ORIGINAL ARTICLE



Image-based hemodynamic simulations for intracranial aneurysms: the impact of complex vasculature

Franziska Gaidzik^{1,2} · Jana Korte^{1,2} · Sylvia Saalfeld^{1,3} · Gábor Janiga^{1,2} · Philipp Berg^{1,4}

Received: 12 July 2023 / Accepted: 6 December 2023 / Published online: 11 January 2024 © The Author(s) 2024

Abstract

Purpose Hemodynamics play an important role in the assessment of intracranial aneurysm (IA) development and rupture risk. The purpose of this study was to examine the impact of complex vasculatures onto the intra-vessel and intra-aneurysmal blood flow.

Methods Complex segmentation of a subject-specific, 60-outlet and 3-inlet circle of Willis model captured with 7T magnetic resonance imaging was performed. This model was trimmed to a 10-outlet model version. Two patient-specific IAs were added onto both models yielding two pathological versions, and image-based blood flow simulations of the four resulting cases were carried out. To capture the differences between complex and trimmed model, time-averaged and centerline velocities were compared. The assessment of intra-saccular blood flow within the IAs involved the evaluation of wall shear stresses (WSS) at the IA wall and neck inflow rates (NIR).

Results Lower flow values are observed in the majority of the complex model. However, at specific locations (left middle cerebral artery 0.5 m/s, left posterior cerebral artery 0.25 m/s), higher flow rates were visible when compared to the trimmed counterpart. Furthermore, at the centerlines the total velocity values reveal differences up to 0.15 m/s. In the IAs, the reduction in the neck inflow rate and WSS in the complex model was observed for the first IA (IA-A δ NIRmean = -0.07 ml/s, PCA.1 δ WSSmean = -0.05 Pa). The second IA featured an increase in the neck inflow rate and WSS (IA-B δ NIRmean = 0.04 ml/s, PCA.1 δ WSSmean = 0.07 Pa).

Conclusion Both the magnitude and shape of the flow distribution vary depending on the model's complexity. The magnitude is primarily influenced by the global vessel model, while the shape is determined by the local structure. Furthermore, intraaneurysmal flow strongly depends on the location in the vessel tree, emphasizing the need for complex model geometries for realistic hemodynamic assessment and rupture risk analysis.

Keywords Circle of Willis · Computational fluid dynamics · Hemodynamics · Intracranial aneurysms · Segmentation

Introduction

The circle of Willis (CoW) and its hemodynamics provide a representation of the global intracranial vascularization

Franziska Gaidzik and Jana Korte have contributed equally to the study.

Franziska Gaidzik franziska.gaidzik@ovgu.de

Research Campus STIMULATE, Otto-von-Guericke-University Magdeburg, Magdeburg, Germany

² Laboratory of Fluid Dynamics and Technical Flows, Forschungscampus STIMULATE, Otto-von-Guericke-University Magdeburg, Universitätsplatz 2, 39106 Magdeburg, Germany structure [1]. Knowledge about cerebral vascularization is beneficial in numerous ways for the medical community. Perosa et al. [2] have investigated the influence of hippocampal vascularization patterns on cognitive performance, but little is known about the relationship between cerebral blood flow, vascular profiles and cognition. The CoW is also a common location for the development of intracranial aneurysms (IAs) that can lead to fatal consequences when ruptured [3].

- ³ Laboratory of Simulation and Graphics, Otto-von-Guericke-University Magdeburg, Magdeburg, Germany
- ⁴ Department of Medical Engineering, Otto-von-Guericke-University Magdeburg, Magdeburg, Germany

High variability in the vascular anatomy necessitates subject-specific investigation of cerebral blood flow. Occlusion or absence of vessel branches can disrupt communication between the internal carotid and vertebral arteries [4]. Recent numerical studies on intracranial hemodynamics have associated parameters such as wall shear stresses (WSS) or oscillatory shear with pathological effects [5, 6]. However, models within these studies usually cover only one inlet and a limited number of outlets in the proximity of the IA. Furthermore, neuroradiologists rarely rely on image-based blood flow simulations due to the lack of patient-specific flow conditions and unrealistic geometry information [7], although numerous studies have investigated the influence of the inlet and outlet flow conditions on the intracranial hemodynamics [8–11].

