REVIEW

Resin glycosides from Convolvulaceae plants

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Abstract Resin glycosides are well known as purgative ingredients, which are characteristic of certain crude drugs such as Mexican Scammony Radix, Orizabae Tuber, and Jalapae Tuber, all of which originate from Convolvulaceae plants. Depending on their solubility in ether, these are roughly classified into two groups-jalapin (soluble) and convolvulin (insoluble). Almost all jalapins hitherto isolated and characterized had common intramolecular macrocyclic ester structures. These are composed of 1 mol of oligoglycoside of hydroxyl fatty acid (glycosidic acid) partially acylated by some organic acids at the sugar moiety, some examples of which are ester-type dimers. On the other hand, convolvulin is regarded as an oligomer of a variety of acylated glycosidic acids. This review describes the isolation and structural elucidation of resin glycosides from some Convolvulaceae plants, including Ipomoea operculata, Pharbitis nil, Quamoclit pennata, Calystegia soldanella, and I. muricata.

Keywords Resin glycoside · Jalapin · Convolvulin · Convolvulaceae · Glycosidic acid · Hydroxyl fatty acid

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Introduction

Resin glycosides are well known as purgative ingredients, which are characteristic of crude drugs such as Mexican Scammoniae Radix (the root of Convolvulus scammonia L.), Orizabae Tuber [the tuber of Ipomoea orizabensis (Pellet) Ledanois], and Jalapae Tuber (the tuber of *I. purga* Hayne). They are commonly found in plants belonging to the Convolvulaceae family [1]. Chemical investigations on these resin glycosides were initiated in the middle of the nineteenth century. The obtained resin glycosides were roughly classified into an ether-soluble group called jalapin or an ether-insoluble group denoted convolvulin [2]. Upon alkaline hydrolysis, both jalapins and convolvulins afforded organic acids such as isobutyric, 2-methylbutyric, (E)-2-methylbut-2-enoic (tiglic), 2-methyl-3-hydroxybutyric (nilic) acids and oligoglycosides of hydroxyl fatty acid (glycosidic acid). Furthermore, the glycosidic acid provided several kinds of monosaccharides (e.g., glucose, rhamnose, and fucose) and a hydroxyl fatty acid (e.g., 11-hydroxyhexadecanoic (jalapinolic), 11-hydroxytetradecanoic (convolvulinolic), and 3,11-dihydroxytetradecanoic (ipurolic) acids) on acidic hydrolysis. Considering this behavior toward acids and bases, Mannich and Schumann speculated that convolvulins from I. purga were oligomers of a glycosidic acid partially acylated by some organic acids at the sugar moiety [3]. However, until our research group isolated pure genuine jalapins, chemical studies had been confined only to the characterization of the component organic acids and glycosidic acids afforded by alkaline hydrolysis of a crude resin glycoside. In 1987, our research group succeeded, for the first time, in the isolation and structural elucidation of four jalapins from Orizabae Tuber [4]. Hitherto, almost all isolated and characterized jalapins have had intramolecular macrocyclic ester



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structures composed of 1 mol of oligoglycoside of hydroxyl fatty acids partially acylated by some organic acids at the sugar moiety, while some examples were ester-type dimers [5-13].

In this review, our studies on the isolation and structural elucidation of resin glycosides from some Convolvulaceae plants, including *I. operculata* (Gomes) Mart., *Pharbitis nil, Quamoclit pennata*, and *I. muricata*, are summarized.

I. operculata

Rhizoma Jalapae Braziliensis (Brazilian Jalap), the root of *I. operculata* (Gomes) Mart., is known to be a substitute for Jalapae Tuber (Vera Cruz Jalap, *I. purga*), and its ethanol extract, Brazilian resin, is a potent laxative. Brazilian Jalap contains both jalapins and convolvulins as active ingredients.

