

# Electrochemical methods for determination of thymine in medical pipefish samples based on HPM $\alpha$ FP/Ppy/GCE-modified electrode

Hui Han · Jin-Zhou Li · Yang Li · Xiao-Zhe Pang

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**Abstract** A sensitive electrochemical method was developed for the voltammetric determination of thymine at a composite film-modified electrode 1-phenyl-3-methyl-4-(2-furoyl)-5-pyrazolone (HPM $\alpha$ FP)/polypyrrole (Ppy)/glassy carbon electrode (GCE). The electrochemical parameters of thymine were investigated by cyclic voltammetry and differential pulse voltammetry. In pH=7.4, one sensitive oxidation peak of thymine with  $E_{pa}$ =0.968 V was observed on the HPM $\alpha$ FP/Ppy-modified electrode. The difference of peak potential ( $\Delta E_{pa}$ ) was 188 mV lower than that for bare GCE. Compared to the bare GCE and Ppy/GCE, the HPM $\alpha$ FP/Ppy/GCE-modified electrode showed an excellent electrocatalytic effect on the oxidation of thymine and displayed a shift of the oxidation potential in the negative direction with significant increase in the peak current. Under the optimum condition, the concentration calibration range and detection limit are  $2 \times 10^{-6}$ – $1 \times 10^{-4}$  and  $4.85 \times 10^{-7}$  M for thymine. This developed method had been applied to the direct determination of thymine in medical pipefish samples with satisfactory results.

**Keyword** Acylpyrazolone · Polypyrrole · Thymine · Modified electrode

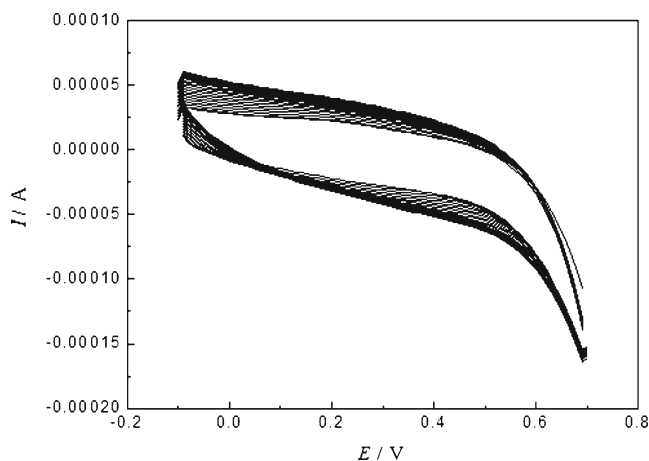
## Introduction

Pyrimidine and purine derivatives play a crucial role in various biological heritage information storage and in

protein biosynthesis processes [1]. In particular, the nucleotides constituting by thymine, cytosine, adenine, and guanine represent the monomer units of deoxyribonucleic acid (DNA), whereas ribose and phosphate groups of the nucleotide units have a structure role. The abnormal changes of these bases in organisms suggest the deficiency and mutation of the immunity system and may indicate the presence of various diseases [2]. Hence, the quantitative analysis of thymine is very essential and has great significance to bio-science and clinical iatrolgy.

Investigations of the redox behavior of the biologically occurring compounds by means of electrochemical techniques have the potential for providing valuable insights into biological redox reactions of such biomolecules. In recent years, a series of methods have been established to indagate the content of the thymine and cytosine in samples. The electrochemical studies have been undertaken concerning their redox behavior at mercury electrode, yet thymine was reported to show no polarographic reduction wave. In addition, the pyrimidine forms sparingly soluble compounds with Hg(II) at the surface of mercury electrode, which allows their determination by cathodic stripping voltammeters [3–6]. Due to low signal-to-noise ratio and their extreme positive oxidation potentials, thymine and cytosine have been assumed to exhibit no electrochemical activity at graphite electrodes [7, 8] or carbon-based electrodes in general [9, 10]. Brett reported oxidation of thymine and cytosine at glassy carbon electrode in alkaline pH solution after ultrasonic pretreatment [11]. However, low-stability response is the main problem when conventional carbon electrodes have been used. Furthermore, purine and pyrimidine bases adsorbed strongly at carbon electrodes [12–14], which reduced reproducibility of the analytical

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**Fig. 1** 0.1 M  $\text{H}_3\text{PO}_4$ - $\text{NaH}_2\text{PO}_4$  (pH=3.0)+0.1 M py, the scan rate: 0.25 V/s and scanning potential range, -0.1~0.7 V

