



Selection and characterization of lead-tolerant sweetpotato cultivars for phytoremediation

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Abstract

Lead (Pb) is one of the most toxic heavy metals (HMs) for plants and the environment. Sweetpotato [*Ipomoea batatas* (L.) Lam], the sixth most important food crop in the world, is tolerant to various environmental stresses, owing to its high antioxidant capacity. In this study, we selected sweetpotato cultivars showing high tolerance to lead (Pb) for phytoremediation-related applications. Young seedlings of 20 sweetpotato cultivars were treated with 30 mM Pb. Daeyumi (KO-12) and Dahomi (KO-5) were selected as Pb-tolerant and -sensitive cultivars, respectively, based on their photosynthetic activity and growth inhibition index (I_{50}). In the Pb treatment, hydrogen peroxide and malondialdehyde contents of KO-12 were 1.5-fold less than those of KO-5. In addition, KO-12 showed a higher ability to accumulate Pb in roots and leaves than KO-5. Expression levels of four Pb-responsive genes, including the metallothionein gene *IbMT1*, were higher in the roots and leaves of KO-12 than in those of KO-5. Interestingly, KO-12 showed greater tolerance to high Pb concentrations than sunflower and rapeseed, which have been well-studied for phytoremediation. Our results suggest that sweetpotato is a suitable biomaterial for the phytoremediation of soils contaminated with HMs, including lead, for sustainable agriculture.

Keywords Lead (Pb) · Sweetpotato · Phytoremediation · Heavy metals · Gene expression

Abbreviations

HMs	Heavy metals
MDA	Malondialdehyde
qRT-PCR	Quantitative real-time polymerase chain reaction
ROS	Reactive oxygen species
TBA	Thiobarbituric acid
TCA	Trichloroacetic acid
MTs	Metallothioneins

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Introduction

Heavy metal (HM) pollution is one of the major environmental problems worldwide (Mitra et al. 2022), and lead (Pb) is one of the most dangerous HMs. Pb pollution of air, water, and agricultural fields has adverse impacts on human health and the environment (Kshyanaprava and Das 2023; Sevak et al. 2021). The main sources of Pb-based environmental pollution include Pb ore mining and smelting, industrial effluents, fertilizers, pesticides, and municipal sewage sludge (Kushwaha et al. 2018).

Several methods are currently available for cleaning the HM-contaminated soils and can be divided mainly into two categories: physicochemical and biological (Zhambakin and Zhapar 2020). The use of physicochemical methods of soil cleaning involves washing and chemical oxidation and reduction, which often leads to the accumulation of secondary pollutants and requires additional manipulations associated with the removal of contaminated soil cover and subsequent waste collection. The biological methods of soil decontamination involve phytoremediation, i.e., remediation of soil using live plants (Daurov et al. 2023). Phytoremediation is a promising technology that interacts harmoniously with the ecosystem (Mocek-Płóćiniak et al. 2023), does not require expensive equipment, and is suitable for the elimination of a wide range of environmental pollutants. Moreover, the use of phytoremediation does not lead to the loss of soil fertility. Hyperaccumulator plants represent a valuable resource for the remediation of metal-contaminated sites as they can carry, absorb, and translocate high levels of HMs that would be toxic to most organisms.

In plants, Pb affects several metabolic processes in various subcellular compartments. Pb toxicity decreases seed germination, plant growth, and root and shoots dry biomass, violates mineral nutrition (Sharma and Dubey 2005), reduces cell division, and inhibits photosynthesis (Nas and Ali 2018). The study of plant responses to external factors reveals the physiological and biochemical mechanisms and adaptation limits of plants (Sapakhova et al. 2023). In addition, Pb has been reported to produce reactive oxygen species (ROS) and increase antioxidant enzyme activity in plants (Samuel et al. 2022). The ROS generated as a result of oxidative stress have various harmful effects on plant cells, such as inhibition of photosynthetic activity, inhibition of adenosine triphosphate (ATP) production, and lipid peroxidation (Pourrut et al. 2011). One of the main consequences of accumulating Pb is the increased production of ROS, which damages cell membranes, nucleic acids, and chloroplast pigments (Afaj et al. 2017; Nas and Ali 2018). Moreover, key members of the peroxidase (POD) family play a critical role in the enzymatic removal of ROS, thus protecting plant cells from oxidative stress (Gülen et al. 2008). Pb toxicity accelerates the production of ROS including hydrogen peroxide (H₂O₂), which, in turn, causes secondary oxidative stress, leading to growth inhibition at the seedling stage (Yang et al. 2010). When plants encounter extreme levels of environmental stress, their malondialdehyde (MDA) levels increase (depending on the nature of the stress). The content of MDA represents the level of lipid peroxidation, and an elevated MDA level indicates stress. In rice, high concentrations of Pb significantly reduced plant yield, adversely affected various biochemical and physiological processes (Xie et al. 2018), and increased the antioxidant level (Mandiwana et al. 2007). These studies show that

strengthening the antioxidant defense system helps plants to overcome HM-induced stress.

The study of gene expression in plants is receiving increasing attention from researchers around the world (Hieno et al. 2019). As with other stress factors, numerous protective genes are expressed when exposed to heavy metals. Abiotic factors, including heavy metals, are the most common stress factors that negatively affect plant growth and development. Plants respond to stress factors at the molecular, cellular and physiological levels through various perceptions and transduction of stress signals, which subsequently activate a variety of defense mechanisms (Rejeb et al. 2014). Recently, there has been great interest in studying the expression of the metallothionein (MT) gene, which plays an important role in heavy metal tolerance in plants (Kim et al. 2010, 2014; Sung et al. 2019). MTs, low-molecular-weight proteins (7–10 kDa) with a high percentage of cysteine (Cys) residues (Bourdineaud et al. 2006; Cobbett and Goldsbrough 2002; Pan et al. 2018), have been widely characterized in various prokaryotic and eukaryotic organisms. Plant MTs are classified into four types according to the arrangement of their Cys residues (Robinson et al. 1993), including the *MT1*, *MT2*, *MT3*, and *MT4* subfamilies (Cobbett and Goldsbrough 2002; Leszczyszyn et al. 2013). MTs play crucial roles in ion homeostasis and tolerance in plants. The *HMA3* gene plays an important role in the transport and regulation of various metals, including lead, zinc, cobalt and other heavy metals (Takahashi et al. 2012; Williams and Mills 2005). Heavy-metal-associated proteins (HMAs) have been extensively studied at the molecular level in *Arabidopsis thaliana* and *Oryza sativa*, where eight and nine *HMA* genes (*AtHMAs* and *OsHMAs*) were discovered, respectively (Takahashi et al. 2012; Williams and Mills 2005). There is also widespread interest in studying the expression of genes encoding peroxidase (*swpb3* and *swpa4*) (Jang et al. 2004; Kim et al. 2010), which have a key role in the oxidation of molecules when exposed to heavy metals in plants.

Sweetpotato [*Ipomoea batatas* (L.) Lam] is an emerging root crop with the potential to ensure global food and nutrition security in the face of the climate crisis (Kwak 2019). Sweetpotato was evaluated as one of 10 superfoods for better health since it contains high levels of low-molecular-weight antioxidants, dietary fiber, minerals including potassium as well as large amounts of carbohydrates (Alam et al. 2020; CSPI 2007; Sun et al. 2019). In the field, sweetpotato plants show high tolerance to environmental stresses including drought and high temperature, owing to their high antioxidant activity (Chauhan et al. 2021; Chen et al. 2016; Wang et al. 2019; Yang et al. 2020). In addition, sweetpotato serves as a cover crop and root crop, reducing soil erosion in the rainy season. Thus, sweetpotato has many characteristics suitable for the phytoremediation of HM-contaminated soils.

Few studies have been conducted to study the phytoremediation capacity of sweetpotatoes on HM-contaminated soil (Izinyon and Seghosime 2013; Kim et al. 2010, 2014; Tamungang et al. 2016). In addition, the antioxidant protection of sweetpotatoes against the toxic effects of ROS under Pb stress has not been sufficiently explained. Thus, this study aimed to investigate the potential of sweetpotatoes as a remediation plant for Pb-contaminated soil.

Materials and methods

Plant materials and growth conditions

Twenty Korean sweetpotato cultivars were used in this study: Daeyumi (KO-12), Dahomi (KO-5), Danjami, Geonhwangmi (KO-7), Hogammi, Jammi, Jeonmi, Jinhongmi, Jinmi, Jinyulmi, Matnami (KO-16), Pungwonmi, Saengmi, Shingunmi, Shinzami, Shinyulmi, Yesmi, Yeonhwangmi, Yeonmi, and Yulmi. These cultivars were obtained from the Bioenergy Crop Research Institute, National Institute of Crop Science, Rural Development Administration, Muan, Korea. Each plant was placed in a square tray (40 cm × 16 cm × 13 cm) and grown at 25 °C under 16 h/8 h light/dark cycle and 50% relative humidity. Stem cuttings were prepared from young seedlings as described previously (Park et al. 2020).