Graf et al. reported that the alkaline hydrolysis of the jalapin fraction from Brazilian Jalap yielded acetic, tiglic, *n*-valeric, trimethylacetic, 2-methylbutyric, isovaleric, and propionic acids as component organic acids, while the convolvulin fraction (named rhamnoconvolvulin) afforded 4-oxodecanoic acid and 3,6:6,9-diepoxydecanoic (exogonic) acid, which are characteristic of Brazilian resin, along with those from the jalapin fraction [14, 15]. Wagner and Kazmaier isolated a major glycosidic acid named operculinic acid from rhamnoconvolvulin and characterized it as 3,12-dihydroxyhexadecanoic acid 12-*O*- α -D-glucopyranosyl-(1 \rightarrow 4)-[*O*- α -L-rhamnopyranosyl-(1 \rightarrow 2)-[*O*- β -D-glucopyranosyl-(1 \rightarrow 3)]-*O*- β -D-glucopyranoside [16].

Separation of crude jalapin and crude convolvulin fractions

The methanol (MeOH) extract of the root of *I. operculata* was partitioned between ether and H_2O . The ether-soluble fraction was subjected to chromatography over MCI gel CHP 20P and Sephadex LH-20 columns to afford the crude jalapin fraction. The H_2O -soluble fraction was partitioned between *n*-butanol and H_2O . The *n*-butanol-soluble fraction was subjected to an MCI gel CHP 20P column to give the crude convolvulin fraction [17].

Jalapin

Component organic acids and hydroxyl fatty acid The crude jalapin fraction was subjected to alkaline hydrolysis to furnish organic acid and glycosidic acid fractions. The former fraction was methylated with diazomethane-ether and then examined by GC–MS, exhibiting two peaks

identical to those of methyl *n*-decanoate and *n*-dodecanoate. The latter fraction furnished an aglycone fraction and a monosaccharide fraction upon acidic hydrolysis. The aglycone fraction was methylated with diazomethane-ether to furnish methyl ester (1) ($[\alpha]_D + 0.9^\circ$) of jalapinolic acid (2) [17].

Determination of the absolute configurations of hydroxyl fatty acids including jalapinolic acid Mosher's method [18] was used to determine the absolute configuration of 1. The ¹H-NMR spectrum of (+)-2-methoxy-2-trifluoromethylphenylacetate (MTPA ester) of the racemate of 1 as well as the ¹H-NMR spectra of the (-)- and (+)-MTPA esters (3, 4; 5, 6; 7, 8) of commercial 2S-pentanol (9), 2Shexanol (10) and 2S-heptanol (11) as model compounds, indicated that Mosher's method is applicable for the determination of the configuration of 1 (Fig. 1). This determination was accomplished using the chemical shift of the terminal methyl group, which is separated by four methylene groups from the asymmetric carbon. The chemical shift difference $(\Delta \delta)$ [$\delta(-)$ -MTPA ester- $\delta(+)$ -MTPA ester] of the H₃-16 signals in the ¹H-NMR spectra of the (-)- and (+)-MTPA esters (12, 13) of 1 indicated the configuration at C-11 of 1 to be S [19, 20]. At the same time, Shibuya et al. determined the configuration of (+)jalapinolic acid, which was obtained from the tuber of Merremia mammosa, to be S by syntheses of S- and Rjalapinolic acid via Sharpless asymmetric epoxidation [21].

In the same manner as **1**, the absolute configuration at C-11 of convolvulinolic acid (**14**), the aglycone of quamoclinic acid A, was determined to be *S* using the $\Delta\delta$ value of (-)- and (+)-MTPA esters (**15**, **16**) of methyl convolvulinolate (**17**) ($[\alpha]_{\rm D}$ + 1.0°) [20].

Ipurolic acid (18), the common aglycone of pharbitic acids C and D, has hydroxyl groups at C-3 as well as at C-11. The ¹H-NMR spectral data of the (-)- and (+)-MTPA esters (19, 20) of commercial methyl 3S-hydroxybutyrate (21), used as model compounds, indicated that the configuration at C-3 could be determined by the $\Delta\delta$ values of the signals originating from the 2-methylene protons and ester methyl protons. Comparison of the ¹H-NMR spectrum of (-)-MTPA ester (22) of methyl ipurolate (23) $([\alpha]_{D} + 1.2^{\circ})$ with that of (+)-MTPA ester (24) suggested that the configurations at C-3 and C-11 of 18 are both S. The influence on their $\Delta\delta$ values by interactions between two MTPA residues at C-3 and C-11 in 22 and 24 was ruled out as follows. Methyl 11-hydroxytetradec-2-enoate, which was prepared from crude pharbitic acid according to Okabe et al. [22], was hydrogenated to furnish 17. In addition, the ¹H-NMR spectra of (-)- and (+)-MTPA esters (25, 26) of methyl 3-hydroxy-11-oxo-tetradecanoate (27), which was obtained by the partial oxidation of 23, showed similar $\Delta\delta$ values to those of **22** and **24** for signals



