

# mTOR-targeted cancer therapy: great target but disappointing clinical outcomes, why?

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**Abstract** The mammalian target of rapamycin (mTOR) critically regulates several essential biological functions, such as cell growth, metabolism, survival, and immune response by forming two important complexes, namely, mTOR complex 1 (mTORC1) and complex 2 (mTORC2). mTOR signaling is often dysregulated in cancers and has been considered an attractive cancer therapeutic target. Great efforts have been made to develop efficacious mTOR inhibitors, particularly mTOR kinase inhibitors, which suppress mTORC1 and mTORC2; however, major success has not been achieved. With the strong scientific rationale, the intriguing question is why cancers are insensitive or not responsive to mTOR-targeted cancer therapy in clinics. Beyond early findings on induced activation of PI3K/Akt, MEK/ERK, and Mnk/eIF4E survival signaling pathways that compromise the efficacy of rapalog-based cancer therapy, recent findings on the essential role of GSK3 in mediating cancer cell response to mTOR inhibitors and mTORC1 inhibition-induced upregulation of PD-L1 in cancer cells may provide some explanations. These new findings may also offer us the opportunity to rationally utilize mTOR inhibitors in cancer therapy. Further elucidation of the biology of complicated mTOR networks may bring us the hope to develop effective therapeutic strategies with mTOR inhibitors against cancer.

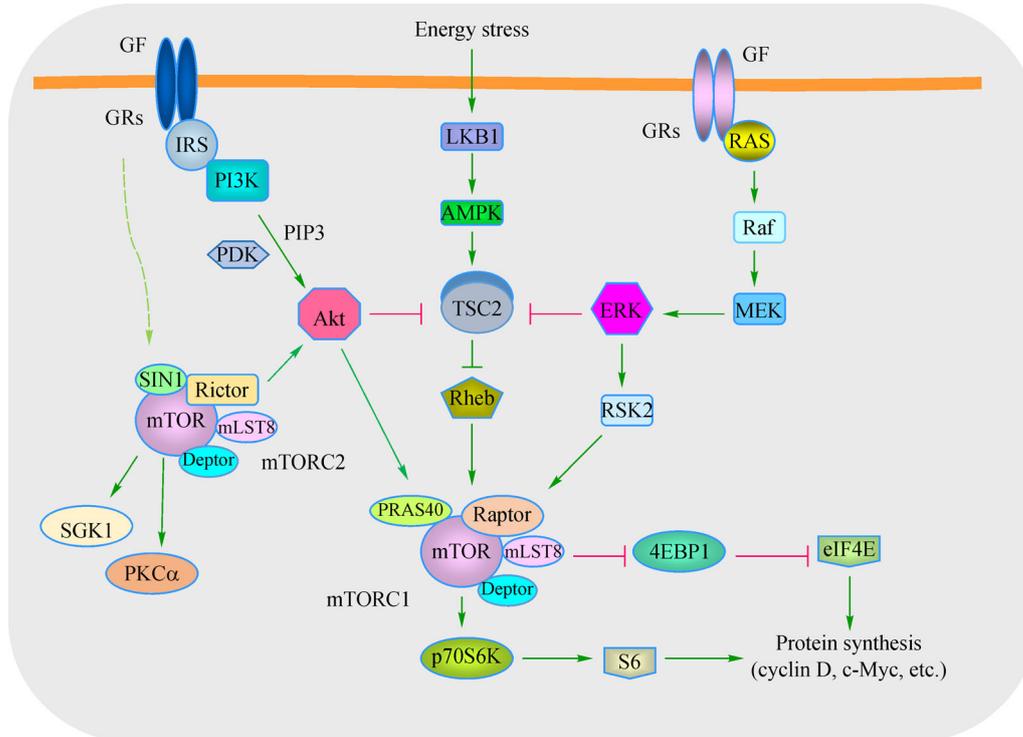
**Keywords** mTOR; cancer therapy; resistance; GSK3; protein degradation; E3 ubiquitin ligase; PD-L1