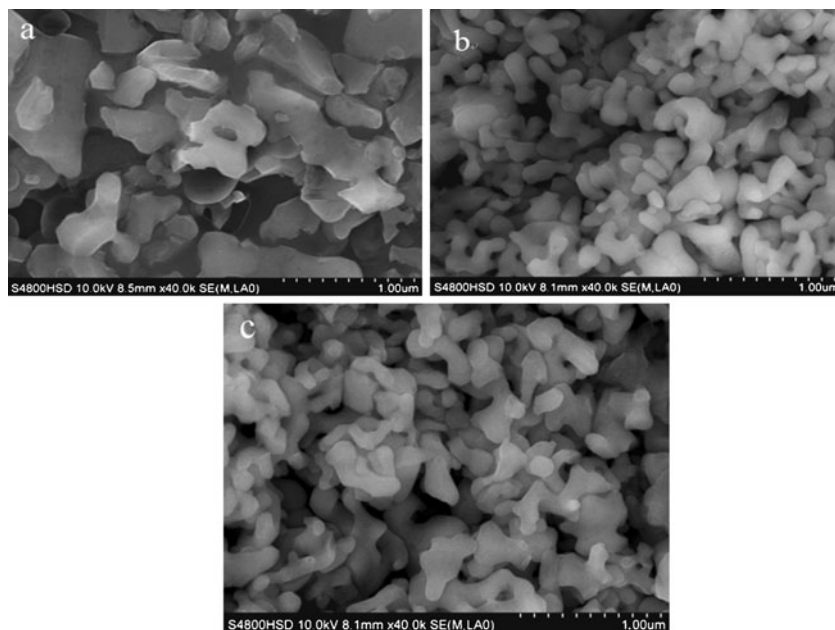
method. After an ultrasonic pretreatment of glassy carbon electrodes, both thymine and cytosine undergo well-defined oxidation [15].

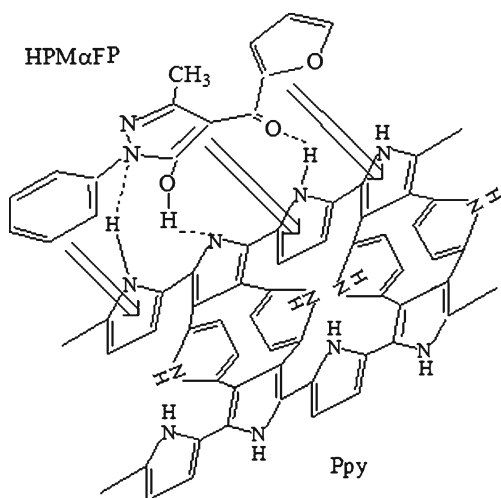
Electropolymerization is a simple but powerful method in targeting different selective modified electrodes with desired matrices. The electroactive polymers material has received considerable attraction in recent years. Numerous conjugated polymers were electrochemically synthesized for their application in the chemical and biochemical sensor devices [16]. These polymers exhibit an interesting enhancement in the electrocatalytic activity towards the oxidation or reduction of several biochemical compounds [17], where some functional groups in the polymers act as catalyst [18–20]. The word “enhanced electrocatalytic activity” could be explained as: both increase in peak current and lower in

over potential [21]. In this work, polypyrrole (Ppy) was prepared as one of the most extensively used conducting polymers [22] because of its ease of fabrication, high conductivity, good biocompatibility, and low cost [23–26]. The existence of surface positive charge on the Ppy film could provide a selective interface for the molecular interaction. The amine group ( $-\text{NH}-$ ) on the pyrrole ring may lead to enhancement of biomolecular sensing [27–29]. In addition, the sensitivity of a Ppy-modified electrode could be significantly improved when it is doped with suitable active materials such as peptide [30] and DNA [31]. Nowadays, this polymer becomes one of the major tools for nanobiotechnological applications [32]. Various approaches have been considered for the synthesis of Ppy, including chemical and electrochemical methods [33]. It was reported that the catalytic activity of Ppy films depends on the synthesis conditions [33, 34].

1-Phenyl-3-methyl-4-(2-furoyl)-5-pyrazolone is an interesting class of  $\beta$ -diketones, containing a pyrazole fused to a chelating arm. The compounds have played an important role in including coordination chemistry and have been widely used in potential antitumor agents areas. The neutral acylpyrazolones may exist with several tautomeric forms as reported [35, 36]. The enolate form of the acylpyrazolones present weakly acidic characteristics, as a surfactant, which has practical significance in polarographic determination of trace metals and makes it suitable for electrochemical analysis [37, 38]. Due to the aromaticity, 1-phenyl-3-methyl-4-(2-furoyl)-5-pyrazolone (HPM $\alpha$ FP) can be adsorbed onto the surfaces of the Ppy through  $\pi$ - $\pi$  force and hydrogen bond to form the composite film [39]. To the best of our knowledge, there are no reports on the fabrication of the

**Fig. 2** Scanning electron microscope images of the surface of **a** Ppy /GCE, **b** HPM $\alpha$ FP/GCE, **c** HPM $\alpha$ FP/Ppy/GCE





**Scheme 1** The interaction between polypyrrole and HPM $\alpha$ FP

HPM $\alpha$ FP/Ppy/GCE composite film-modified electrode and its response on thymine.

