UROLOGY - REVIEW



The effects of radiation on myeloid lineage immune cells within the rodent urinary bladder: a systematic review

Jessica Smith¹ · Rimaz Toto¹ · Christian Moro¹

Received: 15 May 2023 / Accepted: 14 August 2023 / Published online: 24 August 2023 © The Author(s) 2023

Abstract

Purpose Radiotherapy is a prominent therapy for many malignant and non-malignant disorders, though it can cause side effects such as radiation-induced cystitis. Current research has highlighted a role for mast cells and macrophages in the prognosis of such radiation-induced toxicities. However, the prognostic value of these immune cells in the pathophysiology of radiation-induced cystitis is not clear. As such, a systematic review was conducted to assess myeloid-lineage immune cells for their prognostic value in radiation-induced cystitis to address this gap in literature.

Methods The protocol was registered in PROSPERO, and searches were performed in PubMed, Embase and Web of Science databases for pre-clinical rodent studies on radiation-induced cystitis.

Results After de-duplication, 153 articles were screened for relevancy by title and abstract. Title and abstract screening deemed 64 studies irrelevant. The remaining 85 studies were full-text screened, yielding seven unique articles for data extraction. Most included studies had an unclear risk of bias. The findings of this systematic review suggest that the prognostic value of myeloid-lineage immune cells in radiation-induced cystitis is still unclear, indicating a need for further research in this field.

Conclusion Although the studies reviewed provide some insight into the role of these immune cells in disease pathology, the limited number of studies and unclear risk of bias further highlights a need for additional, high-quality research in this area. In summary, this systematic review highlights a need to understand the involvement of immune cells in radiation-induced cystitis pathophysiology and lay the groundwork for further research in this area. **Trial registration** PROSPERO registration: CRD42022345960

Indiregistration PROSPERO registration: CRD42022545900

Keywords Radiation-induced cystitis \cdot Haematopoietic stem cell transplantation \cdot Radiotherapy \cdot Cystitis \cdot Urinary bladder \cdot Immune cells

Introduction

Radiation-induced cystitis is an inflammatory condition that can develop as a side-effect of abdominopelvic radiation exposure [1]. The incidence of radiation-induced cystitis is suggested to range from 23% to 80%, with this variability likely being a result of variable doses of radiotherapy across medical subspecialties [1]. External-beam radiation therapy (EBRT) is a common type of radiation therapy, involving the use of a linear accelerator to deliver a focussed beam of radiation to malignant tissue. Radiation-induced cystitis

Christian Moro cmoro@bond.edu.au can also be induced by myeloablative therapy, which is used a conditioning regiment for hematopoietic stem cell transplantation (HSCT) in the treatment of various hematological diseases by delivering high-dose radiation to the entire body [2].

Radiation-induced cystitis is thought to develop triphasically. The first phase of radiation-induced cystitis is characterized by acute inflammation, involving a loss of the urinary bladder's glycosaminoglycan and urothelium cell layers, infiltration of immune cells, vasodilation, and hyperplasia of bladder endothelium. The acute phase is then followed by a symptom-free phase, lasting from months to years [3, 4]. The third and final phase constitutes a chronic inflammatory response. The chronic phase is best characterized by collagen deposition, loss of smooth muscle epithelia, hemorrhaging, thinning of the urothelial wall and an influx

¹ Faculty of Health Sciences and Medicine, Bond University, Queensland 4226, Australia

of pro-inflammatory immune cells. However, the extent of immune cell distribution and prevalence during acute and chronic phases of radiation-induced cystitis has not been well established, and more importantly, whether these characteristics apply to myeloablative therapy considering its immunosuppressive nature. Though current research has highlighted a possible role for urinary bladder mast cells [3, 5] and macrophages [6] in the development of radiationinduced cystitis, their specific role is unclear.

