



Micro/nano-plastics occurrence, identification, risk analysis and mitigation: challenges and perspectives

Boda Ravi Kiran · Harishankar Kopperi · S. Venkata Mohan

Received: 23 July 2021 / Accepted: 29 December 2021 / Published online: 27 January 2022
© The Author(s), under exclusive licence to Springer Nature B.V. 2022

Abstract Micro/nanoplastics (MP/NPs) are emerging global pollutants that garnered enormous attention due to their potential threat to the ecosystem in virtue of their persistence and accumulation. Notably, - United Nations Environment Programme (UNEP) yearbook in 2014 proposed MPs as one among ten emergent issues that the Earth is facing today. MP/NPs can be found in most regularly used products (primary microplastics) or formed by the fragmentation of bigger plastics (secondary microplastics) and are inextricably discharged into the environment by terrestrial and land-based sources, particularly runoff. They are non-degradable, biologically incompatible, and their presence in the air, soil, water, and food can induce ecotoxicological issues and also a menace to the environment. Due to micro size and diverse chemical nature, MP/NPs easily infiltrate wastewater treatment processes. This communication reviews the current understanding of MP/NPs occurrence, mobility, aggregation behavior, and degradation/assimilation in terrestrial, aquatic (fresh & marine),

atmospheric depositions, wetlands and trophic food chain. This communication provide current perspectives and understanding on MP/NPs concerning (1) Source, occurrence, distribution, and properties (2) Impact on the ecosystem and its services, (3) Techniques in detection and identification and (4) Strategies to manage and mitigation.

Keywords Plastic pollution · Microfiber plastics · Ecological risk · Wastewater treatment · Circular economy

Abbreviations

AS	Acrylonitrile styrene copolymer
Cd	Cadmium
Co	Cobalt
Cr	Chromium
FeSO ₄	Iron (II) sulfate
H ₂ O ₂	Hydrogen peroxide
HBrCDs	Hexabromocyclododecanes
HDPE	High density polyethylene
HNO ₃	Nitric acid
KI	Potassium iodide
KOH	Potassium hydroxide
LDPE	Low density polyethylene
LC-MS	Liquid chromatography-mass spectrometry
NaCl	Sodium chloride
NaI	Sodium iodide
PA	Polyamide

B. R. Kiran · H. Kopperi · S. Venkata Mohan (✉)
Bioengineering and Environmental Sciences Lab,
Department of Energy and Environmental Engineering
(DEEE), CSIR-Indian Institute of Chemical Technology
(CSIR-IICT), Hyderabad 500007, India
e-mail: svmohan@iict.res.in; vmohan_s@yahoo.com

H. Kopperi · S. Venkata Mohan
Academy of Scientific and Innovative Research (AcSIR),
Ghaziabad 201002, India

PAH	Polycyclic aromatic hydrocarbon
PBB	Polybrominated diphenyl
PBDEs	Polybrominated diphenyl ethers
Pb	Lead
PC	Polycarbonate
PCB	Polychlorinated biphenyl
PE	Polyethylene
PE-PP	Polyethylene and polypropylene copolymer
PEST	Polyester
PET	Polyethylene terephthalate
PMMA	Polymethyl methacrylate
PP	Polypropylene
PS	Polystyrene
PU	Polyurethane
Py–GC/MS	Pyrolysis–gas chromatography/mass spectrometry
PVC	Polyvinyl chloride
Ti	Titanium
TiO ₂	Titanium oxide
WWTP	Wastewater treatment plant
Zn	Zinc
ZnCl ₂	Zinc chloride

1 Introduction

Plastics have diversified to infiltrate all human activity and utilities by replacing wood and metals due to their affordability and versatility. The world's first synthetic plastic was produced by Leo Baekeland in 1907. The manufacture of plastics and use was phenomenal between 1950 and 2015, with terrestrial emissions accounting for 80% of plastic waste globally (Geyer et al. 2017). Plastic pollution has become a ubiquitously distributed and severe transboundary threat to natural ecosystems (Thompson 2015). Packaging materials (39.5%), building materials (20.1%), fishing gears (10%), automotive parts (8.6%), electronic utilities (5.7%), and agro-industry components (3.4%) account for the majority of plastic depositions, with the remainder consisting of household, medical waste, and sports equipment (Horton et al. 2017; Plastics Europe 2017). Recent Covid-19 pandemic upsurged the consumption of single-use plastics, namely personal protective equipment (PPE) kits, facemasks, and gloves, contributing to an additional 3.5% of global solid

waste fraction (Patricio Silva et al. 2021). An estimated 4.90 trillion tonnes of plastics are dumped into oceans alone (O'Neill and Lawler 2021).

Microplastics (MPs; <5 mm in diameter) and nanoplastics (NPs; <100 nm in length) are becoming major concerns (Mariano et al. 2021), which can be found in the majority of regularly used products (primary) or can be formed by the fragmentation of bigger plastic litter (secondary) (Kokalj et al. 2021). Primary MPs are mass-produced from manufacturing and domestic applications, which includes exfoliating facial scrubs, toothpaste, detergents, personal care products, abrasive cleaning agents, plastic powder for molding, and synthetic clothing (nylon/polyester) (Wang et al. 2020a; Birch et al. 2020; Sun et al. 2019). Paints, adhesives, electronics, etc., are responsible for NPs release (Kihara et al. 2020). Secondary MP/NPs are formed by fragmentation of macroplastics (200–1000 µM) through shear forces (Auta et al. 2017) which account for 70–80% of all plastic released into the environment, while primary MPs contribute only 15–30% (Mariano et al. 2021). MP/NPs infiltrate terrestrial, freshwater, and marine ecosystems from the equator to the poles and surface water to deep-sea sediments (Wang et al. 2016).

In general, MP/NPs enter the water bodies through domestic waste, sewage treatment plants (STPs), industrial effluents, stormwater, estuaries, and riverine transport, surface runoff, wind currents, and disposal practices (Liu et al. 2021; Sutton et al. 2016). MP/NPs are present as fragments, pellets, fibres, films, granules, and Styrofoam that vary with surface-mass area ratios (Koelmans et al. 2019). The chemical variation found in STPs includes polyethylene, polystyrene, and polypropylene which tend to float while polyethylene terephthalate (PET) and polyvinylchloride (PVC) are more likely to sink due to their density (Wu et al. 2017; Avio et al. 2017; Carr et al. 2016). The presence of MP/NPs in the air, water, and food can cause ecotoxicological issues (Zhang et al. 2020). Influx of MP/NPs causes growth reduction, fecundity, reduced immunity, and malformation in animals and human reproductive systems (Wong et al. 2020; Guzzetti et al. 2018). MP/NPs also act as vectors to adsorb tenacious organic pollutants, trace metals, and harmful additives that are multiple times higher than natural sediments (Jiang et al. 2020). Presence of MP/NPs in WWTPs, impact microbial communities, inhibit sludge hydrolysis and accumulate acids (Zhang and Chen 2020).

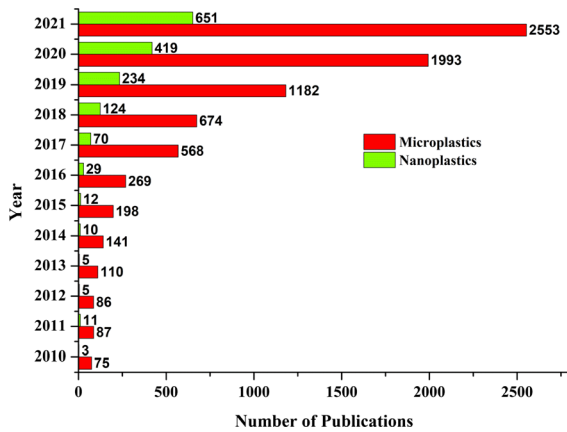


Fig. 1 The cumulative total of annual research papers published on microplastics and nanoplastics. Keyword used = Microplastics and nanoplastics. Data source: www.sciencedirect.com

Plastic occurrence, accumulation and persistence as MP/NPs in terrestrial and marine environments is becoming a major global concern. The issue of plastics was recognized by the United Nations (UN) Sustainable Development Goals (SDGs), specifically Goal 14—“Conserve and sustainably use the oceans, seas, and marine resources for sustainable development (Walker 2021). Using the keywords “microplastics” and “nanoplastics” as database search in indexed journals (ISI Science Direct), the publication number (MPs—7136/NPs -1369) is soaring exponentially, which elucidates growing interest in the scientific community (Fig. 1). This review aims to provide current perspectives on MP/NPs concerning (1) Source, occurrence, distribution, and properties (2) Impact on the ecosystem and its services, (3) Techniques in detection and identification of MP/NPs and (4) Remediation and management strategies to mitigate MP/NPs. The current understanding in the literature is highlighted, and future perspectives are indicated.

2 Occurrence and distribution of Micro/nanoplastics

2.1 Terrestrial system

The terrestrial domain is more susceptible to MP/NPs and the annual inputs from land exceeds the total MP/NPs floating in the global ocean (Yee et al.

2021; Wang et al. 2021; Hu et al. 2019). Soil bound MP/NPs originate from multiple sources, which includes sewage sludge/biosolids, domestic waste, irrigation, land fillings, fertilization, plastic film mulch, greenhouse materials, atmospheric deposition, tire abrasion and garden organic waste (Koutnik et al. 2021; Guo et al. 2020; Wang et al. 2020b). MP/NPs penetrate vertically via water infiltration, facilitated by tilling and action of soil microorganisms (O’Connor et al. 2019). The major source of MP/NPs in the soil is from WWTPs as 95% of MPs are retained in biosolids, and treated biosolids, in addition, are used as fertilizer in agriculture (van den Berg et al. 2020; Horton et al. 2017). Furthermore, a single-use facial scrub could release up to 10^6 primary NPs into the domestic sewage system (Shen et al. 2019). Around 306.9 tonnes of microbeads are discharged into the environment, of which 80% are derived from STPs annually (Cheung and Fok 2017). Biosolids alone introduce approximately 44,000 to 430,000 tonnes of MP/NPs into North American and European grasslands each year (Nizzetto et al. 2016). Textiles contribute 35% of the MP/NPs, specifically in the oceans, in the form of synthetic microfibers (Xu et al. 2020). A single garment produces about 1900 microfibrils, and around 700,000 microfibrils can be released from a single load of laundry, according to the International Union for Conservation of Nature (IUCN) (Boucher and Friot 2017).

Plastics have a lower density than soil minerals, and once they enter the soil, it forms soil aggregates and alter soil physicochemical properties. It effects water-binding capacity, soil density, porosity, soil integrity, soil surface cracking, and organic matter and influences the nutrient cycling process in the soil ecosystem (de Souza et al. 2019; Liu et al. 2017). Polyethylene, polystyrene, and polyvinylchloride nanoplastics (20 nm to 150 nm) indirectly influence soil productivity and directly alter the soil microorganisms and fauna communities (Wahl et al. 2021). Organic fertilizers and compost act as vectors for MP/NPs into the soil (Bläsing and Amelung 2018). Plastic mulch films are used extensively as soil conditioners in agriculture (Gao et al. 2019). Farmlands in China used ~ 1.47 million tonnes of plastic mulch with film content reaching 502 kg ha^{-1} soil in all provinces and regions (Zhang et al. 2016). Farmlands reported $62.5 \text{ particles kg}^{-1}$ in deep soil, $78.0 \text{ particles kg}^{-1}$ in shallow soil, and $1.9 \pm 0.87 \text{ particles kg}^{-1}$ in garden

soils (Liu et al. 2018). A study on the rice-fish co-culture ecosystem showed MPs concentration in soil ranges between 10 to 78 particles kg^{-1} (Lv et al. 2019; Liu et al. 2018). MPs congregations in Sydney's industrial zones ranged from 300 to 67,500 mg kg^{-1} (Fuller and Gautam 2016). The dispersal of plastic debris in coastal belts, vegetable farmlands, and riparian forest zones found MPs concentration ranged from 7100 to 42,960 particles kg^{-1} (Zhang and Liu 2018). Due to its large mass-surface ratio and hydrophobic nature when combines with other persistent organic pollutants (POPs), heavy metals, and antibiotics, they serve as vectors for toxic chemicals (Wang et al. 2020b; Hodson et al. 2017). MPs-POP mixtures threaten the terrestrial environment by adsorbing various POPs (Yang et al. 2019a, b). Microplastics absorb pesticides and hinder the chemical degradation of polyethylene film residues, resulting in pesticides accumulation (Ramos et al. 2015). MPs affinity to adsorb antibiotics was also reported (Li et al. 2018).

Soil act as a preliminary sink and most MP/NPs are deposited in landfills. Between 1950 and 2015, global trash production was expected to total 6300 million tonnes, with landfills and other environmental compartments accounting for 79% of the total (Patricio Silva et al. 2021; Ng et al. 2018). A study in Finland enumerated MP/NPs particles up to 4.5 items L^{-1} with size >1 mm in diameter (Kilponen 2016). Plastic trash blowing from landfills, if not adequately buried, can affect the soil and reach aquatic habitats (Peng et al. 2017). The particle and mass concentrations of MP/NPs in the untreated leachate were 235.4 ± 17.1 item/L and 11.4 ± 0.8 g/L, respectively, with particles size less than 50 μm (Sun et al. 2021). Fresh Kills landfills leached most of the medical waste on New York beaches in 1987–1988 (Hale et al. 2020). Natural catastrophes (hurricanes, tsunamis, and wildfires), construction and demolition landfills frequently aid in the deposition of plastic debris (Brand et al. 2018). PE and PP were the predominant polymer types in municipal solid waste (MSW) landfills with sizes ranging from 100 to 1000 μm (He et al. 2018).

2.2 Air

Micro/nanoplastics are released into the air due to wastewater treatment plant operation and (Chang et al. 2020) mechanical activities such as severe wind

events, sea surface spray and wave breaking which often entrain them into the atmosphere (Brahney et al. 2021). Sea-spray releases MP/NPs from the maritime environment into the atmosphere, resulting in a global extrapolated value of 136,000 tonnes blowing on shore per year (Allen et al. 2020). Fibre, film, foam, and fragment morphologies have been dominant in atmospheric deposition and air masses (Zhang et al. 2020). MPs have a higher density than air (1.225 g/L at 15 °C at sea level), and their density diminishes as altitude rises (Revel et al. 2018). The commonly distributed MP/NPs in environment are 0.91–0.93 g/cm^3 of low-density polyethylene, 0.94 g/cm^3 of high-density polyethylene, 0.85–0.83 g/cm^3 of polystyrene, 1.02–1.05 g/cm^3 of polyamide, 1.37 g/cm^3 of polyester, 1.38 g/cm^3 of PVC, 1.2 g/cm^3 of polycarbonate and 1.42 g/cm^3 of polyformaldehyde (Rezaei et al. 2019). MP/NPs suspended in atmospheric air correspond to synthetic fibres and, upon inhalation, cause respiratory disorders (Wright et al. 2020). A significant amount of MPs are ejected from tyres (100,000 metric tonnes) and brakes (40,000 tonnes) each year which get transported through the air (Evangelidou et al. 2020). Car tyres and brake systems are responsible for 5–10% of worldwide ocean plastic pollution and 3–7% of PM_{2.5} in the air (Evangelidou et al. 2020). According to a study, each person in the United States generates 4.7 kg of tire wear microplastics per year⁻¹, equating to 1.8 million metric tonnes per year⁻¹ (Hale et al. 2020). Road dust has been recognized as an important source of MP/NPs in urban/sub-urban areas. The atmospheric deposition of MP/NPs is higher in indoor environments, and about 33% of microfibrils contain petrochemicals (Wu et al. 2019). On normal exposure, > 900 MP/NPs particles are ingested by a child per year (Abbasi et al. 2019).

2.3 Aquatic system

In the early 1970s, the first report on MPs occurrence in marine system was published (Carpenter and Smith 1972). MPs enter into fresh and marine environments mainly through natural erosion and human activities (industrial and domestic wastewaters, aquaculture, and tourism) (Guo et al. 2020; Birch et al. 2020). “River to the ocean” is one of the major pathways for plastic reservoirs (Wu et al. 2019). Eight rivers in Asia (Yangtze, Yellow, Indus, Hai He, Ganges, Mekong, Amur, and Pearl), and two rivers in Africa (the Nile

and the Niger) are the sources of 90% of the plastics that reach the sea every year (Sadeghi et al. 2021). Every year river-fed estuaries dump 1.15 to 2.41 million tonnes of plastic trash into the sea (Lebreton et al. 2018). The North-western Pacific Ocean and Arabian Bay are widely polluted by MP/NPs with concentrations ranging from 640 to 42,000 items km^{-2} and 4.38×10^4 items per km^{-2} (Xu et al. 2019). An estimated 15–51 trillion MP/NPs particles (93–236 thousand metric tons) were found in oceans (Naik et al. 2019). Plastic pollution, dumping, and trash eventually goes downstream into estuaries and coastal seas. Changing rainfall, wind speed, waves, and oceanic currents contribute to the transfer of pollutants into seawater (Cózar et al. 2014). Agricultural runoff, soil erosion, and atmospheric deposits also transfer MP/NPs in marine ecosystems (Hale et al. 2020). Field studies conducted on the banks of Ciwalengke River, Indonesia, showed the presence of MP/NPs as primary fibres with concentrations ranging from 5.85 particles per litre in surface water and 3.03 particles per 100 g of sediment in sizes ranging from 50 to 2000 μm (Alam et al. 2019). The abundance of MPs ranged from 112 to 234 particles kg^{-1} dry weight in lake Bolsena shoreline sediments of central Italy (Lake Chiusi) (Fischer et al. 2016). Microalgae also can colonize MP/NPs and increase their specific density through biofouling and hetero-aggregation (Mateos-Cárdenas et al. 2021). In 2011, Japan's Tsunami injected 5 million tonnes of plastic debris into the Pacific, which is approximately the amount of plastic waste entering the ocean each year (Murray et al. 2018).