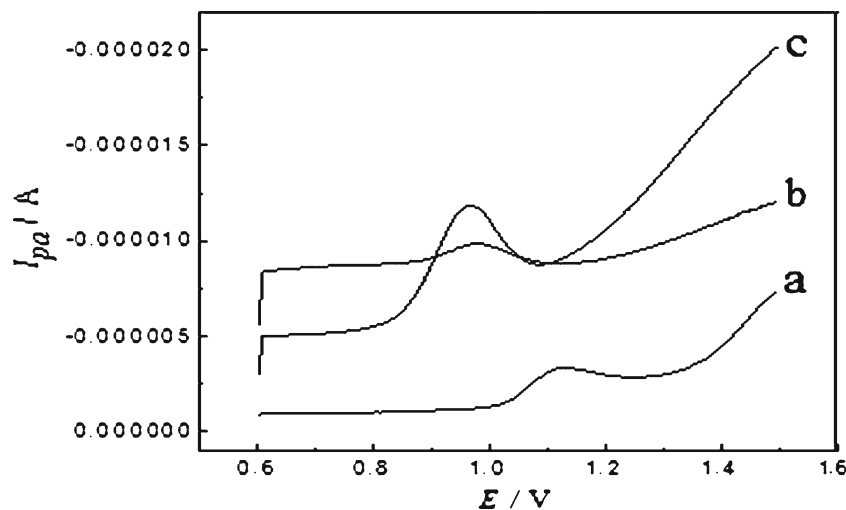
In our previous work, HPM $\alpha$ FP-modified GCE (HPM $\alpha$ FP/GCE) was applied to determination of amino acid and purine [40, 41]. In order to extend of the application of HPM $\alpha$ FP-modified electrode in the bioelectrochemistry, we are reporting a novel electrochemical strategy to prepare the composite film glassy carbon electrode of HPM $\alpha$ FP and Ppy, which shows the great capability of sensitive and selective determination of thymine.

## Experimental

### Apparatus

All the electrochemical measurements that contain cyclic voltammetric and differential pulse voltammetry (DPV)

**Fig. 3** DPV of  $1.0 \times 10^{-4}$  M thymine (pH=7.4) at *a* bare GCE, *b* Ppy-modified GCE, *c* Ppy/HPM $\alpha$ FP GCE



were performed in an analytical system model CHI-650A electrochemical workstation (Shanghai Chenhua Instrument Company, China). A conventional three-electrode system was used with a platinum wire as an auxiliary electrode and a saturated calomel electrode as reference electrode. The working electrode was either an unmodified GCE or a polymer film modified electrode ( $\varphi$ 4 mm) for the electrochemical measurements.

### Materials

Thymine was obtained from National Institute for the control of pharmaceutical and biological products (Shanghai, China, [www.reagent.com.cn](http://www.reagent.com.cn)). A  $1 \times 10^{-2}$  M stock solution of thymine was done with twice distilled water in a refrigerator at approximately 277 K and remained stable for at least 1 month. Working standard solutions were prepared by suitable dilution of the stock standard solution. Pyrrole (analytical reagent, for short py, made in Shanghai reagent company, China) was made with 0.1 % water solutions.

HPM $\alpha$ FP was synthesized according to the method proposed by Dong [42 (yield, 73 %; mp, 373.5–374.5 K) with further purification, and a standard solution (0.01 M) of HPM $\alpha$ FP was prepared by anhydrous ethanol solution. Other chemicals were of analytical reagent grade and used without further purification. Of the phosphate buffer solution (PBS) of various pH values, 0.10 M was used as supporting electrolyte. All solutions were prepared with double-distilled water and experiments were conducted at the room temperature ( $298 \pm 2$  K).

### Preparation of the modified electrode

Prior to modification, the bare GCE was polished successively with 1.0 and 0.3  $\mu$ m alumina slurry on chamois leather, respectively. Electrode was rinsed with double-



**Scheme 2** The interaction between thymine and HPM $\alpha$ FP/Ppy/GCE

distilled water. After that, the electrode was put into nitric acid (1:1), anhydrous ethanol, in order to remove adhered alumina and rinsed thoroughly with distilled water. Glass carbon electrodes after polishing and ultrasonic cleaning were put into 0.5 M H<sub>2</sub>SO<sub>4</sub> solution, then it was polarized to deal with cyclic voltammetry scanning, until a reproducible voltammogram was obtained (about 10 min). The electrode pretreated was put in pure water for standby. The electrode set into system: 0.1 M H<sub>3</sub>PO<sub>4</sub>–NaH<sub>2</sub>PO<sub>4</sub> (pH=3.0)+0.1 M py mixed solution, connecting three electrodes system and electrochemistry workstation, controlling the scan rate of 0.25 V/s and scanning potential range of –0.1~0.7 V. The blue–purple polymer film was obtained by cyclic scanning 20 times (the continuous cyclic voltammograms of polymerization process; Fig. 1). Ppy prepared in this conditions was very steady, the properties of Ppy has not evidently changes after putting in PBS (pH=3.0) after 4 weeks. Finally, the GCE was carefully coated with 1  $\mu$ L of HPM $\alpha$ FP solution and left for the solvent to evaporate at room temperature in the air. When the modified electrode completely dried, it can be used. The HPM $\alpha$ FP coated on the Ppy/GCE was also very steady in the PBS solution.

#### Experimental procedure

PBS 0.1 M (pH=7.4) with certain amount of thymine was transferred into a cell, and the three-electrode system was installed in it. High-purity N<sub>2</sub> was used to remove oxygen. Differential pulse voltammetry was recorded between 0.6

and 1.5 V. All experiments were carried out at room temperature (298 $\pm$ 2 K).