Fig. 1 Structures of 1–35, and values of ¹H-NMR chemical shift difference (ppm) [$\Delta\delta$: δ (–)-MTPA – δ (+)-MTPA ester (×10⁻³)] for MTPA esters (in CDCl₃, 3–8, 12, 13, 15, 16, 19, 20, 22, 24–26, 32,

due to 2-methylene protons and ester methyl protons. Thus, the configurations at C-3 and C-11 of **18** were both determined to be S [20].

The methyl ester (28) ($[\alpha]_D + 0.2^\circ$) of 3,11-dihydroxyhexadecanoic acid (29), the aglycone of pharbitic acid B, the methyl ester (30) of 3,12-dihydroxyhexadecanoic acid (31), and the aglycone of operculinic acid H, were converted into (–)- and (+)-MTPA esters (32, 33; 34, 35). The $\Delta\delta$ values for signals due to H₃-16, Ha-2, Hb-2, and ester methyl protons were analogous to those of 22 and 24. Therefore, the absolute configurations of 29 and 31 were concluded to be 3*S*, 11*S*, and 3*S*, 12*S*, respectively [20, 23]. The absolute configuration of 31 was compatible with that clarified by the enantioselective syntheses performed by Jakob and Gerlach [24].

33: 400 MHz; **34**, **35**: 600 MHz). Data were taken and reproduced from references Ono et al. [20, 23]

Mosher's method was thus found to be useful for the determination of the absolute configuration of a minute amount of hydroxyl fatty acids.

Component glycosidic acids The glycosidic acid fraction in MeOH was treated with diazomethane-ether. The concentrated reaction mixture was subjected to silica gel column chromatography and preparative HPLC on octadecyl silica (ODS), silica gel, and octyl silica columns to yield seven methyl esters of glycosidic acids, named operculinic acids A–G (**36–42**) [17, 19, 25]. Their structures were determined by NMR spectroscopy, negative-ion FAB-MS, and GC analyses of trimethylsilyl ethers of the thiazolidine derivatives [26] of monosaccharides provided on their Fig. 2 Structures of component glycosidic acids (36–42) of the crude jalapin fraction prepared from the root of *I. operculata*



acidic hydrolyses (Fig. 2). Among them, operculinic acid G (42) might be an artifact produced from operculinic acid A (36) during treatment with diazomethane [27].

Isolation of resin glycosides The crude jalapin fraction was subjected to silica gel and ODS column chromatography and HPLC on silica, ODS, and octyl silica columns to yield operculins I–XVIII (**43–60**) [19, 28–30]. Preparative recycling HPLC was a useful method for the isolation of two constitutional isomers, in which the *n*-decanoyl group and *n*-dodecanoyl group were mutually interchanged between the same positions (**49** and **50**; **51** and **52**) [28, 29].

Structures of resin glycosides Operculin I (43) is composed of 2 mol of *n*-dodecanoic acid and 1 mol of 36 and is intramolecularly linked with a hydroxyl group of the sugar moiety to form a macrolactone structure, which was elucidated by using ¹H- and ¹³C-NMR spectra, negative-ion FAB-MS, as well as analyses of the hydrolysates produced by alkaline hydrolysis. The site of each ester linkage in jalapinolic and *n*-dodecanoic acids were fixed by analyses of the acylation shifts observed in the ¹H-NMR spectrum of 43 as well as peak assignments in the EI-MS of the peracetate of 43 and in the negative ion FAB-MS of 43. From these data, the structure of 43 was defined as 11Sjalapinolic acid 11-O- β -D-glucopyranosyl-(1 \rightarrow 3)-O-[4-O-n-dodecanoyl- α -L-rhamnopyranosyl- $(1 \rightarrow 4)$]-O-(2-O*n*-dodecanoyl)- α -L-rhamnopyranosyl- $(1 \rightarrow 4)$ -O- α -Lrhamnopyranosyl- $(1 \rightarrow 2)$ - β -D-fucopyranoside, intramolecular 1, 2"-ester, as shown in Fig. 3. [19, 28].