2.4 Wastewater systems

Anaerobic digestion is a prevalent approach for handling sewage sludge (in WWTPs), wherein particulate organic waste gets converted to soluble substrates is the first step. Polyethylene, polyamide, polyvinyl chloride, and polystyrene in activated sludge impede anaerobic digestion and have varying effects depending on their physicochemical nature (Zhang and Chen 2020). Additives or toxic chemicals released from MP/NPs rupture microbial cells or inhibit key enzymes related to acidogenesis (hydrolysis and fermentation) and methanogenesis and hinder anaerobic digestion (Azizi et al. 2021). MP/NPs also penetrate the exopolysaccharide matrix, alter protein

secondary structures, and damage phospholipids (Azizi et al. 2021). Following MP/NPs exposure, changes in the activity of key enzymes such as protease, cellulase, glucosidase, acetate kinase, butyrate kinase, coenzyme F420, etc. were observed to cause changes in the digester performance (Azizi et al. 2021). The effects of various doses of polyvinyl chloride and polystyrene (10–200 particles g^{-1}) on waste-activated sludge's anaerobic digestion negatively affected hydrolysis rate and biogas generation (Li et al. 2020). After exposing digesters to polystyrene nanoparticles, marked variations in pH, volatile fatty acids, and ammonia nitrogen profiles were reported (Zhang et al. 2020). Microbial communities also resulted in a considerable decrease in the digester upon short and long-term exposure to MP/NPs (Azizi et al. 2021).

3 Micro/nanoplastics degradation

Plastics can disseminate micro- and nanoplastics either via biodegradation or non-biodegradation processes (Yee et al. 2021). Plastics are long-chain organic compounds, and their degradation is determined by the change in physical properties (shape, size, porosity, surface area, and crystallinity) and their ability to interact with other pollutants (Cai et al. 2017; Campanale et al. 2020). Once disposed of, plastic waste undergoes chemical, biological, and environmental changes, breaking large pieces of plastics into micro and nanoplastics (Allen et al. 2020). Thermal degradation, physical deterioration, photodegradation, thermo-oxidative degradation, biodegradation, sand friction, and hydrolysis are some of the weathering mechanisms that degrade larger plastic polymers to MPs (Hale et al. 2020; Wu et al. 2019) (Fig. 2). NPs are formed when MPs are exposed to light (visible and UV rays) at moderate temperatures (30 °C) (Jiang et al. 2020). Hydrolysis and photodegradation are natural chemical reactions that use water molecules and UV-visible light to break down chemical bonds in polymers and turn them into monomeric forms (Yee et al. 2021). The fragmentation of polystyrene down to the nanoscale occurred in four weeks inside a weathering chamber (Kihara et al. 2020). Throughout the backbone of plastic polymers, heteroatoms (O, N, and S) serve as sites for hydrolytic/enzymatic processes and facilitate breakdown into low molecular weight

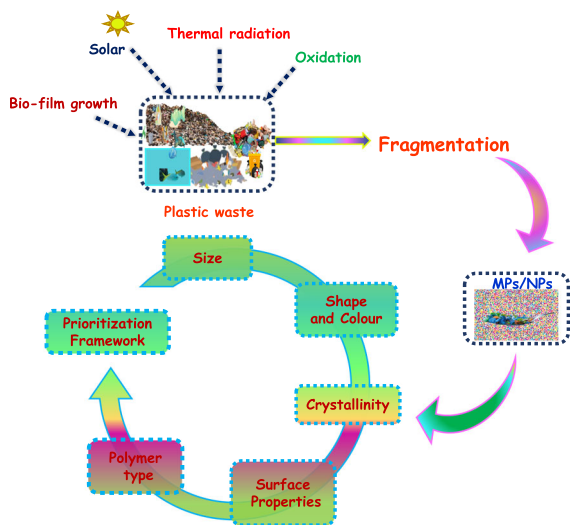


Fig. 2 Micro and nanoplastics fragmentation in terrestrial and aquatic systems and associated physico-chemical properties

fragments that microbial cells can assimilate (Ng et al. 2018). The chemical structure of polymers changes at the molecular level, such as chain scission, cross-linking, and the inclusion of oxygen-containing functional groups (esters, ketones, and alcohols) into the surface of plastic particles lowering their hydrophobicity (Yee et al. 2021). Non-biodegradation methods break down polymeric structures, changing mechanical characteristics and increasing surface zone, ensuring increased physico-chemical reactions and interfaces with microbes (Lucas et al. 2008). Microorganisms such as bacteria, fungi, and other eukaryotic colonizers secrete extracellular enzymes and mediate biodegradation (Naik et al. 2019; Enfrin et al. 2019). Biofilm formation also promotes the formation of hetero-aggregates where MP/NPs become sticky because of extracellular polymeric substance matrix (Wu et al. 2019). High saline content in marine environments, combined with naturally present microbes, promotes plastic disintegration faster than in terrestrial ecosystems (Ng et al. 2018). The structural integrity of the plastics is further weakened by subsequent interactions with wind, waves, solar UV radiation, temperature, additives, environmental conditions, and other abrasive interactions released through delamination (Jiang et al. 2020).

4 Properties of micro/nanoplastics

Microplastics' physical properties are generally associated with particle size, shape, and color, whereas chemical properties associate with crystallinity, surface properties, toxic metals, and additives adhered on MP/NPs surfaces (Hildebrandt et al. Lambert et al. 2017).

4.1 Size

Micro/nanoplastics sizes are often determined by the sampling and analysis methods. MPs come in various sizes, but the most common are 1 μm to 5 mm, whereas NPs are <100 nm in length (Murray and Örmeci 2020; Hartmann et al. 2019). The National Oceanic and Atmospheric Administration proposed these dimensions. The Joint Group of Experts later adopted the Scientific Aspects of Marine Environmental Protection (GESAMP 2016) and the United Nations Environmental Program (UNEP 2014). Currently, the two most common MP/NPs classification methods are sieve retention and microscopic imaging. The dimensions 25 mm, 100 mm, and 500 mm were the most frequently identified sizes in WWTP influent and effluents (Mintenig et al. 2017). Samples from the Atlantic Ocean revealed MPs in size 20 to 40 μm , accounting for 64% of the total detected particles (Simon et al. 2018; Enders et al. 2015), where atmospheric depositions accounted for microfibers of ~ 5000 μm in size (Cai et al. 2017). The size and ubiquitous distribution of MP/NPs in the aquatic and marine environment facilitate their availability to pelagic and benthic habitats (Campanale et al. 2020). Several species of zooplankton (*Calanus finmarchicus*, *Euphausia pacifica*, and *Neocalanus cristatus*), bivalves, macro-sized invertebrates, and fish ingest a wide range of MP/NPs ranging from 0.5 nm to 816 μm (Botterell et al. 2019; Lehtiniemi et al. 2018).

4.2 Shape

The shape is one of the important indicators used for microplastics classification. MP/NPs come in a wide range of shapes (fragment, foam, paint, pellet, foil, sphere, fibre, film, line, bead, flake, sheet, granule, and nurdle) (Campanale et al. 2020; Zhang et al. 2020; Koelmans et al. 2019). The primary form of microplastics, degradation, erosion process and the

duration time in the environment all influence the shape of MP/NPs (Murray and Örmeci 2020). Fibres and fragments are the dominant shapes and account for 52.7% of wastewater and urban atmospheric depositions (Annenkov et al. 2021). Fibre particles form due to mechanical abrasion and chemical weathering, whereas fragmented microplastics may form due to greater plastic items exposed to fatigue or UV light (Zhou et al. 2020). The shape of MP/NPs influences pollutant transport in the environment. Films are thin and have a bigger surface area than fragments of the same mass for atmospheric conveyance (Allen et al. 2020). Microfibres and fragments are dominant shapes found in international waters (Wang et al. 2020a).

4.3 Colour

Colour is one of the most important parameters for identifying plastic debris and potential contaminations in sample preparation. Various MP/NPs have been documented including red, orange, yellow, brown, tan, off-white, white, grey, blue, and green (Murray and Örmeci 2020; Rochman et al. 2019). Particles with dark, transparent, white, or translucent may be under-represented (Hartmann et al. 2019). Blue and red fibres are the most commonly reported colours (Bergmann et al. 2019). Polypropylene has been clear and transparent, while polyethylene and polyethylene with low-density have been assigned opaque colors (LDPE) (Zhang et al. 2020). Microplastic discoloration can occur during weathering and sample preparation (H_2O_2 -oxidative digestion), which should be considered when reporting and interpreting data (Allen et al. 2020). Marine organisms commonly ingest bright color NPs particles similar to natural foods. Zooplankton, euphausiid, copepods, and fish larvae ingest MPs predominantly in blue, red, green, and black (Botterell et al. 2019).

4.4 Chemical additives

MPs and NPs are mostly additives and basic polymeric materials generated from plastics and chemicals absorbed from the environment (Table 1). Plastic toxicity is associated with built-in chemicals (i.e., monomers, solvents, catalysts, additives-dyes, plasticizers) (McGivney et al. 2020). Low molecular weight additives entrenched into the polymer matrix are weak and leaches readily into surrounding water bodies

(flame retardants from electronic items; PET oligomers from bottles and food trays; Pb from unplasticized PVC pipes; nonylphenol from food contact materials and Sb from PET water bottles) (Schwinghammer et al. 2020; Dobslaw et al. 2020). During plastic production, additives are added for desired colour, transparency and to improve the working liability of the product, which makes the polymer more resistant to degradation from physical (temperature, ozone, and light radiation), mechanical, electrical, and biological (fungi, bacteria) agents (Hale et al. 2020). Inert or reinforcing fillers, dyes, UV stabilizers, plasticizers, lubricants, and flame retardants are additives. In addition, wood, graphite, glass fibres, rock flour, kaolin, cotton flakes, jute, clay or linen, cellulose pulp are also used as additives (Campanale et al. 2020; Dobslaw et al. 2020). These additives upgrade the plastic properties, but most are toxic and potential soil, water, and air contaminants. Ingestion or inhalation of these chemicals disrupts the endocrine system and causes hormonal imbalance, asthma, reproductive problems, metabolic disorders, and neuro-developmental conditions (Schwinghammer et al. 2020). Bisphenol A, phthalates, heavy metals, and brominated flame retardants are most common additives (Li et al. 2021).

4.5 Crystallinity

Crystallinity is ordered structural linkages that influence the plastics' density, permeability, and swelling behavior (Botterell et al. 2019; Lambert et al. 2017). The crystallinity of MP/NPs changes with residence time in the environment. The breakdown of the amorphous region in the polymer promotes overall crystallinity and reduces the size of MP/NPs (Campanale et al. 2020). The materialization of crystallites might differ in toxicity and will alter MP/NPs from their counterparts. It eventually influences particle shape, size, surface area, and chemical properties like adsorption of pollutants, additives and consecutively affects ingestion rate (Botterell et al. 2019; Gao et al. 2019).

4.6 Surface properties

The surface property primarily involves surface area and surface chemistry. With the decrease in particle size, MP/NPs surface area increases, and particles at

Table 1 Various Chemical additives, polymers and metals with plastics and their effect on the human health

Additives	Effect on human health	References
Plasticizers	1,2-benzenedicarboxylic acid	Neuronal toxicity
	Chlorinated paraffins, Dicyclohexyl phthalate (DCHP)	Breast cancer
	di-C7-11-branched and linear alkyl esters (DHNUP)	Cardiovascular
	Butyl benzyl phthalate (BBP)	Kidney diseases
	Diethyl phthalate (DEP)	Metabolic and mental disorders
	Diheptyladipate (DHA)	Neuro-degenerative disorder
	Formaldehyde, 4,4'-methylenedianiline (MDA)	
	Dipentyl phthalate (DPP)	
	Diisobutyl phthalate (DiBP)	
	Diisoheptylphthalate (DIHP)	
Heavy metals—Zinc, Cadmium, Tin, Lead, Titanium, Barium		
Biocides	Arsenic trioxide	Metal–estrogen
	Triclosan	Mutagen
	Triphenyltin hydroxide	Carcinogen
	Butyltin trichloride	Brain damage
	Dimethyltin dichloride	Congenital disabilities
	Dibutyltin dichloride	Lung, skin, liver, bladder, kidneys,
	Tetrabutyltin	Gastrointestinal damage
	Tributyltin chloride	
Heavy metals- Antimony, Copper, Mercury, Arsenic, Tin		
Flame retardants	2,2',4,4'-Tetrabromodiphenyl ether (BDE 47)	Carcinogen
	3,3'-5,5'-Tetrabromobisphenol (TBBPA)	Apoptosis
	1,2,3,4,5,6-Hexabromocyclohexane	Genotoxicity
	Tris(2-chlorisopropyl)phosphate (TCPP)	Osteomalacia and bone fractures
	Boric acid	DNA methylation
	Decabromodiphenylethane (DBDPE)	
	2,2',4,4',5,5'-Hexabromobiphenyl ether (BDE 153)	
	Heavy metals—Antimony, Bromine, Aluminum, Zinc, Bromine	
Stabilizers, Antioxidants and Organic pigments	Bisphenol A	Metabolism changes
	Fatty acid amides	DNA methylation
	2-tert-Butyl-4-methoxyphenol	Anemia
	Triglycidylisocyanurate (TGIC)	Neurological disorder
	2-t-butyl-4 hydroxyanisole (BHA)	Cardiovascular and endocrine deficits
	Tris(2,4-di-tert-butylphenyl) phosphate	Hypertension
	Butylated hydroxytoluene (BHT)	Miscarriages
	Tris-nonyl-phenyl phosphate (TNPP)	Disruption of nervous
	4-Nonylphenol	Brain damage
	Irganox 1010	Infertility
4-Octylphenol		
Heavy metals—Aluminum, Manganese, Barium, Cobalt, chromium, Lead, Titanium, Tin, Cadmium, Aluminum		

the nanoscale have a marked impact (Gao et al. 2019). Although the surface area is not widely recognized, microbeads can be calculated using the spherical equivalent diameter (Botterell et al. 2019). Nanoscale particulates are assumed using geometrical estimates, resulting in a sevenfold interpolation of the surface area (Campanale et al. 2020). The plastic surface is affected by photo- and oxidative degradation processes that generate new functional groups when reacted with OH radicals, O and N oxides, and other photo-generated radicals (Schwinghammer et al. 2020). These pathways disrupt the plastic surface area, and microscopic particles are released upon ingestion, chemical leaching, and formation of angular-shaped NPs from primary microbeads (Lambert et al. 2017). Furthermore, surface chemistry significantly impacts particle-biota interactions as the microbial community utilizes the oxygenated binding sites (McGivney et al. 2020; Campanale et al. 2020).

5 Impact on the environment

5.1 Soil

Micro/nanoplastics are a serious threat to soil biota because they inhibit plant growth, organism reproduction, and soil biodiversity (Hale et al. 2020). Soil serves as a habitat for terrestrial animals, and the ecotoxicological effects of MP/NPs on soil fauna are exuberantly increasing (Fig. 3). MP/NPs are ingested by soil invertebrates and poultry, providing a potential gateway to enter humans and animals due to their small size (Cox et al. 2019). Species like nematodes, snails, mice, isopod, collembolan, and chicken are studied with different MP/NPs (HDPE, PVC, PE, PS, PA, and PET), of which earthworm studies are predominant (Ng et al. 2018). The toxicity of MP/NPs on soil biota includes growth suppression, energy metabolism disturbance, immunological responses, locomotion reduction, gastrointestinal disorders, gut microbiota dysbiosis, metabolic abnormality, and mortality (Wang et al. 2020b). In vertebrates, leaching from MP/NPs additives such as bisphenol A and phthalates disrupt the endocrine system through the estrogenic effect (Zhang et al. 2020). Polystyrene MPs showed damage to cholinergic, GABAergic neurons, and oxidative stress kinase (Lei et al. 2018). *Lumbricus terrestris* (earthworm), when exposed to 28% PE-

MP in dry plant litter, experienced growth reduction and eventually cell death (Huerta Lwanga et al. 2016). A cross-section study reported the existence of MPs debris in the gastrointestinal tract and gut system of domestic live-stock and terrestrial birds (Omidi et al. 2012). Sunlight, oxygen availability, high temperature, soil microbes, and terrestrial biota in the top soil layer function as plastic garbage degradative habitat (Wang et al. 2020a). Antibiotic resistance genes are transported, deposited, and dissipated in soil by MP/NPs (Sun et al. 2019). Microbial enzymes (dehydrogenase and fluorescein diacetate hydrolysis) tend to increase plastic mulch residues (Wang et al. 2016). MP/NPs can affect gene expression and alter the microbial cellular structure. They penetrate cells through endocytosis (Zhang and Chen 2020). MP/NPs with exopolysaccharides (EPS) in wastewater systems are mediated by functional groups such as carbonyl, amide groups, and side chains of lipids or aminoacids (Feng et al. 2018). PS-NPs showed a change in the secondary structure of proteins in EPS (Catarino et al. 2021).