Pb treatments

To select sweetpotato cultivars with high tolerance to Pb, 10-day-old seedlings of 20 sweetpotato cultivars were treated with 300 mL of water containing 30 mM Pb(NO₃)₂ for 2 weeks. Plant phenotypes of each sweetpotato cultivar were evaluated and scored on a scale of 0 (complete damage) to 10 (no damage), according to leaf damage by Pb stress. To compare the 50% inhibition (I_{50}) concentrations of Pb between two Pb-tolerant cultivars (KO-12 and KO-16) and two Pb-sensitive cultivars (KO-7 and KO-5), 14-day-old seedlings were treated with different concentrations of Pb (0, 1, 5, 10, 15, and 30 mM) for 30 days. After 30 days of Pb treatment, the leaves and roots of plants were separated and weighed to determine their fresh weight. To compare the phytoremediation capacity of sweetpotato with that of other crops, young plants of sunflower and rapeseed cultivars were treated with Pb (as described above) for 2 weeks. All experiments were carried out in a randomized complete block design with four replications.

Photosynthetic activity and chlorophyll content measurements

Chlorophyll contents and photosynthetic activity were measured in the second and third leaves (from the top) of

two Pb-tolerant and two Pb-sensitive cultivars at 2 weeks after Pb treatments (0, 1, 5, 10 mM). To measure photosynthetic activity, the photochemical yield (Fv/Fm) of leaves was determined using a portable chlorophyll fluorometer (HANSATECH, Serial Number 1947, England) after 30 min of dark adaptation. The chlorophyll content of leaves was measured using a portable chlorophyll meter (Junior-PAM Chlorophyll Fluorometer; Heinz Walz, Germany).

MDA and H₂O₂ quantification

The leaves and roots of Pb-treated plants were separated and immediately frozen in liquid nitrogen. The content of MDA, which indicates the level of lipid peroxidation, was measured using the modified thiobarbituric acid (TBA) method (Wang et al. 2009). Root extracts were assayed with 20% trichloroacetic acid (TCA) and 0.5% TBA, and the absorbance of extracts was quantified at 532 and 600 nm. The H₂O₂ content was measured using the potassium iodide method (Junglee et al. 2014), and the absorbance of extracts was recorded at 390 nm. Samples were extracted using 0.1% TCA extraction buffer in both assays. The samples were homogenized with extraction buffer and centrifuged for 5 min.

Quantification of Pb in plant tissues

To measure the content of Pb content in plant tissues, young seedlings of KO-12 (Pb tolerant) and KO-5 (Pb sensitive) cultivars were treated with 10 mM Pb for 10 days. Leaves and roots of the harvested plants were dried in an oven at 60 °C. First, 25 mL of HNO₃ was added to 0.1 g (dry weight) of the plant sample. The sample was left stationary for 24 h and then decomposed by heating at 120 °C for 2 h using a heating block. Pb content was determined using a flame atomic absorption spectrophotometer, equipped with a hollow cathode lamp and an air-acetylene flame (AnalytikJena AG, novAA 350, Jena, Germany), at 283.3 nm wavelength (Toishimanov et al. 2023).

Gene expression analysis

The leaves and roots of KO-12 and KO-5 plants treated with 20 mM Pb were harvested at 0.5, 1, and 3 days after treatment (DAT). Total RNA was extracted from the leaf and root samples using TRI Reagent (Molecular Research Center, USA, TR 118), according to the manufacturer's protocol. To remove genomic DNA contamination, the total RNA was treated with RNase-free DNase I (Thermo Fisher Scientific, USA). The purity and quantity of DNase I-treated total RNA were confirmed using a NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific, USA). First-strand cDNA synthesis was conducted in a 13-μL reaction mixture containing 0.1 ng to 5 μg of total RNA and 0.6 μg (100 pmol) of

oligo(dT)₁₈ primer (Thermo Scientific RevertAid Reverse Transcriptase), according to the manufacturer's instructions. The mixture was incubated at 65 °C for 5 min. Then, a 7- μ L reaction mixture was prepared that contained 4 μ L of 5 \times reverse transcription reaction buffer, 2 μ L of 10 mM dNTP mix, and 1 μ L (200 units) of Moloney Murine Leukemia Virus (M-MuLV) reverse transcriptase. The mixture was incubated at 42 °C for 60 min, followed by a reaction stop by heating the mixture at 70 °C for 10 min.

The expression levels of four Pb-responsive genes encoding metallothionein (*IbMT1*), ATPase 3 homolog (*IbHMA3*), and two PODs (*swpb3* and *swpa4*) (Kim et al. 2010, 2014) by measured by quantitative real-time PCR (qRT-PCR). The qRT-PCR was performed on the CFX Connect™ Real-Time System (Bio-Rad, Hercules, CA, USA) using EvaGreen fluorescent dye and gene-specific primers (Supplemental Table 1) under the following conditions: initial denaturation at 95 °C for 15 min, followed by 40 cycles of denaturation at 95 °C for 20 s, annealing at 58 °C for 40 s, and extension at 72 °C for 20 s. The melting curves were verified to confirm primer specificity. The ubiquitin (*UBI*) gene was used as an internal reference, and relative gene expression was quantified using the $2^{-\Delta\Delta CT}$ method.

Statistical analysis

One-way analysis of variance (ANOVA) was conducted to identify statistically significant differences ($p < 0.05$). Tukey's honestly significant difference (HSD) post-hoc test was used to perform multiple pairwise comparisons. Principal component analysis (PCA) was completed using the *fviz-pca* function of the *factoextra* R package Ver. 1.0.7 in RStudio. Pearson's correlation analysis was completed using the *mcov* function, and *corrplots* were prepared using the *corrplot* package Ver. 0.92 in RStudio.

Results

Selection of Pb-tolerant and -sensitive sweetpotato cultivars

To select sweetpotato cultivars with high Pb tolerance, the appropriate Pb concentration was determined using the Yulmi cultivar (KO-1). Ten-day-old seedlings were treated with 300 mL of water containing different concentrations of Pb (0, 20, 30, 50, and 100 mM Pb(NO₃)₂) for 2 weeks. The sweetpotato seedlings showed leaf damage in a Pb dosage-dependent manner (Fig. 1). No obvious damage to leaves was observed in the 20-mM Pb treatment for 2 weeks, but clear leaf damage was evident in the 30-mM Pb treatment after 1 week. Seedlings treated with high Pb concentrations (50 and 100 mM) showed severe damage

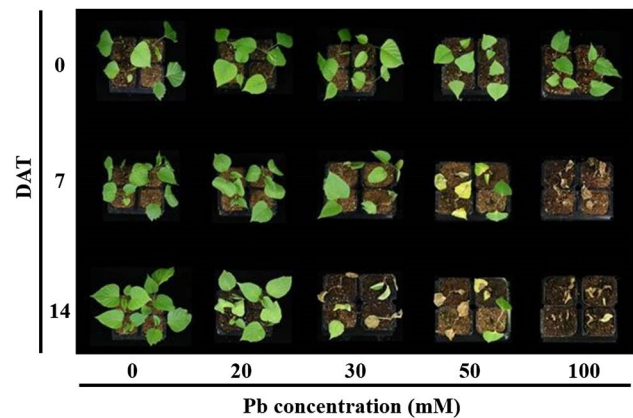


Fig. 1 Phenotypic evaluation of the sweetpotato cultivar Yulmi treated with various Pb concentrations for 14 days. DAT days after treatment

Table 1 Evaluation of the Pb tolerance of 20 sweetpotato cultivars based on leaf damage

Cultivar	Score [†]	Cultivar	Score [†]
KO-1 (Yulmi)	1	KO-19 (Saengmi)	7
KO-3 (Shinjami)	8	KO-20 (Shingunmi)	3
KO-4 (Pungwonmi)	8	KO-21 (Yeonmi)	5
KO-5 (Dahomi)	1	KO-22 (Yesmi)	2
KO-7 (Geonhwangmi)	1	KO-23 (Jammi)	5
KO-10 (Shinyulmi)	2	KO-24 (Jeonmi)	7
KO-11 (Yeonhwangmi)	7	KO-27 (Jinmi)	8
KO-12 (Daeyumi)	10	KO-28 (Jinyulmi)	3
KO-15 (Danjami)	3	KO-29 (Jinhongmi)	5
KO-16 (Matnami)	9	KO-32 (Hogammi)	6

[†]Leaf damage caused by 14-day exposure to 30 mM Pb was scored on a scale from 0 (complete damage) to 10 (no damage)

from one week onward. Thus, we determined 30 mM as the optimal Pb concentration for evaluating the Pb tolerance of 20 sweetpotato cultivars.