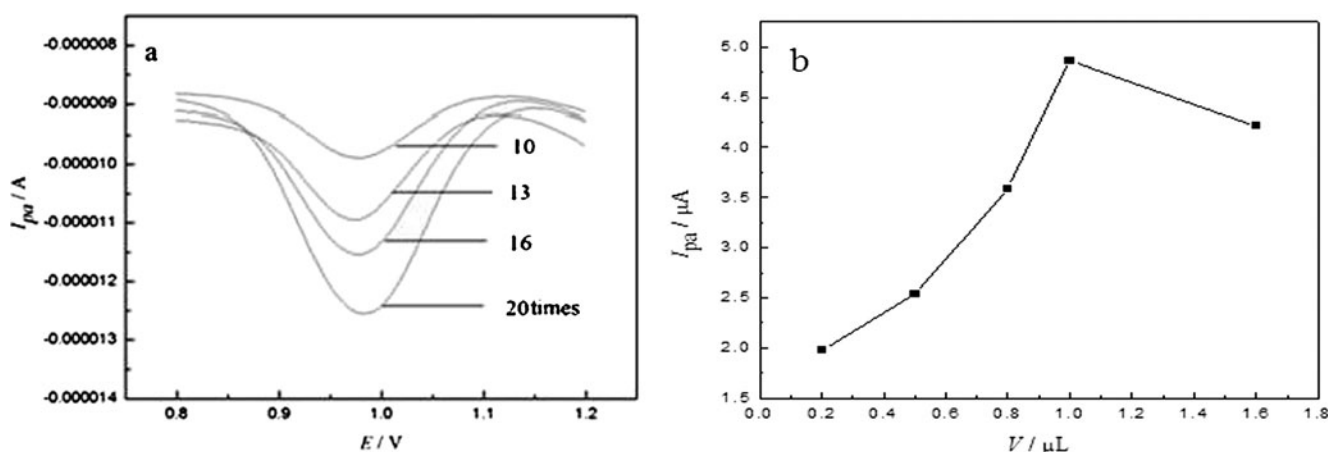
#### Preparation of medical pipefish solution

The medical pipefish (*Syngnathus acus*; Haikou, China) was ground into powder in agate mortar. The sample powder (1.450 g) was weighed and extracted with 25 mL double-distilled water by ultrasonic wave for 90 min. The clear treated solution (10 mL) was transferred to a 25 mL volumetric flask and diluted to the mark with double-distilled water as stock for the experiments.

## Results and discussion

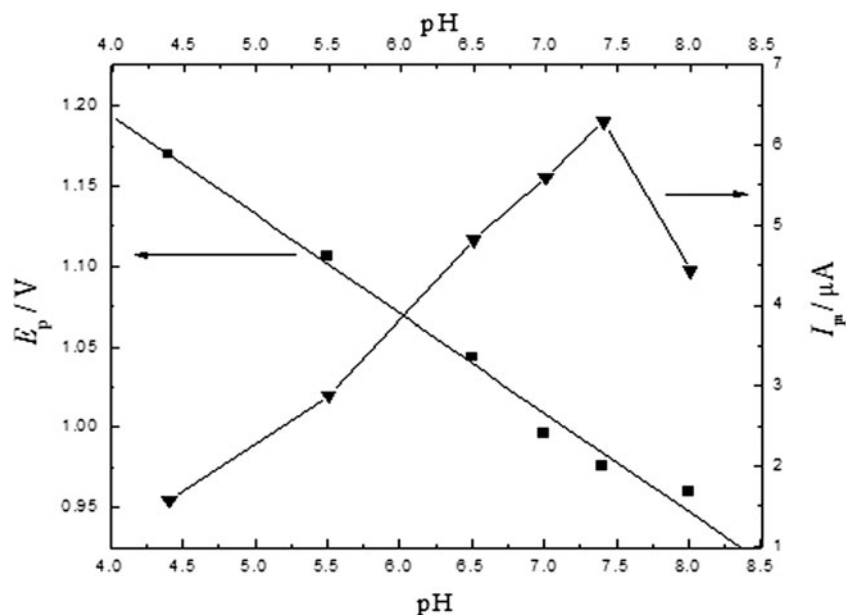
#### The microstructure of the modified electrode

The microscopic structure of modified electrodes was characterized using scanning electron microscope images. Figure 2 exhibits typical images of polypyrrole, HPM $\alpha$ FP, and HPM $\alpha$ FP/Ppy films coated onto GCE. As it can be clearly seen, Ppy-modified electrode (Fig. 2a) surface shows a uniform formation and arranged flakes of polymer layers. The surface area of the modified electrode was substantially increases. HPM $\alpha$ FP (Fig. 2b) was dispersed over the glassy carbon surface, and HPM $\alpha$ FP adsorbed on the surface of Ppy at the HPM $\alpha$ FP/Ppy film (Fig. 2c). The reasons were concluded in the following aspects: (1) At the polypyrrole and HPM $\alpha$ FP, the active functional groups maybe existed, such as –NH–, carbonyl, and hydroxy. The intermolecular effect (hydrogen bond) was formed between Ppy and HPM $\alpha$ FP; (2) Some aromaticities was consistent in HPM $\alpha$ FP containing benzene, parazole, and furfuran. The  $\pi$ – $\pi$  interactions between the aromaticity of HPM $\alpha$ FP and pentagonal structure



**Fig. 4** a DPV of different Ppy amounts from 10 to 20 times; b influence of different HPM $\alpha$ FP amounts on the peak of current ( $I_{pa}$ ) of  $1.0 \times 10^{-4}$  M thymine in PBS (pH7.4)

**Fig. 5** Influence of pH on the peak current ( $I_{pa}$ ) and peak potential ( $E_{pa}$ ) of thymine



of Ppy were attributed to the easy arrival of HPM $\alpha$ FP to the surface of Ppy [39] (as shown in Scheme 1).