## Background

The mammalian target of rapamycin (mTOR) and its mediated signaling pathways are critical for maintaining cell homeostasis through regulation of various biological functions, such as cell growth, metabolism, survival, and immune response. These functions are regulated by mTOR, a serine–threonine kinase, primarily through forming two distinct complexes, namely, mTOR complex 1 (mTORC1) and mTOR complex 2 (mTORC2) [1–3]. Determination of mTOR kinase substrate specificity is dependent on which partner protein it interacts with. mTORC1 is composed of mTOR and four other associated proteins such as raptor, mLST8, PRAS40, and DEPTOR; signaling of mTORC1 is crucial for regulating cap-dependent translation initiation, an essential process for synthesis of many oncogenic proteins, such as cyclin D1,

c-Myc, Mcl-1, and Snail, through phosphorylating p70 ribosomal protein S6 kinase (p70S6K) and eukaryotic translation initiation factor 4E (eIF4E) binding protein 1 (4E-BP1). mTORC2 contains mTOR, rictor, mLST8, DEPTOR, mSin1, and protor and phosphorylates Akt, serum and glucocorticoid-inducible kinase (SGK), and protein kinase C (PKC) [2–4] (Fig. 1). The biological functions of mTORC2, particularly those related to regulation of oncogenesis other than its regulation of cytoskeleton and cell survival, have not been fully understood in comparison with mTORC1 [2], although mTORC2 is involved in the positive regulation of cancer development [5–7]. At present, limited information is known about the upstream regulators of the mTORC2 axis; meanwhile, mTORC1 serves as a convergence point of phosphoinositide 3-kinase (PI3K)/Akt and mitogen-activated protein kinase (MAPK)/MEK/ERK signaling pathways, which are often hyperactivated in many types of cancers [8] (Fig. 1). Nonetheless, the mTOR axis is dysregulated in cancers and thus has emerged as an attractive cancer therapeutic target [9].

Rapamycin and its analogs (rapalogs) are specific



**Fig. 1** mTOR complexes and their regulation by various upstream signaling pathways. The PI3K/Akt survival pathway functions upstream to positively regulate the activity of mTORC1 by phosphorylating tuberous sclerosis complex 2 (TSC2); PRAS40, ERK1/2, and p90RSK2 (RSK2) also positively regulate mTORC1 through respective phosphorylation of TSC2 and raptor, thus linking Ras to the positive regulation of mTORC1 signaling. The tumor suppressor, liver kinase B1 (LKB1), inhibits mTORC1 signaling through activation of AMPK and TSC2. Whether the same upstream signals that regulate mTORC1 also regulate mTORC2 is unclear. mTORC2 is known to phosphorylate Akt, SGK1, and PKC $\alpha$ , whereas mTORC1 primarily phosphorylates p70S6K and 4EBP1. GF, growth factor; GRs, growth factor receptors; IRS, insulin receptor substrate.

allosteric inhibitors of mTOR and represent first-generation mTOR inhibitors. Some rapalogs (e.g., everolimus/RAD001 and temsirolimus/CCI-779) are approved by the Food and Drug Administration (FDA) for treatment of certain cancers such as metastatic renal cell carcinoma, pancreatic neuroendocrine tumors, and postmenopausal hormone receptor-positive advanced breast cancer, but the single-agent activity of rapalogs in many other tumor types has been limited [10]. For the past decades, great efforts have been made to develop second-generation ATP-competitive mTOR kinase inhibitors (TORKinibs), such as INK128, Torin 1, and AZD8055, which suppress mTORC1 and mTORC2 activity, as potential cancer therapeutic agents [11,12], and third-generation bivalent mTOR inhibitors that specifically target mTOR resistance mutations [13] in hopes of developing more efficacious mTOR inhibitors based on the rationale that inhibition of mTORC1 and mTORC2 may achieve better therapeutic efficacy. As a consequence, some TORKinibs have been tested in clinical trials [11,12]. TORKinibs not only represent novel potential anticancer agents but are also valuable research tools for understanding the biology of mTOR signaling. However, TORKinibs have not demonstrated clinical effectiveness against different types of

cancer, although the scientific rationale for targeting mTORCs is very strong.

Thus, the intriguing question is, despite such a great target, why are cancer cells insensitive or not responsive to mTOR-targeted cancer therapy, particularly TORKinib-based cancer therapy, in clinics? The current review will focus on discussing possible mechanisms for the resistance of cancer cells to mTOR-targeted cancer therapy.