In pre-clinical literature, mast cells have been associated with the development of fibrosis in various tissues in response to radiation exposure [7-11]. Specifically, mast cells are believed to release various chemokines, such as transforming growth factor- β (TGF- β), which promotes fibroblast recruitment and proliferation [12], causing fibrosis. Urinary bladder fibrosis has been previously associated with underactive bladder and impaired bladder compliance disorders [13, 14]. Clinical presentations of radiationinduced cystitis range from increased urinary frequency and urgency, dysuria, nocturia, increased risk of bladder infections, hematuria and suprapubic pain, as reviewed by Zwaans, Nicolai [15]. Mast cells are also believed to disrupt vasculature by increasing the permeability of blood vessels [16] as well as playing a direct role in chemical cascades that result in radiation-associated toxicities [9]. Similarly, macrophages are also suggested to play a role in the pathophysiology of radiation-induced cystitis, namely in promoting fibrosis and inflammation, as reviewed by Helissey, Cavallero [6]. Other myeloid-lineage immune cells, such as monocytes, neutrophils, eosinophils and basophils, may be involved, but less is known of their role in the development of radiation-induced inflammation. Hence, this systematic review aims to establish the potential prognostic value of myeloid-derived immune cells in radiation-induced cystitis by reviewing current pre-clinical literature on their involvement in tissue injury and repair of the irradiated urinary bladder.

Materials and methods

This systematic review followed the Preferred Reporting Items for Systematic Reviews and Meta-Analyses statement (PRISMA) guidelines [17]. The protocol for the systematic review was prospectively developed and registered in PROS-PERO (CRD42022345960).

Search strategy

Embase, PubMed, and Web of Science online databases were searched for published articles from database inception to 16 November 2022. Language and year of publication restrictions were not imposed in the systematic review. Studies were identified using the following terms: radiation, radiotherapy, hematopoietic stem cell transplantation, urinary bladder, urothelium, cystitis, rodents, immunomodulation, fibrosis, inflammation, radiation effects, radiation fibrosis syndrome, leukocytes, macrophages, monocytes, mast cells, neutrophils, eosinophils and basophils. The search string used to search the PubMed database is found below:

("Radiation, Ionizing"[tiab] OR "Radiation"[tiab] OR "Radiotherapy"[tiab] OR "Haematopoietic stem cell transplantation"[tiab]) AND ("Urinary Bladder"[tiab] OR "Urothelium"[tiab] OR "Cystitis"[tiab] OR "Bladder"[tiab]) AND ("Rodent"[tiab] OR "Mice"[tiab] OR "Rats"[tiab] OR "Guinea Pigs"[tiab]) AND ("Immunomodulation"[tiab] OR "Fibrosis"[tiab] OR "Inflammation"[tiab] OR "Radiation Effects"[tiab] OR "Radiation Exposure"[tiab] OR "Radiation Injuries"[tiab] OR "Radiation Fibrosis Syndrome"[tiab] OR "Leukocytes"[tiab] OR "Mast Cells"[tiab] OR "Basophils"[tiab] OR "Macrophages"[tiab] OR "Monocytes"[tiab]).

Eligibility criteria

Studies were included in this systematic review if they satisfied the following criteria: (i) included radiotherapy, (ii) assessed myeloid-lineage immune cell function, prevalence or distribution, and (iii) evaluated fibrosis, inflammation or immunosuppression in irradiated rodent urinary bladders. Only original experimental animal studies were included. Animals with infectious diseases or studies without healthy control animals were excluded. Reviews, systematic reviews, meta-analyses and conference abstracts were excluded from this systematic review. Human studies were also excluded from this review but were included in the introduction and discussion to provide a contextual clinical application.

Study screening, data extraction and quality assessment

Two independent reviewers performed study screening, and disagreements were resolved by the third reviewer. After duplicate removal, title and abstracts were screened to exclude irrelevant studies. One author retrieved full-text articles, and two authors screened full-text articles against the eligibility criteria for inclusion in this systematic review.

After full-text article screening, two reviewers extracted data from eligible studies. Information relating to the population (total animal numbers, animal species, strain, sex and age) and intervention (type of radiation, dosage, target tissue, adjuvant treatments) was extracted, in addition to information relating to primary and secondary outcome measures. The primary outcome measure was to assess the impact of radiation on myeloid-lineage immune cells in relation to their function, prevalence or distribution. The secondary outcome measure was to assess the impact of radiation on the urinary bladder with regard to inflammation, fibrosis and immunosuppression. Where possible, data relating to the mean, standard error of mean, or standard deviation were extracted from the main text. Only data from the last time point were extracted when outcome measures were assessed at multiple times. Estimates were cross-checked, and variances in data extraction exceeding 10% were resolved through discussion between all reviewers. Extracted data were then entered into Covidence by one reviewer and checked by another reviewer independently.