Despite being acknowledged as a critical aspect for achieving realistic simulations [12, 13], previous studies on patient-specific hemodynamics have been constrained by limitations imposed on the complexity of the considered vascular geometry. Blood flow simulations for IAs typically include only the parental vessel branch and a short part of the adjacent structures [14]. More complex vessel trees (e.g., the CoW) were investigated, but with restricted regions of interest and a limited number of outlets [13, 15–17]. However, advancements in medical image acquisition allow for higher resolution and accurate segmentation of small vessel branches, enabling consideration of a larger number of outflow cross sections.

Following a sophisticated segmentation to generate a patient-specific complex CoW geometry (Fig. 1), this study analyzes the effect of the model complexity on the intravessel and intra-aneurysmal hemodynamics. High-resolution 7T time-of-flight (ToF) MRI data are used to create a patient-specific 3D surface mesh with 60-outlet vessel cross sections. Blood flow simulations are conducted on this complex geometry model and a trimmed version. Additionally, a pathological variant with two patient-specific intracranial aneurysms is created to examine flow alterations in a clinically relevant scenario. This assessment aims to evaluate the impact of model complexity on the accurate representation of flow- and shear-related parameters in intra-vessel and intra-aneurysmal flow.

Materials and methods

Reconstruction of the complex circle of Willis vasculature

The 3D surface segmentation was extracted from highresolution (0.32 mm isotropic) ToF imaging data from a healthy volunteer. The data were acquired with a 7 Tesla whole-body MRI system (Siemens Healthineers, Erlangen,



Fig. 1 Visualization of the complex (1: red) and trimmed (2: blue) physiological CoW models (top) and the pipeline to create the pathological model: The vessel part was cropped from the CoW, and the IA vessel was placed accordingly. The patient-specific IAs were cropped and scaled to fit the CoW vessel diameter (middle). Afterward, the edited IAs were joined to the CoW model and smoothed to fit. Visualization of the final pathological models (bottom), complex (3: red) and trimmed version (4: blue)

Germany) including prospective motion correction to prevent image blurring, hence preventing the loss of small vessels [18]. For the 3D models, a multiscale vessel enhancement filtering [19] was applied to the 7T ToF MRI data. Six scales $(\sigma = 1...6)$ were extracted and combined into an intermediate dataset by using the maximum of the six filter responses. Next, seed points were placed manually in the three inlet arteries within the intermediate dataset. All voxels with a maximum of 2% intensity changes (with respect to the seed points) are added to a mask starting from the three seed points. Based on this segmentation mask, marching cubes are applied to extract a 3D surface model. Laplacian smoothing is carried out to smooth the 3D surface model. Afterward, manual post-processing is applied to exclude artifacts or unconnected surface parts. Finally, the inlets and outlets of the 3D vessel model were cut perpendicular to the vessel centerline and extruded to reduce the influence of boundary conditions. The resulting, complex 60-outlet geometry was trimmed to a surface model including only the 10 most prominent outlet cross sections in order to generate a comparison that is equivalent to the current state-of-the-art computational

Table 1 Model parameters including the inlet cross-sectional area,mean volumetric inflow rate, number of outlets and sum of outlet cross-sectional areas of each of the three branches (ICAI, ICAr and BA)

	ICAr	ICAl	BA
Inlet area (mm ²)	8.76	2.07	2.00
Mean volumetric inflow rate (l/s)	3.434×10^{-3}	3.065×10^{-3}	2.332×10^{-3}
	Trim./Comp	Trim./Comp	Trim./Comp
Number of outlets	4/17	2/9	4/34
Summarized outlet area (mm ²)	21.76/16.96	8.86/26.85	15.17/27.47

The outlet parameters are divided into trimmed (trim.) and complex (comp.) model and vary among the three inlet branches

fluid dynamics (CFD) studies in a CoW geometry. Model parameters of the three branches right and left internal carotid artery (ICAr and ICAl) and the basilar artery (BA) are given in Table 1.