### Induced activation of survival signaling pathways during mTOR-targeted cancer therapy

mTORC1 functions downstream of the PI3K/Akt signaling pathway. In 2005, we first showed that rapalogs induce Akt phosphorylation (e.g., Ser473) while inhibiting mTORC1 signaling in cancer cells [14]. This observation was subsequently confirmed in other studies in different types of cancer cells [15–17], in xenograft tissues [18] and in human cancer tissues [15,19,20]. One potential mechanism underlying rapalog-induced Akt activation is the relief of mTORC1/p70S6K-mediated feedback inhibition of insulin receptor substrate-1 (IRS-1)/PI3K as suggested [15,21]. However, other studies do not support this model [17,22].

We have suggested that rapalogs induce protein phosphatase 2 (PP2A)-dependent DNA-protein kinase (DNA-PK) activation, resulting in increased Akt phosphorylation [23].

Although mitogen-activated protein kinase (MAPK)/extracellular signal-regulated kinase (ERK) positively regulates mTORC1 activity [9], studies show that mTORC1 inhibition can also induce feedback activation of MAPK/ERK signaling in cell culture models, mouse tumor models, and, most importantly, cancer tissues from patients [24,25], thereby adding another layer of complexity to the already complicated mTOR signaling network. Similar to activation of PI3K/Akt signaling, mTORC1 inhibition-induced MAPK/ERK activation is thought to be a consequence of p70S6K inhibition and subsequent PI3K activation and in turn activates Ras, causing the activation of MAPK/ERK signaling [24].

Beyond the activation of Akt and ERK survival signaling pathways, rapalogs paradoxically increase eIF4E phosphorylation (Ser209) in various types of cancer cells while inhibiting mTORC1 signaling as we previously reported [14]. Similar to Akt phosphorylation, rapalog-induced eIF4E phosphorylation is rapid and sustained [14,26]. We have suggested an underlying mechanism involving PI3K-dependent, MAPK interacting protein kinase (Mnk)-mediated eIF4E phosphorylation but independent of MAPK signaling pathways [14,26].

Given that the blockage of the activation of these pathways enhanced the therapeutic efficacies of rapalogs against the growth of cancer [9], induced activation of PI3K/Akt, MAPK/ERK, and Mnk/eIF4E during rapalog-based cancer therapy will provide survival advantage for cancer cells, eventually blunting the therapeutic efficacies of rapalogs.

### **Critical role of glycogen synthase kinase-3 (GSK3) in mTOR-targeted cancer therapy**

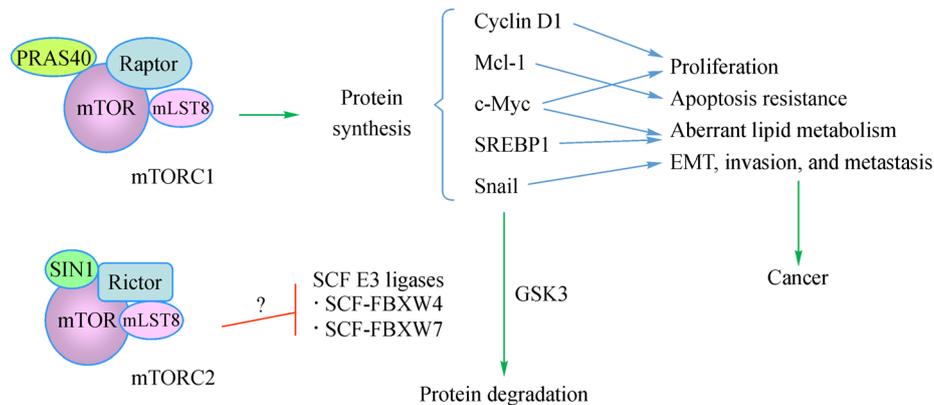
GSK3 is a ubiquitous serine/threonine kinase that is present in mammals in two isoforms, namely,  $\alpha$  and  $\beta$  [27]. GSK3 has a key role in regulating multiple cellular functions, including glycogen metabolism, cell survival, and cell death [27]. However, the involvement of GSK3 in the regulation of oncogenesis is complex: it acts paradoxically as a tumor suppressor in some cancer types while potentiating the growth of cancer cells in other types [28,29]. Phosphorylation at the S21 ( $\alpha$ ) and S9 ( $\beta$ ) of GSK3 by several kinases, such as Akt, p90 ribosomal S6 kinase (p90RSK), PKC, and p70S6K can cause the inactivation of GSK3 [30–32].