5.1.1 Plant and macrophytes

Plants metabolize a wide range of pollutants (MPs, NPs, and POPs) either by avoiding its uptake or detoxifying by various enzymatic activities in the plant cell (Dobslaw et al. 2020; Ng et al. 2018) (Fig. 3). In the plant root zone, soil microorganisms and animals participate in MP/NPs degradation and protection from toxic effects (de Souza et al. 2019). MP/NPs alter soil fertility and significantly affect growth, biomass, root traits, nutrient uptake, and soil microbial activities (Bianco and Passananti 2020; Zhang et al. 2020). MP/NPs adsorption and internalization have been observed in plants used for human consumption (*Triticum aestivum*, *Allium cepa*, *Allium fistulosum*, *Lactuca sativa*, and *Zea mays*) and macrophytes (*Lemna minor* and *Fucus vesiculosus*) (Mateos-Cárdenas et al. 2021). Polystyrene (PS), polyethylene (PE), polypropylene (PP), cotton, and rayon fibres were commonly found adhering to plant surfaces (Mateos-Cárdenas et al. 2021). *Triticum aestivum* and *Lactuca sativa* internalize 200 nm PS nanobeads and 2.0 μm polymethylmethacrylate microbeads, respectively (Li et al. 2020). PS nanobeads (10 nm to 200 nm) was internalized in the root maturation zone and translocated to leaf vessels via



Fig. 3 Micro/nanoplastics distribution across different food trophic levels and their interaction with biological processes in terrestrial, fresh and marine systems

vascular tissues in *Zea mays* L. (Sun et al. 2021), *C. pteridoides* (Yuan et al. 2019), and *Vigna radiata* (Chae and An 2017). Apple, broccoli, and carrots showed significant quantities of MPs per gram (Conti et al. 2020). In a study, co-culturing *Lolium perenne* (ryegrass) with *Aporrectodea rosea* (rosy-tipped earthworm) with different MPs (PE, HDPE, and fibres) showed reduced plant biomass, pH, and inhibited growth (Boots et al. 2019). Various plant species and soil types have different reactions. Plants and macrophytes act as potential vectors for trophic transfer of MP/NPs into natural food webs. Duckweed is toxic to amphipods (Mateos-Cárdenas et al. 2021), *F. vesiculosus* is toxic to *Littorina littorea* (Gutow et al. 2019), and *V. radiata* is toxic to *Achatina fulica* (Mateos-Cárdenas et al. 2021). High doses of MPs are used on a laboratory scale to identify and impact soil biota easily. However, MPs levels in human-impacted soil are unlikely to be greater than 0.1% dry weight (Sun et al. 2021).

5.1.2 Mangroves

Mangrove wetland is an intertidal ecosystem that acts as a barrier for retaining land-based pollutants, including MP/NPs (Helcoski et al. 2020; Li et al.

2020). Nor and Obbard (2014) first reported MPs in mangroves with concentrations ranging from 12 to 62.7 items per kg. Cienaga Grande de Santa Marta (CGSM), one of the coastal lagoons declared as RAMSAR wetland, consists of MP/NPs concentrations oscillated between 31 and 2863 particles per kg of dry sediment (Garcés-Ordóñez et al. 2019). The plastic debris is always accumulated on the leaf's surface, trunk, pneumatophores and transported by ocean currents and storm runoff from the land (Zhou et al. 2020). Fibres, foams, and films are dominant MP/NPs detected in mangrove sediments worldwide (Duan et al. 2020). MP/NPs inhibit the growth of mangrove seedlings, loss of foliage, and negatively affect mollusks, crabs of the *Uca* genus, and crustacean habitats that live in soils associated with mangroves (do Sul et al. 2014). Four species of juvenile fish (viz. *Ambassis dussumieri*, *Terapon jarbua*, *Mugil sp.*, and *Oreochromis mossambicus*) growing in KwaZulu-Natal mangroves unveiled fibres (68%) and fragments (21%) with 0.79 ± 1 particle per fish (Naidoo et al. 2020). Similar findings were found in each fish's gills, stomach, and intestine in the Zhanjiang mangrove wetlands, where 2.83 ± 1.84 items per individual MPs were reported (Huang et al. 2020). *Gerreidae* sps. (*Eugerres brasiliensis*,

Eucinostomus melanopterus, and *Diapterus rhombeus*) found in estuaries and mangroves ingested MPs and showed a reduction in gut weight and false satiation (Ramos et al. 2012).

5.2 Aquatic biota

Plastic pollution is an upsurging global concern in aquatic systems, spreading even to the most remote habitats (Fig. 3). Plastic debris ranging from microscopic to high density is prevailing in benthic and pelagic habitats of rivers, seas, and remote regions of Arctic to Antarctica (Bianco and Passananti 2020). MP/NPs have been identified as a major concern for biodiversity loss, climate change, and threatening living biota.

5.2.1 Fresh water systems

In recent years, freshwater systems such as ponds, lakes, estuaries, and rivers have attracted as much attention as oceans in terms of MP/NPs flux (Wong et al. 2020) through domestic waste, industrial effluents, and sewage disposal sites (Meng et al. 2020; Hu et al. 2019). Particle qualities (size, shape, and density) and physical forces (flow velocity, water depth, sediment topography, tidal cycles, and urban runoff), and anthropogenic activity (e.g., dam release) influence MP/NPs movement in freshwater systems (Helcoski et al. 2020; Ziajahromi et al. 2017). MP/NPs have been found in freshwater systems across the continents (Lahens et al. 2018). The Danube delivers 4.2 tonnes of plastic each day to the Black Sea (Lechner et al. 2014). Microbeads, raw plastic (pellets and flakes), films, fibres, and fragments are types of MP/NPs found across the freshwater streams, where microbeads and fibres are dominant (Li et al. 2021). Plastic debris accumulation in aquatic habitats alters light penetration and affects biogeochemical cycles in the water column (Chen et al. 2020). NPs absorbed on the surface of microalgae cause a shading effect and decrease the fluidity of cell membranes (Zhu et al. 2021). Internalized NPs reduce carbohydrate metabolism and decrease cellular esterase activity, electron transport rate, and lipid reserves, thus modulating algal energy metabolism (Zhu et al. 2021). Large plastic particles, such as fishing ropes and nets, entangle turtles, birds, and mammals, whereas small particles are ingested directly, causing a gut

obstruction (Horton et al. 2017). MP/NPs in freshwater systems severely impact the ecosystem as they are the major reliable sources of food and drinking water for humans (Senathirajah et al. 2021). NPs last longer in the body than MPs and are transported to the digestive gland. Trophic transfer of NPs was investigated, and increased uptake causes tissue inflammation, reduced lipid stores, nutrient uptake, and membrane instability in digestive cells (O'Neill and Lawler 2021). *Daphnia magna* and *Danio rerio* (Zebra fish) are widely used biological and toxicological research models to assess MP/NPs in freshwater systems (Jiang et al. 2020). When exposed to 0.1, 1, and 10 ppm of polystyrene NPs, Zebrafish embryos, accumulation in embryonic tissues and choroid membranes was observed, thereby influencing the inter or trans generational toxicity (Pitt et al. 2018). Freshwater fauna (*Daphnia magna*, *Gammarus pulex*, and *Lumbriculus variegatus*) ingested MPs and translocated them from cells to oil storage droplets and faces (Imhof et al. 2013). A field study in French water streams showed 7 out of 11 gobies (*Gobio gobio*) contained MPs in their tissues (Sanchez et al. 2014). Polystyrene NPs of sizes 20 nm to 39.4 nm affect the biomass, photosynthetic efficiency, and mortality rate of *Chlorella* sp., *Daphnia magna*, *Raphidocelis subcapitata*, and *Scenedesmus obliquus* (Chae and An 2017). Organisms predate freshwater species at higher trophic levels, and MP/NPs consumed have cascade impacts on the ecosystem's health (Nizzetto et al. 2016; Li et al. 2015).

5.2.2 Marine systems

Marine litter is a huge environmental and economic issue around the world. Plastic garbage reaches the ocean at a pace of 4.8 to 12.7 million tonnes per year, with 80% of it coming from land-based sources worldwide (Mofijur et al. 2021; Raju et al. 2018). Rivers, lakes, sewage effluents, and anthropogenic activities are major gateways to transport plastic debris to oceans (Xu et al. 2019). The European Union's Marine Strategy Framework Directive (MSFD), the OSPAR commission, the Stockholm Convention, and International Pellet Watch have all focused on marine litter problems to protect and conserve its resources (Stockholm convention, UNEP 2018; OSPAR 2014). Aerial surveys in the Pacific Ocean discovered 79 thousand tonnes of plastic

floating over a 1.6 million km² area, dubbed the “Great Plastic Garbage Patch.” (Lebreton et al. 2018). The density of sea water is 1.02–1.07 g/cm³, while that of plastic is 0.8–1.5 g/cm³, which tends MPs to float on water surfaces (Wang et al. 2020a). PS, PP, PE, PU, PVC, and PET are among the most common plastics contributing to marine pollution (Annenkov et al. 2021; Birch et al. 2020). Nine sediment samples at 2340–5570 m depth analyzed in the Arctic region showed widespread NPs (42–6595 MPs kg⁻¹) (Bergmann et al. 2019). MPs will outnumber fish in oceans by 2050, according to World Economic Forum 2016.

Once plastic debris enters the marine environment, they interact with various marine species across trophic levels (Wang et al. 2020a; Guzzetti et al. 2018). MP/NPs have been found in a wide variety of marine species, including cetaceans, copepods, chaetognaths, shrimps, echinoderms, zooplankton, corals, fishes, turtles, seabirds, and mammals, causing cascading effects in the marine food web due to their small size (Botterell et al. 2019). Phagocytosis and pinocytosis are two pathways for marine organisms’ uptake of MP/NPs (Allen et al. 2020). They are transported through several physiological routes and translocate mainly to the stomach, intestine, gills, digestive tract, and sometimes to the liver (Allen et al. 2020). Ingestion of MPs causes deleterious effects in marine taxa, mainly impairment, reduced feeding behavior, inhibited growth, nutrient uptake, decreased immune response, oxidative stress, cellular toxicity, pseudo-satiety sensation, fertility reduction, malnutrition, and lastly, death (Gonçalves and Bebianno 2021). *Tigriopus japonicus*, when exposed to polystyrene microbeads, exhibited a decrease in fecundity across two generations (Lee et al. 2013). *Brachionuskoreanus* ingestion with fluorescent-labeled PS nanobeads (50 nm, 500 nm, and 6 µm) causes increased oxidative stress and decreased growth rate, fecundity, and body size (Jeong et al. 2016). Adult oysters and blue mussels on ingestion with 50 nm NPs lead to a substantial decrease in embryo-larvae development and fertilization, which results in complete stagnation (Rist et al. 2019). Similarly, MPs act as vectors for *Aeromonas salmonicida* (a fish bacterial pathogen), which can form biofilms and transfer to the food chain upon consumption of infected fish (Viršek et al. 2017). To investigate the anthropogenic impacts, 41 fishes were sampled from Giglio Island after refloating

project of Coasta Concordia Wreck. Out of 41, 85% of examined fish, i.e., benthonic species *Phycis phycis*, *Scorpaena* sp., and *Uranoscopus scaber* showed 77%, 84%, and 86% plastic particles. In comparison, benthopelagic *Spondyliosoma cantharus* exhibited 100% plastic particles in the gastrointestinal tract dominated by fragments, lines, and films (Avio et al. 2017). Microorganisms and fisheries habiting coral reefs are eight times more likely to be affected by plastics. MP/NP loads correlate to terrestrial dispersal into the season coral reefs. The amount of plastic objects entangled on coral reefs in the Asia–Pacific region is predicted to reach 11.1 billion, with the number expected to rise by 40% by 2025 (Lamb et al. 2018). Due to the high surface-volume ratio, hydrophobicity, and lipophilicity, MP/NPs can incorporate heavy metals (Ni, Pb, Zn, Cu, Cd, and Ti), PCBs, PAHs, POPs, phthalates, and bisphenol A on their surface (Prata 2018). Co-contamination of mercury and microplastic in marine environments causes behavioral changes and reduced swimming velocity in *Dicentrarchus labrax* (Barboza et al. 2018). *Oryzias latipes* (Japanese medaka fish), when exposed to polyethylene and chemical pollutants (PCBs, PAHs, and PBDEs), showed symptoms of tumor formation, cellular necrosis, down regulation of choriogenin (chgH) in males, and vitellogenin (VTgI) gene expression in females (Guzzetti et al. 2018). Perfluorooctane sulfonic acid and benzo[a]pyren) interact with polyethylene MPs, and are known to induce cancer in humans (O’Donovan et al. 2020). MP/NPs also function as medication delivery vectors entering marine water as medical waste or human faces (Campanale et al. 2020).

5.3 Food web

Micro/nanoplastics enter the food chain because of their wide availability in aquatic and terrestrial environments. MP/NPs were observed in honey, beer, poultry, salt, sugar, teabags, milk, salmon, seaweed, shrimps, and bivalves among other things (Markic et al. 2020; Cox et al. 2019; Santillo et al. 2017). According to reports, the average MP/NPs in food products is as follows: seafood (1.48 items g⁻¹); sugar (0.44 items g⁻¹); honey (0.10 items g⁻¹); salt (0.11 items g⁻¹); alcohol (32.27 items L⁻¹); bottled water (94.37 items L⁻¹); and tap water (4.23 items L⁻¹) (Yee et al. 2021; Zhang et al. 2020). Plastic teabags,

bottled water, and seafood are among the sources of exposure (Kosuth et al. 2018), and unusually high MP/NPs in fruits and vegetables have also been observed (Conti et al. 2020). On average, human beings consume 39,000 to 52,000 MP/NPs particles per year, whereas individuals who drink bottled water, in addition, consume 90,000 particles (Mason et al. 2018). MP/NPs contamination in drinking water bottles resulted in $40.1 \text{ mg kg}^{-1} \text{ body-weight day}^{-1}$ for adults and $87.8 \text{ mg kg}^{-1} \text{ body-weight day}^{-1}$ for children (Zuccarello et al. 2019). MPs occurrence was also reported in salt between 56 and 103 particles kg^{-1} of salt (Seth and Shriwastav 2018).

6 Health risk

Plastic usage in daily human activities (drinking water bottles, soft drinks, food packaging, and medical device, etc.) increases their abundance and human health risks (Jiang et al. 2020). The major routes for MPs intake are inhalation, skin perfusion, and ingestion, whereas NPs enter organisms through physical piercing and endocytosis/phagocytosis (O'Neill and Lawler 2021). The most prevalent route of MP/NPs exposure is through contaminated food, associated with gastrointestinal consequences, including reduced epithelial permeability, localized inflammatory processes, and changes in gut microbiota composition (Campanale et al. 2020). Most European countries rely on shellfish and crustaceans as their food diet and are estimated to ingest 1800 MPs/year per person (Barboza et al. 2018). MPs were identified in the tissues of commercially cultivated bivalves, *M. edulis*, and *Crassostrea gigas*, with quantities ranging from 360.07 to 470.16 particles per gram, collected from mussel farms and supermarkets (Zhang et al. 2020; van Cauwenberghhe and Janssen 2014). Rapid bronchospasm, diffuse interstitial fibrosis, inflammatory and fibrotic changes in bronchial and peribronchial tissue, and interalveolar lesions are caused by inhaling MP/NPs (Mariano et al. 2021). The accumulation of NPs by primary producers *Chlamydomonas reinhardtii* and *Daphnia magna* starts the trophic transmission of NPs in the food web (Zhu et al. 2021). The trophic transfer occurs when primary producers are eaten by secondary (*Oryza sinensis*) and tertiary (*Zacco temminckii*) consumers before being passed to humans (Zhu et al. 2021). NPs transferred from the

primary producers to top consumers make morphological alterations and strongly affect their behavior and metabolism (Zhu et al. 2021).

