

Transition metal complexes of triazole-based bioactive ligands: synthesis, spectral characterization, antimicrobial, anticancer and molecular docking studies

Yogesh Deswal¹ · Sonika Asija¹ · Deepak Kumar² · Deepak Kumar Jindal³ · Gourav Chandan⁴ · Vivek Panwar² · Sonia Saroya¹ · Naresh Kumar¹

Received: 27 August 2021 / Accepted: 27 October 2021 / Published online: 23 November 2021 © The Author(s), under exclusive licence to Springer Nature B.V. 2021

Abstract

In the present research work, four new heterocyclic Schiff base ligands (1-4) were synthesized by the condensation of 4-(4-amino-5-mercapto-4*H*-1,2,4-triazol-3-yl) phenol with salicylaldehyde derivatives in 1:1 molar ratio. The synthesized Schiff base ligands were allowed for complexation with Co(II), Ni(II), Cu(II), Zn(II) metal ions. The structure of the newly synthesized compounds (1-20) was elucidated with

🖂 Sonika Asija sonika@gjust.org Yogesh Deswal deshwalyogesh72@gmail.com Deepak Kumar guptadeepak002@gmail.com Deepak Kumar Jindal deepakjindal44@yahoo.com Gourav Chandan cggourav687@gmail.com Vivek Panwar vivekpanwar345@gmail.com Sonia Saroya soniasaroya214@gmail.com Naresh Kumar nareshjanagal91@gmail.com 1 Department of Chemistry, Guru Jambheshwar University of Science and Technology, Hisar 125001, Haryana, India 2 Department of Pharmaceutical Chemistry, School of Pharmaceutical Sciences, Shoolini University, Solan 173229, India

- ³ Department of Pharmaceutical Sciences, Guru Jambheshwar University of Science & Technology, Hisar 125001, Haryana, India
- ⁴ School of Biotechnology, Faculty of Applied Sciences and Biotechnology, Shoolini University, Solan 173229, India

the help of various spectral and physicochemical techniques. Spectroscopic data confirm the tridentate nature of ligands which coordinate to the metal via deprotonated oxygen, azomethine nitrogen and thiol sulphur. Conductivity data showed the non-electrolytic nature of complexes. Furthermore, the synthesized compounds were evaluated for their *in-vitro* antimicrobial activity against four pathogenic bacterial strains and two pathogenic fungal strains. The observed results of microbial activity reveals that compound **3** and its complexes (**13–16**) were found most potent against the pathogenic strains. In addition, the anticancer activity of all the synthesized compounds was evaluated against human carcinoma cell lines i.e. HCT-116, DU145 and A549 using MTT assay. Among the tested compounds **12**, **19**, and **20** were found to show promising potency against the cancer cell lines. To rationalize the preferred modes of interaction of most potent compounds with the active site of human EGFR protein (PDB id: 5XGM), molecular docking studies were performed.

Graphical abstract



Keywords Triazole Schiff base ligands \cdot Metal complexes \cdot Antimicrobial \cdot Anticancer \cdot Molecular docking \cdot EDAX

Introduction

Cancer emerges as a serious health problem in all populations and ranks as the second most leading cause of death all over the world. The International Agency for Research on Cancer (IARC) in 2020 estimated 19.3 million new cancer cases and almost 10.0 million cancer deaths worldwide [1]. The projected death toll due to cancer is increasing at an alarming rate. It is estimated that 29.4 million people will have cancer in 2040 (WHO, 2020) [2]. Recent studies have shown that cancer patients are more likely to be

infected with COVID-19 and serious side effects such as ARDS, liver injury, sepsis, renal insufficiency and myocardial injury are commonly observed in cancer patients with COVID-19 [3]. The current therapeutic strategies used in the treatment of cancer involve chemotherapy, radiotherapy, surgery and immunotherapy either alone or in combination, but the constitutional treatment of cancer is based on chemotherapy which incorporates the use of various chemical entities of natural and synthetic origin [4]. The serendipitous discovery of cis-platin drug has established the importance of metal complexes in cancer cure [5]. But the clinical application of cis-platin is limited due to significant side effects such as bone marrow suppression, hearing problems, vomiting and kidney damage etc. [6, 7]. Therefore the development of novel metal-based drugs with no toxicity or very less toxicity for the treatment of cancer is nowa-days attracting the attention of biochemists.

The coordination chemistry of Schiff base ligands still evokes much current interest and possesses a variety of applications [8–14]. Furthermore, it is also reported that the activity of Schiff base ligands gets increased when administered as metal complexes [15]. Schiff base complexes possess remarkable properties which make these compounds effective as stereospecific catalysts towards hydrolysis, oxidation, reduction and other transformations of inorganic and organic chemistry [16–20]. Heterocyclic ligands containing nitrogen as donor atom, especially 1,2,4-triazole have received much attention due to their huge applications in analytical chemistry [21–24]. 1,2,4-triazole possesses a wide variety of pharmacological activity including antimicrobial, anti-cancer, anti-diabetic, anti-tubercular, anxiolytics, anticonvulsants, etc. [25–30]. Schiff base ligands having triazole ring can be considered as "privileged structures" and are widely used due to their novel structural features. Transition metal complexes have effectively established them as good antimicrobial and anticancer agents [31, 32]. Therefore, the formation of transition metal chelates of triazole derived Schiff base ligands as new metallodrugs for the treatment of various diseases is found to have great potential.

Keeping in view the immense biological activity exhibited by 1,2,4-triazole motif and the potential chemistry of transition metals, we find it lively to join the chemistry of both moieties in a single molecular framework. In the recent study, we report the synthesis of some new Schiff base ligands derived from the condensation of 4-(4-amino-5-mercapto-4*H*-1,2,4-triazol-3-yl)phenol with salicylaldehyde derivatives. Using these triazole containing Schiff base ligands Co(II), Ni(II), Cu(II), Zn(II) complexes were synthesized. Characterization of the synthesized compounds was done by ¹H NMR, ¹³C NMR, FTIR, HRMS, XRD, ESR, TGA and SEM. The work was further extended to investigate the antimicrobial and anticancer activities of the synthesized compounds. Molecular docking studies on the most potent compounds were carried out to check the binding conformations.

Experimental

Materials

The entire chemicals employed in this research work were procured from Sigma Aldrich. The chemicals viz., 3-ethoxysalicylaldehyde (97%), 5-bromosalicylaldehyde

(98%), 5-nitrosalicyladehyde (98%), 3,5-dibromosalicylaldehyde (98%), 4-hydroxybenzhydrazide (\geq 98%), Hydrazine hydrate (50–60%) and metal acetates solids i.e. Co(CH₃COO)₂·4H₂O (98%), Ni(OCOCH₃)₂·4H₂O (98%), Cu(CH₃COO)₂·H₂O (\geq 98%), Zn(CH₃COO)₂·2H₂O (\geq 98%) were used as such in the present study. The organic solvents were of analytical grade and used after drying with standard procedures [33]. The progress of the reaction and the purity of synthesized compounds were monitored regularly by TLC (thin layer chromatography).

Instrumentation

FTIR spectra were recorded on Perkin Elmer BX II spectrometer by using KBr matrix in the wavenumber range of 4000-400 cm⁻¹. ¹H and ¹³C NMR spectra of the compounds were recorded on Bruker Avance III (400 MHz) spectrometer by using DMSO as a solvent. Thermo Scientific FLASH 2000 CHN elemental analyser was used for calculating the percentages of C, H, and N. Melting points were measured by the capillary method using Thermo scientific apparatus and were reported uncorrected. Molar conductance was measured in DMF by utilising Systronics conductivity type bridge model-306. The mass spectra were obtained using SCIEX Triple TOF 5600 instrument by using methanol and DMSO as a solvent. The absorption spectra were recorded in DMF on a Varian cary-5000 spectrometer at room temperature. The X-ray analysis was carried out using a Rigaku table top X-ray diffractometer with a scan rate of 2 min⁻¹ in the 2 θ range of 10°–80°. TGA was recorded using SDT Q600 instrument at a heating rate of 0.1 to 100 °C/min. The electron spin resonance (ESR) studies of copper complexes were carried out on JES-FA200 spectrometer using tetracyanoethylene (TCNE) as an internal standard. Scanning electron microscopy (SEM) with energy dispersive X-ray analysis (EDAX) was carried out on a JEOL 7610F plus instrument and the micrographs were recorded in a dynamic nitrogen atmosphere.

Synthesis

Synthesis of 4-(4-amino-5-mercapto-4H-1,2,4-triazol-3-yl)phenol precursor

The precursor was prepared as per the earlier reported method (Sahoo et al.) [34]. The identity of the synthesized compound was established after comparing its spectral data with the literature reported values.

