagonistic relationship between copper and zinc; overdose of zinc may reduce copper absorption, while a high dose of copper may reduce zinc absorption [15].

ESR and CRP are widely used laboratory markers of systemic inflammation [16, 17]. The result show that blood circulating copper was positively correlated with CRP (r = 0.45, P < 0.01) and ESR (r = 0.58, P < 0.01), while blood circulating iron was negatively correlated with CRP (r = -0.46, P < 0.01) and ESR (r = -0.56, P < 0.01) in



Fig. 4 Linear association between related parameters in RA patients. A Linear relationship between DAS28-CRP and hemoglobin; B Linear relationship between DAS28-CRP and iron; C Linear relationship between DAS28-CRP and copper; D Linear relationship between

copper and CRP; **E** Linear relationship between copper and ESR; **F** Linear relationship between iron and zinc; **G** Linear relationship between hemoglobin and iron; **H** Linear relationship between CRP and iron; **I** Linear relationship between ESR and iron

RA. Copper and iron were associated with the inflammation of RA. However, ESR is under the influence of several factors, such as age, sex, anemia, and plasma proteins [18]. CRP has also been associated with the inflammation of comorbidities such as cardiovascular disease, diabetes, metabolic syndrome, pulmonary diseases, and depression [17]. DAS28-CRP and DAS28-ESR were calculated to assess the disease activity of RA [19]. The two DAS28 variants are not interchangeable especially for low activity of RA [20]. In view of CRP is more sensitive to short-time changes in inflammation [21], DAS28-CRP was used in this study.

Copper level was higher in patients with active RA compared to patients in remission and was positively associated with DAS28-CRP, a score used to evaluate the disease activity of RA. A corresponding study also reported that serum copper concentrations were positively related to disease activity, as assessed by the disease activity score [15]. Chakraborty et al. found a significant positive correlation between serum copper level, erythrocyte sedimentation rate, and morning stiffness in patients with RA [7]. Thus, serum copper level may be considered a potential biomarker of disease activity in RA. Generally, patients with RA have a high blood copper level. Penicillamine, a heavy metal (copper) chelator, is used for the treatment of RA in our hospital. It was thought that D-penicillamine may inhibit cell growth in a variety of cell types by chelating copper ions. Harada et al. suggested that D-penicillamine might help in the regression of rheumatoid synovial hyperplasia via Fas-mediated apoptosis [22].

This study showed that the blood zinc level in patients with RA, particularly those with active RA, was statistically higher than that of controls. However, there were no statistical differences between the zinc levels of patients with active RA and patients in remission, as well as between patients in remission and controls. There was no correlation between blood zinc levels and RA disease activity (r = -0.092, P > 0.05). This result was similar to that of a previous study conducted on juvenile idiopathic arthritis (JIA), a heterogeneous group of autoimmune diseases that arise before the age of 16 years. Hala Salah El Din Talaat et al. measured the serum levels of zinc in patients with JIA of different subtypes. They found that the mean serum zinc level in patients with JIA was significantly higher than that in the controls (P=0.022). There was no statistically significant difference between the mean serum zinc levels in the JIA and control groups (P = 0.693) [23].

Chronic zinc deficiency in a chick model triggered a decrease in gene expression of pro-inflammatory cytokines (IL-1 $\beta$ , IL-6, TNF) [24]. These cytokines are vital for the pathogenesis of RA. Zinc intake increased plasma zinc concentrations and decreased CRP and IL-6 levels, when compared with a placebo [25]. Zinc plays an important role in the immune system by affects both innate and adaptive immune cells [3], although there is no strong correlation between blood zinc levels and RA disease activity.

Our study revealed that blood iron levels in patients with RA were significantly lower than those in controls, and it was negatively correlated with disease activity, which was consistent with the findings of Stefanova et al. [26]. Iron is an important trace element for the synthesis of hemoglobin, and iron deficiency can cause anemia. This study showed that blood iron levels were highly positively correlated with hemoglobin levels. Anemia in RA may be caused by the overexpression of hepcidin, which leads to local deposition of iron resulting in a decrease in iron content in peripheral blood. Studies have shown that iron can promote the production of pro-inflammatory cytokines by T cells and, thus, directly participate in the inflammatory response. Iron deposition has been observed in the central nervous system of patients with inflammatory diseases, such as multiple sclerosis, Alzheimer's disease, and Parkinson's disease, and in the synovial fluid of patients with RA[6, 27].