GSK3 phosphorylates the turn motif of p70S6K and cooperates with mTORC1 to control the activity of p70S6K and cell proliferation [33]. This finding may provide a rationale for co-targeting mTOR and GSK3 to treat cancer. However, our studies based on this rationale

generated opposed outcomes, that is, GSK3 inhibition actually antagonizes the ability of mTOR inhibitors (rapalogs and TORKinibs) to inhibit the growth of non-small cell lung cancer (NSCLC) cells *in vitro* and *in vivo*, implying that the presence of GSK3 activity is in fact crucial for mTOR inhibitors to exert their anticancer activity [34–36]. Using a panel of different NSCLC cell lines, we reported that the high basal levels of p-GSK3 are significantly correlated with decreased sensitivity of cancer cells to rapalogs and TORKinibs [34,35]. These findings collectively suggest that the presence of activated GSK3 is critical for cancer cells to respond to mTOR-targeted cancer therapy. Considering that other studies show that GSK3 inhibits the mTOR pathway through the phosphorylation of tuberous sclerosis complex 2 (TSC2) in a manner dependent on AMPK-priming phosphorylation [37], where GSK3 is a direct target of S6K1 and is activated upon mTORC1 inhibition [38], however, the real role of GSK3 in mTOR signaling networks is obviously unclear. In our own studies, the inhibition of mTOR with rapalogs and TORKinibs, despite causing strong suppression of p70S6K, does not alter GSK3 phosphorylation [34,35,39].

The intriguing question is why GSK3 is so important for mTOR-targeted cancer therapy. Our initial speculation is that GSK3 inhibition might interfere with the ability of mTOR inhibitors to suppress mTORC1 signaling because it antagonizes the activities of rapalogs and TORKinibs [34,35]. Interestingly, GSK3 inhibition does not interfere with the ability of mTOR inhibitors to suppress mTORC1 signaling and cap-dependent binding or eIF4F complex formation. As such, GSK3 inhibition is unlikely to antagonize the therapeutic efficacies of mTOR inhibitors through interfering with their functions in suppressing the mTORC1 signaling. In an effort to find out the real underlying mechanisms, we have revealed that mTORC2 positively regulates the stabilization of several oncogenic proteins, including cyclin D1, Mcl-1, c-Myc, sterol regulatory element binding protein 1 (SREBP1), and Snail, through inhibiting GSK3-dependent and Skp, Cullin, and F-box containing complex (SCF) E3 ubiquitin E3 ligase-mediated proteasomal degradation [35,36,39, 40]. For the first time, these findings have connected mTORC2 to the stabilization of these oncogenic proteins by inhibiting their degradation; this finding represents another layer of regulation of oncogenic proteins in addition to mTORC1-mediated regulation of protein translation (Fig. 2).

One well-known important function of GSK3 is to phosphorylate these oncogenic proteins mentioned above, resulting in their degradation through SCF E3 ubiquitin ligases, including F-box/WD repeat-containing protein 4 (FBXW4), FBXW7, and  $\beta$ -transducin repeats-containing protein ( $\beta$ -TrCP) [41–46]. Hence, GSK3-mediated phosphorylation is a prerequisite for triggering the degradation



**Fig. 2** Diagram for different layers of regulation of several oncogenic proteins by mTORC1 and mTORC2 at translational and posttranslational levels. Currently, how mTORC2 negatively regulates GSK3-dependent and E3 ubiquitin ligase-mediated protein degradation remains unclear.

of these oncogenic proteins. As such, inhibition of GSK3 will compromise the ability of mTORC2 inhibition to induce the degradation of these oncogenic proteins, which are critical for positive regulation of cell survival, cell cycle progression, and cell invasion/metastasis, as demonstrated in our recent publications [35,36,39,40]. As a consequence, the effects of mTOR inhibitors, such as suppression of growth, epithelial–mesenchymal transition (EMT) and migration of cancer cells, will be compromised as a result of GSK3 inactivation. Thus, our findings have demonstrated the critical role of mTORC2 inhibition in the response of cancer cells to mTOR inhibitors.