Synthesis of Schiff base ligands (1-4)

To a methanolic solution of 4-(4-amino-5-mercapto-4H-1,2,4-triazol-3-yl)phenol (0.624 g, 3 mmol) was added 3-ethoxysalicylaldehyde (0.498 g, 3 mmol)/5-bromo-salicylaldehyde (0.603 g, 3 mmol)/5-nitrosalicylaldehyde (0.624 g, 3 mmol)/3,5-dibromosalicylaldehyde (0.839 g, 3 mmol) (Scheme 1). The reaction mixture was refluxed for 4–5 h at room temp. The progress of the reaction was monitored regularly by TLC. The coloured products so obtained were filtered, dried and



Compound. No.	R ₁	\mathbf{R}_2	M(II)	
1.	OCH ₂ CH ₃	Н	-	_
2.	Н	Br	-	
3.	Н	NO_2	-	
4.	Br	Br	-	
5.	OCH ₂ CH ₃	Н	Со	
6.	OCH ₂ CH ₃	Н	Ni	
7.	OCH ₂ CH ₃	Н	Cu	
8.	OCH ₂ CH ₃	Н	Zn	
9.	Н	Br	Со	
10.	Н	Br	Ni	
11.	Н	Br	Cu	
12.	Н	Br	Zn	
13.	Н	NO_2	Со	
14.	Н	NO_2	Ni	
15.	Н	NO_2	Cu	
16.	Н	NO_2	Zn	
17.	Br	Br	Со	
18.	Br	Br	Ni	
19.	Br	Br	Cu	
20.	Br	Br	Zn	

Scheme 1 Synthesis of Schiff base ligands (1–4) and their transition metal complexes (5–20) (i) methanol, reflux, 4–5 h. (ii) methanol, metal acetates, stirring, 2–3 h

recrystallized with the mixture of methanol and chloroform (1:1, v/v) to obtain pure products.

2-ethoxy-6-(((3-(4-hydroxyphenyl)-5-mercapto-4H-1,2,4-triazol-4-yl)imino) methyl)phenol, \mathbf{HL}^{1} (1) Pale yellow, Yield: 81%, M.p.: 94–96 °C, Conductivity (Ohm⁻¹ cm² mol⁻¹) in DMF: 14.09, Anal. calcd for C₁₇H₁₆N₄O₃S (%): C: 57.29, H: 4.52, N: 15.72, Anal. found (%): C: 57.02, H: 4.32, N: 15.16. HRMS *m/z*: calcd for C₁₇H₁₇N₄O₃S [M+H]⁺: 357.1016, found: 357.1021. ¹H NMR (400 MHz, DMSO) δ : 14.04 (s, 1H, S–H), 10.10 (s, 1H, O–H^b), 9.93 (s, 1H, O–H^a), 9.67 (s, 1H, CH=N), 7.70 (d, *J*=8.0 Hz, 2H), 7.41 (d, *J*=7.5 Hz, 1H), 7.24 (d, *J*=7.5 Hz, 1H), 7.17 (d, *J*=8.0 Hz, 1H), 6.88 (d, *J*=8.1 Hz, 2H), 4.17 (q, *J*=8 Hz, 2H, CH₂), 1.52 (t, *J*=8.0 Hz, 3H, CH₃). ¹³C NMR (100 MHz, DMSO) δ : 163.71, 162.38, 160.02,

149.28, 149.03, 147.81, 130.46, 120.04, 119.74, 119.05, 117.20, 116.63, 115.93, 64.83, 15.02. IR (KBr, cm⁻¹): 3288 (O–H^b), 3121 (O-H^a), 2362 (S–H), 1603 (C=N).

4-bromo-2-(((3-(4-hydroxyphenyl)-5-mercapto-4H-1,2,4-triazol-4-yl)imino) methyl)phenol, **HL**² (2) Orange, Yield: 79%, M.p.: 97–99 °C, Conductivity (Ohm⁻¹ cm² mol⁻¹) in DMF: 13.53. Anal. calcd for $C_{15}H_{11}BrN_4O_2S$ (%): C: 46.05, H: 2.83, N: 14.32, Anal. found (%): C: 46.01, H: 2.62, N: 14.29. HRMS *m/z*: calcd for $C_{15}H_{12}BrN_4O_2S$ [M+H]⁺: 390.9859, found: 390.9889. ¹H NMR (400 MHz, DMSO) δ : 14.04 (s, 1H, S–H), 10.83 (s, 1H, O–H^b), 10.11 (s, 1H, O–H^a), 9.96 (s, 1H, CH=N), 7.89 (d, *J*=2.1 Hz, 1H), 7.68 (d, *J*=8.7 Hz, 2H), 7.59 (dd, *J*=8.8, 2.4 Hz, 1H), 6.99 (s, 1H), 6.89 (d, *J*=8.7 Hz, 2H). ¹³C NMR (100 MHz, DMSO) δ : 162.31, 161.03, 160.07, 158.13, 149.49, 136.99, 130.59, 129.20, 121.01, 119.55, 116.53, 115.92, 111.33. IR (KBr, cm⁻¹): 3275 (O-H^b), 3166 (O–H^a), 2363 (S–H), 1615 (C=N).

2-(((3-(4-hydroxyphenyl)-5-mercapto-4H-1,2,4-triazol-4-yl)imino)methyl)-4-nitrophenol, HL^3 (3) Yellow, Yield: 87%, M.p.: 100–102 °C, Conductivity (Ohm⁻¹ cm² mol⁻¹) in DMF: 13.97. Anal. calcd for C₁₅H₁₁N₅O₄S (%): C: 50.42, H: 3.10, N: 19.60, Anal. found (%): C: 50.29, H: 3.09, N: 19.07. HRMS *m/z*: calcd for C₁₅H₁₂N₅O₄S [M+H]⁺: 358.0605, found: 358.0629. ¹H NMR (400 MHz, DMSO) δ : 14.08 (s, 1H, S–H), 12.09 (s, 1H, O–H^b), 10.21 (s, 1H, O–H^a), 10.12 (s, 1H, CH=N), 8.64 (d, *J*=2.9 Hz, 1H), 8.30 (dd, *J*=9.1, 2.9 Hz, 1H), 7.70 (d, *J*=8.8 Hz, 2H), 7.18 (d, *J*=9.2 Hz, 1H), 6.90 (d, *J*=8.8 Hz, 2H). ¹³C NMR (100 MHz, DMSO) δ : 164.07, 162.27, 160.11, 159.38, 149.67, 140.47, 130.70, 129.51, 123.08, 119.60, 117.91, 116.47, 115.91. IR (KBr, cm⁻¹): 3297 (O–H^b), 3135 (O–H^a), 2364 (S–H), 1610 (C=N).

2,4-dibromo-6-(((3-(4-Hydroxyphenyl)-5-mercapto-4H-1,2,4-triazol-4-yl)imino) methyl)phenol, HL^4 (4) Yellow, Yield: 82%, M.p.: 93–95 °C, Conductivity (Ohm⁻¹ cm² mol⁻¹) in DMF: 12.98. Anal. calcd for C₁₅H₁₀Br₂N₄O₂S (%): C: 38.32, H: 2.14, N: 11.92, Anal. found (%): C: 38.20, H: 2.08, N: 11.86. HRMS *m/z*: calcd for C₁₅H₁₁Br₂N₄O₂S [M+H]⁺: 468.8964, found: 468.8997. ¹H NMR (400 MHz, DMSO) δ : 14.14 (s, 1H, S–H), 12.28 (s, 1H, O–H^b), 10.15 (s, 1H, O–H^a), 10.00 (s, 1H, CH=N), 8.02 (d, *J*=2.4 Hz, 1H), 7.91 (d, *J*=2.4 Hz, 1H), 7.66 (d, *J*=8.8 Hz, 2H), 6.90 (d, *J*=8.8 Hz, 2H). ¹³C NMR (100 MHz, DMSO) δ : 163.01, 162.57, 160.19, 154.87, 149.47, 138.86, 130.98, 130.59, 122.28, 116.23, 116.03, 113.76, 111.88. IR (KBr, cm⁻¹): 3241 (O–H^b), 3130 (O–H^a), 2363 (S–H), 1609 (C=N).

Synthesis of transition metal complexes (5–20)

The metal complexes (**5–20**) were prepared by combining hot methanolic solution (20 mL) of synthesized Schiff base ligands (**1–4**) with a methanolic solution (20 mL) of metal acetates in 1:1 molar ratio. The reaction mixture was stirred vigorously for 2–3 h at room temperature. The different coloured products so obtained were filtered and finally dried over vacuum to obtain pure complexes (Scheme 1).

 $Co(L^1)(H_2O)(CH_3COO)$ (5) Light red, Yield: 70%, M.p.: 160–163 °C, Conductivity (Ohm⁻¹ cm² mol⁻¹) in DMF: 13.05. Anal. calcd for $C_{19}H_{20}CoN_4O_6S$ (%): C: 46.44, H: 4.10, N: 11.40, Anal. found (%): C: 46.40, H: 4.01, N: 11.29. HRMS *m/z*: calcd for $C_{19}H_{21}CoN_4O_6S$ [M+H]⁺: 492.0508, found: 492.0514. IR (KBr, cm⁻¹): 3422 (O–H^a, M–H₂O), 2347 (S–H), 1745 (M–CH₃COO), 1593 (C=N), 525 (M–N), 475 (M–O), 435 (M–S).

 $Ni(L^1)(H_2O)(CH_3COO)$ (6) Brown, Yield: 72%, M.p.: 170–172 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 12.76. Anal. calcd for C₁₉H₂₀NiN₄O₆S (%): C: 46.46, H: 4.10, N: 11.41, Anal. found (%): C: 46.34, H: 3.79, N: 11.11. HRMS *m/z*: calcd for C₁₉H₂₁NiN₄O₆S [M+H]⁺: 491.0530, found: 491.0520. IR (KBr, cm⁻¹): 3393 (O–H^a, M–H₂O), 2345 (S–H), 1736 (M–CH₃COO), 1593 (C=N), 539 (M–N), 482 (M–O), 440 (M–S).