**Electrochemical behavior of the modified electrode**

The differential pulse voltammograms of thymine at a bare GCE-, Ppy/GCE-, and HPM $\alpha$ FP/Ppy-modified GC electrode in phosphate buffer (pH=7.4) were shown in Fig. 3. It can be seen that the thymine oxidation peak at the bare GCE and Ppy/GCE was weak and broad due to slow electron transfer, while the response was considerably improved at the HPM $\alpha$ FP/Ppy/GCE. The peak potential at above three electrodes were at different position, at the bare GCE, an irreversible oxidation peak appeared at about 1.156 V (Fig. 3a). At the Ppy/GCE, the oxidation peak is located at 0.984 V (Fig. 3b). The oxidation peak potential ( $E_{pa}$ ) was negatively shifted for 0.172 V and the oxidation peak current ( $I_{pa}$ ) increased clearly. Both the oxidation peak current and potential of thymine were increased and moved negatively 0.968 V slightly at HPM $\alpha$ FP/Ppy/GCE-modified electrode (Fig. 3c).

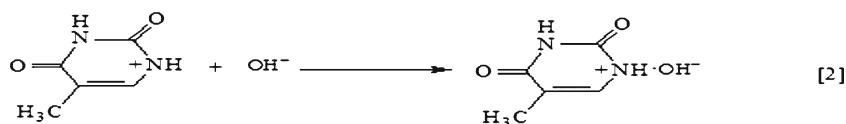
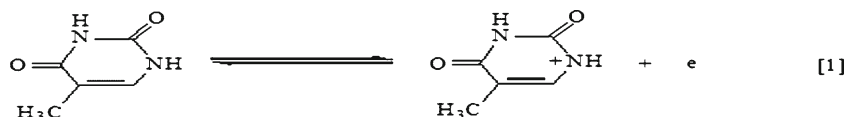
From the comparison of voltammograms of thymine, it can be observed that the peak current at HPM $\alpha$ FP/Ppy/GCE was more than two times higher than the sum of response currents at bare GCE or Ppy/GCE. The above results indicated that the

composite film-modified electrode exhibited the synergistic effect including high conductivity, fast electron transfer rate, and inherent catalytic ability [43]. This phenomenon was probably due to HPMaFP contained some oxygen functional groups such as carbonyl, which was propitious to thymine oxidation, these oxygen functional groups could form hydrogen bonds with the amidogen of thymine. Moreover, the hydrogen bond maybe existed between Ppy and HPM $\alpha$ FP or thymine and Ppy, which can remarkably increase the accumulation amount of thymine (as shown in Scheme 2).

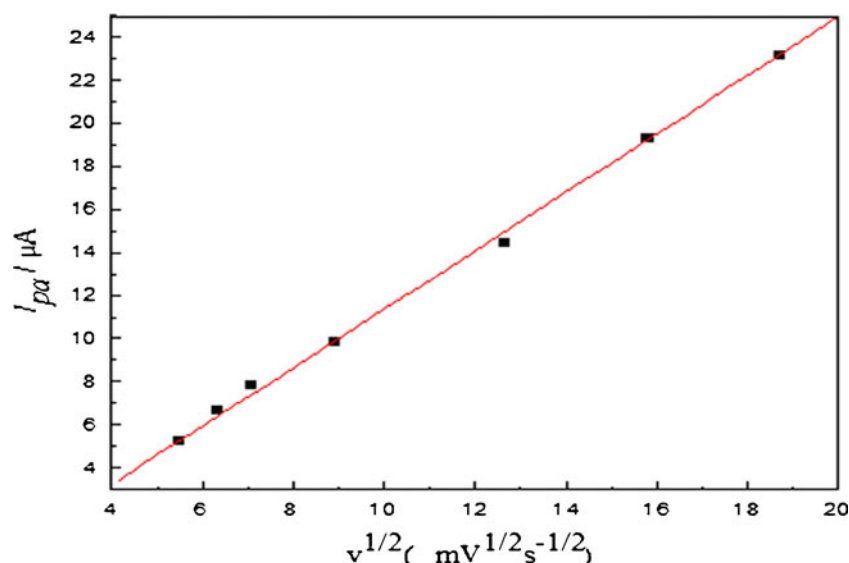
**Effect of Ppy–HPMaFP amount**

The relationship between the amount of Ppy–HPM $\alpha$ FP dispersion on the GCE surface and the oxidation peak current of thymine was investigated. The results showed that the variation of oxidation peak current of thymine was as a function of the amount of Ppy–HPM $\alpha$ FP (Fig. 4). When the amount of Ppy increased from 10 to 20 times, the oxidation peak current increased notably (Fig. 4a). However, the oxidation peak current showed conversely gradual decline when the amount of Ppy exceeds 20 times. The reason is that too thick polypyrrole film may be more easily abraded from the surface of GCE. So, the polymerization

**Scheme 3** The mechanism of thymine oxidation at HPM $\alpha$ FP/Ppy/GC electrode



**Fig. 6** Plot of  $I_{pa}$  vs. square root of sweep rate in pH 7.4 PBS. The concentration of thymine is 1 mM



scanning polypyrrole intercalative 20 times was selected. On the other hand, the oxidation peak current gradually increased with the augment of quantity of HPM $\alpha$ FP. The results were shown in Fig. 4b. When the HPM $\alpha$ FP was beyond 1.0  $\mu\text{L}$ , the peak current decreased. This was due to the surface of Ppy/GCE casted too thick to impede the charge exchange. To sum up, 20 times polypyrrole and 1.0  $\mu\text{L}$  HPM $\alpha$ FP was selected in the subsequent analytical experiments and the maximum electrocatalytic response on thymine was achieved.