Rapalogs primarily inhibit mTORC1 with weak or inactive activity against mTORC2. The challenge is that TORKinibs and rapalogs induce GSK3-dependent and SCF E3 ligase-mediated degradation of cyclin D1, Mcl-1, c-Myc, SREBP1, and Snail as consequences of mTORC2 inhibition. Rapalogs inhibit mTORC2 in different systems *in vitro* and *in vivo*; however, this effect is largely due to chronic or prolonged rapamycin treatment that can eventually disrupt mTORC2 assembly as reported in literature [18,47–53]. Our own data showed that acute treatment with rapamycin (for 1 h) disrupted the assembly of not only mTORC1 (mTOR with raptor) but also mTORC2 (mTOR with rictor) in different cancer cell lines [34]. In fact, a previous study did show that rictor was dissociated from mTOR at 0.5–2 h post rapamycin treatment in few cell lines such as PC-3, BJAB, and Jurkat [47]. Few studies specifically determine the effect of acute rapamycin treatment on mTORC2 assembly in various types of cancer cell lines. We believe that rapalogs, such as rapamycin, have an acute inhibitory effect against the assembly of mTORC2, at least in some cancer cell lines. These findings elucidate why GSK3 inhibition also antagonizes the therapeutic efficacy of rapalogs.

Under the condition of acute treatment with rapamycin,

Akt S473 phosphorylation, a commonly used readout of mTORC2 activity, clearly increased in the tested cell lines [34], consistent with what we initially reported [14]. However, we observed the reduced levels of SGK1 (S422) in every tested cell line under the same conditions [34]. The disruption of the mTORC2 assembly is associated with the suppression of SGK1 S422 phosphorylation. Whether SGK1 S422 phosphorylation serves as a better readout of mTORC2 activity than Akt S473 phosphorylation needs further investigation.

The mutation of *PTEN* and *PIK3CA* activates the PI3K/Akt signaling pathway, leading to the inactivation of GSK3 by phosphorylation, whereas the mutation of *K-Ras* can also inhibit GSK3 activity via ERK/p90RSK-mediated GSK3 phosphorylation [30]. Given the high frequency of these gene mutations or pathway activation in many types of cancers, GSK3 should be highly phosphorylated (i.e., inactivated) in these cancers. Indeed, GSK3 phosphorylation was detected in  $\geq 50\%$  of tissue samples of several human cancers including breast cancer [54], mantle cell lymphoma [55], gastric cancer [56], hepatocellular carcinoma [57], and NSCLC [35]. Hence, many cancer types, particularly those with inactivated GSK3, are insensitive to mTOR-targeted cancer therapy in clinics.

### Potential negative impact of E3 ubiquitin ligase mutation on cancer response to mTOR-targeted therapy

An essential process in proteasomal protein degradation is the ubiquitination of targeted proteins mediated by E3 ubiquitin ligases. In the above-mentioned degradation of cyclin D1, c-Myc, Mcl-1, and SREBP1 induced by mTOR inhibitors, we demonstrated that Mcl-1, c-Myc, and SREBP1 are ubiquitinated by FBXW7 (FBW7) [39,40], whereas cyclin D1 is ubiquitinated by FBX4 (FBXW4)

[35]. Therefore, mTOR inhibitors will be unable to induce the degradation of these proteins in cancers with inactivation of the E3 ligases due to gene mutations, deletion, or other mechanisms, resulting in limited therapeutic efficacies. *FBXW7* has the highest mutation frequency among F-Box and WD repeat domain-containing SCF E3 ubiquitin ligases. The mutation of *FBXW7* may reduce the protein's ability to form SCF complexes and/or alter the complex conformation, leading to a non-functional complex [58]. Data from the Catalogue Of Somatic Mutations In Cancer (COSMIC) database revealed the overall *FBXW7* mutation rate of 2.54 across all human tumors, among which, endometrium, large intestine, cervix, small intestine, and stomach are the top five tumor types [58]. The Cancer Genome Atlas (TCGA) database search found an overall 5% *FBXW7* mutation rate across all human tumor types. Endometrial carcinoma has the highest *FBXW7* mutation rate of up to 20%. Several other types of cancer, including colorectal adenocarcinoma, cervical squamous cell carcinoma, cervical adenocarcinoma, esophagogastric adenocarcinoma, bladder urothelial carcinoma, head and neck squamous cell carcinoma, undifferentiated stomach adenocarcinoma, esophageal squamous cell carcinoma, and melanoma, have high frequencies of *FBXW7* mutations, ranging from 15% to 5% (Fig. 3A). *FBX4* mutations are relatively low in human cancers with an overall rate of around 0.8% across all human tumors. Endometrial carcinoma, adrenocortical carcinoma, pancreatic carcinoma, and melanoma have the highest frequencies of *FBX4* mutations with rates below 2.5% (Fig. 3B).