Generation of the pathological model

Two patient-specific IAs were manually incorporated into the healthy CoW model to analyze the impact of model complexity in a clinically relevant scenario. These IAs were selected from a previous international study (MATCH) [20]. The first IA (A) is located at the right middle cerebral artery (MCA) and the second IA (B) at the left MCA. The fusion process involved transforming the IAs to match their exact positions within both CoW models and scaling the IAs to ensure their parent vessel diameter corresponded to the CoW vessel diameter (see Fig. 1).

Numerical simulations

Four hemodynamic simulations were performed using the finite volume solver STAR CCM + (StarCCM + 2021.3 v16.6, Siemens Product Lifecycle Management Software Inc., Plano, TX, USA) based on the subject-specific geometry models. Blood was assumed to be Newtonian and incompressible, with a dynamic viscosity of 4 mPa·s and a density of 1055 kg/m³. Laminar flow and time-resolved simulations with a temporal discretization of 1 ms were conducted over three cycles, with the last cycle considered for evaluation. Spatial discretization was set at a base size of 0.125 mm, resulting in different cell counts for each model (physiological: complex model 4.7 million cells, trimmed model 2.5 million cells; pathological: complex model 6.3 million cells, trimmed model 2.6 million cells).

Each model included three inlets located at specific arteries: basilar artery (BA), right internal carotid artery (ICAr) and left internal carotid artery (ICAl), respectively. Inlet boundary conditions for the simulations were determined using flow rates extracted from volunteer phase-contrast magnetic resonance imaging (PC-MRI) measurements [16, 21]. Figure 2 shows the measured volume flow rates at each inlet (BA: blue, ICAl: green, ICAr: red). As outlet boundary condition, the splitting method based on Murray's law with exponent n = 2 was applied [11].

Data analysis

The evaluation was performed using Ensight (ANSYS Inc., Canonsburg, PA, USA) and MATLAB (MATLAB R2022a, The MathWorks, Natick, USA). Time-averaged velocity values were compared on representative cross sections in the major CoW vessels and their centerline velocities (*CL-v*). To assess the shape of the flow distribution in the plane-wise evaluation, the velocity distribution was normalized by the corresponding area (*A*) and volumetric flow rate (*Q*).

$$V_{\rm Norm} = \frac{V \times A_{\rm plane}}{Q_{\rm plane}}$$

To assess the intra-saccular blood flow within the IAs, wall shear stresses (WSS) at the IA wall and the neck inflow rates (NIR) were evaluated. For a comparative basis, the chosen parameters' time-averaged WSS (TAWSS), oscillatory shear index (OSI) and velocity (ν) were normalized by their spatial mean within the corresponding IA.

Results

Hemodynamics in the complex and trimmed circle of Willis models

Cross sections representing the qualitative velocity distribution in the right ICA show slower values across all planes in the complex models (Fig. 3). Both models show higher velocity magnitudes in the middle cerebral artery (MCA) compared to the anterior cerebral artery (ACA). While differences between the models are noticeable in MCA planes with higher velocities, the lower velocity values in the ACA show fewer absolute differences between the models. The shape of the normalized velocity distribution is similar in the MCA section of the right ICA for both models, but differences are observed in all three ACA planes. In plane ACAr.1, the flow profile is broader, and in plane ACAr.2, a more complex shape of the flow profile is evident in the trimmed model. Additionally, the profile in ACAr.3 is attached to the vessel **Fig. 2** Volume flow rates (l/s) over time (s) used as inlet boundary conditions for all models. The physiological trimmed and complex models are shown below, with color-coded arrows pointing at the corresponding inlets



wall on the right in the complex model but more confined and directed toward the bottom in the trimmed model.

In the left ICA, the qualitative velocity distributions are more consistent (Fig. 3b). However, notable shape differences are observed after normalization. While the right side showed differences in the ACA vessel, the left side exhibits more prominent differences in the MCA. In plane MCAl.1, the complex model has a round flow shape, while the trimmed model shows a more compound "two-peak" shape. Additionally, the flow in planes MCA1.2 and MCA1.3 is more directed toward the right in the complex model. The presence of four additional outlets in the left MCA compared to only one additional outlet in the right MCA results in higher flow profile differences on the left side when comparing complex and trimmed versions. When comparing the left and right side of the CoW geometry, an inconsistent distribution of the flow between MCA and ACA on both sides can be seen. While for the left side of the ACA and MCA the flow is nearly equally distributed (1.2 and 1.3 ml/s for the complex model), the right side shows a larger difference between ACA and MCA flow (0.31 and 1.6 ml/s for the complex model). The result of this uneven distribution is a higher MCA flow on the right side of the CoW when compared with the left counterpart, which leads to an asymmetric flow distribution. Time-dependent volumetric flow rates and mean flow rates through the planes are given in Supplementary Material S1.