To select sweetpotato cultivars with high tolerance to Pb, 10-day-old seedlings of 20 cultivars were treated with 30 mM of Pb(NO₃)₂ for 14 days. Even though seedlings of all cultivars survived in the 30-mM Pb treatment, differences in leaf phenotypes were observed among the 20 cultivars. The Pb-induced leaf damage was scored on a scale of 0–10 for each cultivar (Table 1). Daeyumi (KO-12) was the most Pb-tolerant cultivar. Matnami (KO-16), KO-4 (Pungwonmi), and KO-27 (Jinmi) also showed high Pb tolerance. However, Dahomi (KO-5), and Geonhwangmi (KO-7) were the most sensitive to Pb. Sensitive cultivars clearly showed leaf chlorosis, stunting, and yellowing (data not shown). Thus, based on plant phenotypes, we tentatively determined KO-12 and KO-16 as

two Pb-tolerant cultivars and KO-7 and KO-5 as two Pb-sensitive cultivars and characterized them further.

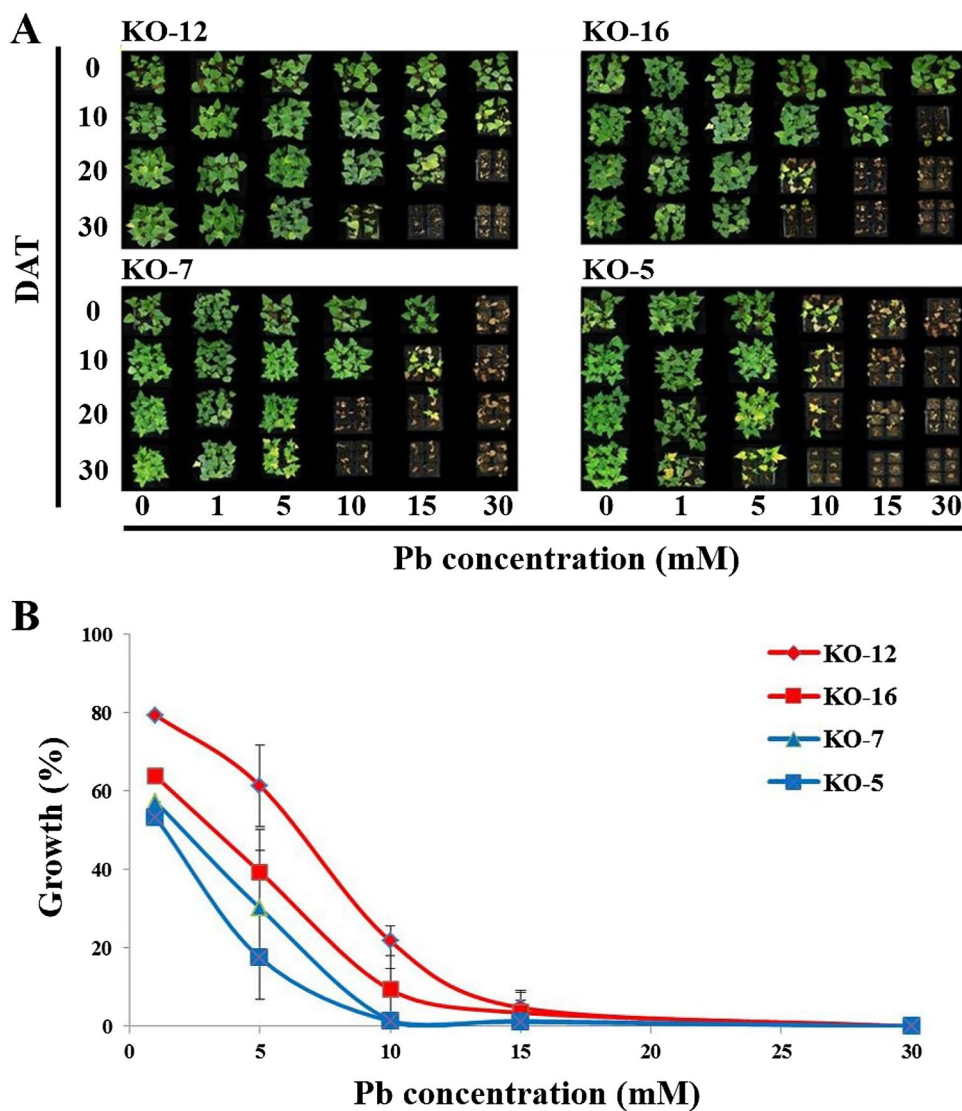
To confirm the phenotypic responses of Pb-tolerant (KO-12 and KO-16) and Pb-sensitive (KO-7 and KO-5) cultivars to Pb exposure, young seedlings were treated with various Pb concentrations (0, 1, 5, 10, 15, and 30 mM) for 30 days. All four cultivars showed clear phenotypic differences at different Pb concentrations (Fig. 2A). The severity of leaf damage in each cultivar and the degree of phenotypic differences among cultivars increased with the increase in Pb concentration and treatment time. KO-12 seedlings appeared quite healthy on day 5 in the 30-mM Pb treatment, whereas KO-5 seedlings showed severe leaf damage on day 1 in the 20-mM Pb treatment. Plant growth was analyzed by measuring the leaf fresh weight of all four cultivars treated with Pb for 30 days (Fig. 2B). The I_{50} values, calculated on the basis of leaf fresh weight, were 6.7, 3.2, 2.0, and 1.3 mM for KO-12, KO-16, KO-7, and KO-5, respectively. The I_{50} value

of KO-12 (6.7 mM) was 5.1 times higher than that of KO-5 (1.3 mM). Therefore, based on the I_{50} value, we designated KO-12 as the most Pb-tolerant sweetpotato cultivar.

Chlorophyll content and photosynthetic activity

To compare the physiological responses of Pb-tolerant (KO-12 and KO-16) and Pb-sensitive (KO-7 and KO-5) cultivars, the chlorophyll content and photosynthetic activity of plants treated with Pb (0, 1, 5, 10 mM) for 15 days were analyzed (Fig. 3A, B). Both the chlorophyll content and photosynthetic activity of sweetpotato cultivars decreased significantly with the increase in Pb concentration. KO-12 (the most Pb-tolerant cultivar) showed a stable chlorophyll content. However, in the 10-mM Pb treatment, chlorophyll contents of KO-16, KO-7, and KO-5 were 1.3-, 1.4-, and 1.7-fold lower than that of KO-12, respectively. Additionally, the photosynthetic activity (Fv/Fm) of KO-12 was 1.2-,

Fig. 2 Phenotypic and growth analyses of two Pb-tolerant and two Pb-sensitive sweetpotato cultivars treated with various concentrations of Pb for 30 days. **A** Plant phenotypes. **B** Relative plant growth determined on the basis of leaf fresh weight (%). Data represent mean \pm SD of four independent replicates. Red line, Pb-tolerant cultivars; blue line, Pb-sensitive cultivars; *DAT* days after treatment



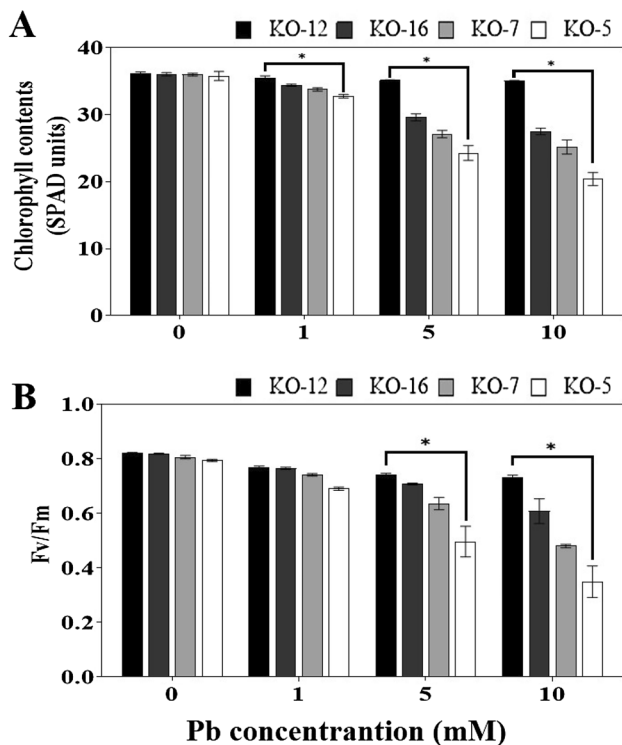


Fig. 3 Physiological analysis of four sweetpotato cultivars treated with different concentrations of Pb stress for 15 days. **A** Chlorophyll contents of leaves (SPAD unit); **B** photosystem II efficiency (Fv/Fm). Data represent mean \pm SD of four independent replicates. Asterisks indicate significant differences among cultivars ($*p < 0.05$; one-way analysis of variance [ANOVA] with Tukey's HSD post-hoc test)

1.5-, and 2.1-fold times higher than that of KO-16, KO-7, and KO-5, respectively, in the 10-mM Pb treatment. These results confirmed that KO-12 exhibits higher chlorophyll content and greater photosynthetic activity than the other sweetpotato cultivars, owing to its high Pb tolerance.