 $Cu(L^1)(H_2O)(CH_3COO)$ (7) Dark brown, Yield: 69%, M.p.: 165–168 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 14.5. Anal. calcd for C₁₉H₂₀CuN₄O₆S (%): C: 46.01, H: 4.06, N: 11.30, Anal. found (%): C: 45.98, H: 3.86, N: 11.30. HRMS *m/z*: calcd for C₁₉H₂₁CuN₄O₆S [M+H]⁺: 496.0472, found: 496.0436. IR (KBr, cm⁻¹): 3422 (O–H^a, M–H₂O), 2346 (S–H), 1743 (M–CH₃COO), 1595 (C=N), 533 (M–N), 471 (M–O), 431 (M–S).

 $Zn(L^1)(H_2O)(CH_3COO)$ (8) Pale yellow, Yield: 75%, M.p.: 164–166 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 13.79, Anal. calcd for C₁₉H₂₀ZnN₄O₆S (%): C: 45.84, H: 4.05, N: 11.25, Anal. found (%): C: 45.22, H: 3.83, N: 11.21. HRMS *m/z*: calcd for C₁₉H₂₁ZnN₄O₆S [M+H]⁺: 497.0468, found: 497.0316. ¹H NMR (400 MHz, DMSO) δ : 13.05 (s, 1H, S–H), 9.89 (s, 1H, O–H^a), 8.09 (s, 1H, CH=N), 7.79 (d, *J*=8.3 Hz, 2H), 7.31 (d, *J*=2.4 Hz, 1H), 7.03 (dd, *J*=8.6, 7.1 Hz, 1H), 6.88 (d, *J*=8.4 Hz, 2H), 6.68 (d, *J*=7.4 Hz, 1H), 3.93 (q, *J*=7.5 Hz, 2H, CH₂), 1.85 (s, 3H, CH₃COO), 1.21 (t, *J*=8 Hz, 3H, CH₃). ¹³C NMR (100 MHz, DMSO) δ : 172.72, 164.34, 162.65, 161.31, 151.34, 149.41, 148.51, 130.72, 122.91, 120.91, 119.25, 117.69, 116.79, 116.09, 65.04, 22.21, 15.34. IR (KBr, cm⁻¹): 3421 (O–H^a, M–H₂O), 2344 (S–H), 1743 (M–CH₃COO), 1596 (C=N), 538 (M–N), 485 (M–O), 436 (M–S).

 $Co(L^2)(H_2O)(CH_3COO)$ (9) Orange red, Yield: 73%, M.p.: 180–182 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 12.02, Anal. calcd for C₁₇H₁₅BrCoN₄O₅S (%): C: 38.80, H: 2.87, N: 10.65, Anal. found (%): C: 38.67, H: 2.65, N: 10.65, HRMS *m/z*: calcd for C₁₇H₁₆BrCoN₄O₅S [M+H]⁺: 525.9351, found: 525.8321, IR (KBr, cm⁻¹): 3400 (O–H^a, M–H₂O), 2349 (S–H), 1730 (M–CH₃COO), 1602 (C=N), 530 (M–N), 458 (M–O), 430 (M–S).

 $Ni(L^2)(H_2O)(CH_3COO)$ (10) Dark red, Yield: 71%, M.p.: 165–168 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 14.7, Anal. calcd for C₁₇H₁₅BrNiN₄O₅S (%): C: 38.82, H: 2.87, N: 10.65, Anal. found (%): C: 38.81, H: 2.09, N: 10.41. HRMS *m/z*: calcd for C₁₇H₁₆BrNiN₄O₅S [M+H]⁺: 524.9373, found: 524.9300. IR (KBr, cm⁻¹): 3400 (O–H^a, M–H₂O), 2349 (S–H), 1746 (M–CH₃COO), 1604 (C=N), 533 (M–N), 464 (M–O), 431 (M–S).

 $Cu(L^2)(H_2O)(CH_3COO)$ (11) Brown red, Yield: 70%, M.p.: 174–176°C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 13.16, Anal. calcd for C₁₇H₁₅BrCuN₄O₅S (%): C: 38.46, H: 2.85, N: 10.55, Anal. found (%): C: 38.01, H: 2.69, N: 10.49. HRMS m/z: calcd for C₁₇H₁₆BrCuN₄O₅S [M+H]⁺: 529.9315, found: 529.9293. IR (KBr,

cm⁻¹): 3377 (O–H^a, M–H₂O), 2346 (S–H), 1740 (M–CH₃COO), 1603 (C=N), 529 (M–N), 462 (M–O), 438 (M–S).

 $Zn(L^2)(H_2O)(CH_3COO)$ (12) Yellow, Yield: 76%, M.p.: 182–184 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 12.61. Anal. calcd for C₁₇H₁₅BrZnN₄O₅S (%): C: 38.33, H: 2.84, N: 10.52, Anal. found (%): C: 38.02, H: 2.35, N: 10.47. HRMS *m/z*: calcd for C₁₇H₁₆BrZnN₄O₅S [M+H]⁺: 530.9311, found: 530.9233. ¹H NMR (400 MHz, DMSO) δ : 13.11 (s, 1H, S–H), 9.94 (s, 1H, O–H^a), 8.07 (s, 1H, CH=N), 7.85 (d, *J*=3.4 Hz, 1H), 7.59 (d, *J*=8.6 Hz, 2H), 7.28 (dd, *J*=8.2, 3.5 Hz, 1H), 6.89 (d, *J*=8.5 Hz, 2H), 6.63 (d, *J*=8.1 Hz, 1H), 1.81 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ : 173.08, 163.61, 162.04, 160.22, 159.44, 149.77, 137.34, 130.70, 129.94, 122.32, 119.87, 116.98, 116.24, 112.72, 22.99. IR (KBr, cm⁻¹): 3412 (O–H^a, M–H₂O), 2349 (S–H), 1749 (M–CH₃COO), 1607 (C=N), 533 (M–N), 488 (M–O), 447 (M–S).

 $Co(L^3)(H_2O)(CH_3COO)$ (13) Dark green, Yield: 71%, M.p.: 160–162 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 13.14. Anal. calcd for C₁₇H₁₅CoN₅O₇S (%): C: 41.47, H: 3.07, N: 14.22, Anal. found (%): C: 41.38, H: 2.59, N: 14.11. HRMS *m/z*: calcd for C₁₇H₁₆CoN₅O₇S [M+H]⁺: 493.0097, found: 493.0079. IR (KBr, cm⁻¹): 3411 (O–H^a, M–H₂O), 2343 (S–H), 1735 (M–CH₃COO), 1605 (C=N), 539 (M–N), 456 (M–O), 427 (M–S).

 $Ni(L^3)(H_2O)(CH_3COO)$ (14) Green, Yield: 74%, M.p.: 163–165 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 13.56. Anal. calcd for C₁₇H₁₅NiN₅O₇S (%): C: 41.49, H: 3.07, N: 14.23, Anal. found (%): C: 41.38, H: 2.89, N: 14.14. HRMS *m/z*: calcd for C₁₇H₁₆NiN₅O₇S [M+H]⁺: 492.0118, found: 492.0092. IR (KBr, cm⁻¹): 3404 (O–H^a, M–H₂O), 2350 (S–H), 1751 (M–CH₃COO), 1604 (C=N), 524 (M–N), 463 (M–O), 431 (M–S).

 $Cu(L^3)(H_2O)(CH_3COO)$ (15) Dark brown, Yield: 66%, M.p.: 178–180 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 14.07, Anal. calcd for C₁₇H₁₅CuN₅O₇S (%): C: 41.09, H: 3.04, N: 14.09, Anal. found (%): C: 41.09, H: 2.49, N: 14.07. HRMS *m*/*z*: calcd for C₁₇H₁₆CuN₅O₇S [M+H]⁺: 497.0061, found: 497.0048. IR (KBr, cm⁻¹): 3402 (O–H^a, M–H₂O), 2351 (S–H), 1753 (M–CH₃COO), 1604 (C=N), 511 (M–N), 467 (M–O), 434 (M–S).

 $Zn(L^3)(H_2O)(CH_3COO)$ (16) Light yellow, Yield: 72%, M.p.: 171–173 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 14.22. Anal. calcd for C₁₇H₁₅ZnN₅O₇S (%): C: 40.94, H: 3.03, N: 14.04, Anal. found (%): C: 40.93, H: 2.67, N: 14.01. HRMS *m/z*: calcd for C₁₇H₁₆ZnN₅O₇S [M + H]⁺: 498.0056, found: 498.0053. ¹H NMR (400 MHz, DMSO) δ : 13.22 (s, 1H, S–H), 10.09 (s, 1H, O–H^a), 8.38 (s, 1H, CH=N), 7.97 (d, *J*=8.5 Hz, 1H), 7.84 (dd, *J*=8.2, 4.1 Hz, 1H), 7.43 (d, *J*=8.2 Hz, 2H), 6.86 (d, *J*=8.9 Hz, 2H), 6.60 (d, *J*=8.1 Hz, 1H), 1.82 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ : 175.38, 167.13, 165.39, 163.66, 161.40, 149.88, 141.60, 137.53, 130.81, 129.83, 120.06, 118.63, 116.80, 116.28, 22.78. IR (KBr, cm⁻¹): 3410 (O–H^a, M–H₂O), 2344 (S–H), 1736 (M-CH₃COO), 1607 (C=N), 516 (M–N), 496 (M–O), 429 (M–S).

 $Co(L^4)(H_2O)(CH_3COO)$ (17) Green, Yield: 73%, M.p.: 177–179 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 14.7. Anal. calcd for C₁₇H₁₄Br₂CoN₄O₅S (%): C: 33.74, H: 2.33, N: 9.26, Anal. found (%): C: 33.57, H: 2.09, N: 9.01. HRMS *m/z*: calcd for C₁₇H₁₅Br₂CoN₄O₅S [M+H]⁺: 602.8384, found: 602.8380.