In addition to mutations, other mechanisms involving microRNA, long noncoding RNAs, and specific oncogenic signaling pathway activation, can inactivate *FBXW7* function in cancer cells [58,59]. We assume that these tumors with inactivated SCF E3 ubiquitin ligases, at least some if not all, may not be sensitive to mTOR-targeted cancer therapy. Further study in this regard is warranted.

### Upregulation of programmed death-ligand 1 (PD-L1) in cancer cells during mTOR-targeted therapy

One critical mechanism accounting for immunosuppression and immune escape of cancer cells is the expression of PD-L1 on cancer cells, which can avoid elimination by immune cells, including T cells, natural killer (NK) cells, and macrophages via interacting with program death-1 (PD-1) [60–62]. Immunotherapy targeting the PD-1/PD-L1 immune checkpoint has shown promising benefits against several types of cancer and changed the landscape of cancer therapy [63–65].

One early report showed that oncogenic activation of the

Akt/mTOR pathway upregulated PD-L1 expression, thereby promoting immune escape in NSCLC; accordingly, inhibition of this signaling pathway decreased the PD-L1 expression [66]. However, our recent study failed to reproduce this finding; rather we generated opposite results, that is, inhibition of PI3K/Akt or mTORC1/p70S6K signaling with different corresponding inhibitors increases the PD-L1 levels in NSCLC and other cancer cell lines expressing basal levels of PD-L1. These data were subsequently confirmed using a genetic approach (e.g., shRNA) [67]. In agreement with our findings, another study reported that everolimus increased the cell surface PD-L1 levels in human and murine renal carcinoma cells [68]. In a recent study with renal cell carcinoma cells, inhibition of mTOR with rapamycin and Torin 1 (a TORKinib) increased PD-L1 expression, conferring resistance of renal cell carcinoma to mTOR inhibition [69]. Hence, we believe that inhibition of the PI3K/mTOR pathway actually elevates PD-L1 levels at least in some cancer cells.

Given that PD-L1 expression in cancer cells is a critical mechanism that contributes to immunosuppression and immune escape through its interactions with PD-1, it is highly likely that elevated PD-L1 levels on the cancer cell surface may protect cancer cells from being eliminated by immune cells by promoting the immune escape of cancer cells during mTOR-targeted cancer therapy. Therefore, whether PD-L1 induction by mTORC1 inhibition contributes to the modest anticancer activity of rapalogs or limited anticancer activity of TORKinibs is a logical and interesting question to address. This process may represent another important mechanism accounting for the limited success in targeting mTOR for cancer therapy, particularly *in vivo* or in human. Further study in this direction is warranted.

### Other factors that may potentially affect the response of cancer cells to mTOR-targeted cancer therapy

In addition to the mechanisms discussed above that may specifically account for the limited efficacy of mTOR-targeted cancer therapy in clinics, other intrinsic features of cancer cells, such as EMT, cancer stem cell (CSC) phenotype, ATP binding cassette (ABC) transporter expression, and mTOR mutations, may affect the response of cancer cells to mTOR inhibitors (see review by Hua *et al.* [70]).

Some of these factors, such as EMT, CSC, and ABC transporters, may not be mechanisms specific to mTOR inhibitors; they can also be cell type- and inhibitor-dependent. For example, while breast cancer cell lines with high EMT feature are relatively insensitive to rapamycin



## Opportunities for enhancing mTOR-targeted cancer therapy

Strategies toward induced induction of survival signaling pathways, including PI3K/Akt, MEK/ERK, and Mnk/eIF4E, for enhancing mTOR-targeted cancer therapy have been discussed in detail in our previous review [9].