Two review authors independently assessed the risk of bias for each included study using the criteria outlined by Systematic Review Centre for Laboratory animal Experimentation [18]. All disagreements were resolved by discussion or by referring to a third author.

Results

Study selection and qualitative assessment

A total of 257 studies were identified in the initial online database search after duplicate removal. 153 studies' titles and abstracts were then assessed for relevancy. From the title and abstract screening, 64 studies were deemed irrelevant. The remaining 85 studies were full text screened against the inclusion and exclusion criteria. A total of seven unique studies were included for data extraction (Fig. 1).

Risk of bias assessment

Study quality was assessed using the Systematic Review Centre for Laboratory animal Experimentation (SYRCLE) guidelines [18]. Most included studies had an unclear risk of bias, illustrated in Fig. 2. No studies stated randomized housing or blinding in performance and detection biases. One study did not explain variance in sample sizes [19], scoring high for risk of bias in "Attrition Bias: Incomplete Outcome Data." All studies had an unclear risk of selective outcome reporting bias. All studies in "Other: Other Sources of Bias" included disclosure of ethics approval or potential conflict of interest statement except for Giglio, Wasén [20].

Study characteristics

Of the seven included studies, five analyzed rat urinary bladders [19-23], and two analyzed the mouse urinary bladder [8, 24]. Five studies were conducted on female urinary bladders [8, 19-21, 24] and two studies were



Fig. 1 PRISMA search strategy flow chart

conducted on male urinary bladders [22, 23]. One author was contacted to clarify the age of rodents analyzed [20]. The most common rodent strain was Sprague–Dawley rats (Table 1).

All studies utilised EBRT to irradiate the urinary bladder. The highest total radiation dose delivered to the urinary bladder was 40 Gy [21]. Three studies delivered a single dose of 20 Gy radiation to the urinary bladder [8, 19, 20], one used 15 Gy [24] and another used 10 Gy [23]. The lowest total radiation dose to the rodent urinary bladder was 300 cGy (3 Gy), delivered twice.

Five of the seven studies analyzed mast cells, and three analyzed macrophages in the irradiated urinary bladder (Table 1). In relation to mast cells, three studies assessed prevalence [8, 19, 22], and three studies assessed distribution across the urinary bladder wall [19, 20, 23]. Three studies assessed macrophage prevalence in the irradiated urinary bladder [19, 21, 24], but not distribution. One study suggested a function for macrophages in the irradiated bladder but did not directly assess macrophage function [21]. No studies examined the effect of radiation on monocytes, neutrophils, eosinophils, or basophils in the irradiated urinary bladder (Table 1).



B



Fig. 2 SYRCLE risk of bias assessment of included studies: **A** indicates the scoring of individual studies, and **B** indicates overall scores, presented as a percentage. Symbols: green tick indicates low risk, the

yellow question mark indicates unclear risk, red cross indicates a high risk of bias