Left and right PCA show qualitatively fever differences not only in the absolute velocity values, but also in the shape of their normalized distributions (Fig. 3c). However, with increasing distance from the BA inlet, the differences increase. The highest difference in the normalized flow rates occurs in plane PCAr.1, with a centered flow in the trimmed model, and a flow attached to the bottom in the complex model.

For the right ICA (Fig. 4a), the *CL-vs* remain higher for the trimmed model over the whole distance. In the ICAr.1, the first four outlets only lead to slight changes of the centerline velocity in the complex model. However, after the fifth outlet, located in the bifurcation separating the ACA and the MCA part of the right ICA, the centerlines of ICAr.1 and ICAr.2 strongly diverge, up to a difference of over 0.1 m/s. The centerline ICAr.3 covers the right ACA vessel. Although the additional outlets (O7 and O8) are smaller than most of the outlets in the MCA branch, the *CL-vs* distal to outlet 8 start to diverge up to a difference of 0.15 m/s. Outlet 8 is located right proximal to the anterior communicating artery



Fig. 3 Qualitative visualization of the intra-vessel flow within trimmed and complex model in vessels originating from the right ICA (a), the left ICA (b) and the BA (c). The position of the planes is displayed on

the left. On the right, the time-averaged velocity magnitude of three representative is displayed (two top rows). Below, the normalized velocity for the same planes is visualized



Fig. 4 Time-averaged centerline velocities in the right ICA (\mathbf{a}), the left ICA (\mathbf{b}) and the BA (\mathbf{c}). The black dots represent positions at which outlets occur in the complex model (sorted by area on the left). The blue dots visualize the positions of the representative planes in Fig. 3. The velocities are plotted over equidistant centerline points with increasing

distance from the inlets. Centerlines (*CL-vs*) are indicated by the letter c and annotated in red or green, while planes are displayed in blue. Outlet, plane and centerline positions are only shown for the complex model, but placed equally in the trimmed version

(Acom), that connects the left and right side of the CoW. While observing mostly higher values in the trimmed centerlines, consistent with the findings in Fig. 3, the left ICA shows higher velocities for the complex centerline between planes MCAl.1 and MCAl.2. At the centerline ICAl.2 in the MCA portion, the larger outlets (O6 and O7) have a higher influence on the flow changes than the smaller outlets (O4 and O5). In the centerline ICAI.3, the highest difference (0.1 m/s) between both models occurs after plane ACAl.1, located distal to the bifurcation separating the left MCA and ACA. The *CL-vs* of the BA (Fig. 4) are more consistent than the ones in the ICAs. However, centerline BA.2 and BA.3 have vessel segments (after O3 and O10) with the complex CLvs being higher than the trimmed values. Furthermore, with increasing distance from the inlet, the BA centerlines start to diverge more, consistent with the findings in Fig. 3. Additionally, the larger outlets (O13, O9, O11 and O4) have a stronger influence on the difference in the CL-vs.

Hemodynamics within the subject-specific IAs of the pathological circle of Willis model

Qualitatively, the comparison of the pathological versions features differences particularly on the IAs surfaces (Fig. 5). Within the complex model, a lower TAWSS compared to the trimmed model occurs in IA-A, but a higher TAWSS occurs in IA-B. OSI decreases for both IAs within the complex model compared to the trimmed one. Looking at the flow field within the velocity planes inside the IAs, no essential differences can be noticed (Fig. 5, right column). Still, the temporal mean velocity v within the complex model is slightly lower.

The average and maximum WSS as well as the NIR curves are shown in Fig. 6 within the complex and trimmed model. Average WSS curves feature an equal distribution for each IA. Nevertheless, after model reduction the average WSS increases for IA-A and decreases for IA-B. This is also the case for the maximum WSS of IA-A. For IA-B, the maximum WSS decreases slightly less, particularly during systole. The resulting NIR are not crucially different in course, however present a similar tendency as the average and maximum WSS: NIR into IA-A increases and into IA-B decreases after model trimming. Mean values of TAWSS, NIR and *v* represent the findings that, with model trimming, the parameter values increase for IA-A and decrease for IA-B. Only the OSI values increase for both IAs after model trimming, as also presented qualitatively (Fig. 5).