The structures of operculins II (44)–XVIIII (60) were determined using similar methods to those used for 43 (Fig. 3).

Compounds **43–60** are the first known resin glycosides to feature fatty acids, *n*-dodecanoic and/or *n*-decanoic acid, as component organic acids.

Convolvulin

Component organic acids Alkaline hydrolysis of the crude convolvulin fraction afforded organic acid and glycosidic acid fractions. The organic acid fraction was esterified with *p*-bromophenacyl bromide, and the product was subjected to chromatographic separation to afford pbromophenacyl isovalerate (61), p-bromophenacyl tiglate (62), and two compounds 63 and 64 (Fig. 4). Both 63 and 64 were identified as the *p*-bromophenacyl esters of the exogonic acid reported by Graf and Dahlke [14] using NMR and FD-MS data. Furthermore, it was elucidated that 63 and 64 are isomers and that interconversion between them occurred because of inversion at the C-6 spiro-center. The absolute configurations at C-3 and C-9 of exogonic acid were identified as S and R, respectively, by applying Mosher's method to methyl 3-hydroxy-6,9-epoxydecanoate (65), methyl 9-hydroxy-3,6-epoxydecanoate (66), and methyl 3,9-dihydroxydecanoate (67), which were provided by hydrogenation of methyl exogonate. It was also found that this acid existed as a mixture of epimers that only differ in the spirocyclic carbon configuration at C-6 [23]. These results were consistent with those of the enantioselective syntheses performed by Lawson et al. [31].

Fig. 3 Structures of resin glycosides (**43–60**) isolated from the crude jalapin fraction prepared from the root of *I. operculata*



Dodeca: *n*-dodecanoyl, Deca: *n*-decanoyl

Component glycosidic acids The glycosidic acid fraction was treated with diazomethane-ether and subsequently subjected to chromatography to isolate the methyl ester (**68**), the alkaline hydrolysis of which afforded a glycosidic acid, named operculinic acid H (**69**). Acidic hydrolysis of **68** afforded an aglycone and a mixture of monosaccharides, which were composed of D-glucose and L-rhamnose. Methylation of the aglycone with diazomethane-ether yielded a methyl 3S, 12S-dihydroxyhexadecanoate (**30**) [23].

The structure of operculinic acid H (**69**) was characterized as shown in Fig. 5 using the NMR data of **68** and its peracetate, negative-ion FAB-MS of **69**, HR-EI-MS of the permethylate of **68**, and structural analyses of the five hydrolysates produced by partial methanolysis of **68** [23]. Despite the large difference between the structures of the sugar moieties in **68** and Wagner's operculinic acid [16], both compounds were considered to be identical. The organic acid and glycosidic acid components of the **Fig. 4** Structures of *p*-bromophenacyl esters (**61–64**) of component organic acids of the crude convolvulin fraction prepared from the root of *I. operculata* and hydrogenated products (**65–67**) of methyl exogonate

ester (68) of operculinic acid H (69), 69 and acylated glycosidic acid methyl esters (70–73) generated from the crude convolvulin fraction from the root of *I. operculata* after treatment with indium(III) chloride in MeOH

Fig. 5 Structures of methyl

convolvulin fraction were different from those of the jalapin fraction obtained from the crude drug.

Structures of acylated glycosidic acid methyl esters Despite numerous attempts, the isolation of pure resin glycosides from the crude convolvulin fraction has been unsuccessful. In consideration of previous results, it was evident that the component resin glycosides of the crude convolvulin fraction possessed at least one carboxyl group. Hence, the fraction was treated with indium(III) chloride in MeOH, which was reported to be a catalyst for mild methyl esterification of carboxylic acids by Mineno and Kansui [32]. The treated fraction exhibited a number of separate spots by TLC on silica gel and was successively separated by Sephadex LH-20 and silica gel column chromatography and HPLC using an ODS column, affording four compounds, referred to as IOM-1 (70)–IOM-4 (73) [33].