| Author | Country | Species (strain) | Age | Sex | Type of radiation | Dose | Primary outcomes | Secondary outcomes |
|-------------------------------------|---------|-----------------------|-------------|--------|--|--|--|---|
| Ito, Yamamoto [21] | Japan | Rats (Fischer F344) | 9weeks | Female | X-irradiation, targeted at the pelvis, filtered through a 2 mm lead plate | 40 Gy total dose in eight 5 Gy fractions (two times a week); 4.9 Gy/min dose rate at 150 kV and 20 mA for 1.021 min | Radiotherapy reduced the prevalence of CD68p and CD163 macrophages (NSD) | The infiltration of mac- rophages into the uri- nary bladder suggested to reflect inflammation; as macrophage preva- lence appeared to reduce (NSD), inflammation was not reported |
| Lombardo, Obradovic [24] | NSA | Mice (C57BL/6 J) | 4 to 6weeks | Female | External beam radiation | Single 15 Gy dose | Increased immunostimu- latory macrophage marker CD86 observed in immunohistochem- istry (NSD) | Radiotherapy appeared to decrease CD4 + T-cells, but not CD8 + T-cells (NSD) |
| Podmolfková, Mukan- yangezi [19] | Sweden | Rats (Sprague-Dawley) | 12 weeks | Female | 6MV linear accelerator, administered in two side-fields | Single 20 Gy dose | Mast cells decrease (NSD) in the submu- cosa, increase in the detrusor muscle (NSD) and no change in the adventitia (NSD). No change in mast cell prevalence when exposed to irradiation (NSD) Marginal increase in CD206+macrophages in the urothelium (NSD) | CD3+lymphocytes decrease in the urothe- lium and lamina propria (NSD) Fibrotic area and collagen deposition significantly increased |
| Chen, Chen [22] | Taiwan | Rats (Sprague-Dawley) | Adult | Male | Brachytherapy targeted towards the bladder surface over the skin | 300 cGy twice, over a 4 h time interval | Mast cells observed to infiltrate the urinary bladder by day 28 after chronic cystitis induc- tion ($p < 0.05$) | Interleukin (IL)-6 and turmour necrosis factor (TNF)- α were signifi- cantly higher in chronic cystitis groups |
| Zwaans, Krueger [8] | USA | Mice (C3H/HeN) | 8 weeks | Female | Small animal radiation research platform (SARRP) | Single 20 Gy dose | No significant influx of mast cells noted in response to radiation | Mild increase in inflam- matory cells (NSD) was detected in response to irradiation Irradiation caused a mod- est increase in fibrosis in the bladder in response to control |

Table 1 Study characteristics summary

| Table 1 (continued) | | | | | | | | |
|-----------------------|--------------|-----------------------------|------------------|------------|---|--|---|--|
| Author | Country | Species (strain) | Age | Sex | Type of radiation | Dose | Primary outcomes | Secondary outcomes |
| Giglio, Wasén [20] | Sweden | Rats (Sprague-Dawley) | Adult | Female | 6MV linear accelerator, targeting the urinary bladder | Single 20 Gy dose | After irradiation, the number of mast cells decreased significantly in the urothelium, but not the detrusor muscle | The number of lympho- cytes in the bladder wall was low and appeared to not be affected by radiation No differences in the expression of IL-1b, IL-4, IL-13 and IL-5 and IFN-c was observed between control and irradiated samples after 14 days. IL-1b and IL-13 expressions in the urothelium decreased in response to bladder irradiation; indicated consistent with signs of an anti-inflammatory response |
| Barcellos, Costa [23] | Brazil | Rats (wistar) | 3-4 months 1 | Male | 10 MeV photon beam generated by a linear accelerator | 2.4 Gy/min for 4.16 min to a total dose of 10 Gy | Mast cell density in the lamina propria was reduced at seven (mean 1.4; full range 0–3) and 15 (1.0; 0–2) days after irradiation com- pared with controls (5.2; 0–9) | |
| Ito, Yamamoto [21] ai | id Lombardo. | , Obradovic [24] included t | the use of addit | tional tre | atments, namely that the t | urinary bladders were also e | xposed to various forms of | f bladder cancer. Data have |

ve tio, ramanou [21] and compared. Obtained us use of additional readictives that us unitary planets were also exposed to various forms of planet cancer. Data have been extracted from irradiated urinary bladders without exposure to cancer or additional therapeutics. Podmolíková, Mukanyangezi [19] was contacted to specify an age for their rodent popula-tion. Barcellos, Costa [23] did not assess any secondary outcomes.

🖄 Springer

Effects of radiation on mast cells and macrophages in the urinary bladder

Podmolíková, Mukanyangezi [19] and Zwaans, Krueger [8] delivered a single dose of 20 Gy to the urinary bladder by external beam radiation technologies (linear accelerator or SAARP) in female Sprague–Dawley rats after two weeks and female C3H/HeN mice after 25 weeks, respectively. Both studies concluded no statistically significant difference (NSD) in total mast cell prevalence in the irradiated urinary bladder. Chen, Chen [22] also reported mast cell prevalence in Sprague–Dawley rats, finding that 28 days post-irradiation, mast cells infiltrated the male urinary bladder (p < 0.05).