MDA and H₂O₂ contents

In the 10-mM Pb treatment, the MDA content of roots started to show a significant difference between KO-12 and KO-5 cultivars from day 1 (Fig. 4). At 10 DAT, the MDA content of KO-5 roots was approximately 1.4-fold higher than that of KO-12 roots. Similarly, the MDA content of leaves started showing a significant difference between KO-12 and KO-5 cultivars from day 5 onward. At 10 DAT, the MDA content of KO-5 leaves was approximately 1.3-fold higher than that of KO-12 leaves.

The H₂O₂ content of KO-5 and KO-12 roots sharply increased from day 3 onward. At 5 and 10 DAT, the H₂O₂ content of KO-5 leaves was approximately 1.2- and 1.5-fold higher than that of KO-12 leaves, respectively. These results indicate that KO-12 exhibits high antioxidant activity to cope with Pb-induced oxidative stress.

Pb accumulation in different tissues

Figure 5 shows that sweetpotato plants accumulate Pb in both roots and leaves. The Pb content of roots of both KO-12 and KO-5 cultivars started increasing from day 1 onward. At 10 DAT, Pb content of KO-12 roots (4600 mg/kg) was approximately 1.2-fold higher than that of KO-5 roots (3797 mg/kg). Once absorbed by the roots, Pb is translocated to leaf tissues. The leaves of KO-12 also accumulated high levels of Pb compared with those of KO-5; at 10 DAT, the level of Pb accumulation in KO-12 leaves (507 mg/kg) was approximately 2.0-fold higher than that in KO-5 leaves (251 mg/kg). These results indicate that the Pb-tolerant KO-12 cultivar can efficiently absorb Pb through the roots and translocate it to leaf tissues.

Expression analysis of Pb-responsive genes

To understand the differences between the Pb-tolerant KO-12 and Pb-sensitive KO-5 cultivars at the molecular level, we examined the expression of four genes involved in HM tolerance in sweetpotato: *IbMT1* (encoding metallothionein), *IbHMA3* (encoding ATPase3 homolog), and *swpb3* and *swpa4* (encoding PODs). All four genes showed differences in expression patterns between the two cultivars (Fig. 6). Pb-tolerant KO-12 showed higher expression levels of the four genes in both leaves and roots than Pb-sensitive KO-5. The expression of *IbMT1* started increasing in both the leaves and roots of KO-12 at 12 h after the Pb treatment. Compared with the KO-5 cultivar, KO-12 showed a significantly higher expression level of *IbHMA3* at 1 DAT in leaves and at 0.5 DAT in roots. The expression levels of *swpb3* and *swpa4* were higher in KO-12 than in KO-5. The *swpb3* gene was highly expressed in the leaves of both KO-12 and KO-5 cultivars at 0.5 DAT and thereafter; however, in the roots of both cultivars, *swpb3* showed a similar level regardless of the Pb treatment duration. The *swpb4* gene showed higher expression in both the leaves and roots of KO-12 than in those of KO-5 starting at 0.5 DAT. In the leaves and roots of the KO-12 cultivar, *swpb4* showed the highest expression level at 1 and 3 DAT, respectively. Our results confirmed that the Pb-tolerant KO-12 cultivar has a Pb tolerance mechanism at the molecular level.

Principal component analysis and correlation analysis

To assess the impact of Pb treatment on tolerant and sensitive cultivars of sweetpotato, PCA was used for multidimensional analysis. The variables that exist closely and in the same quadrant are positively correlated. The first two components, such as PC1 (87.8%) and PC2 (8.4%), made the most significant contribution, constituting 96.2% of

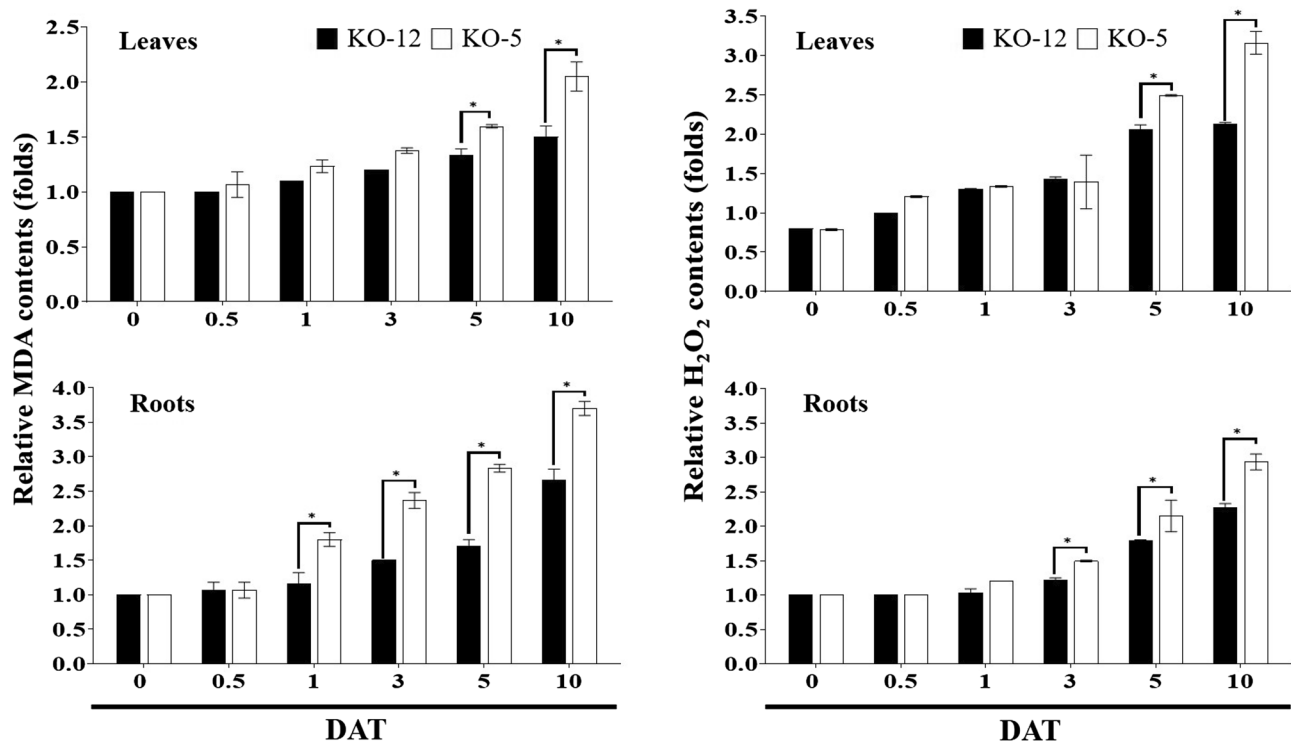


Fig. 4 Relative malondialdehyde (MDA) and hydrogen peroxide (H₂O₂) contents of the leaves and roots of KO-12 (Pb-tolerant) and KO-5 (Pb-sensitive) cultivars. Data represent mean \pm SD of four inde-

pendent replicates. Asterisks indicate significant differences among cultivars (* p < 0.05; ANOVA with Tukey's HSD post-hoc test). DAT Days after treatment

the total variance among physiological and biochemical parameters, thereby confirming the trend of the results (Supplemental Fig. 1A). This segregation of treatments indicates that Pb treatment had a significant enhancing effect on the physiological and biochemical parameters of sweetpotato. Pb treatment at high concentrations showed a strong positive correlation with MDA and H₂O₂. Conversely, a notable negative correlation of PC1 variables comprised photosynthetic activity and chlorophyll, which are associated with PC2 (Supplemental Fig. 1B). Pearson's correlation analysis was conducted among tolerant and sensitive varieties of sweetpotato (Supplemental Fig. 1C). The correlation analysis revealed that physiological indicators (Pn and Chl) positively correlated with each other; in the meantime, there were negatively correlated with biochemical parameters MDA and H₂O₂ (Supplemental Fig. 1C). Similarly, the concentration of Pb in roots and leaves also demonstrated a negative correlation of biochemical parameters with plant physiological parameters, indicating a close connection between Pb concentrations and physiological parameters, and the resilience of sweetpotato to Pb.