IR (KBr, cm⁻¹): 3368 (O–H^a, M–H₂O), 2342 (S–H), 1742 (M–CH₃COO), 1596 (C=N), 537 (M–N), 480 (M–O), 441 (M–S).

 $Ni(L^4)(H_2O)(CH_3COO)$ (18) Brown, Yield: 76%, M.p.: 163–165 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 13.09. Anal. calcd for $C_{17}H_{14}Br_2NiN_4O_5S$ (%): C: 33.76, H: 2.33, N: 9.26, Anal. found (%): C: 33.54, H: 2.12, N: 9.27. HRMS *m/z*: calcd for $C_{17}H_{15}Br_2NiN_4O_5S$ [M+H]⁺: 602.8478, found: 602.8464. IR (KBr, cm⁻¹): 3399 (O–H^a, M–H₂O), 2348 (S–H), 1759 (M–CH₃COO), 1596 (C=N), 536 (M–N), 457 (M–O), 428 (M–S).

 $Cu(L^4)(H_2O)(CH_3COO)$ (**19**) Brown red, Yield: 73%, M.p.: 162–164 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 14.08. Anal. calcd for C₁₇H₁₄Br₂CuN₄O₅S (%): C: 33.49, H: 2.31, N: 9.19, Anal. found (%): C: 33.02, H: 2.02, N: 9.01. HRMS *m/z*: calcd for C₁₇H₁₅Br₂CuN₄O₅S [M+H]⁺: 607.8420, found: 607.8357. IR (KBr, cm⁻¹): 3432 (O–H^a, M–H₂O), 2346 (S–H), 1745 (M–CH₃COO), 1596 (C=N), 533 (M–N), 469 (M–O), 432 (M–S).

 $Zn(L^4)(H_2O)(CH_3COO)$ (20) Orange yellow, Yield: 78%, M.p.: 167–169 °C, Conductivity: (Ohm⁻¹ cm² mol⁻¹) in DMF: 12.05. Anal. calcd for C₁₇H₁₄Br₂ZnN₄O₅S (%): C: 33.39, H: 2.31, N: 9.16, Anal. found (%): C: 33.21, H: 2.12, N: 9.11. HRMS *m/z*: calcd for C₁₇H₁₅Br₂ZnN₄O₅S [M+H]⁺: 608.8416, found: 608.7999. ¹H NMR (400 MHz, DMSO) δ : 13.13 (s, 1H, S–H), 10.00 (s, 1H, O–H^a), 8.18 (s, 1H, CH=N), 7.76 (d, *J*=2.8 Hz, 1H), 7.64 (d, *J*=2.5 Hz, 1H), 7.58 (d, *J*=8 Hz, 2H), 6.89 (d, *J*=8.2 Hz, 2H), 1.84 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ : 173.75, 163.84, 163.21, 160.76, 157.10, 149.57, 138.86, 130.98, 130.59, 122.93, 116.23, 116.03, 113.99, 112.54, 24.28. IR (KBr, cm⁻¹): 3422 (O–H^a, M–H₂O), 2346 (S–H), 1739 (M–CH₃COO), 1598 (C=N), 533 (M–N), 458 (M–O), 430 (M–S).

Pharmacology

Antimicrobial activity assay

The *in-vitro* antimicrobial activity of all the synthesized compounds (**1–20**) was assessed against four bacterial strains and two fungal strains. Norfloxacin and Fluconazole were used as positive control for bacteria and fungi [35]. DMSO was used as a negative control. The synthesized compounds were assessed against two Grampositive bacteria *Bacillus subtilis* (NCIM 2063) and *Staphylococcus aureus* (MTCC 2901), two Gram-negative bacteria *Pseudomonas aeruginosa* (MTCC 424) and *Escherichia coli* (MTCC 732). The antifungal activity was assessed against two fungal strains namely *Aspergillus niger* (MTCC 9933) and *Candida albicans* (MTCC 227). The pathogens were collected from the Department of Bio and Nano Technology, GJUS&T, Hisar.

Compounds concentration

The stock solutions of concentration 1000 μ g/mL were prepared by dissolving 5 mg of the sample compounds in 5 mL of DMSO. 1 mL of the above prepared stock solution was mixed with 9 mL of DMSO to get a stock solution of concentration 100 μ g/mL [36].

Subculture and preservation of pathogens

The bacteria were subcultured on the NB (nutrient broth) and the fungi were subcultured on the PDB (potato dextrose broth) which were prepared by dissolving NB (1.3 g) and PDB (2.4 g) in 100 mL of distilled water. The resulting mixture was autoclaved at 15 psi for 30 min. The prepared cultures were incubated at 37 °C for 24 h in case of antibacterial culture and at 27 °C for seven days in case of antifungal culture [35].

Determination of MIC values

The serial dilution method was employed for carrying out the antimicrobial activity [37-40]. In this method, 1 mL of stock solution was added to the test tube containing 1 mL of broth and diluted serially up to 3.125 µg/mL concentrations. After dilution, the subcultured bacteria and fungi were impregnated in each test tube. Then, the test tubes were kept in an incubator for a fixed time to develop the culture. Pathogens growth was checked visually and spectrophotometrically. The MICs (minimum inhibitory concentration) value for tested compounds was determined. The obtained values were compared to those of reference drugs. The overall process was repeated twice to obtain more accurate and precise results.

Anticancer activity assay

The *in-vitro* anticancer activity of the synthesized compounds (1–20) was assessed on the basis of measurement of inhibition of cell lines in 96-well microtitre plates by cell mediated reduction of tetrazolium salt to water insoluble formazan crystals using Paclitaxel as a standard. The cytotoxic evaluation was performed against a panel of three cell lines namely: HCT-116 (ATCC No. CCL-247) derived from human colon cancer cell line, DU145 (ATCC No. HTB-81) derived from human prostate cancer cell line, A549 (ATCC No. CCL-185) derived from human alveolar adenocarcinoma epithelial cell line, using the MTT assay [41]. To study the effects of synthesized compounds on the viability of human cancer cell lines, the absorbance was noted at 595 nm using a micro plate reader (InfiniteM200, Tecan, Mannedorf, Switzerland). The effect of test samples on the procreation of cells was expressed in the form of % inhibition and is calculated as

From the dose–response curves, IC_{50} values of the compounds were reported as average of three independent experiments. The IC_{50} value is expressed in the form of mean \pm standard deviation.

Molecular docking

The crystal structure of the human epidermal growth factor receptor (EGFR) was retrieved from the RCSB protein data bank (https://www.rcsb.org) and was encoded with PDB code 5XGM [42]. The complexes bound to the receptor, such as non-essential water molecules and any inhibitors were removed before the docking run. Hex 8.0.0 which is an interactive molecular graphics program (http://www.loria.fr) and is used to perform rigid docking to compute possible interactions of 5XGM protein with potent compounds. During docking the following parameters were used: correlation type – shape + electrostatics, FFT mode—3D, post processing- mm energies, grid dimension – 0.6, receptor range – 180, ligand range – 180, twist range – 360, distance range – 40. After docking, the detailed protein–ligand visualization and comparison of the docked sites of target proteins and ligands were done by Chimera 1.14 and Ligplot software [43, 44].

Results and discussion

Chemistry

Schiff base ligands (1–4) were synthesized by the condensation of 4-(4-amino-5-mercapto-4*H*-1,2,4-triazol-3-yl)phenol with salicylaldehyde derivatives. The solid products so obtained have different colours owing to the presence of a chromophoric group in the molecule. Moreover, the treatment of synthesized compounds with corresponding metal acetates leads to formation of new complexes (**5–20**). Various spectral (¹H, ¹³C NMR, FTIR and HRMS) and physicochemical techniques (XRD, ESR, TGA and SEM) were utilised for the characterization of synthesized compounds. The synthesized complexes (**5–20**) are coloured solids which are soluble in polar aprotic solvents like DMF, DMSO. The melting point of synthesized complexes was higher than those of Schiff base ligands, which means the complexes are more stable as compared to ligands. The molar conductivity values of synthesized compounds lie between 12 - 15 (Ohm⁻¹ cm² mol⁻¹), which indicate that the complexes are non-electrolytic in nature.