The ¹H-NMR spectra of **70–73** showed that they consisted of mixtures of epimers that differed in the spirocyclic carbon configuration only at C-6 of the exogonoyl residue (Exg). Since HPLC using a naphthylethyl group-bonded silica (π -nap) column of **70–73** indicated that interconversion of the epimers of Exg in **70–73** was facile, structural analyses were carried out using a mixture of the epimers.

The structures of **70–73** were determined as shown in Fig. 5 using their NMR spectra, including HMBC and ${}^{1}\text{H}{}^{-1}\text{H}$ TOCSY spectra, negative-ion FAB-MS, and the







structural analyses of their hydrolysates produced by alkaline hydrolysis and partial deacylation [33].

Compounds **70–73** were all regarded as the methyl ester monomers of the acylated glycosidic acid. Furthermore, negative-ion FAB-MS and HR-negative-ion ESI-TOF–MS of the crude convolvulin fraction exhibited intense ion peaks at m/z 1659 and 1659.8010 (calculated for $C_{77}H_{127}O_{38}^{-}$, 1659.8011), corresponding to the values of an $[M - H]^{-}$ ion peak of the demethylated derivatives of **72** and **73**, respectively. In addition, no distinct peak was detected in the region of m/z 1700–4000 in the negative-ion FAB-MS of the crude convolvulin fraction. Therefore, a part of the crude convolvulin fraction might be a mixture of monomers composed of free carboxylic acid forms corresponding to **72** and **73** [33].

Thus, we consider that methyl esterification of carboxylic acids via indium(III) chloride catalysis is a useful tool for structural investigations involving convolvulins.

Pharbitis nil

Pharbitis Semen, the seed of *Pharbitis nil* Choisy, is used as a crude purgative drug, and its resin glycoside is a typical Mayer's convolvulin. Asahina et al. reported that alkaline hydrolysis of the crude glycoside named pharbitin gave (+)-2-methylbutyric acid, tiglic acid, nilic acid, and a glycosidic acid named pharbitic acid, which was composed of ipurolic acid, D-glucose, and L-rhamnose [34–36]. In 1970, Okabe et al. isolated two glycosidic acids, named pharbitic acids C (**74**) and D (**75**), along with valeric, tiglic, nilic, and (+)-2-methylbutyric acids, as components of the alkaline hydrolysis products of pharbitin [22, 37–39].

Component organic acids

Alkaline hydrolysis of pharbitin furnished organic and glycosidic acid fractions. The GC of the organic acid fraction revealed the presence of 2-methylbutyric, tiglic, and nilic acid in a molar ratio of approximately 17:1:11. Furthermore, the organic acid fraction was acylated with *p*-bromophenacyl bromide followed by chromatographic separation to yield *p*-bromophenacyl 2*S*-methylbutyrate and *p*-bromophenacyl nilate, which was found to be a mixture of the 2*R*, 3*R*- and the 2*S*, 3*S*-forms with a ratio of approximately 6:5 based on the ratio of H₃-5 signal intensities obtained from the ¹H-NMR spectrum of its (–)-MTPA ester [40, 41].

Component hydroxyl fatty acids and monosaccharides Acidic hydrolysis of the glycosidic acid fraction yielded hydroxyl fatty acid and monosaccharide fractions. Methylation of the hydroxyl fatty acid fraction yielded methyl 3*S*,11*S*-ipurolate (**23**) and methyl 3*S*,11*S*-dihydroxyhexadecanoate (**28**). The component monosaccharides were determined to be L-rhamnose, D-quinovose, and D-glucose [40].