On the other hand, MPs are unlikely to be absorbed via the skin since stratum corneum diffusion is limited to nano-sized polymers with a diameter of less than 100 nm (Gonçalves and Bebianno 2021). Personal and cosmetic goods, particularly in the body, include nanoplastics, as face washes are administered topically to the skin. Urea, glycerol, and -hydroxyl acids, all common constituents in body lotions, improved the nanoparticle's capacity to penetrate the skin barrier (Jatana et al. 2016). Oral administration of MP/NPs produces redox imbalance, disruption of energy homeostasis, and neurotoxicity in the gut, intestine, and kidney (Deng et al. 2017). Endocytic mechanisms allow polystyrene and PVC particles (150 nm) to penetrate the gut wall and end up in lymph nodes and the blood vascular system (Xu et al. 2019). A significant reduction in testosterone, luteinizing hormone, and follicle-stimulating hormone was reported with mice given 40 nm polystyrene NPs for 35 days (Amereh et al. 2020). Recent research found 20 MPs (50–500 μm) per 10 g of human feces, with nine distinct plastic types (most common of which were polypropylene and polyethylene terephthalate) (Shen et al. 2019). Organochlorines are well-known endocrine-disrupting compounds that increase cytotoxicity and inflammatory response (Prata 2018). NPs affect neustonic/planktonic habitats and act as surface receptors for accumulating organochlorines (OCs) and cause toxicological effects in organisms such as *Balaenoptera physalus* and *Cetorhinus maximus* (Fossi et al. 2014).

7 Sampling and analysis

Organizations, namely ICES, GESAMP, UNEP-MEDPOL and JPL Oceans, developed standardized methodologies on MP/NPs monitoring (GESAMP 2016).

7.1 Sampling

Wastewater streams, surface water, bottled water, sediments, untreated and treated tap water were collected by pumping, trawling, followed by sieving to separate MP/NPs particles of the desired size range

(Yang et al. 2019a, b; Koelmans et al. 2019; Raju et al. 2018). Auto-sampler, surface filtration, separate pumping, and containers are employed in collecting sampling for MP/NPs quantification and characterization from wastewater streams (Fig. 4). Separate pumping and filtration processes are efficient methods for collecting wastewater due to high solids, organic matter, and sampling volume ranging from hundreds to cubic metre litres (Song et al. 2018). Neuston nets, manta trawls, etc., are often used for sampling MP/NPs from aqueous samples (Song et al. 2018). Samples above 500 μm are fractionated with a stainless-steel filter and sorted under a stereomicroscope (Yang et al. 2019a, b; Koelmans et al. 2019). For identification, samples with size fractions less than 500 μm underwent enzymatic and oxidative purification. Sand samples from the shoreline identified MPs when sieved through 5 mm mesh followed by 2 mm (Ceccarini et al. 2018). Post-treated sewage sludge and anaerobically digested sludge were passed through 250 μm sieve to trap MPs and later filtered through 212, 63, and 45 μm sieves for particle size fractionation (Mahon et al. 2017). Samples from various WWTPs passed through the customized filtration device with an electric pump and identified MPs with

a particle size of $> 300 \mu\text{m}$, 100–300 μm , and 20–100 μm (Talvitie et al. 2017; Li et al. 2021; Yang et al. 2019a, b; Cabernard et al. 2018; Song et al. 2018; Ceccarini et al. 2018; Dyachenko et al. 2017). Hermsen et al. (2018) suggested using fixatives like ethanol, formalin, or methyl aldehyde in sampling containers avoids retention of MPs. Ethanol and formalin have less effect on polymer, but it tends not to lose their characteristics (Sun et al. 2019). For NPs separation, magnetic field flow fractionation (MFFF), gel electrophoresis, and size-exclusion chromatography (SEC) were used (Li et al. 2021; Nguyen et al. 2019).

7.2 Density separation

Density separation is also used to isolate MP/NPs from wastewater to limit their number in WWTPs (McGivney et al. 2020; Schwinghammer et al. 2020). This method separates MP/NPs from sludge, sediments, and other inorganic materials that have not been degraded during pre-treatment/breakdown (chemical or enzymatic) processes (Hermsen et al. 2018). The density of most plastics is almost equal to water, and MP/NPs particles suspended in water tend to float, making their removal process easy (Enfrin et al. 2019). High-density solutions like sodium chloride (NaCl), sodium iodide (NaI), potassium formate (K(HCOO)), zinc chloride (ZnCl_2), sodium tungstate dehydrates and sodium polytungstate are used in the density separation of MP/NPs (Table 2). NaI is expensive and hazardous to the environment according to the GHS classification (Enfrin et al. 2019). NaCl is recommended for sediment samples due to its low cost and non-toxic nature. Due to their lower density, most plastics tend to float on the surface and can be reduced up to 80% (Li et al. 2021; Zhao et al. 2017). ZnCl_2 is one of the inexpensive methods with a good recovery rate (Li et al. 2021). K(HCOO), after utilization, can be filtered and reusable (Mahon et al. 2017). However, sodium polytungstate is relatively expensive, whereas sodium tungstate dihydrate is recommended in standardized protocols for sediments (McGivney et al. 2020; Schwinghammer et al. 2020). Canola oil was also used to separate MPs (fibres and fragments) with 92–97% recovery (Stock et al. 2019). The dual-density isolation mechanism for separating MP/NPs was also reported (Zhao et al. 2017). First, a higher-density medium is used to separate plastic particles

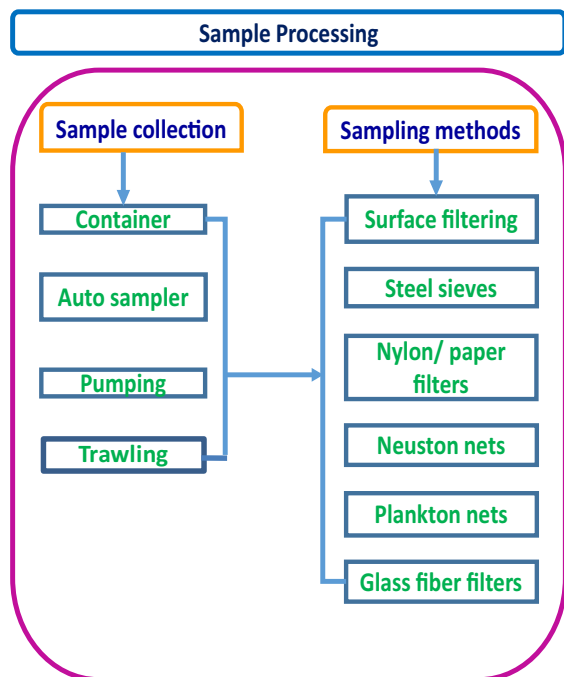


Fig. 4 Schematic representation of sample collection and processing routes of micro/nanoplastics in the environment

Table 2 Various processes involved in density separation and pre-treatment methods of micro/nanoplastics in the environment

	Type of Polymer	Solution	Density (g/cm ³)	Advantage	Disadvantage	References
Density Separation	PP, PE	Water	1	Simple; Easy to operate	Limited with high-density plastics and less recovery rates with more errors	Lv et al. (2021); Li et al. (2021); Stock et al. (2019); Enfrin et al. (2019); Sun et al. (2019); Munno et al. (2018); Lares et al. (2018); Zhao et al. (2017); Coppock et al. (2017); Mahon et al. (2017); Löder et al. (2017); Dehaut et al. (2016); Avio et al. (2017)
	Microplastics with a density below 1.2 g/cm ³ (eg: PA, PE, PP etc.)	NaCl	1.2	Economically feasible and Non-toxic, Easy available and eco-friendly	Limited with high-density plastics and less recovery rates with more errors	
	Nylon, PVC, PET	Sodium polytungstate	1.4	Non-toxic and Highly efficient to extract certain high-density plastics	Not specified	
	PVC, PS, PET, PE,PUR, PP	NaI	1.6	Recovery rates are very high > 99%	Cross contamination (Reacts with cellulose matter) Not eco-friendly and expensive	
	Almost all plastics	ZnCl ₂	1.7	Easy and applicable for almost all plastics	High economic inputs and Toxic	
	Microplastics with a density below 1.8 g/cm ³ (eg: PA, PE, PP etc.)	Potassium formate	1.9	More microplastics can be extracted	Not specified	
	Digestion	Reagents		Effect on polymer		
Pre-treatment	Oxidative	Fenton reagent; K ₂ S ₂ O ₈ ; 5% NaClO; 30–35% H ₂ O ₂		Decolouration of polyethylene, PET and polyester Degrades NY6 and NY66 and difficult in complete digestion of biomaterials at any temperature. Digestibility rates are < 95%		
	Acid	37% HCl; H ₂ SO ₄ ; 65% HNO ₃		Digestibility rates are > 95%. Higher concentrations of acids cause degradation of polymers and significant decolouration of polymers. Acids are Corrosive to NY6 and NY66 and melts PET as well		

Table 2 continued

Digestion	Reagents	Effect on polymer
Alkali	10% KOH; NaOH	Higher concentrations of NaOH causes partial destruction of the nylon fibers, melting of the polyethylene, and colour change of plastic particles. KOH at 40 °C is measured to be the destructive treatment of all polymers
Enzymatic	Proteinase K; Trypsin; Cellulose; Chitinase; Lipase rtc	Low concentration of trypsin, Proteinase K etc. has higher digestion. Enzymatic digestion efficiencies are 88% and does not effect on microplastics. The optimized concentrations can digest > 95% in the phytoplankton of seawater samples without damaging any MPs fragments

such as polyethylene (PE), polypropylene (PP), and polystyrene (PS) from heavier components (minerals). Second, adding calcium fluoride to a low-density solvent (pure methanol) enables selective sedimentation of plastic enabling effective recovery via centrifugation. These solvents don't harm the plastic polymers, except polypropylene, since the density is relative to solvent (Fries et al. 2013). The density separation is not ideal for larger WWTPs due to MP/NPs, and the procedure must be done in the static mode because rushing water would disperse the particles (Stock et al. 2019).

7.3 Pre-treatment

Wastewater in general consists of suspended matter, sediments, solid biomass, and organic material. Sediments and solid biomass can be easily separated by picking or filtration, but the complexity comes with suspended and organic matter. Heating of the sample at higher temperature damages MP/NPs due to degradation or mechanical friction (Sun et al. 2019). Prior digestion, samples need to be treated due to high concentrations of organic matter or inorganic solids materials. The biofilm formation on MPs with smaller size (0.063 mm) is an alternative to the digestion process and enables recovery to some extent. However, this method has many drawbacks like inefficiency for large-scale samples, cost-effectiveness, time-consuming, and limited recovery (Felsing et al. 2018; Rummel et al. 2017). Organic content in the waste samples is pre-treated/digested mostly by chemical digestion and enzymatic digestion.

7.3.1 Chemical

The chemical digestion methods can be classified into acid, alkali, and oxidation treatments. Acidic digestion was performed to reduce organic matter in the samples using H_2SO_4 , HNO_3 , and HCl (Li et al. 2021; Avio et al. 2017). A maximum of 98% degradation was noticed using HNO_3 compared to other acids. It has a disadvantage with polymers PS and PE since these particles get agglutinated during dissolution. HCl is less preferred due to its inefficiency in destroying bulk organic matter (Cole et al. 2014), while H_2SO_4 deform and degrade the plastic along with the organic compounds. Alkali digestion was carried out using strong base solutions like $NaOH$ or KOH . The use of 1 M $NaOH$ shows the efficiency of $\geq 90\%$ organic digestion (Cole et al. 2014). Further increase in molarity, polymers like PE, PET, PVC, and cellulose acetate start degrading into simple forms (Munno et al. 2018). Upon exposure for a longer time, $NaOH$ degrades the MPs, but KOH retains them from degradation except for cellulose acetate (Dehaut et al. 2016). Oxidizing digestion is one of the commonly used methods for digesting organic matter. $NaClO$ and H_2O_2 (30%) are efficient oxidizers (Li et al. 2021; Sun et al. 2019; Lares et al. 2018). Fenton reagent ($FeSO_4/H_2O_2$) is generally referred to as an advanced oxidizing agent used in digesting soil organic matter, compost, and large volumes of wastewater (Tagg et al. 2017; Masura et al. 2015). The reagent degrades MPs with less than 5 mm and facilitates biological samplings (gut or tissues) (Tagg et al. 2017). MPs can be recovered up to 70% when digested with 30% H_2O_2 for seven days (Liu et al. 2019). Further increase in H_2O_2 concentration,

deformation of plastics was observed. So, using a lower concentration of H₂O₂ and Fenton reagent is recommended with an exposure time of 12–24 h (Munno et al. 2018; Zhao et al. 2017). Recently, a metal-doped particle approach was adopted for NPs extraction. Pd-doped nanoplastic particles were spiked into ultrapure water with microwave-assisted acid digestion to assess the presence of NPs (Hildebrandt et al. 2020).

7.3.2 Enzymatic

Enzymatic degradation is the alternative method for chemical digestion for removing organic matter. It digests biological/aquatic tissues and organic matter in the waste samples. Enzymes such as cellulose, chitinase, protease, trypsin, papain, collagenase, and lipase were used in various studies (Sun et al. 2019; Courtene-Jones et al. 2017). Enzymatic digestion using protease showed >97% degradation (Cole et al. 2014). Löder et al. (2017) conducted experiments using a combination of enzymes. It modified the process and incubation conditions, where the whole degradation procedure of organic matter could not take more than one week. Utilizing enzymes for MP/NPs digestion is safe, but this process is limited to low quantity samples as it is an expensive process. This process does not apply to large-scale treatment as each enzyme requires its optimum pH for its functioning and digestion of samples (Stock et al. 2019; Löder et al. 2017).

7.4 Identification

Detection and predicting the formation of MP/NPs in wastewater is complex to perform (Zhao et al. 2017). Various analytical protocols to identify the size, quantity, and types of plastic particles have been used (Table 3). Sieve pattern and size create a primary cut-off for particles present in the samples (Li et al. 2021; Lv et al. 2019; Magnusson and Norén 2014). The simple method for identifying plastic matter is the ‘hot-point test’ where heated needle, when placed in the sample, plastic gets melted and masks over the needle, which will not permit identifying the nature and type of plastic (Hebner and Maurer-Jones 2020). Transmission electron microscopy (TEM), Field emission scanning electron microscopy (FE-SEM), dynamic light scattering (DLS), and multi-angle light

scattering (MALS) with pyrolysis detect particles of size (10 nm) and provide finite pictures/patterns. Energy-dispersive X-ray spectroscopy (EDX) coupled with FE-SEM reveals the elemental composition of MP/NPs (Li et al. 2021; Sun et al. 2019; Long et al. 2019; Corcoran et al. 2009). Fourier transform infrared spectroscopy (FTIR), focal plan array-based systems (FPA), micro-FTIR, and Raman spectroscopy are used to identify functional groups of polymers (Sun et al. 2019; Duemichen et al. 2015; Fries et al. 2013), while the structural architect of plastics can be identified using thermogravimetric analysis-gas chromatography-mass spectrometry (TGA-GC-MS), pyrolysis-gas chromatography-mass spectrometry (GC/MS) and thermogravimetric analysis (TGA) with thermal desorption-gas chromatography-mass spectrometry (TED-GC/MS) (Li et al. 2021; Lv et al. 2019; Duemichen et al. 2015). However, these techniques are sensitive to analyzing MP/NPs with impurities. Prior pre-treatment processes like wet peroxide oxidation, enzymatic digestion, and oxidizing agents facilitate the detection of MP/NPs. For nanoplastics, identification through these methods becomes more challenging when the sample size exceeds 500 mg, and the weight is less than 10 mg.