FTIR spectra

The FTIR spectra of synthesized compounds (1-20) were recorded over the wavenumber range of 4000–400 cm⁻¹. Comparison between infrared absorption

Compound. no.	ν(Ο–H ^a /Ο– H ^b)	ν(C=N)	ν(S–H)	ν(M–N)	ν(M–O)	ν(M–S)	ν(M– CH ₃ COO)	ν(O–H ^a /M– H ₂ O)
1	3122, 3288	1603	2362				_	_
2	3166, 3275	1615	2363	_	_	_	_	_
3	3135, 3297	1610	2364	_	_	_	_	_
4	3130, 3241	1609	2363	_	_	_	_	_
5	_	1593	2347	525	475	435	1745	3422 br
6	_	1593	2345	539	482	440	1736	3393 br
7	_	1595	2346	533	471	431	1743	3422 br
8	_	1596	2344	538	485	436	1743	3421 br
9	_	1602	2349	530	458	430	1730	3400 br
10	_	1604	2349	533	464	431	1746	3400 br
11	_	1603	2346	529	462	438	1740	3377 br
12	_	1607	2349	533	488	447	1749	3412 br
13	-	1605	2343	539	456	427	1735	3411 br
14	-	1604	2350	524	463	431	1751	3404 br
15	-	1604	2351	511	467	434	1753	3402 br
16	-	1607	2344	516	496	429	1736	3410 br
17	-	1596	2342	537	480	441	1742	3368 br
18	-	1596	2348	536	457	428	1759	3399 br
19	-	1596	2346	533	469	432	1745	3432 br
20	_	1598	2346	533	458	430	1739	3422 br

 Table 1 Characteristic IR bands (cm⁻¹) for the compounds (1–20)

frequencies of ligands and their respective metal complexes helps in understanding the binding modes of the ligand with the metal. The most explicit IR bands recorded for the synthesized compounds with the probable assignment are given in Table 1. The band appearing at 1615–1603 cm⁻¹ in the IR spectra of ligands is due to ν (C=N) group which shifted to lower frequency region 1607–1593 cm⁻¹ on complexation with metal atom, these shifting in spectra justified that lone pair of nitrogen is involved in bonding with the metal centre [45-47]. A band at 3297-3241 cm⁻¹ is observed due to $\nu(O-H^b)$ group which gets disappeared in complexes due to deprotonation of phenolic O-H on complexation with the metal atom. The bands appearing in ligands in the range of 2364–2362 cm⁻¹ are due to ν (S–H) group [48] which gets shifted to slightly lower values in complexes, due to the participation of sulphur in bonding with the metal atom. Thiol sulphur coordinates to metal without deprotonation. Some new bands appears at 496–456 cm⁻¹ [due to ν (M–O)] and 447–427 cm⁻¹ [due to ν (M-S)] [49]. The coordinated acetate molecule in complexes was ascertained by the presence of a band in the range of $1759-1730 \text{ cm}^{-1}$ [50]. A new broad band near 3432-3368 cm⁻¹ in complexes is due to the coordinated water molecule and ν (O–H^a) group. Furthermore, some new bands in complexes were observed in the range 539–511 cm⁻¹ which are due to ν (M–N) [51]. The above results imply that ligands behave in a tridentate manner and the metal complexes have pentacoordinated geometry.

¹H NMR spectra

¹H NMR spectra of the synthesized Schiff base ligands (1-4) and their corresponding zinc (II) complexes were recorded in DMSO- d_6 at 400 MHz. The spectra of compound (2) and its corresponding zinc complex (12) are depicted in Fig. 1. The ¹H NMR spectra of synthesized ligands showed one sharp singlet in the range of δ 12.28–10.10 ppm (due to O-H^b proton) [52] which got disappeared in complexes due to involvement of oxygen in complexation. Another singlet for ligands was observed in the range of δ 14.14–14.04 ppm (due to S–H proton) [52], which shift to slightly lower values on complexation with metal atom. The ligands showed another singlet in the range of δ 10.21–9.93 ppm (due to another O-H^a proton), which remained very less changed in complexes. The azomethine (H-C=N) peak of ligands appeared in the range of δ 10.12–9.67 ppm. In case of complexes, it undergoes upfield shifting up to 8.07 ppm, which clearly indicates the confinement between metal ion and imine nitrogen. The upfield shifting is due to chelation [53]. The aromatic protons in ligands appeared at δ 8.64–6.88 ppm which remained unaffected or very less affected in complexes with a slight change in chemical shift values. In the spectra of complexes, a new singlet of three protons appeared at δ 1.84–1.81 ppm, which further confirms the presence of acetate group in complexes. The observed NMR data were found to be in good correlation with the expected values.

¹³C NMR spectra

In the ¹³C NMR spectra of ligands, (1–4) the peak for azomethine carbon atom appeared at δ 159.38–149.28 ppm which showed downfield shifting on complexation, suggesting that nitrogen atom is involved in coordination with the metal ion [50]. In compound (1) extra peaks at δ 64.83 ppm and δ 15.02 ppm were observed due to the presence of an ethoxy group in the aliphatic region. Two peaks at δ 164.07–162.31 ppm and δ 162.57–161.03 ppm were observed due to carbon atom attached to oxygen and sulphur which was shifted downfield in complexes, thus indicating the involvement of oxygen and sulphur in complexation. But the peaks appearing at δ 160.19–160.02 ppm in ligands remains unchanged on complexation, which showed that the O-H^a group attached with this carbon is not involved in complexation. Two new peaks arising at δ 175.38–172.72 ppm and δ 24.28–22.21 ppm, due to the presence of a coordinated acetate group in complexes. The ¹³C NMR data confirm the authenticity of the purposed structures.

Mass spectra

Mass spectrometry plays a valuable role in determining the molecular weight and stoichiometry of the newly synthesized compounds. The molecular ion peak of ligand (3) appeared at m/z 358.0629 which was same as the estimated value. Similarly, for ligand (4) the molecular ion peak appeared at m/z 468.8997, while the signals at m/z 470.8977 and 472.8955 were assigned to (M+H+Br⁷⁹) and



Fig. 1 1 H NMR spectra of compounds A HL² (2) and B [Zn(L²)(H₂O)(CH₃COO)] (12)

 $(M+H+Br^{81})$ isotopes. In case of complex (14) molecular ion peak appeared at m/z 492.0092. In case of complex (18) molecular ion peak appeared at m/z 602.8464 and the other signals observed at m/z 604.8502 and 606.8515 were due to $(M+H+Br^{79})$ and $(M+H+Br^{81})$ isotopes [54]. The mass spectral data results were found in good

agreement with theoretical values and support the formation of $[M(L^{1-4})(H_2O)(CH_3COO)]$ type complexes in 1:1 stoichiometry.

Electronic spectra

The electronic transitions involved in the ligands (1–4) and their corresponding transition metal complexes (5–20) were recorded in DMF over the range 200–800 nm. The synthesized ligands showed absorption bands around 265–296 nm due to intraligand $\pi \rightarrow \pi^*$ transitions. Moreover, the absorption bands appearing in the range of 319–370 nm are due to the occurrence of $n \rightarrow \pi^*$ transitions. These bands shift to a slightly higher wavelength on complexation. For all the complexes $n \rightarrow \pi^*$ transitions are red shifted, due to coordination of azomethine nitrogen to the central metal ion. For the Ni(II) complexes, electronic spectrum shows bands in the range of 757–798, 607–630, 345–388 nm which are typical for a pentacoordinated square pyramidal Ni(II) complex and could be assigned to d-d transitions. For Cu(II) centres the visible spectra exhibits broad band in the range of λ_{max} at 628–655 nm which are specifically consistent for five-coordinate geometry and are correlated to d-d transition of Cu(II) ions. Zinc(II) complexes do not show any d-d transitions. In case of complexes, some new bands appeared in the range of 416–447 nm, which were attributed to ligand to metal charge transfer [55].

Molar conductivity studies

The molar conductivity studies of synthesized compounds (**1–20**) were undertaken using a digital conductivity meter at room temperature and using DMF as a solvent. The DMF solution of synthesized complexes (10^{-3} M) exhibited lower molar conductance values ($12-15 \ \Omega^{-1} \ \text{cm}^2 \ \text{mol}^{-1}$), which supports the non-electrolytic nature of complexes [56].

X-ray diffraction studies

Powder X-ray diffraction method is one of the primary techniques used by solidstate chemists to explore the physicochemical parameters of synthesized materials. The prepared ligand (**2**) and their metal complexes (**9–11**) were analysed by the use of this technique over the 2θ range 10° – 80° at a wavelength of 1.5406Å. The obtained X-ray diffraction patterns are shown in Fig. 2. Schiff base ligand (**2**) showed the peaks at 15.42°, 16.46°, 18.14°, 20.64°, 24.18°, 25.92°, 27.28°, 29.92°, 35.98°, 41.34°. The two very important characteristic peaks appearing in Schiff base ligand at 2θ =18.14° and 20.64° confirms the formation of Schiff base imine [57]. The observed diffraction pattern confirms that the compound is semi-crystalline in nature. The peaks for cobalt complex (**9**) were observed at 12.18°, 15.12°, 18.04°, 22.78°, 29.38°, 32.46°, 35.22°, 38.48°, 41.88°. The peaks for nickel complex (**10**) existed at 17.08°, 18.18°, 19.82°, 21.44°, 22.28°, 23.4°,



Fig. 2 XRD diagrams of **A** [HL²] (2), **B** [Co(L²)(H₂O)(CH₃COO)] (9), **C** [Ni(L²)(H₂O)(CH₃COO)] (10) and **D** [Cu(L²)(H₂O)(CH₃COO)] (11)

24.02°, 26.76°, 28.28°, 32.16°, 33.5°, 35.72°, 36.14°, 40.3°. Copper complex (**11**) showed the peaks at 12.74°, 16.42°, 19.64°, 21.7°, 23.28°, 25.82°, 26.96°, 27.92°, 29.38°, 30.9°, 32.3°, 35.2°, 37.6°, 39.32°, respectively. The diffraction pattern of the ligand is completely different from metal complexes, thus indicating the quantum confinement of ligands by metal ions [58]. The XRD data observed for complexes confirm the dominant amorphous nature with very less crystallinity.