Comparison of the Pb tolerance of sweetpotato, sunflower, and rapeseed

We compared the phytoremediation capacity of sweetpotato cultivar KO-12 with that of rapeseed and sunflower, which have been well-studied for the clean-up of HM-contaminated soils (Eapen and Dsouza 2005; Eapen et al. 2007; Prasad 2015). Young rapeseed and sunflower plants at a similar growth stage as sweetpotato (KO-12) plants were treated with Pb(NO₃)₂ for 14 days as described above. Rapeseed and sunflower plants died at 30 mM Pb, whereas sweetpotato plants survived both 30 and 50 mM Pb treatments (Supplemental Fig. 2A). To perform a quantitative comparison among the three plant species, the I_{50} value of each plant species was calculated based on plant growth at different Pb concentrations (Supplemental Fig. 2B). The I_{50} values of sweetpotato, sunflower, and rapeseed were 9.5, 7.3, and 6.2 mM Pb, respectively. Thus, sweetpotato shows high potential as a suitable root crop for the phytoremediation of soils contaminated with Pb and possibly other HMs.

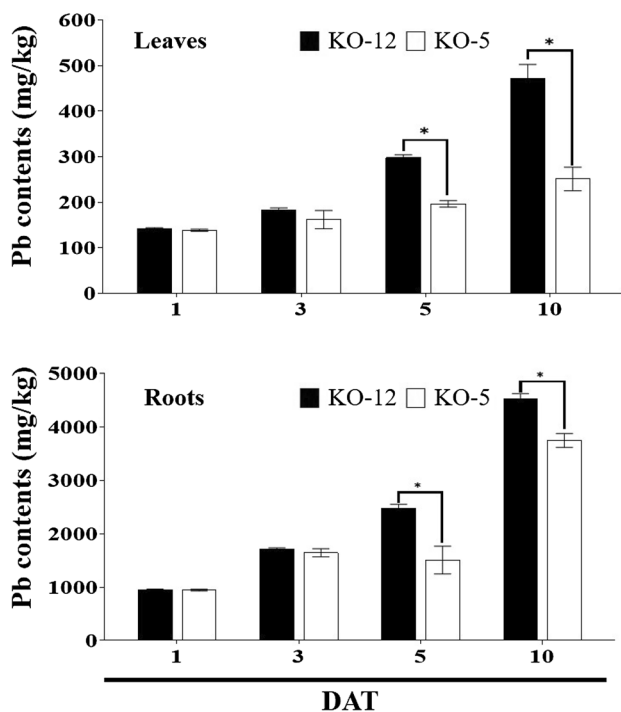


Fig. 5 Pb contents of the leaves and roots of KO-12 and KO-5 cultivars treated with 10 mM Pb. Data represent mean \pm SD of four independent replicates. Asterisks indicate significant differences among cultivars at a given time point (* $p < 0.05$; one-way ANOVA with Tukey's HSD post-hoc test). DAT Days after treatment

Discussion

Sweetpotato offers many advantages over other starchy crops on marginal lands, including HM-contaminated soils, for sustainable agriculture (Mohammad 2021; Mohanraj 2018; Ziska et al. 2009). Moreover, sweetpotato shows high water use efficiency and reduce soil erosion during the rainy season, and therefore can be used as a cover crop. The minimum requirement for sweetpotato cultivation is a frost-free period lasting at least 4 months.

A high concentration of HMs such as Pb can cause a number of toxic symptoms in plants, such as growth retardation (stunting), poor photosynthesis (chlorosis), and root blackening. Pb can inhibit photosynthesis, disrupt mineral nutrition and water balance, and affect membrane structure and permeability (Nas and Ali 2018).

According to our scientific knowledge, sweetpotato, a root crop, can be suitable for phytoremediation, because its high antioxidant activity can help cope with oxidative stress caused by HMs. Although the presence of HMs in sweetpotato storage roots has been investigated in terms of food safety (Zhang et al. 2018), an intensive analysis of sweetpotato plants for phytoremediation is lacking. In this study, we screened the responses of 20 sweetpotato cultivars to Pb treatment with the aim to identify cultivars with high

Pb tolerance. Finally, we selected two Pb-tolerant cultivars (KO-12 and KO-16) and two Pb-sensitive cultivars (KO-7 and KO-5) for further analysis (Table 1). In our experiments, as the Pb concentration and the treatment time increased, the total biomass of plants decreased (Fig. 2B). A similar result has previously been reported in other plant species upon Pb exposure (Zulfiqar et al. 2019).

Among the various physiological processes that determine plant growth and productivity, photosynthesis is the most important. The main reason for the decrease in the content of green pigments in plants under HM toxicity is the suppression of chlorophyll synthesis (Mukhtar et al. 2010; Nawaz et al. 2017). We confirmed that Pb-tolerant cultivars showed higher growth and greater photosynthetic activity than Pb-sensitive cultivars (Fig. 3). The Pb-tolerant cultivar KO-12, when treated with 10 mM for 10 days, showed more stable chlorophyll content and photosynthetic activity than the Pb-sensitive cultivar KO-5. A similar decrease in photosynthetic activity after exposure to HMs, including Pb, has been observed in other plant species (Giannakoula et al. 2021; Mukhtar et al. 2010). Moreover, it has been reported that changes in photosynthetic activity due to Pb exposure lead to decreased growth rate and total plant biomass (Beat et al. 2022).

Based on the obtained results, we can infer that elevated concentrations of Pb are potentially toxic to sweetpotatoes, with high concentrations leading to chlorosis and poor plant growth. Such a trend is observed in other studies investigating the effects of heavy metals on various plants (Dalyan et al. 2020; Zulfiqar et al. 2019). At the same time, some plant genotypes exhibit greater tolerance to different stress conditions compared to others. In our case, the Pb-tolerant cultivar of sweetpotato (KO-12) was able to grow and thrive under elevated lead concentrations in the soil and thus could potentially serve as a phytoremediant.

The I_{50} values were calculated for the Pb-tolerant cultivars KO-12 and KO-16 and Pb-sensitive cultivars KO-7 and KO-5 based on leaf fresh weight. The I_{50} value of KO-12 (6.7 mM) was 5.1 times higher than that of KO-5 (1.3 mM). We speculated that all sweetpotato cultivars exhibit relatively high tolerance to Pb stress, regardless of their genotype, because of their high antioxidant capacity (Kim et al. 2020). However, a comparison of the I_{50} values of the four cultivars indicated that the KO-12 cultivar is the most tolerant to Pb stress. Interestingly, our previous study showed that KO-12 is also highly tolerant to other HMs including cadmium, arsenic, copper, cobalt, and nickel.

Lipid peroxidation is a sign of oxidative stress caused by various abiotic stresses. MDA level is indicative of the level of lipid peroxidation, indicating that high MDA level reflects damage to cell membranes in plants (Morales and Munné-Bosch 2019). MDA has been well characterized as an evaluative trait of HM-stressed plants (Lamhamdi et al.