Discussion

Flow- and shear-related hemodynamics play a significant role in the formation and growth of intracranial aneurysms (IAs). However, the acceptance of recent image-based blood flow simulations for assessing intra-aneurysmal flow is limited due to the presence of partly unrealistic boundary conditions. To our knowledge, this is the first study focusing on the impact of a highly complex CoW model in the intracranial hemodynamics and featuring a detailed comparison between an extensively segmented, subject-specific model with 60 outlets and a trimmed model version. By manually adding patient-based IAs, the resulting differences in intra-aneurysmal hemodynamics could be investigated directly, visualizing possible clinical relevant flow changes. The highly complex CoW model is an advancement on previous studies that focused only on the major vessel branches [4, 15, 22].

Intra-vessel flow analysis revealed lower velocities due to the outlet-related flow reduction in the complex model. However, certain vessel parts, such as MCA1.1, MCA1.2 and PCAl.1, showed higher velocities. These parts are located near bifurcations (e.g., the bifurcation separating left MCA and ACA) separating major CoW vessels. The CL-vs in the ICAl.1 (MCA branch) and ICAl.3 (ACA branch) at the described bifurcation indicate that the flow increases in the MCA branch to the same extent as it decreases in the ACA branch. Thus, depending on the outlet splitting algorithm used, the reduction or inclusion of different outlets can lead to a change in flow distribution between the major CoW vessel branches. These results are reflected in the analysis of the two patient-specific IAs. For IA-A, located at the right MCA, the flow is lower within the complex model. IA-B is located at the left MCA and reveals higher hemodynamics (AWSS, maximum WSS, NIR, v) within the complex model, only the OSI decreases here. However, OSI is not necessarily affected by the volumetric flow rate [23].

Nevertheless, the overall lower volumetric flow rates through vessels located on the right and further back left arteries leads to the assumption of a general overestimation of the flow in a trimmed model. This overestimation is usually more present with increasing distance from the inlet vessels as the number of outlets neglected also increases (see Fig. 4c). Differences in model complexity lead to differences in flow magnitude, which is particularly relevant for evaluating pathologies in the distal part of the CoW, such as IAs. These findings can be related to Castro et al. [24], who investigated the impact on hemodynamics when parent arteries are truncated. In previous studies correlating IA geometry and hemodynamics to IA rupture, the model complexity was not taken into account [25, 26]. Thus, specific findings suggesting that the formation and rupture of IAs are solely caused by specific hemodynamic thresholds could have led to misleading conclusions.

Furthermore, independent from the absolute flow values, differences in the flow distribution were noted. The shape of the normalized velocity distribution (Fig. 3) strongly depends on the location analyzed, since higher differences appear **Fig. 5** Qualitative visualization of hemodynamics within the IAs A (**a**) and B (**b**). TAWSS normalized by the spatial mean TAWSS inside the according IA, OSI normalized by the spatial mean OSI within the according IA, velocity plane through IA colored by v normalized by spatial mean v within the according IA



close to bifurcations and outlets—additionally shown by the centerline graphs. With higher outlet diameters, the possible influence onto the flow pattern is increased. This is in line with Ren et al. [13] and Alnas et al. [12], who determined that blood flow and WSS is affected by anatomical variations of the CoW. Locations with higher WSS are related to IA occurrence and are thought to carry a higher risk of IA formation. Within this study, the TAWSS distribution in IA-A showed higher variability when more outlets are taken into account, representing a more complex geometry. This is in line with Shen et al. [27] and Valen-Sendstad et al. [14] stating that the intra-aneurysmal flow mainly depends on the parent vessel flow and local geometry.