Electron spin resonance studies

The ESR spectra of copper complexes give information about the geometry and bonding sites. The solid-state X-band ESR spectrum of copper complex (7) was recorded in powder form at room temperature (300 K) and the micrograph obtained is presented in Fig. 3. The values of g_{\parallel} and g_{\perp} in case of Cu(II) complex lies at 2.15 and 2.09 which holds in good agreement with the condition $g_{\parallel} > g_{\perp} > 2.0023$. Since complexes are pentacoordinated, therefore two basic configurations with five coordinated geometry are square pyramidal and trigonal bipyramidal. The two configurations are characterised by the ground state dx^2-y^2 and dz^2 , respectively. For systems with $g_{\parallel} > g_{\perp}$ the geometry is square pyramidal, while systems with $g_{\parallel} < g_{\perp}$ the geometry is trigonal bipyramidal [59, 60]. From the observed results it can be concluded that there exists a square pyramidal geometry around the Cu(II) metal ion with unpaired electron lying mainly in the dx^2-y^2 ground state orbital. The $g_{average}$ and G values were computed from the given equations



Fig. 3 The X-band ESR spectrum of complex $[Cu(L^1)(H_2O)(CH_3COO)]$ (7)

$$g_{average} = 1/3 [g_{\parallel} + 2g_{\perp}]$$

 $G = (g_{\parallel} - 2.0023)/(g_{\perp} - 2.0023)$

From the above calculations, the value of $g_{average}$ and G was found to be 2.11 and 1.68. According to Hathway and Billing [61, 62], if the value of G>4.0 then we can consider negligible exchange interaction between Cu(II) centres in the solid state and if the value of G<4.0 there will be significant exchange interactions. Here, the value of G was found to be 1.68, which indicates that exchange interactions are operative. By using the g-vlaue, magnetic moment can be calculated using the following equation.

$$\mu_{\rm eff} = \$ \sqrt{\left(g_{\parallel}^2 + 2g_{\perp}^2 \right)}$$

The calculated value of magnetic moment for complex 7 was found to be 1.82.

Thermal studies

Thermal analysis of copper complex (15) was carried out over the temperature range 0 to 900 °C. The percentage weight loss was recorded in the temperature range of

22–790 °C, which gives information about the stability and volatility of the complex. The thermogram showed a total 55.10% weight loss at 790 °C temperature which was observed in three steps: (1) about 7% weight loss in the range of 75–100 °C which was due to trace amount of moisture present in sample (2) 20% weight loss in the range of 150–272 °C was due to loss of coordinated water molecule (3) about 18% weight loss in the range of 272–459 °C which might be due to complete decomposition of ligand around metal ion. Finally, the complex was converted into metal oxide. The endothermic band observed in the range of 150–272 °C, confirmed the presence of water molecule in the complex [63].

$$\begin{bmatrix} CuC_{17}H_{15}N_{5}O_{7}S].H_{2}O \xrightarrow{75-100 \ ^{\circ}C} & [CuC_{17}H_{15}N_{5}O_{7}S] \xrightarrow{150-272 \ ^{\circ}C} & [CuC_{17}H_{13}N_{5}O_{6}S] \\ & -H_{2}O & \\ & -H_$$

SEM and EDAX

The SEM images of the ligand (3) and its zinc (II) complex (16) was viewed at different magnifications and the micrographs obtained are presented in Fig. 4. The



Fig.4 A SEM image of $[HL^3]$ (3), B EDAX image of $[HL^3]$ (3), C SEM image of $[Zn(L^3)(H_2O)$ (CH₃COO)] (16) and D EDAX image of $[Zn(L^3)(H_2O)(CH_3COO)]$ (16)

micrograph of ligand exhibited a rough surface with numerous territorial patches. On the other hand, the micrograph of zinc complex indicated an ice rock like shape which may be due to the tightening of voids on the outer surface by coordination of ligand with the metal ion. The SEM-micrographs indicated that ligand has completely different surface morphology from complex, which is another evidence for coordination of ligand to metal ion.

Moreover, Energy dispersive X-ray analysis (EDAX) of ligand and its Zn(II) complex was also performed to check the elemental composition of prepared materials. EDAX spectra shown in Fig. 4 clearly show that complexation successfully occurred.

Antimicrobial activity

To investigate the antimicrobial action of synthesized compounds (1-20), four bacterial and two fungal strains were used. The serial dilution method was used for

Compound. no.	MIC in µmol/mL							
	Gram + ve bacteria		Gram –ve bacteria		Fungi			
	B. subtilis	S. aureus	P. aeruginosa	E. coli	A. niger	C. albicans		
1	0.0702	0.0702	0.0702	0.0702	0.0702	0.0702		
2	0.0641	0.0641	0.0641	0.0641	0.0641	0.0641		
3	0.0350	0.0350	0.0350	0.0350	0.0175	0.0175		
4	0.0532	0.0532	0.0532	0.0532	0.0266	0.0266		
5	0.0510	0.0510	0.0510	0.0510	0.0510	0.0510		
6	0.0510	0.0510	0.0510	0.0510	0.0510	0.0510		
7	0.0505	0.0505	0.0505	0.0505	0.0505	0.0505		
8	0.0503	0.0503	0.0503	0.0503	0.0503	0.0503		
9	0.0476	0.0476	0.0476	0.0476	0.0476	0.0476		
10	0.0476	0.0476	0.0476	0.0476	0.0476	0.0476		
11	0.0472	0.0472	0.0472	0.0472	0.0472	0.0472		
12	0.0470	0.0470	0.0470	0.0470	0.0470	0.0470		
13	0.0254	0.0254	0.0254	0.0254	0.0254	0.0254		
14	0.0255	0.0255	0.0255	0.0255	0.0255	0.0255		
15	0.0252	0.0252	0.0252	0.0252	0.0126	0.0126		
16	0.0251	0.0251	0.0251	0.0251	0.0125	0.0125		
17	0.0414	0.0414	0.0414	0.0414	0.0414	0.0414		
18	0.0414	0.0414	0.0414	0.0414	0.0414	0.0414		
19	0.0411	0.0411	0.0411	0.0411	0.0252	0.0252		
20	0.0409	0.0409	0.0409	0.0409	0.0251	0.0251		
Norfloxacin	0.0391	0.0195	0.0391	0.0391				
Fluconazole					0.0408	0.0204		

Table 2 Results of antimicrobial assay of compounds (1-20) in terms of MIC (µmol/mL)

High active value is marked as bold



Fig. 5 Graph showing antimicrobial activity of the compounds (1-20) and standard drugs

screening the antimicrobial activity. Norfloxacin and Fluconazole were used as reference drugs and the MIC values were calculated in μ mol/mL. The obtained activity data are presented in Table 2 and Fig. 5.

By comparing the activity of ligands with their respective metal complexes, it was found that the metal complexes exhibit more potency as compared to ligands. This is due to the chelation of metal which increases the hydrophobic and lipophilic character as discussed in tweedy chelation theory [64]. The results of the activity data are summarized below:

Among the synthesized ligands (1–4), HL^3 (MIC=0.0175–0.0350 µmol/mL) was the most active compound. The higher activity of HL^3 is due to the presence of strong electron withdrawing group (nitro group), while due to the presence of electron donating group (ethoxy group) HL^1 showed least potency. The observed results indicate that compounds having electron withdrawing groups attached were found to be more active than compounds having electron donating groups attached [65]. The order of activity of ligands is $HL^3 > HL^4 > HL^2 > HL^1$.

Among the transition metal complexes, zinc complexes showed good activity as compared to other metal complexes. The general order of the activity of complexes follows the trend $[Zn(L^{1-4})(H_2O)(CH_3COO)] > [Cu(L^{1-4})(H_2O)(CH_3COO)] > [Ni(L^{1-4})(H_2O)(CH_3COO)] > [Co(L^{1-4})(H_2O)(CH_3COO)] [66].$

Compounds 3, 13, 14, 15 and 16 were found to be active against all bacterial strains except *S. aureus* with MIC values in the range of $(0.0251-0.0350 \ \mu mol/mL)$, as compared to standard drug Norfloxacin $(0.0195-0.0391 \ \mu mol/mL)$. The compounds 3, 4, 13, 14, 15, 16, 19 and 20 were active against fungal strains with MIC values in the range of $(0.0125-0.0266 \ \mu mol/mL)$ as compared to standard drug Fluconazole (MIC = $0.0204-0.0408 \ \mu mol/mL)$. Also, it was found that compounds were more active against *A. niger* as compared to *C. albicans*.

Anticancer assay

The synthesized compounds (1–20) were screened for *in-vitro* anticancer activity against three cancer cell lines i.e. HCT-116 (human colon cancer), DU145 (prostate cancer) and A549 (lung cancer) by using MTT colorimetric assay. The cytotoxicity of the synthesized compounds was represented in terms of median growth inhibitory

Table 3 Anticancer assay of compounds (1–20) and standard drug against three cancer cell lines	Compound no.	IC ₅₀ values (µN	(1±SD)	SD)		
		HCT-116	DU145	A549		
	1	85.42 ± 2.05	92.34 ± 3.14	90.39 ± 2.06		
	2	83.09 ± 2.10	98.21 ± 1.21	96.08 ± 2.10		
	3	>100	>100	>100		
	4	>100	>100	>100		
	5	>100	>100	>100		
	6	>100	>100	>100		
	7	81.61 ± 2.11	>100	>100		
	8	>100	>100	>100		
	9	68.74 ± 1.14	85.76 ± 1.23	81.61 ± 2.09		
	10	55.05 ± 1.07	65.59 ± 2.10	66.62 ± 1.08		
	11	44.46 ± 1.13	47.59 ± 1.24	48.60 ± 1.12		
	12	8.66 ± 1.11	12.66 ± 1.2	9.75 ± 1.2		
	13	68.95 ± 1.05	72.99 ± 2.29	73.04 ± 1.05		
	14	88.10 ± 2.13	79.54 ± 2.87	73.06 ± 3.07		
	15	38.09 ± 1.05	36.34 ± 3.45	67.97 ± 2.02		
	16	41.04 ± 1.07	47.24 ± 2.43	56.42 ± 1.09		
	17	99.12 ± 2.08	>100	>100		
	18	81.63 ± 1.17	83.49 ± 1.19	85.02 ± 1.15		
	19	19.49 ± 1.05	25.63 ± 2.07	$\textbf{28.41} \pm \textbf{1.09}$		
	20	6.89 ± 1.04	10.45 ± 1.19	8.30 ± 1.12		
	Paclitaxel	4.32 ± 1.04	3.16 ± 0.91	3.69 ± 0.98		

High active value is marked as bold

concentration (IC_{50}) and values were compared to standard anticancer drug Paclitaxel. The obtained results are presented in Table 3.