7.4.1 Microscopy

Identification of MPs using a microscope can be classified mainly by Optical/visual microscopy, Scanning electron microscope (SEM), and Transmission electron microscopy (TEM). The plastic particles were identified based on surface, and chemical structures as MPs do not shine and exhibit ductility/ malleability under physical stress (Zhao et al. 2017). MPs collected from various wastewater streams were passed through a filtered mesh and categorized into wires, needles, fibres, clusters, fragments, irregular partials, etc., with two or three-dimensional shapes (Hebner and Maurer-Jones 2020). However, visual observation of MPs is difficult due to the lower magnification factor of microscopes. MPs are size constrained and have a high error ratio (Uurasjärvi et al. 2020; Lv et al. 2019). Filtered MPs from the secondary wastewater treatment plants were observed under stereo-microscope, confirming bead-shaped MPs with 87% recovery (Dyachenko et al. 2017). Waste sludge samples free of organic matter observed fibres and particles when visualized under a digital optical microscope with

Table 3 Various methods of identification and characterization of micro/nanoplastics in environmental samples

Processes	Technique	Range/ Limits	Advantage	Drawbacks	References
Pre-concentration	Membrane filtration/ Ultra-filtration	> 10 nm 5–50 nm	Easy and cheap process, Process larger volumes No sample damage/ aggregation	Frequent interaction with membrane. Low flow rates with small pores and less volumes	Li et al. (2021); Schwinghammer et al. (2020); McGivney et al. (2020); Uurasjärvi et al. (2020); Lv et al. (2019); Schwaferts et al. (2019); Xu et al. (2019); Lv et al. (2019); Erni-Cassola et al. (2017); Ziajahromi et al. (2017); Shim et al. (2016); Fries et al. (2013)
	Dialysis	5–50 nm	Performed at mild condition	Slow process and requires large volume of counter dialyzing medium	
	Ultracentrifugation and Analytical ultracentrifugation	1 nm–1 µm	Simple and easy operation Can provide more information on MPs/NPs Very effective in separation	Operated at harsh conditions and no separation from particulate matrix Difficult to obtain complete separation	
Separation techniques	Solvent evaporation	All particulates	Easy and economic	Requires more energy Difficult to remove dissolved matter	
	Asymmetric Flow Field Fractionation	1 nm–1 µm	There is no stationary phase and done by online monitoring	Difficult in operation, Interaction with membrane and Steric inversion	
	Size Exclusion Chromatography	1–100 nm	Fixed with Coupled detectors	Operated at small range of samples and stationary phase	
	High Performance Liquid Chromatography	1–40 nm	Fixed with Coupled detectors	Operated at small range of samples and stationary phase	
	Capillary Electrophoresis	5–500 nm	Fast and high separation rates Fixed with Coupled detectors	External charge required, Electrolyte/surface modification Might damage sample by Interaction with capillary/clogging	

Table 3 continued

Processes	Technique	Range/ Limits	Advantage	Drawbacks	References
Characterization technique	Hydrodynamic Chromatography	5 nm– 1.2 μm	Minimal interaction with stationary phase	Application is very less	
	Dynamic Light Scattering (DLS) (Size (dh), PSD, aggregation behaviour)	1 nm–3 μm conc. 10^{-6} to 10^{-1}	Easy, fast and economic Process is In- situ, Non-invasive, Aggregation and direct coupling	Large particles, Polydispersity with complex matrix, Non-spherical particles	
	Electrophoretic Light Scattering (ELS) (Surface charge, stability)	1 nm–3 μm	Fast and cheap Non-invasive	Involves Electro-osmotic effect and Sensitive to environment	
	MALS (Size (dg), PSD)	10–1000 nm	Fixed with Coupled detectors	Prior sample preparation is done, requires neat and clear samples	
	Laser Diffraction (LD) (Size)	10 nm– 10 mm	Measures large size range Process is easy, fast and automated	Only spherical model	
	Nanoparticle Tracking Analysis (NTA) (Size (dh), PSD, number concentration)	30 nm–2 μm	Better with polydisperse samples, complex, media, Particle corona	Complexity in operation	
	FPA-FT-IR (Vibrational spectrum, Pigments, Additives, Ageing)	> 10 mm	Non-destructive and Automated process	Not applicable for single sub μ - and nanoplastic Easy interference from water takes place	
	ATR-FT-IR; AFM-IR (Spectrum, imaging)	> 50 nm	Advanced, Simple and fast performing method AFM-IR Spectrum, imaging > 50 nm, High resolution and Chemical imaging	Slow and covers small area	
	Raman Microspectroscopy (Fingerprint spectrum, Pigments, Additives)	> 0.5 mm, Bulk	Non-destructive, Easy sample preparation, Fast and no interference from water	Works on Fluorescence	
XPS (Binding energies of orbitals)	Bulk	Surface scanning and measurement	UHV and Laborious		

Table 3 continued

Processes	Technique	Range/ Limits	Advantage	Drawbacks	References
	Py-CG-MS (Mass Polymer type Additives)	Bulk Limit: ng-mg	Measurement with matrix, Fast, Higher sample masses can be done	Dependent on type of polymer, Dry sample needed Prior sample preparation is necessary	
Imaging studies	Optical Microscopy and Fluorescence Microscopy (Size, shape, morphology)	> 1 μm	Non-destructive, Cheap, easy to handle Operated at sub-diffraction variants	Limited Diffraction, Environmental plastic is not fluorescent	
	AFM (Size, shape, topography, aggregation)	> 0.1 nm	High resolution and AFM-IR TERS and liquid	High resolution, AFM-IR, TERS and done in liquid	
	STM; SEM (Size, shape, surface morphology, aggregation)	> 1 nm	High resolution imaging	Conductive samples, Slow and covers small area Prior sample preparation is involved and has charge effect	
	TEM (Size, shape, aggregation, imaging)	< 1 nm	High resolution imaging, Precise size information can be produced	Quantification is difficult Prior sample preparation is involved and expensive process	
	EDS (Elemental composition)	nm range	Complementary to SEM, measurement of all elements	Few Elemental information is not given	
	Confocal Laser Scanning Microscope (CLSM); Near-field Scanning Optical Microscopy (NSOM) (Size, shape, location in)	> 1 μm	Non-destructive Fluorescence imaging	Covers small area and Diffraction limit	
	Environmental scanning Electron Microscopy (ESEM) (Size, shape, imaging, surface morphology)	> 0.1 nm	Wet samples can be done at environmental conditions	Reduced in resolution	

magnification up to 1000X using built-in software (Lares et al. 2018). Particles with shapes such as round, flat and oval fibres have a twisted, layered, and

tapered nature. Hebner and Maurer-Jones (2020) conducted experiments to observe MPs photo-degradability in simulated moving water systems and noticed

cracks in polymer linear or structural alignments irrespective of their thickness upon exposure to UV light. Distinguishing the difference between transparent, pale-colored MPs and natural polymers (cellulose and fibrils) microscopy requires higher magnification (Dris et al. 2015; Magnusson and Norén 2014). Dyes such as Rose Bengal, Oil red EGN, Eosin B, Hostasol Yellow3G, fluorescein, safranin, isophosphate, and Nile Red are used for staining MP/NPs (Lv et al. 2019; Ziajahromi et al. 2017; Shim et al. 2016). Nile Red is used to identify MPs, i.e., polyethylene, polypropylene, polystyrene, polycarbonate, polyurethane, nylon particles, and poly (ethylene–vinyl acetate) with sizes ranging from 20 μm to 1 mm. Its fluorescence is stable for two months and is mostly used for marine and environmental pollutants (Lv et al. 2019; Ziajahromi et al. 2017). Rose-Bengal solution (4,5,6,7-tetrachloro-20,40,50,70-tetraiodo-fluorescein) is used to differentiate natural fibres and MPs since the stain gets adsorbed only by natural polymers and allows easy visual separation (Lv et al. 2019). Karakolis et al. 2019 carried out their experiments by staining more than ten fluorescent dyes on plastics, and excitation and emission colours were studied using fluorescence microscopy. Application of dyes could identify MPs in the micrometre to nano scale level (Karakolis et al. 2019).

Scanning electron microscopy (SEM) is used to characterize the surface topography of MPs. The pre-treated polymers samples were first coated with gold (Long et al. 2019; Corcoran et al. 2009). SEM analysis showed the dominant shapes like fragments, pellets, and fibres with average particle diameters of 11 μm from sediments and 130 μm from water suspensions, including polypropylene, polyethylene, polystyrene, and polyethylene terephthalate (Long et al. 2019). Transmission electron microscopy (TEM) effectively visualizes NPs for less amorphous particles; simplified sample preparation to electron microscopy and heavy-metal stains may be necessary (Hebner and Maurer-Jones 2020). By combining the Brownian motion properties, the degradation studies of NPs can be analyzed effectively (Hildebrandt et al. 2020).

7.4.2 Elemental composition

The elemental analysis gives the individual composition of elements, whereas CHNSO analysis predicts only carbon, hydrogen, nitrogen, sulfur, and oxygen

ratios present in the sample. However, detection of MPs could be difficult if sampling matter contains impurities or organic compounds and can be analyzed only by comparing with standard polymers (Li et al. 2021; Sun et al. 2019). Energy-dispersive X-ray spectroscopy (EDS) was used to assess the elemental composition of MPs based on diffraction and reflection of radiation emitted from MP/NPs surfaces (Sun et al. 2019; Fries et al. 2013; Dubaish and Liebezeit 2013). Fries et al. (2013) applied this technique to identify the inorganic plastic additives (IPAs) present in the samples. The elements such as aluminum, zinc, sulfur, barium, and titanium dioxide nanoparticles (TiO_2 -NPs) were detected in marine (Fries et al. 2013). TiO_2 -NPs are formed due to the degradation of polymer– TiO_2 composites in the marine environment.

7.4.3 Fourier transforms infrared (FTIR) spectroscopy

Fourier transforms infrared (FTIR) spectroscopy is used to detect MP/NPs chemical constituents/moieties (carbon-linked substituents) of MP/NPs in wastewater, sediments, and biota samples. It aids in the identification of plastic polymer particles by comparing their characteristic infrared spectra of carbon attached functional groups to reference spectral libraries, where each characteristic peak corresponds to chemical connections between atoms (Schwinghammer et al. 2020; McGivney et al. 2020; Zhang et al. 2016; Murphy et al. 2016). Cincinelli et al. 2017 identified rayon, polyamide, and semi-synthetic cellulose in gastrointestinal fish tracks using FTIR. Uurasjärvi et al. (2020) identified polyethylene, polypropylene, polymethyl methacrylate, polyvinylchloride, polyethylene terephthalate, and polystyrene in surface waters of lake samples. Lares et al. (2018) followed FTIR spectroscopic approach to confirm the presence of plastics and non-plastic organic particles (cellulose) in wastewater sludge. However, it is labor-intensive and needs sample preparation before identification. Attenuated total reflection FTIR spectroscopy (ATR) μ -FTIR can detect irregular shaped and scattered MPs effectively than FTIR (Hidalgo-Ruz et al. 2012). Hebner and Maurer-Jones. (2020) conducted their studies to assess the photo-oxidation of plastic fragments collected from the North Pacific Gyre sample using (ATR) μ -FTIR spectra and compared them against the spectral

library. Carbonyl content was seen at the maximum value of $1700\text{--}1800\text{ cm}^{-1}$. Chen et al. (2019) analyzed synthetic PVC MPs exposed to light and UV radiation using ATR-FTIR. PVC polymer has a characteristic absorption peak at $600\text{--}700\text{ cm}^{-1}$ region associated with C–Cl stretching vibration. Upon UV irradiation, C–Cl bonds at 613 and 711 cm^{-1} were decreased and replaced by C=C or C=O bonds resulting in dehydrochlorination of PVC. Similarly, Dyachenko et al. (2017) examined MPs from secondary wastewater treatment plant (WWTP) effluent using (ATR) FTIR spectrometer combined with continuum microscope in transmission mode. Polyoctadecyl methacrylate, amides, polyacrylonitrile, polyethylene, and non-plastic cellulosic cotton were detected compared with library references. Focal plane array (FPA) detector-based micro-FTIR imaging could detect MPs with sizes smaller than $20\text{ }\mu\text{m}$ (Schwinghammer et al. 2020; McGivney et al. 2020; Lv et al. 2019; Tagg et al. 2017). Mintenig et al. (2017) used FPA-based transmission micro-FTIR to identify MPs in wastewater and sludge samples, limiting fibre size ($10\text{--}20\text{ }\mu\text{m}$) and lateral resolution. Xu et al. (2019) collected 68 influent and 72 effluent samples from WWTPs and, upon analysis, found 112 plastics of 14 different types, which includes polyethylene (PE), polyamide (PA), polypropylene (PP), polystyrene (PS), polyethylene terephthalate (PET), rayon, polyvinyl chloride (PVC), poly methylmethacrylate (PMMA), rubber, polyethylene and polyether urethane (PU), polypropylene copolymer (PE-PP), acrylonitrile styrene copolymer (AS) and polyacrylate.

7.4.4 Raman spectroscopy

Raman spectroscopy analyzes MPs based on the inelastic scattering of light and delivers the evidence of chemical compounds in the form of molecular vibrations by a spectrum. Plastic particles of size less than $1\text{ }\mu\text{m}$ are detected with better response and reliability to other non-polar functional groups, and it is unaffected towards signals caused by H_2O and atmospheric CO_2 (Schwinghammer et al. 2020; McGivney et al. 2020; Li et al. 2018; Zhao et al. 2017; Hidalgo-Ruz et al. 2012). Raman spectroscopy linked with a microscope favors robust and non-presumptive identification of plastic components with various sizes (Lv et al. 2019). Raman spectroscopy

was performed with excitation at $455\text{--}532\text{ nm}$ on gold-coated polycarbonate filters or silicon filters (Kelkar et al. 2019; Schymanski et al. 2018). Several studies often used excitation at near-infrared (NIR, 785 nm) with laser intensity of $5\text{--}7\text{ mW}$ and integration times of $15\text{--}30\text{ s}$ in the wavelength range between 200 and 3000 cm^{-1} to identify MPs without deformation and compared with Raman library (Li et al. 2021; Cabernard et al. 2018; Zhao et al. 2017; Hidalgo-Ruz et al. 2012). Cole et al. (2014) suggested coherent anti-Stokes Raman scattering (CARS) microscopy based on confocal laser scanning microscopy (CLSM) and a synchronized dual-wavelength picosecond laser source. It can localize polymeric units in biological tissues with subcellular precision and gives spectral signals, but this process is limited in sensing the fluorescence samples. In such cases, samples free from fluorescence are recommended for Raman measurements (Sun et al. 2019; Li et al. 2021). Micro-Raman spectroscopy for High-density polyethylene (HDPE), Polypropylene, Polystyrene polymers before and after chlorine disinfection during sterilization and detected changes in Raman intensities by total loss of existing Raman peaks and emergence of new chemical bonds resulting aggressive effect of chlorine during water treatment (Kelkar et al. 2019). HDPE has prominent Raman peaks at 1064 cm^{-1} , 1130 cm^{-1} , 1295 cm^{-1} , and 1416 cm^{-1} . A shift was noted with increased chlorine dosage, and a new chlorine carbon bond peak at 678 cm^{-1} was detected. Polypropylene did not show any degradation effect on chlorination, and no new bonds formation was seen in Raman spectra. While in Polystyrene, new intense Raman peaks at 400 cm^{-1} , 445 cm^{-1} , and 348 cm^{-1} were observed due to the oxidative stress created by excessive chlorination. Long et al. (2019) performed micro-Raman spectroscopic analysis for WWTPs and reported the presence of plastics such as 21% polyethylene, 31.6% polypropylene, 10.1% polystyrene, 7.5% polyethylene terephthalate, and 9.2% propylene copolymer. Zada and co-workers used Stimulated Raman scattering (SRS) microscopy and worked on coherent interaction of two separate laser beams with vibrational levels to identify MPs. Different polymers such as polypropylene, high-density polyethylene, polyamide 6, 6, and polyethylene terephthalate are common in environmental or consumer products. (Li et al. 2021; Schwinghammer et al. 2020; McGivney et al. 2020; Zada et al. 2018). Zhao

et al. (2017) identified 11 chemical matrices including 6 polymers, 5 plastic additives, and pigments 100 to 500 nm in size from marine sediments using Raman microscopy.

7.4.5 Mass spectroscopy

Some of the commonly reported analyses for MP/NPs identification are liquid chromatography (LC), gas chromatography coupled with mass spectrometry (GC–MS), liquid chromatography–mass spectrometry (LC–MS), and Pyrolysis–gas chromatography–mass spectrometry GC–MS (Py–GC/MS) (Sun et al. 2019; Nguyen et al. 2019). Among them, the Pyrolysis GC/MS technique gained much attention in recent years as GC–MS, LC–MS analytical methods consist of multiple steps such as solvent extraction, filtration, and sample preparation before chromatographic injection, which are time-consuming processes (Li et al. 2021; Sun et al. 2019; Lv et al. 2019). In the Pyrolysis GC/MS technique, samples were analyzed by injecting MP/NPs directly into the pyrolyzer, preventing contamination during sample preparation by improved analytical efficiency (Nguyen et al. 2019; Schwinghammer et al. 2020; McGivney et al. 2020). Hermabessiere et al. (2018) worked extensively on Py–GC/MS method and determined the limit of detection (LOD) values for eight commercial plastics, i.e., polyethylene (PE), polystyrene (PS), polypropylene (PP), polycaprolactam (PA-6), polyethylene terephthalate (PET), polycarbonate (PC), poly(methyl methacrylate) (PMMA), and unplasticized polyvinyl chloride (uPVC). They developed LOD standards ranging from 1 µg and applied them to environmental samples from beach sediments, surface seawater, and organisms. The average of all obtained masses was calculated by using intensity (h), time (t), and response factor (R) (Hermabessiere et al. 2018).

Response factor (R) = Weight/Height

Limit of detection (LOD) = $3 \times R \times H_{20FWHM}$

Fries et al. (2013) conducted experiments on MPs and bound additives in the marine environment using Py–GC/MS. Pyrolysis results selected from standard polymers showed the presence of polystyrene, polypropylene, polyethylene, polyamide, chlorinated and chlorosulfonated polyethylene, diethyl phthalate, diethylhexyl phthalate, 2,4-di-tert-butylphenol, etc.

from the analyzed samples. However, environmental samples contain a mixture of complex structured molecules where the difficulty arises in determining the exact polymer upon pyrolysis. Duemichen et al. (2015) used thermogravimetric analysis (TGA) with thermal desorption–gas chromatography–mass spectrometry (TED–GC/MS) for handling the complex structured environmental samples of soil suspended solids of wastewater and identified mostly PE along with PS, PET, PS, and polyamide 6. NPs with lower concentrations can be analyzed by using Thermal desorption coupled with gas chromatography–mass spectrometry (TDS–GC–MS) with greater sensitivity of masses less than ~50 µg. This may be applicable for simple matrices like drinking water, where separation is straightforward (Nguyen et al. 2019).