Regarding mast cell distribution, Podmolíková, Mukanyangezi [19] reported no statistical difference in mast cell number in the lamina propria (NSD), detrusor muscle (NSD) and adventitia of the urinary bladder (NSD). Barcellos, Costa [23] and Giglio, Wasén [20] reiterated these findings in male Wistar rats exposed to 10 Gy and female Sprague–Dawley rats exposed to a single dose of 20 Gy, respectively. Barcellos, Costa [23] reported no significant changes in mast cell density in the lamina propria at sevenand 15-day time points following radiation exposure (NSD) compared with controls. Similarly, Giglio, Wasén [20] reported a significant decrease of mast cells in the urothelium (p < 0.05) but no observable changes in the detrusor muscle of the irradiated urinary bladder after 14 days (NSD).

Ito, Yamamoto [21] exposed 40 Gy of radiation to the urinary bladder of female Fischer F344 rats, finding that the prevalence of CD68b⁺ and CD163⁺ macrophages did not change (NSD) seven days following irradiation. At 20 Gy, Podmolíková, Mukanyangezi [19] reported no changes in CD206⁺ macrophages in the irradiated urothelium (NSD). Additionally, Lombardo, Obradovic [24] observed no changes in the CD86⁺ macrophage marker (NSD).

Inflammation, fibrosis and immunosuppression following radiation exposure

Four studies assessed inflammation in the urinary bladder following radiation exposure [8, 20–22]. Giglio, Wasén [20] reported that mRNA expressions of inflammatory markers: IL-1 β , IL-4, IL-5 and IL-15 in irradiated urinary bladders were similar to that of control mRNA levels 14 days postirradiation, suggesting that inflammatory protein expression did not change as a result of radiation exposure. Dissimilarly, semi-quantitative data analysis indicated that IL-1 β and IL-13 expression was down-regulated 14 days following irradiation exposure compared to controls (NSD) [20]. Zwaans, Krueger [8] also assessed inflammation by assessing the prevalence of polymorphonuclear cells, lymphocytes, plasma cells and macrophages. A mild increase in these inflammatory cells was reported in response to a single dose of 20 Gy radiation, but was ultimately not statistically significant. Similarly, Ito, Yamamoto [21] assessed inflammation by the infiltration of macrophages into the urinary bladder, in which no changes were observed in macrophages seven days following irradiation (NSD). Supporting both Zwaans, Krueger [8] and Ito, Yamamoto [21], an inflammatory response was also reported by Chen, Chen [22] in the urinary bladders of male rats exposed to 300 cGy twice. In this study, Chen, Chen [22] reported an increase in IL-6 and TNF- α inflammatory markers in irradiated urinary bladders compared to controls, suggesting an inflammatory response.

Two studies assessed fibrosis in the urinary bladder following radiation exposure [8, 19]. Zwaans, Krueger [8] reported a modest increase in fibrosis in the urinary bladder of C3H/HeN mice in response to 20 Gy radiation, but was not statistically significant. In contrast, Podmolíková, Mukanyangezi [19] reported a significant increase in the total fibrotic area and collagen deposition of the rat urinary bladder in response to 20 Gy irradiation (p < 0.05).

Three of the seven included studies reported on other immune cells' prevalence in the urinary bladder [19, 20, 24]. Studies conducted by Podmolíková, Mukanyangezi [19] and Lombardo, Obradovic [24] both reported a decrease in lymphocytes in the urinary bladder. Specifically, Podmolíková, Mukanyangezi [19] reported a decrease in CD3⁺ lymphocytes in the urothelium and lamina propria of the irradiated urinary bladder; however, no statistical significance was observed. Lombardo, Obradovic [24] reported a decrease in the number of CD4⁺ T-cells (p < 0.05) but no change to CD8⁺ T-cells in response to irradiation (NSD). Giglio, Wasén [20] reported that the prevalence of lymphocytes in the irradiated bladder was not affected by irradiation, where no statistical significance was found between control and irradiated groups.

Discussion

Radiation-induced cystitis, a known side effect of radiation exposure to the urinary bladder, can significantly impact patient outcomes [2, 25, 26]. The lack of evidence-based research and therapeutics available in treating this disease remains a significant issue that needs to be addressed. Current pre-clinical research has demonstrated a potential role for macrophages [6] and mast cells [7–11] in the pathophysiology of radiation-induced toxicities; however, a clear link has not been established for the involvement of these immune cells, and other myeloid-derived immune cells, in the prognosis of radiation-induced cystitis.