Based on the cytotoxicity data, it may be concluded that compound 12 displayed the highest potency towards all cancer cell lines with IC₅₀ value of 8.66 (HCT-116), 12.66 (DU145) and 9.75 (A549) µM. Compound 20 also showed good inhibition and its IC₅₀ values are 6.89 (HCT-116), 10.45 (DU145) and 8.30 (A549) µM. However, these compounds do not completely touch the potential of the known anticancer drug Paclitaxel. Compounds 10, 11, 15, 16 and 19 displayed moderate activity, while the rest of compounds show very less inhibition towards the cancer cell lines.

Structure-activity relationship

The SAR of compounds (1-20) was also studied to check the effect of substituents on biological activity. The synthesized compounds having different substituents attached were found to be active against the three cancer cell lines (except compounds 3-6 and 8), but particularly the compounds 12 and 20 were found to be more active against the cancer cell lines. Compound 1 having ethoxy substitution on the phenyl ring was very less active. On the other hand, its complexation with different

metals (compounds 5-8) is ineffective for the enhancement of anticancer activity. Compound 2 having bromo substitution on the phenyl ring itself shows very less activity, but its complexation with nickel (compound 10) and copper metal (compound 11) metal makes them moderately active. The complexation of compound 2 with cobalt metal (compound 9) improved its activity to some extent. However, the complexation of compound 2 with zinc metal (compound 12) results in a significant increase of activity, thus making it the most outstanding compound of the series. Compound 3 having nitro substitution on the phenyl ring was negligibly active, but its complexation with copper (compound 15) and zinc metal (compound 16) resulted in significant improvement of its anticancer activity up to a moderate level. However, its complexation with cobalt (compound 13) and nickel metal (compound 14) resulted in no remarkable increase in anticancer activity. The anticancer activity of compound 4 was almost negligible, but its complexation with copper (compound 19) and zinc metal (compound 20) resulted in a significant increase in anticancer activity. However, its complexation with cobalt (compound 17) and nickel (compound **18**) has no effects on its anticancer activity. After accessing the biological potential, it was observed that the activity of metal complexes gets enhanced on complexation. The higher activities of complexes may be due to an increase in their lipophilicity on complexation [66].



Molecular docking studies

Among the synthesized compounds **12**, **19**, and **20** showed good inhibition against tested cancer cell lines that are closely associated with Human Epidermal Growth Factor Receptor (EGFR) which is a cell surface protein molecule and found to be

Compound no.	Binding energy (E_{total}) , kcal/mol	Interacting amino acids with covalent bonding	Interacting amino acids with non- covalent bonding
12	-268.27	No covalent bond interaction	Leu718A, Gly719A, Val726A, Ala743A, Met793A, Gly796A, Cys797A, Leu844A
19	-248.39	Gln820A, Gln849A	His773A, Val819A, Lys823A, His850A, Ser969A
20	-262.34	No covalent bond interaction	Pro772A, Lys852A, Asp1014A

Table 4 Binding energy score of compounds with 5XGM protein

associated with various types of cancer. Many drugs have been developed as a targeted approach to inhibit over-expressed EGFR, but these drugs are not able to inhibit its over-expression due to drug resistance. Drug resistance is a major challenge in targeted therapy of EGFR-mutated non-small cell lung cancers (NSCLCs). However, the efficacy of the tyrosine kinase inhibitors used in targeted therapy is often limited due to the emergence of drug-resistant mutations in the targeted kinases [42, 67, 68]. Resistance to osimertinib, a third generation EGFR tyrosine kinase inhibitor used to treat NSCLCs with specific mutations, caused by EGFR at position 797 is currently a new challenge in the targeted therapy of EGFR-mutated NSCLCs. Therefore, mutated EGFR was chosen for molecular docking studies to support the observed *in-vitro* anticancer activity and to add more insight into the possible mechanism of action.

To understand the probable binding modes and estimate the binding energies, three compounds (12, 19 and 20) were docked into EGFR-mutated NSCLCs binding cavity. The docking studies were performed using Hex 8.0.0 and the results are shown in Table 4. Further, the most suitable docking pose of the protein receptor with preferred target ligands was selected and then analysed using Chimera 1.14 and Ligplot software. The obtained docking poses are shown in Fig. 6A-C. The binding energy value of compound 12 is -268.27 kcal/mol, and it shows good interaction with eight amino acid residues of the protein structure i.e. Leu718A, Gly719A, Val726A, Ala743A, Met793A, Gly796A, Cys797A, and Leu844A. Most interesting feature revealed by the docking of this compound is its interaction with Cys797A site, which is responsible for the mutated and drug-resistant behaviour of EFGR, thereby making this compound of utmost importance. The binding energy value of compound 20 is -262.34 kcal/mol and it shows interaction with three amino acid residues of the protein structure i.e. Pro772A, Lys852A, and Asp1014A. Similarly, compound 19 shows appreciable binding energy (-248.39 kcal/mol), but the values are slightly lower as compared to compounds 12 and 20. Compound 19 shows interaction with amino acid residues i.e. Gln820A, Gln849A via covalent bonding, while it shows non-covalent bonding interactions with His773A, Val819A, Lys823A, His850A, Ser969A amino acid residues. The results of docking studies were found to be in line with the results of *in-vitro* anticancer studies. Based on docking results, it may be concluded that compound **20** is having highest anticancer potential.



Fig. 6 Interaction view of 5XGM protein with compounds 12 (A), 19 (B) and 20 (C)

D Springer

Conclusion

In the present study, four new Schiff base ligands (1-4) and their transition metal (II) complexes (5–20) were synthesized and characterized by various spectral and physicochemical techniques. The obtained spectroscopic data confirm that the synthesized ligands coordinate to metal in a tridentate manner via azomethine nitrogen, deprotonated oxygen and thiol sulphur. The antimicrobial assessment data showed that metal complexes exhibit more potency as compared to ligands. Compound 3and its metal complexes (13-16) showed reliable activity against the tested pathogenic strains. Furthermore, the results of anticancer activity of the compounds were encouraging and compounds 12, 19 and 20 showed significant anticancer activity against the three tested cancer cell lines (HCT-116, DU145, and A549). The docking results correlate very well with the IC₅₀ data and enabled the importance of interactions for depicting the binding energies of the potent compounds. Based on the results in the current manuscript, the compounds 12, 19 and 20 may be considered as good cytotoxic drugs and this class of compounds could be a better model to develop new leads in the treatment of cancer. The procured results also showed that structural modifications in the synthesized compounds may provide products with different pharmacological activities.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s11164-021-04621-5.

Acknowledgements The author Mr. Yogesh Deswal (SRF) expresses their thanks to CSIR, New Delhi, for providing the financial support in the form of JRF-SRF fellowship with Reference No.: 09/752(0087)/2018-EMR-I dated 31/12/18. The authors are also highly thankful to Department of Chemistry and Dr. APJ Abdul Kalam Central Instrumentation Laboratory (CIL) of GJUS&T, Hisar, Haryana, for providing the needed research facilities to pursue this research work.

Declarations

Conflict of interest The author declares no conflicts in publication of this paper.

References

- H. Sung, J. Ferlay, R.L. Siegel, M. Laversanne, I. Soerjomataram, A. Jemal, F. Bray, CA: Cancer J. Clin. 71, 209 (2021)
- 2. World Health Organisation (WHO) (2020).
- 3. F. Yang, S. Shi, J. Zhu, J. Shi, K. Dai, X. Chen, J. Med. Virol. 92, 2067 (2020)
- 4. N. Hoti, D. Zhu, Z. Song, Z. Wu, S. Tabassum, M. Wu, J. Pharmacol. Exp. Ther. 311, 22 (2004)
- 5. Z.H. Siddik, Oncogene 22, 7265 (2003)
- 6. J. Reedijk, Eur. J. Inorg. Chem. 2009, 1303 (2009)
- 7. L. Kelland, Nat. Rev. Cancer. 7, 573 (2007)
- 8. A.M. Abu-Dief, I.M. Mohamed, Beni-Seuf Univ. J. Appl. Sci. 4, 119 (2015)
- L.H. Abdel-Rahman, A.M. Abu-Dief, M. Basha, A.A. Hassan Abdel-Mawgoud, Appl. Organomet. Chem. 31, e3750 (2017).
- L.H. Abdel-Rahman, M.S.S. Adam, A.M. Abu-Dief, H. Moustafa, M.T. Basha, A.S. Aboraia, B.S. Al-Farhan, H.E.S. Ahmed, Appl. Organomet. Chem. 32, e4527 (2018).