In the context of analyzing IA hemodynamics and disease progression, it is indispensable to incorporate realistic and patient-specific geometries. The findings of this study emphasize the importance of aneurysm location with respect to the complexity needed in numerical simulations. Moreover, the location of the IA is decisive if a high complex segmentation is crucial. The IAs analyzed in this study are located at the right and left MCA and show influence from the model complexity. As IAs statistically occur mostly at the anterior circulation (ICA junction to PCA, MCA trifurcation, ACA complex) [28], this should be taken into consideration when studying the risks of IA formation and rupture, particularly when correlating these risks with hemodynamics. Previous studies have demonstrated varying results in relation to this aspect [29, 30]. It is important to note that the assessment of intra-aneurysmal hemodynamics has clinical relevance with respect to treatment planning, risk assessment and outcome prediction.

This study has several limitations. First, the inlet boundary condition derived from PC-MRI measurements features Fig. 6 WSS plots over time for spatial mean (average), maximum (max) and NIR inside IA-A (dashed lines) and B (solid lines) comparative for the complex (red) and trimmed (green) model. Spatially and temporally averaged resulting OSI, TAWSS, NIR and v in bar plots (bottom right)



low resolution and therefore may result in an underestimation. Nevertheless, the waveform can be assumed to be as subject-specific as possible due to the data acquisition occurring during MRI scans. Second, only one case is taken into account within this study. However, this case provides a complex structure which has not been published in the literature before, and the segmentation process required considerable time and effort. Finally, although the IAs featured are certainly patient-specific, they are added onto a healthy CoW model. Moreover, the inflow rates are waveforms from the same healthy subject. Nevertheless, the addition of the IAs was performed by ensuring the preservation of the IA neck and by scaling the IA to the equivalent parent vessel diameter.

As can be derived from the results, the model complexity influences the flow magnitudes and the flow distribution. A complex model is advantageous and should be used in imagebased blood flow simulations of IAs if possible, especially in the vicinity of and in proximity to the IA. Specifically, local differences in complexity mainly influences the shape of the flow distribution. However, global complexity influences the magnitude of velocities in the model considered.

Future work will include a larger number of cases to strengthen the findings of this study. Furthermore, this can be accomplished by adding IAs in different locations to discover the impact of the altered flow on the intra-aneurysmal hemodynamics within the ICA or ACA, for instance. Accessing the imaging data of a patient-specific pathological CoW would provide the opportunity to analyze IA hemodynamics in a realistic and not artificially created pathological model, which would ensure the pathological inflow conditions as well.

Conclusion

In this study, the influence of geometric complexity on intravessel and intra-aneurysmal flow was shown and, for the first time, analyzed in detail in a 60-outlet CoW model. Both flow magnitude and the shape of the flow distribution depend on the number of outlets in the model. The first is mainly influenced by the global vessel model, whereas the latter depends on the local structure. Moreover, the location of IAs is critical with respect to the sensitivity of flow- and shearrelated parameters in the aneurysm to complexity.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s11548-023-03045-3.

Acknowledgements The authors would like to express their gratitude to Dr. Michael Markl (Department of Radiology and Biomedical Engineering, Northwestern University Feinberg School of Medicine, Chicago, IL) and his group for developing and providing the 4D PC-MRI sequence and the flow tool used to extract the flow data measured by 4D PC-MRI. We would like to thank the BMMR group of the Ottovon-Guericke University Mageburg, especially Dr. Hendrik Mattern and Daniel Stucht, for performing the respective ToF and PC-MRI measurements.

Funding Open Access funding enabled and organized by Projekt DEAL. We thank the German Federal Ministry of Education and Research within the Research Campus STIMULATE (Grant No. 13GW0473A) and the German Research Foundation (SPP2311, Project Number: 465189657) for financial support.

Availability of data and material Processed data and code are available upon reasonable request from the corresponding author.

Declarations

Conflict of interest The authors declare that they have no conflict of interest.

Ethical approval All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards.