This systematic review revealed that the effect of radiation on myeloid-derived immune cells in radiation-induced cystitis is poorly defined. In other literature, a role for the urinary bladder mast cell [3, 5] in the pathophysiology of radiation-induced cystitis has been suggested. In the healthy urinary bladder, mast cells are critical regulators of immunity [27–29] distributed throughout all layers of the urinary bladder wall [30]. In the context of radiation-induced toxicities, mast cells have been associated with the development of radiation-induced fibrosis around various tissues of the body [7–11]. Specifically, mast cells are believed to release various chemokines, such as TGF- β , which cause fibrosis by promoting fibroblast recruitment and proliferation [12]. Mast cells are also believed to disrupt vasculature by increasing the permeability of blood vessels [16] as well as playing a direct role in chemical cascades that result in radiation-induced inflammation [9].

In the reviewed studies, several investigated mast cells in the irradiated urinary bladder [8, 19, 20] [22, 23]. Two studies indicated no statistical difference in the infiltration of mast cells between irradiated and non-irradiated groups [8, 19]. These studies suggest that mast cells are resistant to radiation exposure in the urinary bladder, consistent with reports on mast cells exhibiting resistance to radiationinduced cytotoxicity [31, 32]. Contrastingly, two studies indicated a decrease [20, 23] and one study indicated an increase in mast cell density following irradiation [22]. Mast cells were reported to decrease in the urothelium [20] and lamina propria [23], which may suggest that mast cells are more susceptible to radiation-induced toxicities in the most apical layers of the urinary bladder wall following irradiation at 14-day, and 7- and 15-day timepoints, respectively. After 28 days, Chen, Chen [22] reported that mast cells infiltrated the bladder, suggesting that this increase in mast cells may be indicative of inflammatory response. Despite this, further research is needed to validate these findings.

In addition to mast cells, macrophages have also been implicated in the development of radiation-induced toxicities through similar mechanisms of inflammation and fibrosis [33]. Macrophages were investigated in three of the included studies [19, 21, 24], in which all three studies concluded no statistical difference in macrophage prevalence following irradiation. Despite no statistical difference between non-irradiated and irradiated groups, Podmolíková, Mukanyangezi [19] and Lombardo, Obradovic [24] reported a marginal increase in CD206⁺ macrophages and increased CD86⁺ macrophage-associated markers in the urinary bladder. Ito, Yamamoto [21], however, reported a reduction in CD68b macrophages, suggesting that CD68b macrophages may be susceptible to radiation.

Though previous research has suggested a role for urinary bladder mast cells and macrophages in the pathophysiology of radiation-induced cystitis, this systematic review has determined that these cells do not have a clear prognostic value in the development of this disease. Firstly, only four studies assessed inflammation following irradiation of the urinary bladder [8, 20–22], two of which identified inflammation in the irradiated urinary bladder [8, 20]. Incongruencies in techniques used to assess cystitis, as well as a lack of assessment in several studies, make it difficult to draw meaningful conclusions about the involvement of mast cells or macrophages in relation to radiationinduced cystitis, a disease characterized by inflammation. Thus, it is recommended that a guideline is developed for the assessment of radiation-induced cystitis in pre-clinical models to allow for more meaningful and accurate comparisons in the assessment of radiation-induced cystitis, with particular regard to its triphasic development [3, 4].

Of particular interest was the lack of any literature on the impact of myeloablative radiotherapies, or the associated HSCT on myeloid lineage immune cells. Despite its curative potential, HSCT and its conditioning regiments have been linked to radiation-induced cystitis. The development of radiation-induced cystitis following HSCT is associated with increased disease morbidity, prolonged hospitalisations [2, 25] and significant increases in nonrelapse-related mortality, according to Galli, Sorà [26]. The prevalence of radiation-induced cystitis, associated with HSCT conditioning regimens, has a reported prevalence ranging between 10% and 20% in paediatric transplant recipients [34] and 17% in adult transplant recipients [35], with the median incidence of cystitis reported to occur within the first 30 days of transplantation [26]. Due to its prevalence and apparent risk to immunocompromised patients, future research on HSCT and myeloablative radiotherapies is highly recommended.