7.4.6 Differential scanning calorimetry (DSC)

Differential scanning calorimetry analysis is used to evaluate polymeric MP/NPs by glass transition temperatures (T_g) and depicts chemical structures upon exposure to higher temperatures. Kelkar et al. (2019) applied DSC for physical changes and conversion of virgin plastics to other forms while sterilizing drinking water and wastewater disinfection using different chlorine concentrations. Initial analyses were performed with high-density polyethylene and polypropylene standards, and their respective glass transition temperatures (T_g) were noted. Concerning the traditional values, experimental samples were analyzed, and types of polymers present were identified with heating from ambient temperature to 300 °C at a heating rate of 5 °C/min and cooled to –80 °C and enthalpy changes were noted. The DSC analysis upon plastic particles exposure to extreme chlorination conditions resulted in a substantial change, denoted by the difference in melting points and characters of stressed plastics. Likewise, Hebner and Maurer-Jones (2020) calculated DSC exotherms heating and cooling cycles of plastics in moving water. The polymer samples identified have an increased crystallinity, which might be due to the photodegradation of polymers. Plastic polymers melting at low temperatures or glass transition temperature (T_g) show that these plastics having lower molecular weight polymers were attributed to polypropylene and polyethylene samples.

8 Interaction energy calculation by DLVO theory and kinetic model

MP/NPs generation and interactions are necessary to assess its surface chemistry, size and shape in WWTPs processes to predict the existence of plastic nature, which is either single or agglomerate state particles in water (Andrady 2017; Li et al. 2019). Derjaguin, Landau, Verwey/Overbeek's (DLVO) theory is used to study the surface energy interactions between suspended solids-polystyrene nanoplastics (PSNPs) and suspended solids-polyethylene MPs (PMPs) particles by applying particle-plate geometry in DLVO calculations (Enfrin et al. 2019). Using theoretical calculations, the interaction energies were treated as particle-particle geometry for PSNPs-PSNPs and plate-plate geometry for PEMP-PEMP. Enfrin et al. (2019) explained homogeneous and heterogeneous interactions between the nano/microplastics in water. In homogenous interaction, the surface interaction energies of colloidal MP/NPs have less than 1 μm in water as per DLVO theory. From the interparticle distances, the electrostatic repulsion and van der Waals attraction energy between the particles results in agglomeration/aggregation rate in the sample. The potential energy between interparticle distance and kinetic energy due to motion forms a huge effect on dense aggregation/agglomeration and other parameters such as surface area, charge, and ionic strength of sample suspension. In heterogeneous interaction, the aggregation occurs between NPs/MPs and organic matter or microorganisms. Zhao et al. (2017) studied the marine organic aggregate encapsulate MPs like PP, PS, and PE resulting in the formation of agglomeration. Here, the secretions of microbial extracellular polysaccharides form an interfacial bonding between the plastic particles and the chemicals in the water create the surface interactions between plastics, resulting in agglomeration. As a result, the dispersion pattern can be improved by inserting nucleophile chloro-halides groups to boost negative charge surface density on MP/NPs. Li et al. (2018) looked at how different monovalent and divalent electrolytic solutions affected the aggregation behavior of polystyrene MPs. The aggregation kinetics were achieved by DLVO theory. They have also studied critical coagulation concentration for the aggregation and interaction energies between polystyrene MPs using a combination of theoretical

calculations and experimental measurements that could be useful for assessing MP/NPs fate in aquatic environments.

9 Removal of microplastics in WWTPs

Wastewater treatment plants (WWTPs) interface natural and mechanical systems. Large quantities of particulate plastic resulting from industrial, domestic, and surface runoff were treated before release into the environment (Frehland et al. 2020; Hu et al. 2019). MPs, unlike suspended solids in standard WWTPs, necessitate the use of settling and filtration equipment to remove MPs (Iyare et al. 2020). Three distinct WWTPs in New York recorded daily particle discharges of 109, 556, 81, 911 particles (Blair et al. 2017). The WWTPs may not completely remove MP/NPs, but treatment processes involving degradation, generation, and advanced treatment sources should be considered. Nevertheless, MP/NPs concentration downstream is higher than those upstream, suggesting the enormous release of particulate plastic into the natural system is conceivable. The removal of MP/NPs by the WWTPs can be observed from primary, secondary, and tertiary treatments.

9.1 Primary treatment

Primary treatment removes suspended solids and most of the MP/NPs in wastewater. It was evident that preliminary (mechanical) treatment could remove 35–59% of the microplastics, while primary treatment could remove 23–53% (Hale et al. 2020). Pre-treatment was able to remove bigger microplastics effectively. Rough grille bars (16–25 mm) and fine grille bars (3–10 mm) used in WWTPs independent MPs of smaller size (Raju et al. 2018; Michielssen et al. 2016). According to Dris et al. (2015), the fraction of massive particles (1000–5000 μm) reduced from 45 to 7% after the preliminary treatment. Surface skimming on primary clarifiers removed light floating MPs, whereas gravity parting removed MPs bound in solid flocs (Simon et al. 2019). Microbeads made of PE were effectively removed by skimming due to their buoyancy and floating on the top of water, grease, and oil particles (Murphy et al. 2016). These findings were consistent with Michielssen et al. (2016) and Sutton et al. (2016), where a considerable decrease of

microbeads was noticed in the outflow of the WWTPs after primary treatment. In terms of MPs morphologies, pre-treatment may be more effective at removing fibres from wastewater than pieces (Ziajahromi et al. 2017). This is most likely because fibres are abundantly captured in flocs and sedimentation (Talvitie et al. 2015; Magnusson and Norén 2014).

9.2 Secondary treatment

Secondary treatment typically includes biological treatment/clarification, which reduces MPs in wastewater by 0.2% to 14% (Sun et al. 2019; Besseling et al. 2017). Plastic debris production in the secondary clarifying tank is likely to be aided by sludge flocs or bacterial extracellular polymers in the aeration tank (Carr et al. 2016). Microplastics have an impact on microbial-mediated processes that affect ammonium synthesis (ammoniation) and reduction, as well as inorganic nitrogen biological conversion efficiency (nitrification and denitrification) (Zhang and Chen 2020). Furthermore, the concentrations of microplastics are positively linked with BOD, DO, and total phosphorous. Chemicals (ferric sulfate) and flocculating agents employed during secondary treatment may help to remove MPs by causing suspended particulate matter to combine and form a “floc” (Murphy et al. 2016). Due to protozoa or metazoan feeding, MPs might be trapped in sludge flocs (Scherer et al. 2018; Jeong et al. 2016). Secondly, prolonged interaction of MPs with wastewater aids secondary discharges and removes MP/NPs escaped from skimming/settling practices. These bio-coatings may operate as wetting agents, altering micro/nano plastics surface characteristics and relative densities (Rummel et al. 2017; Fazey and Ryan 2016). Large MPs particles can be removed even more thoroughly during subsequent treatment, resulting in a low abundance. The secondary treatment eliminates additional fragments than fibres due to their abundance and neutral buoyancy (Ziajahromi et al. 2017; Talvitie et al. 2015, 2017).

9.3 Tertiary treatment

The tertiary treatment provides further polishing to wastewater and removes microplastics by 0.2–2% relative to the influent (Sun et al. 2019). In this stage, MP/NPs suspended in the wastewater hinder coagulation, air flotation, microfiltration and affects the

disinfection process (chlorine and UV disinfection) (Zhang and Chen 2020). Membrane-related technologies such as disc-filter (DF), rapid sand filtration (RSF), dissolved air flotation (DAF), and membrane bioreactor (MBR) are utilized in the treatment of tertiary effluents (Mintenig et al. 2017). The performance of several membranes in removing MP/NPs found that MBR showed relatively higher removal efficacy (99.9%) trailed by RSF (97%) and DAF (96%) (Talvitie et al. 2017). The exclusion of DF varied from 40 to 98.5%. Similar studies were conducted by Schneiderman (2015) in WWTPs and found an absence of microbeads with membrane filters while other advanced filters did. MPs concentrations were considerably lower following ultrafiltration and reverse osmosis (Ziajahromi et al. 2017). In some cases, fibres with fractions (20–190 mm) were found to be abundant in the final effluent of tertiary treatment (Ziajahromi et al. 2017). This is most likely since fibres can readily pass a filter or membrane longitudinally. MPs were reported to be removed from WWTPs in 98–99.9% of cases (Simon et al. 2018; Lares et al. 2018; Carr et al. 2016).

10 Strategies to reduce MP/NPs

Most interventions to prevent plastic leakage are related to removing MP/NPs from wastewater infrastructure (Pico et al. 2019). One of the effective solutions is to eliminate MP/NPs at the source itself. Improved solid waste infrastructure and management will reduce plastic debris dispersion into environmental matrices. The “GoJelly Project,” which employs jellyfish mucus to trap microplastics, is one example of microplastic prevention technology (Schmaltz et al. 2020). The “Cora Ball” and “Fibre Free” are laundry balls used to capture synthetic fibres in the washing machine. At the same time, the “Lint LUV-R” and “Showerloop” are household water systems that filter out microfibers (Schmaltz et al. 2020). Ban on the usage of microbeads alone prevents >2.9 trillion pieces of MPs from entering water bodies per year (Prata et al. 2019). Ban or imposing taxes on non-biodegradable plastics, single-use plastic cutlery, packaging materials, and labeling have been passed recently in various global jurisdictions (Karasik et al. 2020). Bio-based polymers and biodegradable plastics derived from renewable starting materials (starch,

cellulose, lignin, etc.) are mostly biodegradable (Amulya et al. 2021; Kopperi et al. 2021). Bio-based polyethylene terephthalate (bio-PET), Bio-based polyethylene (bio-PE); Polyethylene furanoate (PEF), Polyhydroxyalkanoates (PHA), Poly(butylene succinate) (PBS), and Polycaprolactone (PCL) accounts for 40% of the global bioplastics used (Hahladakis et al. 2020; Prata et al. 2019). Swallowing biodegradable MP/NPs also affects health and the environment (Sadeghi et al. 2021).

Mitigation can be achieved through legislation (regulations on the management of plastic waste and products in various industries), technical (bio-innovations including bioplastics), and social awareness (reducing the use of single-use plastics or disposable plastics and adopting recycling habits) (Allouzi et al. 2021). The United States put forth a microbead ban in 2015 (H.R.1321 Microbead—Free Waters Act of 2015), and the European Union launched the single-use plastics Directive and proposed a ban on intentionally added MPs (EU 2018, Directive (EU) 2019/904) (Catarino et al. 2021). Between 2000 and 2019, near to 28 policies were come into force established on reducing and managing plastic pollution around the globe (Karasik et al. 2020). The Antarctic Treaty, London Convention and Protocol amendments, G20 Action Plan on Marine litter, International Convention for the Prevention of Pollution from Ships (MARPOL) Annex V, and the United Nations Environment Assembly (UNEA) on marine plastic pollution (Karasik et al. 2020). As of 2019, 39 regional policy documents have been adopted in Europe (Schmaltz et al. 2020). In India, the 'Plastic Waste Management Rules, 2016' was implemented to remove all disposable plastic products by 2022 and prohibit the production of plastics below 50 μm (Ministry of Environment and Forest, India 2016; Veerasingam et al. 2020). Malaysia has implemented a "No Plastic Bag" campaign by imposing a minimum charge per plastic bag (Allouzi et al. 2021).

10.1 Circular economy (CE) approach

Recycling techniques cannot be a comprehensive solution to plastic reduction. Operational issues, infrastructure for collection and sorting, and lack of green methods are inherent limitations of recycling (Bishop et al. 2020). Circular Economy (CE) may be a viable option for reducing the harmful impacts of

mass-produced plastics, rising consumption, and the usage of plastic items for short periods (Sandhiya and Ramakrishna, 2020). The end of the product life is one of the most significant contrasts between linear and circular economies. In the linear economics model, the management of a "post-use" product takes the form of dumping, whereas, in the CE model, end-of-life is determined as the product's "first life" (Sadeghi et al. 2021). CE focuses on repair and reconstruction with the aim of continuous use. Managing a finite supply of natural resources and renewing natural systems, closing loops, and waste design are the essential principles of CE (Chae and Hinestroza 2020; Venkata Mohan et al. 2019). Economic considerations are impending for relooping low-value plastic products in the consumption cycle (Fadeeva and Van Berkel 2021). Retailers are crucial in establishing a consumption loop by establishing a viable link between producers and consumers (Fadeeva and Van Berkel 2021). Therefore, manufacturers should create products that cater to consumer preferences while using few virgin resources, reducing toxicity, and increasing recyclability. Adapting technology change in process control, equipment modification, onsite reuse, and recycling helps achieve production loss and curtailing plastic waste at the point source (AliAkbari et al. 2021; Venkata Mohan and Katakojwala, 2021). Mechanical, physical, chemical, biological, and energy recovery are the main recycling technologies employed for recovery (Shamsuyeva and Endres 2021). Recycling can be achieved through the "close-loop" and "open-loop" approaches. In closed-loop, plastic's inherent properties stay unchanged, allowing it to be reused as a primary material (bottle-to-bottle recycling), whereas recycle is used to produce new plastic products (bottle-to-fiber recycling) in an open-loop system (Shamsuyeva and Endres 2021). The recycling technologies are well established for managing single-polymer plastic waste (PET, PE, PP, PS, etc.) and mixed plastic waste (Hahladakis et al. 2020). Recycling rates for PET exceed 10%, while PS and PP are not yet started (AliAkbari et al. 2021). Recycling plastics will reduce GHG emissions from 8 GtCO₂e to 4.9 GtCO₂e by 2050 (Zheng and Suh 2019). From an environmental standpoint, the use of microbial polymers and plant-based fibres should be encouraged. The use of biodegradable materials, sustainable management of landfills, designing efficient recycling methods, providing support, incentive laws, and creating

awareness among stakeholders can help gain traction in adopting CE (Sadeghi et al. 2021). Reprocessing and recycling also create jobs and save energy while downing resource consumption and waste (Chae and Hinestroza 2020).

11 Conclusion and future perspectives

Misuse and mismanagement of plastics have led to the accumulation of MP/NPs in the environment, posing a risk to the ecosystem and living beings. They are ubiquitous and released into different environmental compartments (terrestrial, aquatic, and atmospheric depositions) via rivers, lakes, stormwater runoff, sewage, sludge, and wastewater treatment plants. The fate and transport of MP/NPs strongly depend on the physicochemical properties of the plastics in soil and water. MP/NPs accumulation in soil affects the soil productivity, microorganism fecundity and inhibits plant growth. The trophic transfer of MP/NPs to human beings leads to cancer, digestive problems, and cardiovascular diseases. Research priority can be given to MP/NPs transport and accumulation in soil microbiome, crop plants, and trophic transfer of plastic-derived contaminants (additives and chemicals) in the food web. In MP/NPs analysis analytical front, developments in simple and precise tools, validations, and implications in detection, characterization, and quantification still need focus. The traditional use of Raman and FT-IR is already reinforced by the emergence of some promising techniques such as GC–MS performed with pyrolysis or TDS and especially new hyperspectral imaging techniques. Assessing and quantifying MP/NPs in the soil-microbiome-plant system still needs interventions in sample preparation, pre-treatment, and pre-concentration to meet the detection limit of the analytical instruments. In the context of NPs, sampling, pre-treatment, and characterization still lack standardization and homogenization of the analyte methods. In addition, it is necessary to establish matching libraries to identify MP/NPs from environmental samples. Particulate plastics from industrial, domestic, and surface runoff are entering wastewater treatment plants (WWTPs) before releasing into the environment. Increasing loads of MP/NPs in sludge digestion system reduces the effectiveness of process and raise operational expenses. Remediation technologies,

prioritizing recycling, sustainable waste management, education and awareness, infusing circular models (reduce, reuse, recycle and recover), influxing bio-based materials (bioplastics), legislation, policy, and road map are some of the essential prerequisites that may be critically considered for the establishment of sustainable practices in plastic management domain.

Acknowledgements The authors would like to thank CSIR-IICT for its support (IICT Manuscript No. IICT/Pubs./2021/212).

Author contributions SVM, BRK and HK Conceptualization, SVM, BRK and HK Writing-original draft preparation, SVM Supervision, SVM Funding acquisition. Authors have read and agreed to the published version of the manuscript.

Declarations

Conflict of interest The authors declare no conflict of interest.