Informed consent Informed consent was obtained from all individual participants included in the study.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- 1. Lee RM (1995) Morphology of cerebral arteries. Pharmacol Ther 66(1):149–173
- Perosa V, Priester A, Ziegler G, Cardenas-Blanco A, Dobisch L, Spallazzi M, Assmann A, Maass A, Speck O, Oltmer J, Heinze H-J, Schreiber S, Düzel E (2020) Hippocampal vascular reserve associated with cognitive performance and hippocampal volume. Brain 143(2):622–634
- Linn F, Rinkel G, Algra A, Van Gijn J (1996) Incidence of subarachnoid hemorrhage: role of region, year, and rate of computed tomography: a metaanalysis. Stroke 27(4):625–629
- Cebral JR, Castro MA, Soto O, Löhner R, Alperin N (2003) Bloodflow models of the Circle of Willis from magnetic resonance data. J Eng Math 47(3):369–386
- Cebral JR, Vazquez M, Sforza DM, Houzeaux G, Tateshima S, Scrivano E, Bleise C, Lylyk P, Putman CM (2015) Analysis of hemodynamics and wall mechanics at sites of cerebral aneurysm rupture. J NeuroIntervent Surg 7(7):530–536
- Janiga G, Berg P, Sugiyama S, Kono K, Steinman D (2015) The computational fluid dynamics rupture challenge 2013—Phase I: prediction of rupture status in intracranial aneurysms. Am J Neuroradiol 36(3):530–536
- 7. Berg P, Saalfeld S, Voss S, Beuing O, Janiga G (2019) A review on the reliability of hemodynamic modeling in intracranial aneurysms:

Why computational fluid dynamics alone cannot solve the equation. Neurosurg Focus 47(1):E15

- Chnafa C, Brina O, Pereira V, Steinman D (2018) Better than nothing: a rational approach for minimizing the impact of outflow strategy on cerebrovascular simulations. Am J Neuroradiol 39(2):337–343
- Saalfeld S, Voss S, Beuing O, Preim B, Berg P (2019) Flowsplitting-based computation of outlet boundary conditions for improved cerebrovascular simulation in multiple intracranial aneurysms. Int J Comput Assist Radiol Surg 14(10):1805–1813
- Valen-Sendstad K, Piccinelli M, KrishnankuttyRema R, Steinman D (2015) Estimation of inlet flow rates for image-based aneurysm CFD models: Where and how to begin? Ann Biomed Eng 43(6):1422–1431
- Murray CD (1926) The physiological principle of minimum work: I. The vascular system and the cost of blood volume. Proc Natl Acad Sci 12(3):207–214
- Alnaes MS, Isaksen J, Mardal KA, Romner B, Morgan MK, Ingebrigtsen T (2007) Computation of hemodynamics in the circle of Willis. Stroke 38:2500–2505
- Ren Y, Chen Q, Li ZY (2015) A 3D numerical study of the collateral capacity of the Circle of Willis with anatomical variation in the posterior circulation. Biomed Eng Online 14:S11
- 14. Valen-Sendstad K, Bergersen AW, Shimogonya Y, Goubergrits L, Bruening J, Pallares J, Cito S, Piskin S, Pekkan K, Geers AJ, Larrabide I, Rapaka S, Mihalef V, Fu W, Qiao A, Jain K, Roller S, Mardal K-A, Kamakoti R, Spirka T, Ashton N, Revell A, Aristokleous N, Houston JG, Tsuji M, Ishida F, Menon PG, Browne LD, Broderick S, Shojima M, Koizumi S, Barbour M, Aliseda A, Morales HG, Lefèvre T, Hodis S, Al-Smadi YM, Tran JS, Marsden AL, Vaippummadhom S, Einstein GA, Brown AG, Debus K, Niizuma K, Rashad S, Sugiyama S-I, Khan MO, Updegrove AR, Shadden SC, Cornelissen BMW, Majoie CBLM, Berg P, Saalfield S, Kono K, Steinman DA (2015) Real-world variability in the prediction of intracranial aneurysm wall shear stress: the 2015 international aneurysm CFD challenge. Cardiovasc Eng Technol 9(2018):544–564
- 15. Sutalo ID, Bui AV, Ahmed S, Liffman K, Manasseh R (2014) Modeling of flow through the Circle of Willis and cerebral vasculature to assess the effects of changes in the peripheral small cerebral vasculature on the inflows. Eng Appl Comp Fluid Mech 8:609–622
- Gaidzik F, Pathiraja S, Saalfeld S, Stucht D, Speck O, Thévenin D, Janiga G (2021) Hemodynamic data assimilation in a subjectspecific Circle of Willis geometry. Clin Neuroradiol 31(3):643–651
- Berg P, Stucht D, Janiga G, Beuing O, Speck O, Thévenin D (2014) Cerebral blood flow in a healthy Circle of Willis and two intracranial aneurysms: Computational fluid dynamics versus four-dimensional phase-contrast magnetic resonance imaging. J Biomech Eng 136:1–9.
- Mattern H, Sciarra A, Godenschweger F, Stucht D, Lüsebrink F, Rose G, Speck O (2018) Prospective motion correction enables highest resolution time-of-flight angiography at 7T. Magn Reson Med 80(1):248–258
- Frangi AF, Niessen WJ, Vincken KL, Viergever MA (1998) Multiscale vessel enhancement filtering. In: Wells WM, Colchester A, Delp S (eds) Medical image computing and computer-assisted intervention—MICCAI'98. Springer, Berlin, pp 130–137
- 20. Berg P, Voss S, Saalfeld S, Janiga G, Bergersen AW, Valen- Sendstad K, Bruening J, Goubergrits L, Spuler A, Cancelliere NM, Steinman DA, Pereira VM, Chiu TL, Tsang ACO, Chung BJ, Cebral JR, Cito S, Pallarès J, Copelli G, Csippa B, Paál G, Fujimura S, Takao H, Hodis S, Hille G, Karmonik C, Elias S, Kellermann K, Khan MO, Marsden AL, Morales HG, Piskin S, Finol EA, Pravdivtseva M, Rajabzadeh-Oghaz H, Paliwal N, Meng H, Seshadhri S, Howard M, Shojima M, Sugiyama S, Niizuma K, Sindeev S, Frolov S, Wagner T, Brawanski A, Qian Y, Wu Y-A, Carlson KD, Dragomir-Daescu D, Beuing O (2018) Multiple aneurysms