- 11. L.H. Abdel-Rahman, A.M. Abu-Dief, A.A.H. Abdel-Mawgoud, J. King Saud Univ. Sci. 31, 52 (2019)
- F.S. Aljohani, A.M. Abu-Dief, R.M. El-Khatib, H.A. Al-Abdulkarim, A. Alharbi, A. Mahran, M.E. Khalifa, N.M. El-Metwaly, J. Mol. Struct. 1246, 131139 (2021).
- 13. L.H. Abdel-Rahman, A.M. Abu-Dief, H. Moustafa, A.A.H. Abdel-Mawgoud, Arab. J. Chem. 13, 649 (2020)
- E.M.M. Ibrahim, L.H. Abdel-Rahman, A.M. Abu-Dief, A. Elshafaie, S.K. Hamdan, A.M. Ahmed, Mater. Res. Bull. 107, 492 (2018)
- R. Antony, S.T. David, K. Saravanan, K. Karuppasamy, S. Balakumar, Spectrochim. Acta A Mol. Biomol. Spectrosc. 103, 423 (2013)
- 16. H. Chakraborty, N. Paul, M.L. Rahman, Transit. Met. Chem. 19, 524 (1994)
- 17. A.A. Alshaheri, M.I.M. Tahir, M.B.A. Rahman, T. Begum, T.A. Saleh, J. Mol. Liq. 240, 486 (2017)
- 18. V. Arun, N. Sridevi, P.P. Robinson, S. Manju, K.M. Yusuff, J. Mol. Catal. A Chem. 304, 191 (2009)
- 19. S. Iyer, G.M. Kulkarni, C. Ramesh, Tetrahedron 60, 2163 (2004)
- 20. G. Desimani, G. Dasi, G. Paita, P. Quadrelle, P. Righille, Tetrahedron 51, 4131 (1995)
- 21. T.T. Calam, Int. J. Environ. Anal. Chem. 99, 1298 (2019)
- 22. M. Finsgar, Corros. Sci. 77, 350 (2013)
- R.M. Izatt, G.C. LindH, R.L. Bruening, P. Huszthy, C.W. McDaniel, J.S. Bradshaw, J.J. Christensen, Anal. Chem. 60, 1694 (1988)
- 24. R.V. Gadag, M.R. Gajendragad, Talanta 25, 418 (1978)
- E. Stingaci, M. Zveaghinteva, S. Pogrebnoi, L. Lupascu, V. Valica, L. Uncu, A. Smetanscaia, M. Drumea, A. Petrou, A. Ciric, J. Glamoclija, Bioorg. Med. Chem. Lett. 30, 127368 (2020).
- H.A. El-Sherief, B.G. Youssif, S.N.A. Bukhari, A.H. Abdelazeem, M. Abdel-Aziz, H.M. Abdel-Rahman, Eur. J. Med. Chem. 156, 774 (2018)
- 27. R.S. Gani, K. Timanagouda, S. Madhushree, S.D. Joshi, M.B. Hiremath, S.B.H. Mujawar, A.K. Kudva, J. King Saud Univ. Sci. **32**, 3388 (2020)
- 28. K.N. Venugopala, M. Kandeel, M. Pillay, P.K. Deb, H.H. Abdallah, M.F. Mahomoodally, D. Chopra, Antibiotics **9**, 559 (2020)
- 29. Y. Pruglo, ScienceRise: Pharm Sci. 3, 19 (2018)
- B. Kaproń, J.J. Łuszczki, A. Płazińska, A. Siwek, T. Karcz, A. Gryboś, G. Nowak, A. Makuch-Kocka, K. Walczak, E. Langner, K. Szalast, Eur. J. Pharm. Sci. 129, 42 (2019)
- A.Z. El-Sonbati, M.A. Diab, S.M. Morgan, A.M. Eldesoky, M.Z. Balboula, Appl. Organomet. Chem. 32, e4207 (2018).
- 32. B. Abulhasanov, U. Yıldız, S. Akkoç, B. Coban, Appl. Organomet. Chem. 34, e5866 (2020).
- 33. A.I. Vogel, fifth ed. (Longmans, Edison, Wesley, London, 1999).
- 34. P.K. Sahoo, R. Sharma, P. Pattanayak, Med. Chem. Res. 19, 127 (2010)
- 35. L. Deswal, V. Verma, D. Kumar, A. Kumar, M. Bhatia, Y. Deswal, A. Kumar, Future Med. Chem. 13, 975 (2021)
- 36. A. Ahlawat, P. Khatkar, V. Singh, S. Asija, Res. Chem. Intermed. 44, 4415 (2018)
- 37. L.H. Abdel-Rahman, A.M. Abu-Dief, M.R. Shehata, F.M. Atlam, A.A.H. Abdel-Mawgoud, Appl. Organomet. Chem. **33**, e4699 (2019).
- 38. A.M. Abu-Dief, H.M. El-Sagher, M.R. Shehata, Appl. Organomet. Chem. 33(8), e4943 (2019).
- A.M. Abu-Dief, L.H. Abdel-Rahman, A.A. Abdelhamid, A.A. Marzouk, M.R. Shehata, M.A. Bakheet, O.A. Almaghrabi, A. Nafady, Spectrochim. Acta A Mol. Biomol. Spectrosc. 228, 117700 (2020).
- H.A. Al-Abdulkarim, R.M. El-khatib, F.S. Aljohani, A. Mahran, A. Alharbi, G.A. Mersal, N.M. El-Metwaly, A.M. Abu-Dief, J. Mol. Liq. 339, 116797 (2021).
- 41. T. Mosmann, J. Immunol. Methods 65, 55 (1983)
- 42. L.L. Kong, R. Ma, M.Y. Yao, X.E. Yan, S.J. Zhu, P. Zhao, C.H. Yun, Biochem. Biophys. Res. Commun. **488**, 266 (2017)
- 43. E.F. Pettersen, T.D. Goddard, C.C. Huang, G.S. Couch, D.M. Greenblatt, E.C. Meng, T.E. Ferrin, J. Comput. Chem. 25, 1605 (2004)
- 44. R.A. Laskowski, M.B. Swindells, J. Chem. Inf. Model. 51, 2778 (2011)
- 45. L.H. Abdel-Rahman, R.M. El-Khatib, L.A. Nassr, A.M. Abu-Dief, M. Ismael, A.A. Seleem, Spectrochim. Acta A Mol. Biomol. Spectrosc. **117**, 366 (2014)
- 46. A.M. Abu-Dief, L.A. Nassr, J. Iran. Chem. Soc. 12, 943 (2015)
- 47. L.H. Abdel-Rahman, A.M. Abu-Dief, R.M., El-Khatib, S.M. Abdel-Fatah, Bioorg. Chem. 69, 140 (2016).

- 48. N. Mehrotra, R.M. Tripathi, F. Zafar, M.P. Singh, IEEE Trans. Nanobiosci. 16, 280 (2017)
- 49. A.A. Olanrewaju, F.S. Fabiyi, C.U. Ibeji, E.G. Kolawole, R. Gupta, J. Mol. Struct. **1211**, 128057 (2020)
- 50. M. Yadav, S. Sharma, J. Devi, J. Chem. Sci. 133, 1 (2021)
- 51. V.V. Dhayabaran, T.D. Prakash, R. Renganathan, E. Friehs, D.W. Bahnemann, J. Fluoresc. **27**, 135 (2017)
- M.M. Slaihim, F.S.R. Al-Suede, M. Khairuddean, M.B.K. Ahamed, A.M.S.A. Majid, J. Mol. Struct. 1196, 78 (2019)
- 53. V. Gomathi, R. Selvameena, Asian J. Chem. 25, 2083 (2013)
- 54. J. Devi, M. Yadav, D. Kumar, L.S. Naik, D.K. Jindal, Appl. Organomet. Chem. 33, e4693 (2019)
- 55. S.S.A. Fathima, M.M.S. Meeran, E.R. Nagarajan, J. Mol. Liq. 279, 177–189.
- 56. W.J. Geary, Coord. Chem. Rev. 7, 81 (1971)
- 57. M. Murugaiyan, S.P. Mani, M.A. Sithique, New J. Chem. 43, 9540 (2019)
- 58. H.F. Abd El-Halim, G.G. Mohamed, M.N. Anwar, Appl. Organomet. Chem. 32, e3899 (2018)
- 59. S. Chandra, A. Kumar, Spectrochim. Acta A Mol. Biomol. Spectrosc. 68, 1410 (2007)
- 60. B. Hathaway, D.E. Billing, Coord. Chem. Rev. 5, 143 (1970)
- 61. B.J. Hathaway, A.A.G. Tomlinson, Coord. Chem. Rev. 5, 1 (1970)
- 62. N. Kavitha, P.V.A. Lakshmi, J. Saudi Chem. Soc. 21, S457 (2015)
- 63. J. Porkodi, N. Raman, Appl. Organomet. Chem. 32, e4030 (2018)
- 64. B.B. Beyene, A.M. Mihirteu, A.T. Ayana, A.W. Yibeltal, Results Chem. 2, 100073 (2020)
- 65. N. Beyazit, D. Çakmak, C. Demetgül, Tetrahedron 73, 2774 (2017)
- 66. J. Devi, S. Sharma, S. Kumar, D.K. Jindal, P.P. Dutta, D. Kumar, Res. Chem. Intermed. 47, 2433 (2021)
- D. Ercan, H.G. Choi, C.H. Yun, M. Capelletti, T. Xie, M.J. Eck, N.S. Gray, P.A. Jänne, Clin. Cancer Res. 21, 3913 (2015)
- C. Hu, A. Wang, H. Wu, Z. Qi, X. Li, X.E. Yan, C. Chen, K. Yu, F. Zou, W. Wang, Oncotarget 8, 18359 (2017)

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.