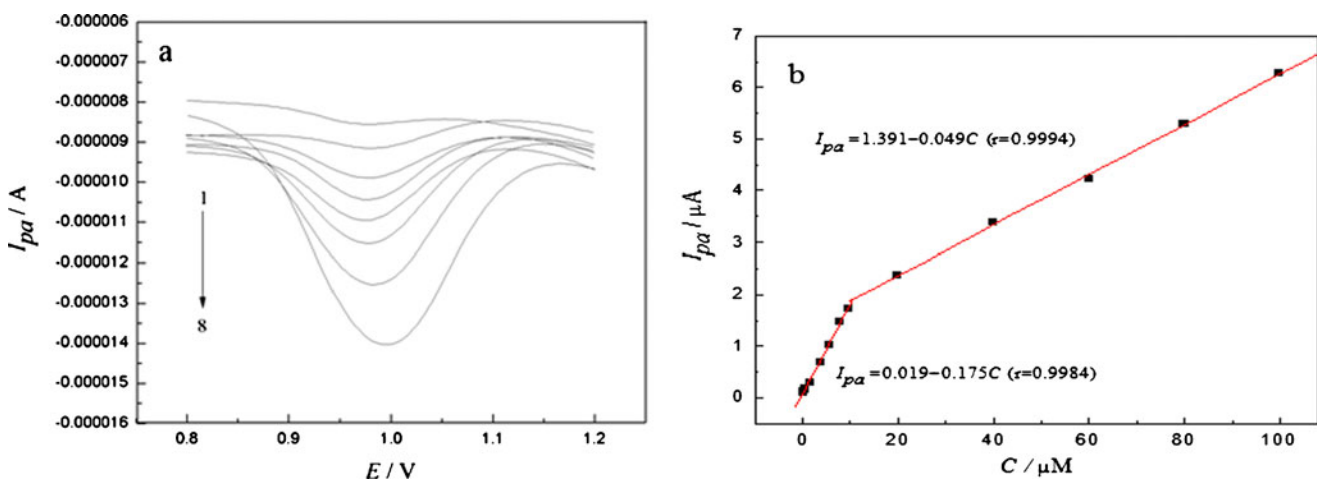
#### Effect of solution pH

Influence of pH on electrochemical behavior of  $1.0 \times 10^{-5} \text{ M}$  thymine at HPM $\alpha$ FP/Ppy/GCE was studied in the pH range from 5.0 up to 8.0. Both the peak current and potentials were depended on pH values of solution. The results were shown in Fig. 5. With the increment of pH, the oxidation peak current of

thymine at the HPM $\alpha$ FP/Ppy/GCE-modified electrode decreased slightly and the peak potential was found to shift negatively. The pH values around 7.4 should provide the highest oxidation currents justifying the selection of pH 7.4 for the subsequent experiments using the HPM $\alpha$ FP/Ppy/GCE film electrode. According to the equation:  $-59\chi/n = -61$ , where  $\chi$  is the hydroxyl ion participating the electrode reaction and  $n$  is the electron-transfer number, the slope of the  $E_{pa}$ -pH is 61 mV. The number of OH's and transferred electrons are 1. So, the loss of electrons was accompanied by the loss of an equal amount of hydroxyl ion and  $\chi = n = 1$ . The reaction mechanism for thymine was shown in Scheme 3 [44].

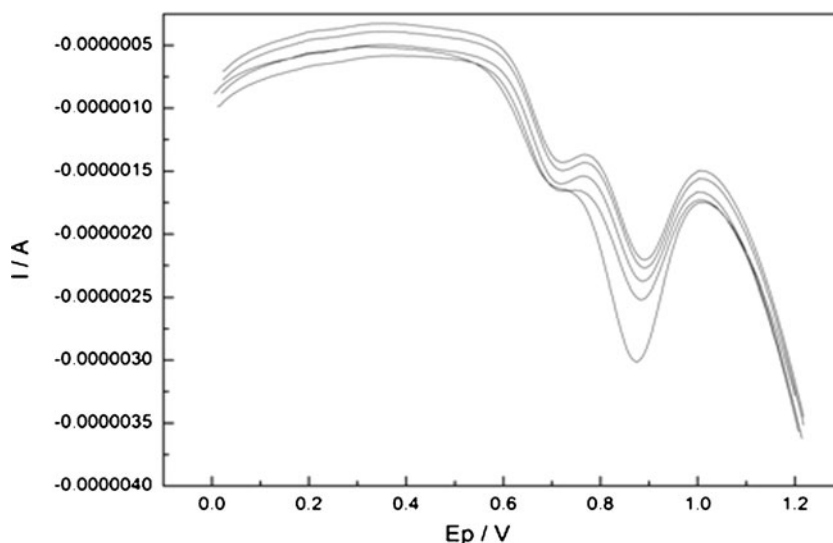
#### Effect of sweep rate

The effect of scan rate on the voltammetric response for the oxidation of  $1.0 \times 10^{-4} \text{ M}$  thymine on the HPM $\alpha$ FP/Ppy/GCE was investigated from 30 to  $350 \text{ mV s}^{-1}$ . The cyclic