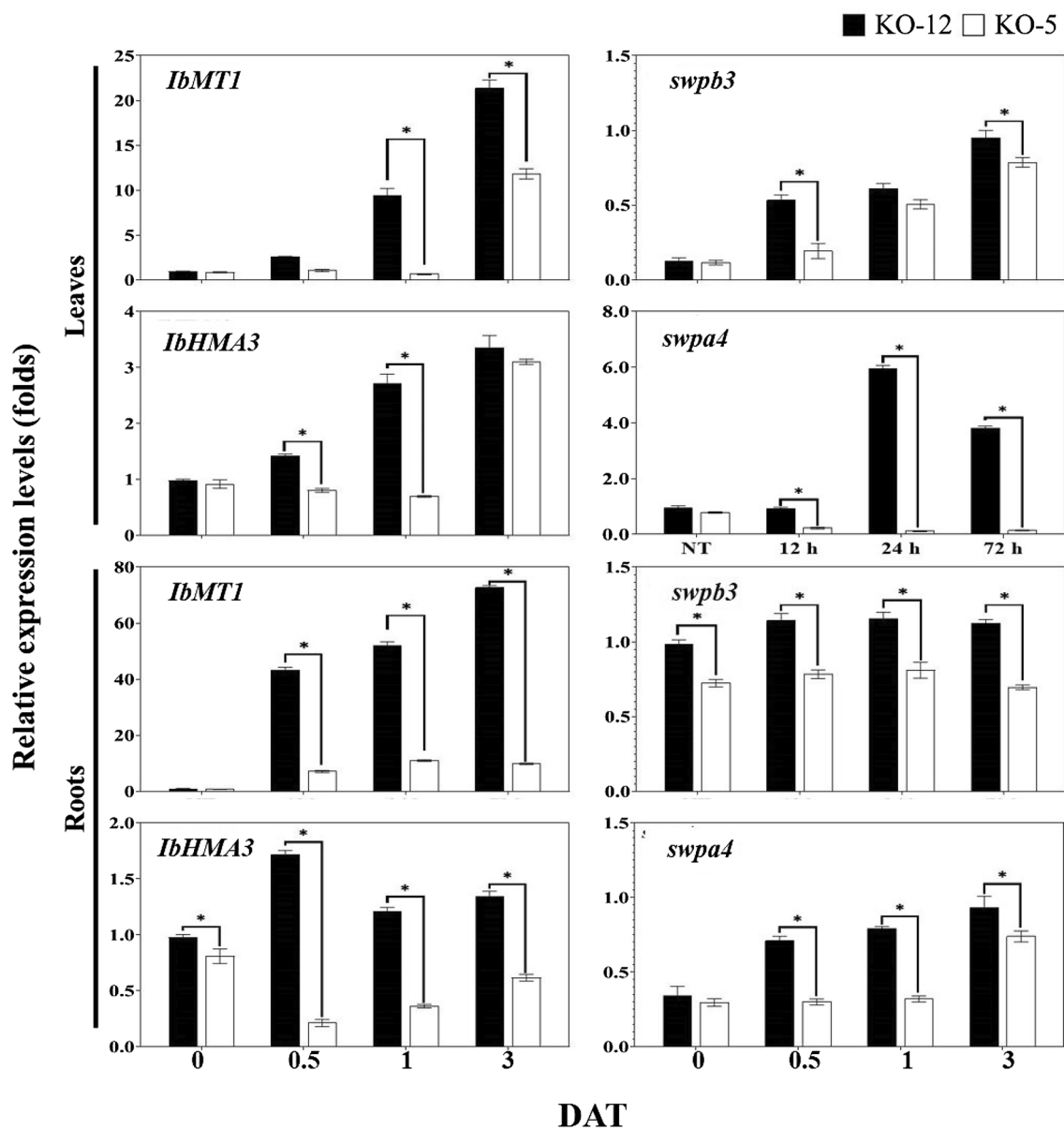


Fig. 6 Expression patterns of Pb-responsive genes in the leaves and roots of KO-12 and KO-5 cultivars treated with 20 mM Pb. Data represent mean \pm SD of four independent replicates. Asterisks indi-

cate significant differences among cultivars at a given time point ($*p < 0.05$; one-way ANOVA with Tukey's HSD post-hoc test). DAT Days after treatment

2013). The MDA content of the roots of KO-12 and KO-5 cultivars significantly increased from 1 day after Pb treatment (Fig. 4). At 10 DAT, the MDA contents of the roots and leaves of KO-5 were approximately 1.4 and 1.3 times higher, respectively, than those of KO-12. Previously published data show that exposure to Pb significantly increases the MDA content of plant cells (Navabpour et al. 2020).

The production of H_2O_2 , the main ROS, increases under HM stress (Sachdev et al. 2021). At 5 and 10 DAT, the H_2O_2 content of KO-5 leaves was approximately 1.2 and 1.5 times higher, respectively, than that of KO-12 leaves, respectively.

A significant increase in H_2O_2 levels in plants subjected to Pb exposure has also been reported previously (Navabpour et al. 2020). Our results indicate that the sweetpotato cultivar KO-5 exhibits high antioxidant activity to cope with Pb-induced oxidative stress.

To tackle the increased levels of environmental pollution, the ability of plants to accumulate HMs is important. Our results showed that sweetpotato plants can accumulate Pb in roots and leaves (Fig. 5). At 10 DAT, the Pb accumulation levels in KO-12 roots and leaves were approximately 1.2 and 2.0 times higher, respectively, than those in the

corresponding KO-5 tissues. A recent study showed higher Pb accumulation in roots than in leaves (Souri et al. 2019).

To understand the differences between the KO-12 (Pb-tolerant) and KO-5 (Pb-sensitive) cultivars at the molecular level, the expression levels of four Pb-responsive genes (*IbMT1*, *IbHMA3*, *swpb3*, and *swpa4*) were analyzed. The expression levels of all four genes were higher in the leaves and roots of KO-12 than in those of KO-5 (Fig. 6). Metallothioneins (MTs) are low-molecular-weight proteins (7–10 kD). The sulfur-containing amino acid cysteine, whose sulfhydryl groups are capable of binding HM ions, accounts for approximately 30% of all amino acids in MTs. To date, four types of MT proteins have been identified (MT1, MT2, MT3, and MT4), which differ in the number and location of cysteine residues (Duan et al. 2019). The expression of *MT1* is more pronounced in plant roots than in stems and leaves. In our study, the *IbMT1* gene was highly expressed in the leaves and roots of KO-12 starting at 12 h in the Pb treatment. Similar work has also been carried out with the *MT* genes in other plant species, with similar results (Kim et al. 2014; Kim and Kang 2018; Nezhad et al. 2013).

The ATPase-related transport protein HMA3 transfers free HM ions through the tonoplast using the energy generated by ATP hydrolysis. In *Arabidopsis thaliana*, overexpression of the *AtHMA3* gene increased tolerance not only to Cd²⁺ but also to Co²⁺, Pb²⁺, and Zn²⁺ (Hermosa et al. 2011). In the current study, the expression level of *IbHMA3* in the KO-12 cultivar was highest at 1 DAT in leaves and at 0.5 DAT in roots. High-level expression of *HMA3* has been reported in the aerial tissues of *Sedum plumbizincicola* (Liu et al. 2017). In other crops, the high expression level of HMA3 in aerial plant parts and roots was associated with the hyperaccumulation of HMs (Talke et al. 2006; Ueno et al. 2011).

POD enzymes protect cells from HM-induced oxidative stress (Kim et al. 2010; Zhang et al. 2023). The expression levels of sweetpotato *POD* genes *swpb3* and *swpa4* have been reported to significantly increase in leaves and roots in response to HM treatment (Kim et al. 2010). Consistently, in our study, *swpb3* and *swpa4* showed higher expression levels in KO-12 than in KO-5. Our results confirmed that the Pb-tolerant KO-12 cultivar is equipped with a Pb tolerance mechanism at the molecular level. Further investigation of the molecular mechanism underlying HM tolerance in sweetpotato is needed for the molecular breeding of sweetpotato cultivars suitable for phytoremediation.

More than 450 hyperaccumulator plant species are currently used for soil remediation (Skuzza et al. 2022). In this study, we compared the phytoremediation capacity of sweetpotato cultivar KO-12 with that of rapeseed and sunflower, which are well-studied for the clean-up of HM-contaminated soils (Eapen and Dsouza 2005; Eapen et al.

2007; Prasad 2015). Sweetpotato plants showed greater tolerance to Pb stress than rapeseed and sunflower plants (Supplemental Fig. 2). The I_{50} value of sweetpotato, sunflower, and rapeseed was 9.5, 7.3, and 6.2 mM Pb. These results suggest that sweetpotato is suitable for the remediation of HM-contaminated soils.

We also compared the Pb tolerance of sweetpotato, sunflower, and rapeseed and analyzed their plant growth inhibition indexes under high Pb concentrations. The results showed that sweetpotato is more tolerant to Pb toxicity than sunflower and rapeseed. Additionally, the plant growth inhibition index of sunflower and rapeseed was 1.3 and 1.5 times higher than that of sweetpotato (Supplemental Fig. 2A, B). The results indicate that sweetpotato is highly Pb tolerant and therefore an excellent candidate for the remediation of HM-contaminated soils. Moreover, sweetpotato storage roots produced on HM-contaminated soils can be used for the extraction of useful biomaterials including bioethanol Ziska et al. 2009). In this study, the results of physiological, biochemical, and molecular analyses suggest that Pb-tolerant sweetpotato cultivar KO-12 (Daeyumi) is a promising biomaterial for the remediation of Pb-contaminated areas. The key genes involved in Pb tolerance in sweetpotato can be found by the comparative transcriptome analysis of the Pb-tolerant and -sensitive cultivars identified in this study. Further studies are needed for the field application of Pb-tolerant sweetpotato cultivars to Pb-contaminated soils and for the characterization of the tolerance of sweetpotato cultivars to other HMs.

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Data availability All datasets supporting the conclusions of this article are included in the article and supplementary files.

Declarations

Conflict of interest The authors declare that they have no conflict of interest.