Component glycosidic acids The glycosidic acid fraction was converted into *p*-phenylphenacyl ester, and chromatography yielded three glycosidic acid esters. The alkaline hydrolysis of each ester furnished a new glycosidic acid, named pharbitic acid B (**76**), along with pharbitic acids C (**74**) and D (**75**), whose structures were corrected by NMR and negative-ion FAB-MS data as well as the EI-MS data for the permethylates of **74** and **75** [40] (Fig. 6). Using NMR and negative-ion FAB-MS, **76** was concluded to be a homolog of

75, in which 3*S*,11*S*-ipuroric acid (**18**) of the aglycone was replaced by 3*S*,11*S*-dihydroxyhexadecanoic acid (**29**) [40].

Structures of acylated glycosidic acid methyl esters Pharbitin was treated with indium(III) chloride in MeOH, in a similar manner to the treatment of the crude convolvulin fraction from *I. operculata*. This treated pharbitin was successively subjected to Diaion HP20, Sephadex LH-20, and silica gel column chromatography and HPLC using an ODS column to afford seven compounds, temporarily named PM-1 (**77**)–PM-7 (**83**) [42].

The structures of **77–83** were determined as indicated in Fig. 6 using similar methods as those used for **70–73** [33]. Compounds **77–83** were all monomers of methyl esters of glycosidic acids esterified with several organic acids at the sugar moiety. In addition, negative-ion FAB-MS of pharbitin exhibited ion peaks at m/z 1551, 1451, and 1305, which corresponded to the values of $[M - H]^-$ ion peaks of the demethylated derivatives of **80**, **78**, and **82**, respectively. However, for pharbitin, no intense ion peaks were observed in the m/z 1580–3500 region. Therefore, a part of the pharbitin was considered to be a mixture of monomers composed of free carboxylic acid forms corresponding to **77–83**. Furthermore, it should be noted that **80** and **81** had 2R,3R-nilic acid and its enantiomer as the component organic acids in each molecule [42].

Quamoclit pennata

Quamoclit pennata is native to tropical regions of South America, and is cultivated primarily as an ornamental plant. This seed contains both jalapins and convolvulins.

Jalapin

Component organic acids and glycosidic acid The component organic acids were elucidated to be 2*S*-methylbutyric, *n*-decanoic, and *n*-dodecanoic acids. On the other hand, the glycosidic acid fraction yielded a new glycosidic acid, named quamoclinic acid A (**84**), whose structure was determined to be 11*S*-convolvulinolic acid 11-*O*- β -D-glucopyranosyl-(1 \rightarrow 3)-*O*-[α -L-rhamnopyranosyl-(1 \rightarrow 4)]-*O*- α -L-rhamnopyranosyl-(1 \rightarrow 4)-*O*- α -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-fucopyranoside on the basis of spectroscopic data and chemical evidence (Fig. 7). It is noteworthy that the aglycone (11*S*-convolvulinolic acid) of **84** differs from jalapinolic acid [43].

Structures of resin glycosides Six resin glycosides with macrolactone structures, named quamoclins I (**85**)–VI (**90**) and one acylated glycosidic acid methyl ester, named quamoclin VII (**91**), were isolated. Their structures were assigned using similar methods as those employed for

operculins. Compounds **85–90** were the first examples of jalapins with an organic acid (2*S*-methylbutyric acid) and fatty acid components (*n*-decanoic or *n*-dodecanoic acids) [43, 44] (Fig. 7). Compound **91** was considered to be an artifact produced from **89** and/or **90** during the extraction and isolation procedures.

Convolvulin

Component organic acids, hydroxyl fatty acids, and monosaccharides The alkaline hydrolysis products of the crude convolvulin fraction were fractionated into organic acid and glycosidic acid fractions. The former fraction was composed of isobutyric, 2S-methylbutyric, tiglic, 2R,3Rnilic, 7S-hydroxydecanoic, and 7S-hydroxydodecanoic acids.

Acidic hydrolysis of the glycosidic acid fraction gave aglycone and monosaccharide fractions. Methylation of the former with diazomethane-ether yielded methyl 7*S*-hydroxydecanoate and methyl 3S,11*S*-ipurolate (**23**). The monosaccharide fraction was composed of D-glucose, D-fucose, D-quinovose, and L-rhamnose [41].