**Fig. 7** a DPV of HPM $\alpha$ FP/Ppy/GCE in 0.10 M PBS (pH 7.4) containing different concentration of thymine in the range of 2–100  $\mu\text{M}$ ; b plot of the peak current vs. the concentration of thymine. Pulse width=0.05 s, amplitude=0.05 V, sample period=0.0167 s, and pulse period=0.2 s

**Fig. 8** DPVs obtained for the increment of 2 μM thymine to 200 μM xanthine in 0.1 M PBS at HPMαFP/Ppy/GC electrode



voltammetric results demonstrated that the peak current varied linearly with the square root of the scan rate according to the equation:  $I_{pa}(A) = -1.923 + 1.333v^{1/2}$ , with an excellent linear correlation coefficient of 0.9990. Such behaviors indicating that the HPMαFP/Ppy composites could efficiently accelerate the electron transfer at the GCE with a diffusion-controlled process (Fig. 6).

**Determination of thymine**

DPV technique was employed to develop a voltammetric method for determination of thymine. Under the optimal conditions, voltammograms at these concentrations were shown in Fig. 7a, and two linear calibration graphs were obtained (Fig. 7b). The initial linear portion increased from  $2 \times 10^{-6}$ – $1 \times 10^{-5}$  M with a linear regression equation of  $I_{pa}(A) = 0.175C + 0.019$  ( $r = 0.9984$ ). The second linear segment is from  $1 \times 10^{-5}$  M up to  $1 \times 10^{-4}$  M with a linear regression equation of  $I_{pa}(A) = 0.049C + 1.391$  ( $r = 0.9994$ ). The detection limit was found to be  $4.85 \times 10^{-7}$  M. This detection limit was lower, and the sensitivities were higher than those obtained by other electrochemical methods.

**Determination of thymine in the presence of xanthine**

The main objective of the present study is to selectively determine thymine in the presence of xanthine. Figure 8 showed the

DPVs for the increment of 2 μM thymine in the presence of 200 μM xanthine in 0.1 M PBS (pH=3.0). During the anodic sweep from 0.1 to 1.2 V, two oxidation peaks at 0.687 V (xanthine) and 0.968 V (thymine) were observed at the HPMαFP/Ppy/GCE. The oxidation current of thymine was linearly decreases by its concentration from 2 to 20 μM with a correlation coefficient of  $r = 0.9996$  ( $I_{pa}(\mu A) = 15.39998C - 27.45901$ ). These results indicated that HPMαFP/Ppy/GCE was highly selective towards the oxidation of thymine and detection of very low concentration thymine was possible in the presence of an excess of xanthine.

**Stability and repeatability of the modified electrode**

In order to investigate the repeatability of HPMαFP/Ppy/GCE, the differential pulse voltammetry values for  $1 \times 10^{-4}$  M thymine were recorded in 6-min intervals. It was shown that the peak of thymine nearly remained the same, and the relative standard (RSD) gave the result of 1.6 %, indicating excellent repeatability of the modified electrode.

The stability of the modified electrode was also investigated over a period of 7 days. The modified electrode was used daily and stored in air. The peak potential for thymine was unchanged, and the current signals showed only less than 3 % decrease of the initial response, which suggested that the HPMαFP/Ppy/GCE possessed good stability for the detection of adenine.

**Table 1** Results of analysis of thymine in medical pipefish samples ( $n=5$ )

Sample	Added ( $\times 10^{-6}$ M)	Expected ( $\times 10^{-6}$ M)	Found ( $\times 10^{-6}$ M)	Recovery (%)	RSD (%)
1	0.0	1.66	1.70	–	1.8
2	1.0	2.66	2.68	100.7	1.7
3	2.0	3.66	3.63	99.2	1.3
4	3.0	4.66	4.69	100.6	1.9
5	4.0	5.66	5.67	100.2	1.1

## Interferences

The effects of interfering ions or biomolecules on the determination of thymine were investigated. Under experimental optimal conditions, the proposed method was used for the determination of thymine at  $1 \times 10^{-4}$  M level, some interference experiments were carried out. The tolerance limit was taken as approximately 5 % relative error of the foreign substance. The experimental results showed that at least a 200-fold excess of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Cl}^-$ ,  $\text{H}_2\text{PO}_4^-$ ,  $\text{HPO}_4^{2-}$ ,  $\text{PO}_4^{3-}$ , a 100-fold excess of DL-serine, L-leucine, L-lysine, a 50-fold excess of cytosine, glucose, lysine, and a 20-fold excess of DL-phenylalanine, citric acid did not cause any observable interference for the determination of thymine.