References

- Abbasi S, Keshavarzi B, Moore F, Turner A, Kelly FJ, Dominguez AO, Jaafarzadeh N (2019) Distribution and potential health impacts of microplastics and microrubbers in air and street dusts from Asaluyeh County. *Iran Environ Pollut* 244:153–164
- Alam FC, Sembiring E, Muntalif BS, Suendo V (2019) Microplastic distribution in surface water and sediment river around slum and industrial area (case study: Ciwalgke River, Majalaya district, Indonesia). *Chemosphere* 224:637–645
- AliAkbari R, Mohammad HG, Nariman N, Elaheh K, Seeram R, Mehdi M, Yousef M (2021) High value add bio-based low-carbon materials: conversion processes and circular economy. *J Cleaner Prod*, p 126101
- Allen S, Allen D, Moss K, Le Roux G, Phoenix VR, Sonke JE (2020) Examination of the ocean as a source for atmospheric microplastics. *PloS one*, 15(5), p e0232746
- Allouzi MMA, Tang DYY, Chew KW, Rinklebe J, Bolan N, Allouzi SMA, Show PL (2021) Micro (nano) plastic pollution: The ecological influence on soil-plant system and human health. *Sci Total Environ*, p 147815
- Amereh F, Babaei M, Eslami A, Fazelipour S, Rafiee M (2020) The emerging risk of exposure to nano (micro) plastics on endocrine disturbance and reproductive toxicity: From a hypothetical scenario to a global public health challenge. *Environ Pollut* 261:114158
- Amulya K, Katakajwala R, Ramakrishna S, Venkata Mohan S (2021) Low carbon biodegradable polymer matrices for sustainable future. *Composites Part C: Open Access*, p 100111
- Andrady AL (2017) The plastic in microplastics: A review. *Mar Pollut Bull* 119(1):12–22

- Annenkov VV, Danilovtseva EN, Zelinskiy SN, Pal'shin VA (2021) Submicro-and nanoplastics: how much can be expected in water bodies? *Environ Pollut* 278:116910
- Auta HS, Emenike CU, Fauziah SH (2017) Distribution and importance of microplastics in the marine environment: a review of the sources, fate, effects, and potential solutions. *Environ Int* 102:165–176
- Avio CG, Cardelli LR, Gorbi S, Pellegrini D, Regoli F (2017) Microplastics pollution after the removal of the Costa Concordia wreck: first evidences from a biomonitoring case study. *Environ Pollut* 227:207–214
- Azizi SMM, Hai FI, Lu W, Al-Mamun A, Dhar BR (2021) A review of mechanisms underlying the impacts of (nano) microplastics on anaerobic digestion. *Bioresour Technol*, p 124894
- Barboza LGA, Vieira LR, Branco V, Carvalho C, Guilhermino L (2018) Microplastics increase mercury bioconcentration in gills and bioaccumulation in the liver, and cause oxidative stress and damage in *Dicentrarchus labrax* juveniles. *Sci Rep* 8(1):1–9
- Bergmann M, Mützel S, Primpke S, Tekman MB, Trachsel J, Gerdts G (2019) White and wonderful? Microplastics prevail in snow from the Alps to the Arctic. *Sci Adv* 5(8):eaax1157
- Besseling E, Quik JT, Sun M, Koelmans AA (2017) Fate of nano-and microplastic in freshwater systems: A modeling study. *Environ Pollut* 220:540–548
- Bianco A, Passananti M (2020) Atmospheric micro and nanoplastics: an enormous microscopic problem. *Sustainability* 12(18):7327
- Birch QT, Potter PM, Pinto PX, Dionysiou DD, Al-Abed SR (2020) Sources, transport, measurement and impact of nano and microplastics in urban watersheds. *Rev Environ Sci Biotechnol* 19(2):275–336
- Bishop G, Styles D, Lens PNL (2020) Recycling of European plastic is a pathway for plastic debris in the ocean. *Environ Int* 142:105893
- Blair RM, Waldron S, Phoenix V, Gauchotte-Lindsay C (2017) Micro-and nanoplastic pollution of freshwater and wastewater treatment systems. *Springer Sci Rev* 5(1):19–30
- Bläsing M, Amelung W (2018) Plastics in soil: analytical methods and possible sources. *Sci Total Environ* 612:422–435
- Boots B, Russell CW, Green DS (2019) Effects of microplastics in soil ecosystems: above and below ground. *Environ Sci Technol* 53(19):11496–11506
- Botterell ZL, Beaumont N, Dorrington T, Steinke M, Thompson RC, Lindeque PK (2019) Bioavailability and effects of microplastics on marine zooplankton: a review. *Environ Pollut* 245:98–110
- Boucher J, Friot D (2017) Primary microplastics in the oceans: a global evaluation of sources (pp 227–229). Gland, Switzerland: Iucn
- Brahney J, Mahowald N, Prank M, Cornwell G, Klimont Z, Matsui H, Prather KA (2021) Constraining the atmospheric limb of the plastic cycle. *Proc Natl Acad Sci* 118(16)
- Brand JH, Spencer KL, O'shea FT, Lindsay JE (2018) Potential pollution risks of historic landfills on low-lying coasts and estuaries. *Wiley Interdiscip. Rev*, 5(1), p e1264
- Cabernard L, Roscher L, Lorenz C, Gerdts G, Primpke S (2018) Comparison of Raman and Fourier transform infrared spectroscopy for the quantification of microplastics in the aquatic environment. *Environ Sci Technol* 52(22):13279–13288
- Cai L, Wang J, Peng J, Tan Z, Zhan Z, Tan X, Chen Q (2017) Characteristic of microplastics in the atmospheric fallout from Dongguan city, China: preliminary research and first evidence. *Environ Sci Pollut R* 24(32):24928–24935
- Campanale C, Savino I, Pojar I, Massarelli C, Uricchio VF (2020) A Practical overview of methodologies for sampling and analysis of microplastics in riverine environments. *Sustainability* 12(17):6755
- Carpenter E, Smith K (1972) Plastics on the sargasso sea surface. *Science* 175:1240–1241. <https://doi.org/10.1126/science.175.4027.1240>
- Carr SA, Liu J, Tesoro AG (2016) Transport and fate of microplastic particles in wastewater treatment plants. *Water Res* 91:174–182
- Catarino AI, Kramm J, Völker C, Henry TB, Everaert G (2021) Risk posed by microplastics: Scientific evidence and public perception. *Curr. Opin. Green Sustain. Chem.*, p 100467
- Ceccarini A, Corti A, Erba F, Modugno F, La Nasa J, Bianchi S, Castelvetro V (2018) The hidden microplastics: new insights and figures from the thorough separation and characterization of microplastics and of their degradation byproducts in coastal sediments. *Environ Sci Technol* 52(10):5634–5643
- Chae Y, An YJ (2017) Effects of micro-and nanoplastics on aquatic ecosystems: Current research trends and perspectives. *Mar Pollut Bull* 124(2):624–632
- Chae Y, Hinestroza J (2020) Building circular economy for smart textiles, smart clothing, and future wearables. *Mater Circ Econ* 2(1):1–4
- Chang X, Xue Y, Li J, Zou L, Tang M (2020) Potential health impact of environmental micro and nanoplastics pollution. *J Appl Toxicol* 40(1):4–15
- Chen C, Chen L, Yao Y, Artigas F, Huang Q, Zhang W (2019) Organotin Release from Polyvinyl Chloride Microplastics and Concurrent Photodegradation in Water: Impacts from Salinity, Dissolved Organic Matter, and Light Exposure. *Environ Sci Technol* 53(18):10741–10752
- Chen X, Chen X, Zhao Y, Zhou H, Xiong X, Wu C (2020) Effects of microplastic biofilms on nutrient cycling in simulated freshwater systems. *Sci Total Environ*, 719, p 137276
- Cheung PK, Fok L (2017) Characterisation of plastic microbeads in facial scrubs and their estimated emissions in Mainland China. *Water Res* 122:53–61
- Cincinelli A, Scopetani C, Chelazzi D, Lombardini E, Martellini T, Katsoyiannis A, Fossi MC, Corsolini S (2017) Microplastic in the surface waters of the Ross Sea (Antarctica): occurrence, distribution and characterization by FTIR. *Chemosphere* 175:391–400
- Cole M, Webb H, Lindeque PK, Fileman ES, Halsband C, Galloway TS (2014) Isolation of microplastics in biota-rich seawater samples and marine organisms. *Sci Rep* 4:4528
- Conti GO, Ferrante M, Banni M, Favara C, Nicolosi I, Cristaldi A, Fiore M, Zuccarello P (2020) Micro-and nano-plastics in edible fruit and vegetables. The first diet risks assessment for the general population. *Environ Res* 187:109677

- Corcoran PL, Biesinger MC, Grifi M (2009) Plastics and beaches: A degrading relationship. *Mar Pollut Bull* 58:80–84
- Courteney-Jones W, Quinn B, Murphy F, Gary SF, Narayanaswamy BE (2017) Optimisation of enzymatic digestion and validation of specimen preservation methods for the analysis of ingested microplastics. *Anal Methods* 9(9):1437–1445
- Cox KD, Covernton GA, Davies HL, Dower JF, Juanes F, Dudas SE (2019) Human consumption of microplastics. *Environ Sci Technol* 53(12):7068–7074
- Cózar A, Echevarría F, González-Gordillo JJ, Irigoien X, Úbeda B, Hernández-León S, Palma ÁT, Navarro S, García-de-Lomas J, Ruiz A, Fernández-de-Puelles ML (2014) Plastic debris in the open ocean. *Proc Natl Acad Sci* 111(28):10239–10244
- De Souza MAA, Lau CW, Kloas W, Bergmann J, Bachelier JB, Faltin E, Becker R, Görlich AS, Rillig MC (2019) Microplastics can change soil properties and affect plant performance. *Environ Sci Technol* 53(10):6044–6052
- Dehaut A, Cassone AL, Frère L, Hermabessiere L, Himber C, Rinnert E, Rivière G, Lambert C, Soudant P, Huvet A, Duflou G (2016) Microplastics in seafood: Benchmark protocol for their extraction and characterization. *Environ Poll* 215:223–233
- Deng Y, Zhang Y, Lemos B, Ren H (2017) Tissue accumulation of microplastics in mice and biomarker responses suggest widespread health risks of exposure. *Sci Rep* 7:46687
- Dobslaw D, Woiski C, Kiel M, Kuch B, Breuer J (2020) Plant uptake, translocation and metabolism of PBDEs in plants of food and feed industry: a review. *Rev Environ Sci Biotechnol*, pp 1–68
- do Sul JAI, Costa MF, Silva-Cavalcanti JS, Araújo MCB (2014) Plastic debris retention and exportation by a mangrove forest patch. *Mar Pollut Bull* 78(1–2):252–257
- Dris R, Gasperi J, Rocher V, Saad M, Renault N, Tassin B (2015) Microplastic contamination in an urban area: a case study in Greater Paris. *Environ Chem Lett* 12(5):592–599
- Duan J, Han J, Zhou H, Lau YL, An W, Wei P, Cheung SG, Yang Y, Tam NFY (2020) Development of a digestion method for determining microplastic pollution in vegetal-rich clayey mangrove sediments. *Sci Total Environ* 707:136030
- Dubaish F, Liebezeit G (2013) Suspended microplastics and black carbon particles in the Jade system, Southern North Sea. *Water Air Soil Pollut* 224(2):1–8
- Duemichen E, Braun U, Sturm H, Kraemer R, Deglmann P, Gaan S, Senz R (2015) A new molecular understanding of the thermal degradation of PA 66 doped with metal oxides: experiment and computation. *Polym Degrad Stab* 120:340–356
- Dyachenko A, Mitchell J, Arsem N (2017) Extraction and identification of microplastic particles from secondary wastewater treatment plant (WWTP) effluent. *Anal Methods* 9(9):1412–1418
- Enders K, Lenz R, Stedmon CA, Nielsen TG (2015) Abundance, size and polymer composition of marine microplastics $\geq 10 \mu\text{m}$ in the Atlantic Ocean and their modelled vertical distribution. *Mar Pollut Bull* 100(1):70–81
- Enfrin M, Dumée LF, Lee J (2019) Nano/microplastics in water and wastewater treatment processes—Origin, impact and potential solutions. *Water Res* 161:621–638
- Engwa GA, Ferdinand PU, Nwalo FN, Unachukwu MN (2019) Mechanism and health effects of heavy metal toxicity in humans. Poisoning in the modern world—new tricks for an old dog? Ozgur Karcioğlu and Banu Arslan, IntechOpen. <https://doi.org/10.5772/intechopen.82511>
- Evangelio N, Grythe H, Klimont Z, Heyes C, Eckhardt S, Lopez-Aparicio S, Stohl A (2020) Atmospheric transport is a major pathway of microplastics to remote regions. *Nat Commun* 11(1):1–11
- Fadeeva Z, Van Berkel R (2021) Unlocking circular economy for prevention of marine plastic pollution: an exploration of G20 policy and initiatives. *J Environ Manage* 277:111457
- Fazey FM, Ryan PG (2016) Biofouling on buoyant marine plastics: An experimental study into the effect of size on surface longevity. *Environ Poll* 210:354–360
- Felsing S, Kochleus C, Buchinger S, Brennholt N, Stock F, Reifferscheid G (2018) A new approach in separating microplastics from environmental samples based on their electrostatic behavior. *Environ Poll* 234:20–28
- Feng LJ, Wang JJ, Liu SC, Sun XD, Yuan XZ, Wang SG (2018) Role of extracellular polymeric substances in the acute inhibition of activated sludge by polystyrene nanoparticles. *Environ Pollut* 238:859–865
- Fischer EK, Paglialonga L, Czech E, Tamminga M (2016) Microplastic pollution in lakes and lake shoreline sediments—a case study on Lake Bolsena and Lake Chiusi (central Italy). *Environ Poll* 213:648–657
- Fossi MC, Coppola D, Baini M, Giannetti M, Guerranti C, Marsili L, Panti C, de Sabata E, Clò S (2014) Large filter feeding marine organisms as indicators of microplastic in the pelagic environment: the case studies of the Mediterranean basking shark (*Cetorhinus maximus*) and fin whale (*Balaenoptera physalus*). *Mar Environ Res* 100:17–24
- Frehland S, Kaegi R, Hufenus R, Mitrano DM (2020) Long-term assessment of nanoplastic particle and microplastic fiber flux through a pilot wastewater treatment plant using metal-doped plastics. *Water Res* 182:115860
- Fries E, Dekiff JH, Willmeyer J, Nuelle MT, Ebert M, Remy D (2013) Identification of polymer types and additives in marine microplastic particles using pyrolysis-GC/MS and scanning electron microscopy. *Environ Sci: Process Impacts* 15(10):1949–1956
- Fuller S, Gautam A (2016) A procedure for measuring microplastics using pressurized fluid extraction. *Environ Sci Technol* 50(11):5774–5780
- Gao H, Yan C, Liu Q, Ding W, Chen B, Li Z (2019) Effects of plastic mulching and plastic residue on agricultural production: A meta-analysis. *Sci Total Environ* 651:484–492
- Garcés-Ordóñez O, Castillo-Olaya VA, Granados-Briceño AF, García LMB, Díaz LFE (2019) Marine litter and microplastic pollution on mangrove soils of the Ciénaga Grande de Santa Marta, Colombian Caribbean. *Mar Pollut Bull* 145:455–462
- GESAMP (2016) Sources, fate and effects of microplastics in the marine environment (part2). <http://www.gesamp.org/publications/microplastics-in-the-marineenvironment-part-2>
- Geyer R, Jambeck JR, Law KL (2017) Production, use, and fate of all plastics ever made. *Sci Adv* 3(7):e1700782
- Gonçalves JM, Bebianno MJ (2021) Nanoplastics impact on marine biota: a review. *Environ Poll*, p 116426