Structures of glycosidic acids The glycosidic acid fraction furnished five new glycosidic acids, named quamoclinic acids B (92)–H (98). These compounds were characterized on the basis of spectroscopic data as well as chemical evidence (Fig. 8). Compounds 95–97 were the first examples of heptaglycosides of glycosidic acid, and 98 was the first octaglycoside of glycosidic acid. Furthermore, 97 and 98 were the first known glycosidic acids to feature bisdesmosides with sugar linkages at the C-3 position of ipurolic acid as well as at its C-11 position [41, 45].

Except for 2*S*-methylbutyric acid, the component organic and glycosidic acids were different from those obtained from the crude jalapin fraction of this seed.

Acylated glycosidic acid methyl esters and acylated methyl glycosides Treatment of the crude convolvulin fraction with indium(III) chloride in MeOH afforded nine acylated glycosidic acid methyl esters, named QM-1 (99), QM-2 (100), QM-3 (101), QM-9 (102), QM-10 (103), QM-11 (104), QM-12 (105), QM-13 (106), and QM-14 (107), and five acylated methyl glycosides, named QM-4 (108), QM-5 (109), QM-6 (110), QM-7 (111), and QM-8 (112). Their structures were elucidated on the basis of spectroscopic data and chemical conversions [46–48] (Fig. 8).

Since two acylated trisaccharides closely related to the resin glycosides were previously reported as natural constituents of the seeds of *Cuscuta chinensis* (Convolvulaceae) [49], **108–112** are presumably formed from the corresponding acylated saccharides with a reducing end during treatment with indium(III) chloride in MeOH. It was

Fig. 7 Structures of component glycosidic acid (84) and resin glycosides (85–91) isolated from the crude jalapin fraction prepared from the seeds of *Q. pennata*



Dodeca: n-dodecanoyl, Deca: n-decanoyl

therefore presumed that one of the reasons for the difficulty in isolating convolvulin from the crude convolvulin fraction of *Q. pennata* was the co-existence of the acylated saccharides with a reducing end, because the α - and β anomers easily reach equilibrium.

Calystegia soldanella

Calystegia soldanella Roem. et Schult. (Convolvulaceae) is distributed widely on the sandy beaches of seas and lakes in temperate regions of the world. The roots of this plant are used for the treatment of arthritis. Gasper reported the

isolation and structural elucidation of two resin glycosides, soldanellines A and B, as chemical constituents of the root [50, 51].

Component organic acids and glycosidic acids

Alkaline hydrolysis of the crude resin glycoside fraction obtained from the MeOH extract of the leaves, stems, and roots gave four new glycosidic acids, named calysolic acids A (113)–D (116), along with one known glycosidic acid, soldanellic acid B (117) [51], and three organic acids, 2*S*-methylbutyric, tiglic, and 2*S*,3*S*-nilic acids. The structures

Fig. 8 Structures of component glycosidic acids (92–98) of the crude convolvulin fraction prepared from the seeds of *Q. pennata* and acylated glycosidic acid methyl esters (99–107) and acylated methyl glycosides (108–112) generated from the crude convolvulin fraction by treatment with indium(III) chloride in MeOH



Fig. 9 Structures of component glycosidic acids (113–117) of the crude resin glycoside fraction prepared from the leaves, stems, and roots *of C. soldanella*



Fig. 10 Structures of resin glycosides (118-135) isolated from the leaves, stems, and roots of C. soldanella

of the new glycosidic acids were characterized on the basis of spectroscopic data and chemical evidence [52] (Fig. 9).

Structures of resin glycosides Seventeen new resin glycosides, named calysolins I–XVII (**118–134**), were isolated from the crude resin glycoside fraction along with soldanelline B (**135**). Their structures were determined on the basis of spectroscopic data and chemical evidence [53–56] (Fig. 10). They consisted of two types, one possessing macrolactone structures (**118–131**, **135**) and the other type

with non-macrolactone structures (132–134). Compounds 132 and 133 were methyl esters of acylated glycosidic acids, and 134 was the free carboxylic acid form of 133. Furthermore, macrolactone structures fall into two types, one having a 22-membered ring (118, 121–125, 128–130, 135) and the other with a 27-membered ring (119, 120, 126, 131). It is possible that 132–134 are artifacts formed from the macrolactone resin glycosides during the extraction and/or isolation procedures. Compounds 121, 124–126, and 128–131 were the first examples of hexaglycosides of resin glycosides with a



Fig. 11 Structure of muricatin IX (136) isolated from the seeds of *Ipomoea muricata*

macrolactone structure. Additionally, **118–135** showed antiviral activity toward herpes simplex type 1 [54–56].