anatomy challenge 2018 (MATCH): Phase I: segmentation. Cardiovasc Eng Technol 9(4):565–581.

- 21. Stucht D, Danishad KA, Schulze P, Godenschweger F, Zaitsev M, Speck O (2015) Highest resolution in vivo human brain MRI using prospective motion correction. PLoS ONE 10(7):e0133921
- Moore S, David T, Chase JG, Arnold J, Fink J (2006) 3D models of blood flow in the cerebral vasculature. J Biomech 39:1454–1463
- Najafi M, Cancelliere NM, Brina O, Bouillot P, Vargas MI, Delattre BM, Pereira VM, Steinman DA (2021) How patient-specific do internal carotid artery inflow rates need to be for computational fluid dynamics of cerebral aneurysms? J NeuroIntervent Surg 13:459–464
- Castro MA, Putman CM, Cebral JR (2006) Computational fluid dynamics modeling of intracranial aneurysms: effects of parent artery segmentation on intra-aneurysmal hemodynamics. Am J Neuroradiol 27:1703–1712
- Wan H, Ge L, Huang L, Jiang Y, Leng X, Feng X, Xiang J, Zhang X (2019) Sidewall aneurysm geometry as a predictor of rupture risk due to associated abnormal hemodynamics. Front Neurol 10(8):841–848
- Zhou G, Zhu Y, Yin Y, Su M, Li M (2017) Association of wall shear stress with intracranial aneurysm rupture: systematic review and meta-analysis. Sci Rep 7:5331
- 27. Shen Y, Molenberg R, Bokkers RP, Wei Y, Uyttenboogaart M, van Dijk JMC (2022) The role of hemodynamics through the Circle of Willis in the development of intracranial aneurysm: a systematic review of numerical models. J Personalized Med 12(6).

- Brown RD, Broderick JP (2014) Unruptured intracranial aneurysms: epidemiology, natural history, management options, and familial screening. Lancet Neurol 13:393–404
- Cebral JR, Mut F, Weir J, Putman CM (2011) Association of hemodynamic characteristics and cerebral aneurysm rupture. Am J Neuroradiol 32(2):264–270
- 30. Detmer FJ, Chung BJ, Mut F, Slawski M, Hamzei-Sichani F, Putman C, Jiménez C, Cebral JR (2018) Development and internal validation of an aneurysm rupture probability model based on patient characteristics and aneurysm location, morphology, and hemodynamics. Int J Comput Assist Radiol Surg 13:1767–1779

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.