Based on the multiple linear regression analysis of the relationship between disease activity and trace element levels, and other clinical parameters, DAS28-CRP was found to be correlated with age, hemoglobin, copper, and RF-IgG levels. Except for hemoglobin, which had a negative correlation, the other three parameters all had a positive correlation with DAS28-CRP. This suggests that lower hemoglobin levels, older age, higher copper, and RF-IgG levels increase the risk of high disease activity in patients with RA. This may explain why RF-IgG can predict RA-associated consequences [28]. According to Elshafie et al. [29], RF-IgG was the strongest marker for poor prognosis because it was associated with a higher total number of affected joints. RF-IgG was associated with young age (p=0.0005) and a lower age of disease onset (P < 0.0001), which was not found in our study.

Our study had some limitations, including the small number of subjects involved in the study and the lack of analyses of the relationship between trace elements, cytokines, and antioxidant enzymes. Further studies should focus on overcoming these limitations.

In conclusion, we found that the zinc, copper, and iron levels in the blood differed significantly in patients with RA and healthy individuals. Blood levels of iron and copper were markedly associated with RA disease activity.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s12011-022-03148-z.

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#### Declarations

**Ethics Approval** This study was approved by the Ethics Committee of Guanghua Hospital of Integrated Traditional Chinese and Western Medicine (approval number: 2018-K-12) and written consent was collected prior to blood drawing of participants.

Conflict of Interest The authors declare no competing interests.

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# References

- Guo S, Xu L, Chang C, Zhang R, Jin Y, He D (2020) Epigenetic regulation mediated by methylation in the pathogenesis and precision medicine of rheumatoid arthritis. Front Genet 11:811
- Li J, Liang Y, Mao H, Deng W, Zhang J (2014) Effects of B-lymphocyte dysfunction on the serum copper, selenium and zinc levels of rheumatoid arthritis patients. Pak J Med Sci 30:1064–1067
- 3. Maywald M, Wang F, Rink L (2018) The intracellular free zinc level is vital for treg function and a feasible tool to discriminate between treg and activated Th cells. Int J Mol Sci 19:3575
- Gao H, Dai W, Zhao L, Min J, Wang F (2018) The role of zinc and zinc homeostasis in macrophage function. J Immunol Res 2018:1–11
- Li S, Yin H, Zhang K, Wang T, Yang Y, Liu X et al (2017) Effector T helper cell populations are elevated in the bone marrow of rheumatoid arthritis patients and correlate with disease severity. Sci Rep 7:4776
- 6. Khalaf W, Al-Rubaie HA, Shihab S (2019) Studying anemia of chronic disease and iron deficiency in patients with rheumatoid arthritis by iron status and circulating hepcidin. Hematol Rep 11:16–19
- Chakraborty M (2015) Serum copper as a marker of disease activity in rheumatoid arthritis. J Clin Diagn Res 9:BC09
- Ala S, Shokrzadeh M, Pur Shoja AM, Saeedi Saravi SS (2009) Zinc and copper plasma concentrations in rheumatoid arthritis patients from a selected population in Iran. Pak J Biol Sci PJBS Pakistan 12:1041–1044
- Guo S, Jin Y, Zhou J, Zhu Q, Jiang T, Bian Y et al (2021) Micro-RNA variants and HLA-miRNA interactions are novel rheumatoid arthritis susceptibility factors. Front Genet 12:747274
- Aletaha D, Neogi T, Silman AJ, Funovits J, Felson DT, Bingham CO et al (2010) 2010 Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. Arthritis Rheum 62:2569–2581
- Egger M, Davey Smith G, Schneider M, Minder C (1997) Bias in meta-analysis detected by a simple, graphical test. BMJ 315:629–634
- Önal S, Nazıroğlu M, Çolak M, Bulut V, Flores-Arce MF (2011) Effects of different medical treatments on serum copper, selenium and zinc levels in patients with rheumatoid arthritis. Biol Trace Elem Res United States 142:447–455
- Strecker D, Mierzecki A, Radomska K (2013) Copper levels in patients with rheumatoid arthritis. Ann Agric Environ Med AAEM Poland 20:312–316
- Xin L, Yang X, Cai G, Fan D, Xia Q, Liu L et al (2015) Serum levels of copper and zinc in patients with rheumatoid arthritis: a meta-analysis. Biol Trace Elem Res United States 168:1–10
- Sahebari M, Ayati R, Mirzaei H, Sahebkar A, Hejazi S, Saghafi M et al (2016) Serum trace element concentrations in rheumatoid arthritis. Biol Trace Elem Res United States 171:237–245
- Bray C, Bell LN, Liang H, Haykal R, Kaiksow F, Mazza JJ et al (2016) Erythrocyte sedimentation rate and C-reactive protein measurements and their relevance in clinical medicine. WMJ 115:6