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References

- Afaj A, Abd J, Noori M, Schüth C (2017) Effects of lead toxicity on the total chlorophyll content and growth changes of the aquatic plant *Ceratophyllum demersum* L. *Int J Environ Studies* 74:119–128
- Alam MK, Rana ZH, Islam SN, Akhtaruzzaman M (2020) Comparative assessment of nutritional composition, polyphenol profile, antidiabetic and antioxidative properties of selected edible wild plant species of Bangladesh. *Food Chem* 1(320):126646
- Beat K, Lars Z, Uwe R, Shizue M, Angelina S, Onno M (2022) Toward predicting photosynthetic efficiency and biomass gain in crop genotypes over a field season. *Plant Physiol* 188:301–317
- Bourdineaud JP, Baudrimont M, Gonzalez P, Moreau JL (2006) Challenging the model for induction of metallothionein gene expression. *Biochimie* 88:1787–1792
- Chauhan VBS, Behera S, Pati K, Bansode VV, Nedunchezhiyan M (2021) Breeding for drought tolerance in sweet potato (*Ipomoea batatas* L.). In: More SJ, Giri NA, Suresh KJ, Visalakshi CC, Tadigiri S (eds) Recent advances in root and tuber crops. Brill-lion Publishing House, New Delhi, pp 65–87
- Chen SP, Lin IW, Chen X, Huang YH, Chang SC, Lo HS, Lu HH, Yeh KW (2016) Sweet potato NAC transcription factor, IbNAC1, upregulates sporamin gene expression by binding the SWRE motif against mechanical wounding and herbivore attack. *Plant J* 86:234–248
- Cobbett C, Goldsbrough P (2002) Phytochelatin and metallothioneins: roles in heavy metal detoxification and homeostasis. *Annu Rev Plant Biol* 53:159–182
- CSPI (The Center for Science in the Public Interest) (2007) 10 best foods. <https://cspinet.org/eating-healthy/what-eat/10-best-foods>
- Dalyan E, Yüzbaşıoğlu E, Akpınar I (2020) Physiological and biochemical changes in plant growth and different plant enzymes in response to lead stress. In: Gupta D, Chatterjee S, Walther C (eds) Lead in plants and the environment. Radionuclides and Heavy metals in the environment. Springer, Cham
- Daurov D, Zhambakin K, Shamekova M (2023) Phytoremediation as a way to clean technologically polluted areas of Kazakhstan. *Braz J Biol* 83:e271684
- Duan L, Yu J, Xu L, Tian P, Hu X, Song X, Pan Y (2019) Functional characterization of a type 4 metallothionein gene (CsMT4) in cucumber. *Hortic Plant* 3:120–128
- Eapen S, Dsouza SF (2005) Prospects of genetic engineering of plants for phytoremediation of toxic metals. *Biotechnol Adv* 23:97–114
- Eapen S, Singh S, D'Souza SF (2007) Phytoremediation of metals and radionuclides. *Environmental bioremediation technologies*. Springer, Berlin Heidelberg New York, pp 189–209
- Giannakoula A, Therios I, Chatzissavvidis C (2021) Effect of lead and copper on photosynthetic apparatus in citrus (*Citrus aurantium* L.) plants. The role of antioxidants in oxidative damage as a response to heavy metal stress. *Plants* 10:155
- Gülen H, Çetinkaya C, Kadioğlu M, Kesici M, Cansev A, Eriş A (2008) Peroxidase activity and lipid peroxidation in strawberry (*Fragaria ananassa*) plants under low temperature. *J Biol Environ Sci* 2:95–100
- Hermosa R, Botella L, Keck E, Jiménez JÁ, Montero-Barrientos M, Arbona V, Gómez-Cadenas A, Monte E, Nicolás C (2011) The overexpression in *Arabidopsis thaliana* of a *Trichoderma harzianum* gene that modulates glucosidase activity, and enhances tolerance to salt and osmotic stresses. *J Plant Physiol* 168:1295–1302
- Hieno A, Naznin HA, Inaba-Hasegawa K, Yokogawa T, Hayami N, Nomoto M, Tada Y, Yokogawa T, Higuchi-Takeuchi M, Hanada K, Matsui M, Ikeda Y, Hojo Y, Hirayama T, Kusunoki K, Koyama H, Mitsuda N, Yamamoto YY (2019) Transcriptome analysis and identification of a transcriptional regulatory network in the response to H₂O₂. *Plant Physiol* 180(3):1629–1646
- Izinyon OC, Seghosime A (2013) Assessment of sweet potato (*Ipomoea batatas*) for phytoremediation of motor oil contaminated soil. *Niger J Technol* 32:371–378
- Jang IC, Park SY, Kim KY, Kwon SY, Kim JG, Kwak SS (2004) Differential expression of 10 sweetpotato peroxidase genes in response to bacterial pathogen, *Pectobacterium chrysanthemi*. *Plant Physiol Biochem* 42(5):451–455
- Junglee S, Urban L, Sallanon H, Lopez-Lauri F (2014) Optimized assay for hydrogen peroxide determination in plant tissue using potassium iodide. *Am J Anal Chem* 5:730–736
- Kim YO, Kang H (2018) Comparative expression analysis of genes encoding metallothioneins in response to heavy metals and abiotic stresses in rice (*Oryza sativa*) and *Arabidopsis thaliana*. *Biosci Biotechnol Biochem* 82:1656–1665
- Kim YH, Lee HS, Kwak SS (2010) Differential responses of sweet potato peroxidases to heavy metals. *Chemosphere* 81:79–85
- Kim SH, Jeong JC, Ahn YO, Lee HS, Kwak SS (2014) Differential responses of three sweet potato metallothionein genes to abiotic stress and heavy metals. *Mol Biol Rep* 41:6957–6966
- Kim HS, Wang WB, Kang L, Kim SW, Lee CJ, Park SC, Park WS, Ahn MJ, Kwak SS (2020) Metabolic engineering of low-molecular-weight antioxidants in sweetpotato. *Plant Biotechnol Rep* 14:193–205
- Kshyanapra R, Das AP (2023) Lead pollution: impact on environment and human health and approach for a sustainable solution. *Environ Toxicol Chem* 5:79–85
- Kushwaha A, Hans N, Kumar S, Rani R (2018) A critical review on speciation, mobilization and toxicity of lead in soil-microbe-plant system and bioremediation strategies. *Ecotoxicol Environ Saf* 147:1035–1045
- Kwak SS (2019) Biotechnology of the sweet potato: ensuring global food and nutrition security in the face of climate change. *Plant Cell Rep* 38:1361–1363
- Lamhamdi M, Bakrim A, Bouayad N, Aarab A, Lafont R (2013) Protective role of a methanolic extract of spinach (*Spinacia oleracea* L.) against Pb toxicity in wheat (*Triticum aestivum* L.) seedlings: beneficial effects for a plant of a nutraceutical used with animals. *Environ Sci Pollut Res* 20:7377–7385
- Leszczyszyn OI, Imam HT, Blindauer CA (2013) Diversity and distribution of plant metallothioneins: a review of structure, properties and functions. *Metallomics* 5:1146–1169
- Liu H, Zhao H, Wu L, Liu A, Zhao FJ, Xu W (2017) Heavy metal ATPase 3 (HMA3) confers cadmium hypertolerance in the cadmium/zinc hyperaccumulator sedum plumbizincicola. *New Phytol* 215:687–698