Only three of the included studies in this systematic review assessed the prevalence of other immune cells in the urinary bladder [19, 20, 24], and found that there were no statistically significant differences in assessed lymphocytes [19, 20]. However, Lombardo, Obradovic [24] reported no change to the number of CD4⁺ and CD8⁺ T-cells in response to irradiation compared to controls (NSD). To this end, it is well known that myeloablative radiotherapy can induce immunosuppression [36]. As all included studies utilized EBRT to evaluate myeloid-lineage immune cells in the irradiated urinary bladder, it is unclear as to whether these findings are applicable to myeloablative radiotherapy. Dually, it is also recommended that future research is done to validate a model of radiation-induced cystitis, in consideration of different species, strains, dosimetry, and modes of radiation. Due to scarcity in the assessment of other myeloid-lineage immune cells, namely monocytes, neutrophils, eosinophils and basophils, future studies should also aim to assess the function, prevalence, and distribution of a variety of immune cells in the irradiated bladder, focusing on how these characteristics may contribute to disease pathology or how they may differ in the context of myeloablative radiotherapy.

Limitations of this systematic review include the small number of identified studies and the inability to make valuable comparisons due to the different species, strains, dosimetry, and techniques used. Additionally, it is important to recognise that only four of the included studies sought to investigate cystitis resulting from radiation exposure [8, 19, 20, 22]. The remaining three studies [21, 23, 24] had discussed inflammation resulting from radiation exposure, but did not explicitly identify cystitis. These studies were still included in this systematic review, however, as they still met the inclusion criteria.

Conclusion

In summary, this systematic review highlights a need for further research into the pathology of radiation-induced cystitis. Further research is needed to identify the role of myeloid-lineage immune cells in the development of radiation-induced cystitis (amongst other radiation-induced toxicities in the urinary bladder), particularly in the context of myeloablative radiotherapy. Specifically, pre-clinical research should further investigate the potential prognostic role of mast cells, macrophages, and other myeloid-lineage immune cells in the development of radiation-induced cystitis, as well as continuing to refine current models of radiation-induced cystitis to aptly reflect the pathophysiology of this disease.

Funding Open Access funding enabled and organized by CAUL and its Member Institutions. Not applicable.

Data availability Data can be made available upon reasonable request by emailing the corresponding author.

Declarations

Conflict of interest The authors declare no conflict of interest.

Patient consent Not applicable.