- Pope JE, Choy EH (2021) C-reactive protein and implications in rheumatoid arthritis and associated comorbidities. Semin Arthritis Rheum 51:219–229
- Fransen J, Welsing PM, Keijzer RMD, Riel PLV (2004) Disease activity scores using C-reactive protein: CRP may replace ESR in the assessment of RA disease activity. Ann Rheum Dis 62:130–139
- Prevoo MML, Van Thof MA, Kuper HH, Van Leeuwen MA, Van De Putte LBA, Riel PLCM (1995) Modified disease activity scores that include twenty eight joint counts development and validation in a prospective longitudinal study of patients with rheumatoid arthritis modified disease activity score. Arthritis Rheum 38:44–8
- 20. Shivacheva TK (2020) DAS28-ESR and DAS28-CRP are they interchangeable in measuring the activity of rheumatoid arthritis in response to treatment with biological agents? Folia Med (Plovdiv). Bulgaria 62:46–51
- Sengul I, Akcay-Yalbuzdag S, Ince B, Goksel-Karatepe A, Kaya T (2015) Comparison of the DAS28-CRP and DAS28-ESR in patients with rheumatoid arthritis. Int J Rheum Dis England 18:640–645
- 22. Harada S, Sugiyama E, Taki H, Shinoda K, Fujita T, Maruyama M et al (2002) D-penicillamine cooperates with copper sulfate to enhance the surface expression of functional Fas antigen in rheumatoid synovial fibroblasts via the generation of hydrogen peroxide. Clin Exp Rheumatol Italy 20:469–476
- Talaat HSED, Mohamed MF, Ahmed AI, Shafie ES. (2019) Serum levels of zinc and copper in cases of juvenile idiopathic arthritis. J Adv Med Med Res. 1–8
- Bañuls-Mirete M, Ogdie A, Guma M (2020) Micronutrients: essential treatment for inflammatory arthritis? Curr Rheumatol Rep 22:87
- 25. Bao B, Prasad AS, Beck FW, Fitzgerald JT, Snell D, Bao GW et al (2010) Zinc decreases C-reactive protein, lipid peroxidation, and inflammatory cytokines in elderly subjects: a potential implication of zinc as an atheroprotective agent. Am J Clin Nutr 91:1634–1641
- 26. Stefanova KI, Delcheva GT, Maneva AI, Batalov AZ, Geneva-Popova MG, Karalilova RV et al (2018) Pathobiochemical mechanisms relating iron homeostasis with parameters of inflammatory activity and autoimmune disorders in rheumatoid arthritis. Folia Med (Plovdiv). Bulgaria 60:124–32
- 27. Wang Z, Yin W, Zhu L, Li J, Yao Y, Chen F et al (2018) Iron drives T helper cell pathogenicity by promoting RNA-binding protein PCBP1-mediated proinflammatory cytokine production. Immunity United States 49:80-92.e7
- 28. Shakiba Y, Koopah S, Jamshidi AR, Amirzargar AA, Masoud A, Kiani A et al (2014) Anti-cyclic citrullinated peptide antibody and rheumatoid factor isotypes in Iranian patients with rheumatoid arthritis: evaluation of clinical value and association with disease activity. Iran J Allergy Asthma Immunol 13:10
- 29. Ossipova E, Oliynyk G, Cerqueira C, Becker S, Ytterberg J, Auer G et al (2013) Identification of novel targets in rheumatoid arthritis synovial tissues using gel electrophoresis and mass spectrometry. Ann Rheum Dis 72:A77.1-A77

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