I. muricata

The seeds of *I. muricata* (L.) Jacq. are used as a laxative and carminative in the folk medicine of India. In previous papers, Noda et al. reported that alkaline hydrolysis of the crude resin glycoside fraction of the seeds of *I. muricata* furnished three organic acids, isobutyric, 2*S*-methylbutyric, and 2*R*,3*R*-nilic acid, along with a glycosidic acid fraction composed of L-rhamnose, D-fucose, D-quinovose, and 11*S*-jalapinolic acid (2). They also discussed the isolation and structural elucidation of three glycosidic acids including muricatic acids A–C [20, 57, 58]. Furthermore, eight genuine resin glycosides, muricatins I–VIII, which possessed characteristic macrolactone structures, were reported [58, 59].

Structure of resin glycoside

A new resin glycoside, named muricatin IX (136), was isolated from the seeds of *I. muricata*. The structure of 136 was determined on the basis of spectroscopic data as well as chemical evidence. It should be noted that 136 is different from all the resin glycosides isolated so far, in that the carboxyl group of its aglycone moiety is linked with a hydroxyl group of the organic acid, which was attached to a sugar moiety by ester linkage, to form a macrocyclic ester ring [60] (Fig. 11).

Conclusions

Resin glycosides were classified into two groups by Mayer in 1855, according to their solubility in ether. These groups consisted of jalapins, which are ether-soluble, and convolvulins, which are ether-insoluble. This classification has since been conventionally used. Almost all ether-soluble resin glycosides hitherto isolated had common intermolecular macrocyclic ester structures composed of 1 mol of a variety of acylated glycosidic acids, of which some examples were ester-type dimers. On the other hand, the crude ether-insoluble resin glycosides fraction of the root of I. operculata, pharbitin, and the crude ether-insoluble resin glycosides fraction from seeds of Q. pennata, which were typical Mayer's convolvulins, show tailing or broad peaks on TLC and HPLC, suggesting that they differ from ether-soluble resin glycosides in chemical structure, and their crude ether-insoluble resin glycoside fractions furnished acylated glycosidic acid methyl ester monomers after treatments with indium(III) chloride in MeOH. In addition, the m/z values of intense ion peaks in the negative-ion FAB-MS of the crude ether-insoluble resin glycoside fractions obtained from the root of I. operculata and pharbitin corresponded to the values of $[M - H]^{-}$ ion peaks of demethylated derivatives of the some of their acylated glycosidic acid methyl esters. However, no intense ion peaks were observed in their dimer region. Therefore, a part of the crude ether-insoluble resin glycoside fraction might be a mixture of acylated glycosidic acid monomers with free carboxylic acid forms. On the other hand, calysolins IV, XI, and XIV, which have a macrolactone structure, hardly dissolve in ether. In addition, muricatin IX is different from all the resin glycosides isolated so far, in that the carboxyl group of its aglycone moiety linked with a hydroxyl group of the organic acid, which is attached to a sugar moiety by an ester linkage, to form a macrocyclic ester ring. Furthermore, the root of I. operculata and the seeds of Q. pennata contain both ethersoluble resin glycosides and ether-insoluble resin glycosides. The component organic and glycosidic acids of ether-soluble resin glycoside fractions of these materials were different from those of the corresponding ether-insoluble resin glycoside fraction, respectively. Therefore, classification of resin glycosides into two groups is considered to be meaningful.

As mentioned above, many findings concerning the structures of resin glycosides have been obtained so far. Taking those findings into account, it would be preferable that the terms 'jalapin' and 'convolvulin' should not be used based on their solubility in ether as classified by Mayer, but rather to describe structural groups, i.e., the resin glycosides having intramolecular cyclic structures and the acylated glycosidic acid monomers and polymers with free carboxylic acid forms, respectively [61].

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