Permission to reproduce material from other sources Not applicable.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- 1. Browne C et al (2015) A Narrative review on the pathophysiology and management for radiation cystitis. Adv Urol 2015:346812
- Li DZ et al (2012) Comparison of total body irradiation before and after chemotherapy in pretreatment for hematopoietic stem cell transplantation. Cancer Biother Radiopharm 27(2):119–123
- Zwaans BM, Chancellor MB, Lamb LE (2016) Modeling and Treatment of Radiation Cystitis. Urology 88:14–21
- Manikandan R, Kumar S, Dorairajan LN (2010) Hemorrhagic cystitis: a challenge to the urologist. Ind J Urol 26(2):159–166
- Shelburne CP et al (2009) Mast cells augment adaptive immunity by orchestrating dendritic cell trafficking through infected tissues. Cell Host Microbe 6(4):331–342
- Helissey C et al (2020) Chronic inflammation and radiationinduced cystitis: molecular background and therapeutic perspectives. Cells 10(1):21
- Brossard C et al (2022) Understanding molecular mechanisms and identifying key processes in chronic radiation cystitis. Int J Mol Sci 23(3):1836
- Zwaans BM et al (2016) Modeling of chronic radiation-induced cystitis in mice. Adv Radiat Oncol 1(4):333–343
- Blirando K et al (2011) Mast cells are an essential component of human radiation proctitis and contribute to experimental colorectal damage in mice. Am J Pathol 178(2):640–651
- Strattan E et al (2019) Mast cells are mediators of fibrosis and effector cell recruitment in dermal chronic graft-vs-host disease. Front Immunol. https://doi.org/10.3389/fimmu.2019.02470
- Zheng H, Wang J, Hauer-Jensen M (2000) Role of mast cells in early and delayed radiation injury in rat intestine. Radiat Res 153(5 Pt 1):533–539
- Conti P et al (2018) Critical role of inflammatory mast cell in fibrosis: Potential therapeutic effect of IL-37. Cell Prolif 51(5):e12475
- 13. Kim SJ et al (2021) Irreversible bladder remodeling induced by fibrosis. Int Neurourol J 25(1):S3-7
- 14. Fry CH et al (2018) Fibrosis and the bladder, implications for function ICI-RS 2017. Neurourol Urodyn 37(4):S7-s12
- Zwaans BM et al (2016) Challenges and opportunities in radiation-induced hemorrhagic cystitis. Rev Urol 18(2):57–65
- Park KR et al (2016) Mast cells contribute to radiation-induced vascular Hyperpermeability. Radiat Res 185(2):182–189
- Shamseer L et al (2015) Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015: elaboration and explanation. BMJ Brit Med J 349:g7647
- Hooijmans CR et al (2014) SYRCLE's risk of bias tool for animal studies. BMC Med Res Methodol 14(1):43
- Podmolíková L et al (2020) Radiation of the urinary bladder attenuates the development of lipopolysaccharide-induced cystitis. Int Immunopharmacol 83:106334
- Giglio D et al (2016) Downregulation of toll-like receptor 4 and IL-6 following irradiation of the rat urinary bladder. Clin Exp Pharmacol Physiol 43(7):698–705
- Ito Y et al (2022) Ascorbic acid-2 glucoside mitigates intestinal damage during pelvic radiotherapy in a rat bladder tumor model. Int J Radiat Biol 98(5):942–957
- Chen YT et al (2018) Extracorporeal shock wave markedly alleviates radiation-induced chronic cystitis in rat. Am J Transl Res 10(3):1036–1052
- Barcellos LM et al (2013) Protective effects of l-glutamine on the bladder wall of rats submitted to pelvic radiation. Micron 47:18–23
- Lombardo KA et al (2022) BCG invokes superior STING-mediated innate immune response over radiotherapy in a carcinogen murine model of urothelial cancer. J Pathol 256(2):223–234

- 25. Silva Lde P et al (2010) Hemorrhagic cystitis after allogeneic hematopoietic stem cell transplants is the complex result of BK virus infection, preparative regimen intensity and donor type. Haematologica 95(7):1183–1190
- Galli E et al (2022) Hemorrhagic cystitis in allogeneic stem cell transplantation: a role for age and prostatic hyperplasia. Supp Care Cancer 30(6):4953–4959
- 27. Wu Z et al (2019) Pyroptosis engagement and bladder urothelial cell-derived exosomes recruit mast cells and induce barrier dysfunction of bladder urothelium after uropathogenic E coli infection. Am J Physiol-Cell Physiol 317(3):544–555
- He S, Walls AF (1998) Human mast cell chymase induces the accumulation of neutrophils, eosinophils and other inflammatory cells in vivo. Br J Pharmacol 125(7):1491–1500
- Atiakshin D, Buchwalow I, Tiemann M (2019) Mast cell chymase: morphofunctional characteristics. Histochem Cell Biol 152(4):253–269
- Choi HW et al (2016) Loss of Bladder Epithelium Induced by Cytolytic Mast Cell Granules. Immunity 45(6):1258–1269
- Soule BP et al (2007) Effects of gamma radiation on FceRI and TLR-mediated mast cell activation. J Immunol 179:3276

- Murakami S et al (2015) Effects of ionizing radiation on differentiation of murine bone marrow cells into mast cells. J Rad Res 56:865
- Meziani L, Deutsch E, Mondini M (2018) Macrophages in radiation injury: a new therapeutic target. Oncoimmunology 7(10):e1494488
- Decker DB, Karam JA, Wilcox DT (2009) Pediatric hemorrhagic cystitis. J Pediatr Urol 5(4):254–264
- Manikandan R, Kumar S, Dorairajan LN (2010) Hemorrhagic cystitis: a challenge to the urologist. Indian J Urol 26(2):159–166
- Sabloff M et al (2021) Total body irradiation for hematopoietic stem cell transplantation: what can we agree on? Curr Oncol 28(1):903–917

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.