As discussed above, GSK3 activity is required for mTOR inhibitors, including rapalogs and TORKinibs, to induce the degradation of certain oncogenic proteins, such as cyclin D1, Mcl-1, and SREBP1, and to inhibit tumor cell growth [34,35,39,40]. Moreover, basal p-GSK3 levels are inversely correlated to cell sensitivity to mTOR inhibitors in human NSCLC cells [34,35]. Hence, the activation of GSK3 is speculated to enhance the anticancer efficacy of mTOR inhibitors, particularly in tumors with low GSK3 activity (or highly phosphorylated GSK3). However, directly targeting GSK3 activation in cancer therapy is difficult considering the complex roles of GSK3 in regulation of cancer development [28,29]. Moreover, GSK3 activation may cause other issues related to neurodegenerative diseases [78,79].

PI3K/Akt and MEK/ERK/p90RSK signaling pathways positively regulate GSK3 phosphorylation [30–32]. Targeting these signaling pathways may indirectly activate GSK3, thereby achieving enhanced therapeutic effects when combined with mTOR-targeted therapy. Some published studies reported the enhanced antitumor effects of mTOR inhibition *in vitro* and *in vivo* when combined with PI3K/Akt inhibition [25,80–83] or with MEK/ERK inhibition [84–87] despite having different rationales and mechanisms. Whether GSK3 activation contributes to these enhanced antitumor effects needs further investigation.

Although rapalogs have long been regarded as immunosuppressants and are used in organ transplantation, many recent studies suggest that the effects of rapamycin on immune function, particularly T cell activation, are highly dose dependent [88–91]. Low doses of rapamycin stimulate memory CD8<sup>+</sup> T cell generation and enhance the memory T cell response to viral infection or cancer in mice [92]. Similarly, everolimus at low doses improves immune function in elderly volunteers, as assessed by their response to influenza vaccination [93]. A human study in patients with renal cancer showed that the therapeutic efficacy of rapalogs is associated with modulation of antitumor T cell immunity. Patients treated with rapalogs showed better clinical responses if they presented a shift toward decreased Treg levels and high expansion of antitumor Th1 or activated CD8<sup>+</sup> T cells [94]. Several works have shown that rapalogs enhance response to various modalities of immunotherapy, including adoptive cell therapy and cancer vaccines [95–98].

In theory, the elevation of PD-L1 in cancer cells might

increase the availability of epitopes for anti-PD-L1 antibody to bind, which could enhance the efficacy of anti-PD-L1 therapy. Therefore, the positive impact of mTOR inhibition on generation and activation of memory CD8<sup>+</sup> T cells and on elevation of PD-L1 in cancer cells may constitute a strong scientific rationale for combining mTOR inhibition with PD-1 or PD-L1 blockade immunotherapy as an effective strategy to enhance therapeutic efficacy. The combination of everolimus or rapamycin with PD-L1 blockade significantly enhanced antitumor activity compared with each single agent treatment against renal and oral cavity cancers [68,99]. The combination of rapamycin with anti-PD-1 antibody also enhanced the reduction of lung tumor burden in a mutant *KRAS*-driven mouse lung cancer model [66]. Another recent study showed that simultaneous inhibition of mTOR with temsirolimus and blockade of PD-L1 enhanced CD8<sup>+</sup> cytolytic function and tumor suppression in a xenografted mouse model of renal cell carcinoma based on the rationale of mTOR inhibition-induced upregulation of PD-L1 expression [69].

## Summary and conclusions

On top of the early findings on induced activation of PI3K/Akt, MEK/ERK, and Mnk/eIF4E survival signaling pathways that blunt the efficacy of mTOR-targeted cancer therapy, recent studies have further shown the essential role of GSK3 in mediating cancer cell response to mTOR inhibitors and the potential negative effect of induced upregulation of PD-L1 in cancer cells on mTOR-targeted cancer therapy. These findings not only suggest the complexity of mTOR networks but also have complicated the story of mTOR-targeted therapy. Full elucidation of the biology of mTOR networks and potential resistance mechanisms in mTOR-targeted cancer therapy will eventually help us to better utilize mTOR inhibitors in a rational manner for effective treatment of cancer. The hope is there, but there is a long way ahead of us.

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## Compliance with ethics guidelines

Shi-Yong Sun declares no conflict of interest. This manuscript is a review article and does not involve a research protocol requiring approval by the relevant institutional review board or ethics committee.

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