## Analysis of thymine in medical pipefish samples

The HPM $\alpha$ FP/Ppy/GCE was used to detect thymine in the medical pipefish (*S. acus*) samples. The preparative procedures for the *S. acus* solution were followed as described in the “Experimental” section. The determination of thymine concentration was performed by the standard addition method. The results were summarized in Table 1 with the recovery and RSD respectively from 99.2 to 100.7 and 1.1 to 1.9. The data showed that the proposed method can be used efficiently for the detection of thymine in medical samples.

## Conclusions

In this work, an easily prepared HPM $\alpha$ FP/Ppy composite film-modified electrode was firstly performed to investigate the electrochemical behavior of thymine in detail. The electrode reaction was a diffusion-controlled irreversible process with one electrons and one hydroxyl ion. The HPM $\alpha$ FP/Ppy/GCE exhibited high electrocatalytic activities towards the oxidation of thymine by significantly decreasing their oxidation overpotentials and enhancing the peak currents, and used for the determination of thymine in the presence of high concentration of xanthine by DPV. Also, the proposed method offers important advantages such as easy fabrication, reproducibility, stability, and antijamming, which could be applied to the determination of thymine in medical pipefish samples with satisfactory results.

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## References

- Shen Q, Wang XM (2009) J Electroanal Chem 632:149
- Liu HY, Wang GF, Hu JS, Chen DL, Zhang W, Fang B (2008) J Appl Polym Sci 107:3173
- Palecek E, Jank B (1962) Arch Biochem Biophys 98:527
- Jank B, Palecek E (1964) Arch Biochem Biophys 105:225
- Dryhurst G, Elving PJ (1969) Talanta 16:855
- Palecek E (1966) J Mol Biol 20:263
- Smith D, Elving PJ (1962) Anal Chem 8:930
- Brabec V (1981) Bioelectrochem Bioenerg 8:437
- Hart JP (1990) Electroanalysis of biologically important compounds. Horwood, Chichester, 46
- Stul K, Pacfikovfi V (1987) Electroanalytical measurements in flowing liquids. Horwood, Chichester, 262
- Brett AMO, Piedade JAP, Silva LA, Diclescu VC (2004) Anal Biochem 332:321
- Yano T, Tryk DA, Hashimoto K, Fujishima A (1998) J Electrochem Soc 143:1870
- Wang HS, Ju HX, Chen HY (2001) Electroanalysis 13:1105
- Rao TN, Sarada BV, Tryk DA, Fujishima A (2000) J Electroanal Chem 491:175
- Brett AMO, Matysik FM (1997) J Electroanal Chem 429:95
- McMahon CP, Rocchitta G, Kirwan SM, Killoran SJ, Serra PA, Lowry JP, O'Neill RD (2007) Biosens Bioelectron 22:1466
- Becerik I, Kadirgan F (2001) Synth Met 124:379
- Selvaraju T, Ramaraj RR (2005) Electroanal J Chem 585:290
- Mao M, Zhang D, Sotomura T, Nakatsu K, Koshihara N, Ohsaka T (2003) Electrochim Acta 48:1015
- Yasuzawa M, Kunugi A (1999) Electrochem Commun 1:459
- Andrieux CP, Haas O, SavGant JM (1986) J Am Chem Soc 108:8175
- Adelaju SB, Wallance GG (1996) Analyst 121:699
- Hepel M (1998) J Electrochem Soc 145:124
- Kim DH, Richardson-Burns SM, Hendricks JL, Sequera C, Martin DC (2007) Adv Funct Mater 17:79
- Malitesta C, Palmisano F, Torsi L, Zamboni PG (1990) Anal Chem 62:2735
- Pernaut JM, Reynolds JR (2000) J Phys Chem B 104:4080
- Ulubay S, Dursun Z (2010) Talanta 80:1461
- Li J, Lin XQ (2007) Sensors Actuators B 124:486
- Li J, Lin XQ (2007) Anal Chim Acta 596:222
- Cui XY, Lee VA, Raphael Y, Wiler JA, Hetke JF, Anderson DJ, Martin DC (2001) J Biomed Mater Res Part A 56:261
- Li J, Wei W, Luo S (2010) Microchim Acta 171:109
- Malinauskas A, Malinauskiene J, Ramanavicius A (2005) Nanotechnology 16:51
- Schmidt HL, Gutberlet F, Schuhmann W (1993) Sensors Actuators B 13:366
- Ramanaviciene A, Ramanavicius A, Thomas DW (2004) Advance biomaterials for medical applications. Kluwer, Dordrecht, p 111
- Nishihama S, Hirai T, Komazawa I (2001) Ind Eng Chem Res 40:3085
- Marchetti F, Pettinari C, Pettinari R (2005) Coord Chem Rev 249:2909
- Ghoneim MM, El-Desoky HS, Amer SA, Rizk HF, Habazy AD (2008) Dyes Pigments 77:493
- Zhang D, Li JZ (2008) Anal Lett 41:2832
- Li FH, Chai J, Yang HF, Han DX, Niu L (2010) Talanta 81:1063
- Li XL, Li JZ (2011) Rev Anal Chem 30:23
- Song Y, Li JZ (2011) Instrum Sci Technol 39:261
- Dong XC, Liu FC, Zhao YL (1983) Acta Chim Sinica 41:848
- Song Y, Li JZ (2011) J Solid State Electrochem 16:693
- Wu SG, Zheng LZ, Rui L, Lin XQ (2001) Electroanal 13:967