- Mandiwana KL, Panichev N, Kataeva M, Siebert S (2007) The solubility of Cr(III) and Cr(VI) compounds in soil and their availability to plants. *J Hazard Mater* 147:540–545
- Mitra S, Chakraborty AJ, Tareq AM, Emran TB, Nainu F, Khuro A, Idris AN, Khandaker MU, Osman H, Alhumaydhi FA, Simal-Gandara J (2022) Impact of heavy metals on the environment and human health: novel therapeutic insights to counter the toxicity. *J King Saud Univ Sci* 34:101865
- Mocek-Plóćiniak A, Mencil J, Zakrzewski W, Roszkowski S (2023) Phytoremediation as an effective remedy for removing trace elements from ecosystems. *Plants* 12:1653
- Mohammad KA (2021) A comprehensive review of sweet potato (*Ipomoea batatas* [L.] Lam): revisiting the associated health benefits. *Trends Food Sci Technol* 115:512–529
- Mohanraj R (2018) Sweet potato: bioactive compounds and health benefits. In: Merillon JM, Ramawat K (eds) *Bioactive molecules in food*. Springer, Cham, pp 919–934
- Morales M, Munné-Bosch S (2019) Malondialdehyde: facts and artifacts. *Plant Physiol* 180:1246–1250
- Mukhtar S, Bhatti HN, Khalid M, Anwar-ul-Haq M, Shahzad SM (2010) Potential of sunflower (*Helianthus annuus* L.) for phytoremediation of nickel (Ni) and lead (Pb) contaminated water. *Pak J Bot* 42:4017–4026
- Nas FS, Ali M (2018) The effect of lead on plants in terms of growing and biochemical parameters: a review. *MOJ Eco Environ Sci* 3:265–268
- Navabpour S, Yamchi A, Bagherikia S, Kafi H (2020) Lead-induced oxidative stress and role of antioxidant defense in wheat (*Triticum aestivum* L.). *Physiol Mol Biol Plants* 26:793–802
- Nawaz F, Naeem M, Akram A, Ashraf MY, Ahmad KS, Zulfiqar B, Sardar H, Shabbir RN, Majeed S, Shehzad MA, Anwar I (2017) Seed priming with KNO₃ mediates biochemical processes to inhibit lead toxicity in maize (*Zea mays* L.). *J Sci Food Agric* 14:4780–4789
- Nezhad RM, Shahpiri A, Mirolohi A (2013) Heterologous expression and metal-binding characterization of a type I metallothionein isoform (OsMTI-1b) from rice (*Oryza sativa*). *Protein J* 32:131–137
- Pan Y, Zhu M, Wang S, Ma G, Huang X, Qiao C, Wang R, Xu X, Liang Y, Lu K, Li J, Qu C (2018) Genome-Wide characterization and analysis of Metallothionein family genes that function in metal stress tolerance in *Brassica napus* L. *Int J Mol Sci* 19(8):2181
- Park SU, Lee CJ, Ki SE, Lim YH, Lee HU, Nam SS, Kim HS, Kwak SS (2020) Selection of flooding stress tolerant sweetpotato cultivars based on biochemical and phenotypic characterization. *Plant Physiol Biochem* 155:243–251
- Pourrut B, Shahid M, Dumat C, Winterton P, Pinelli E (2011) Lead uptake, toxicity, and detoxification in plants. *Rev Environ Contam Toxicol* 213:113–136
- Prasad MNV (2015) *Phytoremediation crops and biofuels. Sustainable agriculture reviews*. Springer, Berlin Heidelberg New York, pp 159–261
- Rejeb IB, Pastor V, Mauch-Mani B (2014) Plant responses to simultaneous biotic and abiotic stress: molecular mechanisms. *Plants* 3(4):458–475
- Robinson NJ, Tommey AM, Kuske C, Jackson PJ (1993) Plant metallothioneins. *Biochem J* 295:1–10
- Sachdev S, Ansari SA, Ansari MI, Fujita M, Hasanuzzaman M (2021) Abiotic stress and reactive oxygen species: generation, signaling, and defense mechanisms. *Antioxidants* 10(2):277
- Samuel C, Amritha B, Deepthi MG, Mohamed NVSA, Praveena B, Rajan Ch, Vladislav L, Gabriel IT, Fedor S, Sivasankar K, Sasikumar S (2022) Bioaccumulation of lead (Pb) and its effects in plants: a review. *J Hazard Mat Lett* 3:100064
- Sapakhova Z, Raissova N, Daurov D, Zhapar K, Daurova A, Zhigailov A, Zhambakin K, Shamekova M (2023) Sweet potato as a key crop for food security under the conditions of global climate change: a review. *Plants* 12:2516
- Sevak PI, Pushkar BK, Kapadne PN (2021) Lead pollution and bacterial bioremediation: a review. *Environ Chem Lett* 19:4463–4488
- Sharma P, Dubey RS (2005) Lead toxicity in plants. *Braz J Plant Physiol* 17:1–19
- Skuza L, Szućko-Kociuba I, Filip E, Bożek I (2022) Natural molecular mechanisms of plant hyperaccumulation and hypertolerance towards heavy metals. *Int J Mol Sci* 23(16):9335
- Souri MK, Hatamian M, Tesfamariam T (2019) Plant growth stage influences heavy metal accumulation in leafy vegetables of garden cress and sweet basil. *Chem Biol Technol Agric* 6:25
- Sun Y, Pan Z, Yang C, Jia Z, Guo X (2019) Comparative assessment of phenolic profiles, cellular antioxidant and antiproliferative activities in ten varieties of sweet potato (*Ipomoea batatas*) storage roots. *Molecules* 24:4476
- Sung YW, Lee IH, Shim D, Lee KL, Nam KJ, Yang JW, Lee JJ, Kwak SS, Kim YH (2019) Transcriptomic changes in sweet potato peroxidases in response to infection with the root-knot nematode *Meloidogyne incognita*. *Mol Biol Rep* 46(4):4555–4564
- Takahashi R, Bashir K, Ishimaru Y, Nishizawa NK, Nakanishi H (2012) The role of heavy-metal ATPases, HMAs, in zinc and cadmium transport in rice. *Plant Signal Behav* 7(12):1605–1607
- Talke IN, Hanikenne M, Kramer U (2006) Zinc-dependent transcriptional control, transcriptional deregulation, and higher gene copy number for genes in metal homeostasis of the hyperaccumulator *Arabidopsis halleri*. *Plant Physiol* 142:148–167
- Tamunggang NEB, Aline D, Alakeh NMN, Gilda TF (2016) Phytoextraction of cadmium by beans and sweet potatoes from soils. *IJRRAS* 28:65–70
- Toishimanov M, Abilda Z, Daurov D, Daurova A, Zhapar K, Sapakhova Z, Kanat R, Stangaliyeva Z, Zhambakin K, Shamekova M (2023) Phytoremediation properties of sweet potato for soils contaminated by heavy metals in South Kazakhstan. *Appl Sci* 13:9589
- Ueno D, Milner MJ, Yamaji N, Yokosho K, Koyama E, Clemencia Zambrano M, Kaskie M, Ebbs S, Kochian LV, Ma JF (2011) Elevated expression of TcHMA3 plays a key role in the extreme Cd tolerance in a Cd-hyperaccumulating ecotype of *Thlaspi caerulescens*. *Plant J* 66:852–862
- Wang WB, Kim YH, Lee HS, Kim KY, Deng XP, Kwak SS (2009) Analysis of antioxidant enzyme activity during germination of alfalfa under salt and drought stresses. *Plant Physiol Biochem* 47:570–577
- Wang W, Yu H, Kim HS, Yang Y, Qiu X, Kwak SS (2019) Molecular characterization of a sweet potato stress tolerance-associated *trehalose-6-phosphate synthase 1* gene (IbTPS1) in response to abiotic stress. *Plant Biotechnol Rep* 13:235–243
- Williams LE, Mills RF (2005) P(1B)-ATPases—an ancient family of transition metal pumps with diverse functions in plants. *Trends Plant Sci* 10(10):491–502
- Xie L, Hao P, Cheng Y, Ahmed IM, Cao F (2018) Effect of combined application of lead, cadmium, chromium and copper on grain, leaf and stem heavy metal contents at different growth stages in rice. *Ecotoxicol Environ Saf* 162:71–76
- Yang Y, Wei X, Lu J, You J, Wang W, Shi R (2010) Lead-induced phytotoxicity mechanism involved in seed germination and seedling growth of wheat (*Triticum aestivum* L.). *Ecotoxicol Environ Saf* 73:1982–1987
- Yang D, Xie Y, Sun H, Bian X, Ke Q, Kim HS, Ji CY, Jin R, Wang W, Zhang C, Ma J, Li Z, Ma D, Kwak SS (2020) IbINH positively regulates drought stress tolerance in sweet potato. *Plant Physiol Biochem* 146:403–410
- Zhambakin K, Zhapar K (2020) Current status and prospects of plant biotechnology in Kazakhstan. *Plant Biotechnol Rep* 14:177–184

- Zhang D, Zhang C, Dong F, Zhou Hong, Zhang Y, Huang Y (2018) Study on the accumulation of cadmium in sweetpotato. 8th International Sweetpotato Symposium “Promoting High-value Added Industry of Sweetpotato for the Fourth Industrial Innovation” (September 5–8, 2018, RDA Auditorium, Jeonju, Korea). Abstract book, pp 138–139
- Zhang H, Sun X, Hwarari D, Du X, Wang Y, Xu H, Lv S, Wang T, Yang L, Hou D (2023) Oxidative stress response and metal transport in roots of *Macleaya cordata* exposed to lead and zinc. *Plants* 12(3):516
- Ziska LH, Runion GB, Tomecek M, Prior SA, Torbet HA, Sicher R (2009) An evaluation of cassava, sweetpotato and field corn as a

potential carbohydrate source for bioethanol production in Alabama and Maryland. *Biomass Bioenerg* 33:1503–1508

Zulfiqar U, Farooq M, Hussain S, Maqsood M, Hussain M, Ishfaq M, Ahmad M, Anjum MZ (2019) Lead toxicity in plants: impacts and remediation. *J Environ Manage* 2250:109557

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