- Guo JJ, Huang XP, Xiang L, Wang YZ, Li YW, Li H, Cai QY, Mo CH, Wong MH (2020) Source, migration and toxicology of microplastics in soil. *Environ Int* 137:105263
- Guo X, Wang J (2019) The chemical behaviors of microplastics in marine environment: A review. *Mar Pollut Bull* 142:1–14
- Gutow L, Bartl K, Saborowski R, Beermann J (2019) Gastropod pedal mucus retains microplastics and promotes the uptake of particles by marine periwinkles. *Environ Poll* 246:688–696
- Guzzetti E, Sureda A, Tejada S, Faggio C (2018) Microplastic in marine organism: Environmental and toxicological effects. *Environ Toxicol Pharmacol* 64:164–171
- Hahladakis JN, Lacoividou E, Gerassimidou S (2020) Plastic waste in a circular economy. In: *Plastic waste and recycling*. Academic Press, pp 481–512
- Hale RC, Seeley ME, La Guardia MJ, Mai L, Zeng EY (2020) A global perspective on microplastics. *J Geophys Res Oceans* 125(1):e2018JC014719
- Hartmann NB, Hüffer T, Thompson RC, Hassellöv M, Verschoor A, Daugaard AE, Rist S, Karlsson T, Brennholt N, Cole M, Herrling MP (2019) Are we speaking the same language? Recommendations for a definition and categorization framework for plastic debris
- He D, Luo Y, Lu S, Liu M, Song Y, Lei L (2018) Microplastics in soils: analytical methods, pollution characteristics and ecological risks. *TrAC Trends Analyt Chem* 109:163–172
- Hebner TS, Maurer-Jones MA (2020) Characterizing microplastic size and morphology of photodegraded polymers placed in simulated moving water conditions. *Environ Sci: Process Impacts* 22(2):398–407
- Helcoski R, Yonkos LT, Sanchez A, Baldwin AH (2020) Wetland soil microplastics are negatively related to vegetation cover and stem density. *Environ Pollut* 256:113391
- Hermabessiere L, Himber C, Boricaud B, Kazour M, Amara R, Cassone AL, Laurentie M, Paul-Pont I, Soudant P, Dehaut A, Duflos G (2018) Optimization, performance, and application of a pyrolysis-GC/MS method for the identification of microplastics. *Anal Bioanal Chem* 410(25):6663–6676
- Hermesen E, Mintenig SM, Besseling E, Koelmans AA (2018) Quality criteria for the analysis of microplastic in biota samples: a critical review. *Environ Sci Technol* 52(18):10230–10240
- Hidalgo-Ruz V, Gutow L, Thompson RC, Thiel M (2012) Microplastics in the marine environment: a review of the methods used for identification and quantification. *Environ Sci Technol* 46(6):3060–3075
- Hildebrandt L, Mitrano D M, Zimmermann T, Pröfrock D (2020) A nanoplastic sampling and enrichment approach by continuous flow centrifugation. *Front Environ Sci*
- Hirai H, Takada H, Ogata Y, Yamashita R, Mizukawa K, Saha M, Kwan C, Moore C, Gray H, Laursen D, Zettler ER (2011) Organic micropollutants in marine plastics debris from the open ocean and remote and urban beaches. *Mar Pollut Bull* 62(8):1683–1692
- Hodson ME, Duffus-Hodson CA, Clark A, Prendergast-Miller MT, Thorpe KL (2017) Plastic bag derived-microplastics as a vector for metal exposure in terrestrial invertebrates. *Environ Sci Technol* 51(8):4714–4721
- Horton AA, Walton A, Spurgeon DJ, Lahive E, Svendsen C (2017) Microplastics in freshwater and terrestrial environments: evaluating the current understanding to identify the knowledge gaps and future research priorities. *Sci Total Environ* 586:127–141
- Hu Y, Gong M, Wang J, Bassi A (2019) Current research trends on microplastic pollution from wastewater systems: a critical review. *Rev Environ Sci Biotechnol* 18(2):207–230
- Huang JS, Koongolla JB, Li HX, Lin L, Pan YF, Liu S, He WH, Maharana D, Xu XR (2020) Microplastic accumulation in fish from Zhanjiang mangrove wetland, South China. *Sci Total Environ* 708:134839
- Huerta Lwanga E, Gertsen H, Gooren H, Peters P, Salánki T, van der Ploeg M, Besseling E, Koelmans AA, Geissen V (2016) Microplastics in the terrestrial ecosystem: implications for *Lumbricusterrestris* (Oligochaeta, Lumbricidae). *Environ Sci Technol* 50(5):2685–2691
- Imhof HK, Ivleva NP, Schmid J, Niessner R, Laforsch C (2013) Contamination of beach sediments of a subalpine lake with microplastic particles. *Curr Biol* 23(19):R867–R868
- Iyare PU, Ouki SK, Bond T (2020) Microplastics removal in wastewater treatment plants: a critical review. *Environ Sci Water Res* 6(10):2664–2675
- Jatana S, Callahan LM, Pentland AP, DeLouise LA (2016) Impact of cosmetic lotions on nanoparticle penetration through ex vivo C57BL/6 hairless mouse and human skin: a comparison study. *Cosmetics* 3(1):6
- Jeong J, Choi J (2020) Development of AOP relevant to microplastics based on toxicity mechanisms of chemical additives using ToxCast™ and deep learning models combined approach. *Environ Int* 137:105557
- Jeong CB, Won EJ, Kang HM, Lee MC, Hwang DS, Hwang UK, Zhou B, Souissi S, Lee SJ, Lee JS (2016) Microplastic size-dependent toxicity, oxidative stress induction, and p-JNK and p-p38 activation in the monogonont rotifer (*Brachionuskoreanus*). *Environ Sci Technol* 50(16):8849–8857
- Jiang B, Kauffman AE, Li L, McFee W, Cai B, Weinstein J, Lead JR, Chatterjee S, Scott GI, Xiao S (2020) Health impacts of environmental contamination of micro-and nanoplastics: a review. *Environ Health Prev Med* 25(1):1–15
- Karakolis EG, Nguyen B, You JB, Rochman CM, Sinton D (2019) Fluorescent dyes for visualizing microplastic particles and fibers in laboratory-based studies. *Environ Sci Technol Lett* 6(6):334–340
- Karasik R, Vegh T, Diana Z, Bering J, Caldas J, Pickle A, Rittschof D, Viridin J (2020) Years of government responses to the global plastic pollution problem: The plastics policy inventory. *NI X* 20:20–105
- Kelkar VP, Rolsky CB, Pant A, Green MD, Tongay S, Halden RU (2019) Chemical and physical changes of microplastics during sterilization by chlorination. *Water Res* 163:114871
- Kihara S, Köper I, Mata JP, Mc Gillivray DJ (2020) Reviewing nanoplastic toxicology: it's an interface problem. *Adv Colloid Interface Sci*, p 102337
- Kilponen J (2016) Microplastics and harmful substances in urban runoffs and landfill leachates: possible emission sources to marine environment
- Koelmans AA, Nor NHM, Hermesen E, Kooi M, Mintenig SM, De France J (2019) Microplastics in freshwaters and

- drinking water: Critical review and assessment of data quality. *Water Res* 155:410–422
- Kokalj AJ, Hartmann NB, Drobne D, Potthoff A, Kühnel D (2021) Quality of nanoplastics and microplastics ecotoxicity studies: refining quality criteria for nanomaterial studies. *J Hazard Mater*, p 125751
- Kopperi H, Amulya K, Venkata Mohan S (2021) Simultaneous biosynthesis of bacterial polyhydroxybutyrate (PHB) and extracellular polymeric substances (EPS): process optimization and Scale-up. *Bioresour Technol* 341:125735
- Kosuth M, Mason SA, Wattenberg EV (2018) Anthropogenic contamination of tap water, beer, and sea salt. *PLoS one* 13(4):e0194970
- Koutnik VS, Alkidim S, Leonard J, DePrima F, Cao S, Hoek EM, Mohanty SK (2021) Unaccounted Microplastics in Wastewater Sludge: Where Do They Go? *ACS ES&T Water* 1(5):1086–1097
- Lahens L, Strady E, Kieu-Le TC, Dris R, Boukerma K, Rinnert E, Gasperi J, Tassin B (2018) Macroplastic and microplastic contamination assessment of a tropical river (Saigon River, Vietnam) transversed by a developing megacity. *Environ Pollut* 236:661–671
- Lamb JB, Willis BL, Fiorenza EA, Couch CS, Howard R, Rader DN, True JD, Kelly LA, Ahmad A, Jompa J, Harvell CD (2018) Plastic waste associated with disease on coral reefs. *Science* 359(6374):460–462
- Lambert S, Scherer C, Wagner M (2017) Ecotoxicity testing of microplastics: Considering the heterogeneity of physico-chemical properties. *Integr Environ Assess Manag* 13(3):470–475
- Lares M, Ncibi MC, Sillanpää M, Sillanpää M (2018) Occurrence, identification and removal of microplastic particles and fibers in conventional activated sludge process and advanced MBR technology. *Water Res* 133:236–246
- Lebreton L, Slat B, Ferrari F, Sainte-Rose B, Aitken J, Marthouse R, Hajbane S, Cunsolo S, Schwarz A, Levivier A, Noble K (2018) Evidence that the Great Pacific Garbage Patch is rapidly accumulating plastic. *Sci Rep* 8(1):1–15
- Lechner A, Keckeis H, Lumesberger-Loisl F, Zens B, Krusch R, Tritthart M, Glas M, Schludermann E (2014) The Danube so colourful: a potpourri of plastic litter outnumbers fish larvae in Europe's second largest river. *Environ Pollut* 188:177–181
- Lee KW, Shim WJ, Kwon OY, Kang JH (2013) Size-dependent effects of micro polystyrene particles in the marine copepod *Tigriopus japonicus*. *Environ Sci Technol* 47(19):11278–11283
- Lehtiniemi M, Hartikainen S, Näkki P, Engström-Öst J, Koistinen A, Setälä O (2018) Size matters more than shape: ingestion of primary and secondary microplastics by small predators. *Food Webs* 17:e00097
- Lei L, Liu M, Song Y, Lu S, Hu J, Cao C, Xie B, Shi H, He D (2018) Polystyrene (nano) microplastics cause size-dependent neurotoxicity, oxidative damage and other adverse effects in *Caenorhabditis elegans*. *Environ Sci Nano* 5(8):2009–2020
- Li J, Yang D, Li L, Jabben K, Shi H (2015) Microplastics in commercial bivalves from China. *Environ Pollut* 207:190–195
- Li J, Zhang K, Zhang H (2018a) Adsorption of antibiotics on microplastics. *Environ Pollut* 237:460–467
- Li S, Liu H, Gao R, Abdurahman A, Dai J, Zeng F (2018b) Aggregation kinetics of microplastics in aquatic environment: complex roles of electrolytes, pH, and natural organic matter. *Environ Pollut* 237:126–132
- Li QC, Lai YJ, Yu SJ, Li P, Zhou XX, Dong LJ, Liu X, Yao ZW, Liu JF (2021) Sequential Isolation of Microplastics and Nanoplastics in Environmental Waters by Membrane Filtration. Followed by Cloud-Point Extraction Analytical Chemistry 93(10):4559–4566
- Li L, Geng S, Li Z, Song K (2020) Effect of microplastic on anaerobic digestion of wasted activated sludge. *Chemosphere* 247:125874
- Li R, Yu L, Chai M, Wu H, Zhu X (2020) The distribution, characteristics and ecological risks of microplastics in the mangroves of Southern China. *Sci Total Environ* 708:135025
- Li Y, Wang X, Fu W, Xia X, Liu C, Min J, Zhang W, Crittenden JC (2019) Interactions between nano/micro plastics and suspended sediment in water: Implications on aggregation and settling. *Water Res* 161:486–495
- Liu H, Yang X, Liu G, Liang C, Xue S, Chen H, Ritsema CJ, Geissen V (2017) Response of soil dissolved organic matter to microplastic addition in Chinese loess soil. *Chemosphere* 185:907–917
- Liu M, Lu S, Song Y, Lei L, Hu J, Lv W, Zhou W, Cao C, Shi H, Yang X, He D (2018) Microplastic and mesoplastic pollution in farmland soils in suburbs of Shanghai, China. *Environ Pollut* 242:855–862
- Liu P, Qian L, Wang H, Zhan X, Lu K, Gu C, Gao S (2019a) New insights into the aging behavior of microplastics accelerated by advanced oxidation processes. *Environ Sci Technol* 53(7):3579–3588
- Liu S, Jian M, Zhou L, Li W (2019b) Distribution and characteristics of microplastics in the sediments of Poyang Lake. *China Water Sci Technol* 79(10):1868–1877
- Liu Q, Li L, Zhao X, Song K (2021) An evaluation of the effects of nanoplastics on the removal of activated-sludge nutrients and production of short chain fatty acid. *Process Saf Environ Prot* 148:1070–1076
- Löder MG, Imhof HK, Ladehoff M, Löscher LA, Lorenz C, Mintenig S, Piehl S, Primpke S, Schrank I, Laforsch C, Gerdt G (2017) Enzymatic purification of microplastics in environmental samples. *Environ Sci Technol* 51(24):14283–14292
- Long Z, Pan Z, Wang W, Ren J, Yu X, Lin L, Lin H, Chen H, Jin X (2019) Microplastic abundance, characteristics, and removal in wastewater treatment plants in a coastal city of China. *Water Res* 155:255–265
- Lucas N, Bienaime C, Belloy C, Queuneudec M, Silvestre F, Nava-Saucedo JE (2008) Polymer biodegradation: Mechanisms and estimation techniques—A review. *Chemosphere* 73:429–442
- Lv W, Zhou W, Lu S, Huang W, Yuan Q, Tian M, Lv W, He D (2019) Microplastic pollution in rice-fish co-culture system: A report of three farmland stations in Shanghai, China. *Sci Total Environ* 652:1209–1218
- Magnusson K, Norén F (2014) Screening of microplastic particles in and down-stream a wastewater treatment plant. IVL Swedish Environmental Research Institute. DOCUMENT NV-05269-13

- Mahon AM, O'Connell B, Healy MG, O'Connor I, Officer R, Nash R, Morrison L (2017) Microplastics in sewage sludge: effects of treatment. *Environ Sci Technol* 51(2):810–818
- Mariano S, Tacconi S, Fidaleo M, Rossi M, Dini L (2021) Micro and nanoplastics identification: classic methods and innovative detection techniques. *Front Toxicol* 3:2
- Markic A, Gaertner JC, Gaertner-Mazouni N, Koelmans AA (2020) Plastic ingestion by marine fish in the wild. *Crit Rev Environ Sci Technol* 50(7):657–697
- Mason SA, Welch VG, Neratko J (2018) Synthetic polymer contamination in bottled water. *Front Chem* 6:407
- Masura J, Baker J, Foster G, Arthur C (2015) Laboratory methods for the analysis of microplastics in the marine environment: recommendations for quantifying synthetic particles in waters and sediments. Silver Spring, MD, NOAA Marine Debris Division, 31 pp. (NOAA Technical Memorandum NOS-OR&R-48). <https://doi.org/10.25607/OBP-604>
- Mateos-Cárdenas A, Scott DT, Seitmaganbetova G, van Pelt FNAM, A K J M, (2019) Polyethylene microplastics adhere to *Lemna minor* (L.), yet have no effects on plant growth or feeding by *Gammarus duebeni* (Lillj.). *Sci Total Environ* 689:413–421
- Mateos-Cárdenas A, van Pelt FN, O'Halloran J, Jansen MA (2021) Adsorption, uptake and toxicity of micro- and nanoplastics: Effects on terrestrial plants and aquatic macrophytes. *Environ Pollut*, p 117183
- McGivney E, Cederholm L, Barth A, Hakkarainen M, Hamacher-Barth E, Ogonowski M, Gorokhova E (2020) Rapid Physicochemical Changes in Microplastic Induced by Biofilm Formation. *Front Bioeng Biotechnol* 8:205
- Meng Y, Kelly FJ, Wright SL (2020) Advances and challenges of microplastic pollution in freshwater ecosystems: a UK perspective. *Environ Pollut*, p 113445
- Michielssen MR, Michielssen ER, Ni J, Duhaime MB (2016) Fate of microplastics and other small anthropogenic litter (SAL) in wastewater treatment plants depends on unit processes employed. *Environ Sci Water Res* 2(6):1064–1073
- Mintenig SM, Int-Veen I, Löder MG, Primpke S, Gerdtz G (2017) Identification of microplastic in effluents of waste water treatment plants using focal plane array-based micro-Fourier-transform infrared imaging. *Water Res* 108:365–372
- Mofijur M, Ahmed SF, Rahman SA, Siddiki SYA, Islam AS, Shahabuddin M, Ong HC, Mahlia TI, Djavanroodi F, Show PL (2021) Source, distribution and emerging threat of micro- and nanoplastics to marine organism and human health: Socio-economic impact and management strategies. *Environ Res* 195:110857
- Munno K, Helm PA, Jackson DA, Rochman C, Sims A (2018) Impacts of temperature and selected chemical digestion methods on microplastic particles. *Environ Toxicol Chem* 37(1):91–98
- Murphy F, Ewins C, Carbonnier F, Quinn B (2016) Wastewater treatment works (WwTW) as a source of microplastics in the aquatic environment. *Environ Sci Technol* 50(11):5800–5808
- Murray A, Örmeci B (2020) Removal effectiveness of nanoplastics (< 400 nm) with separation processes used for water and wastewater treatment. *Water* 12(3):635
- Murray CC, Maximenko N, Lippiatt S (2018) The influx of marine debris from the Great Japan Tsunami of 2011 to North American shorelines. *Mar Pollut Bull* 132:26–32
- Naidoo T, Thompson RC, Rajkaran A (2020) Quantification and characterisation of microplastics ingested by selected juvenile fish species associated with mangroves in Kwa-Zulu-Natal, South Africa. *Environ Pollut* 257:113635
- Naik RK, Naik MM, D'Costa PM, Shaikh F (2019) Microplastics in ballast water as an emerging source and vector for harmful chemicals, antibiotics, metals, bacterial pathogens and HAB species: a potential risk to the marine environment and human health. *Mar Pollut Bull* 149:110525
- Ng EL, Lwanga EH, Eldridge SM, Johnston P, Hu HW, Geissen V, Chen D (2018) An overview of microplastic and nanoplastic pollution in agroecosystems. *Sci Total Environ* 627:1377–1388
- Nguyen B, Claveau-Mallet D, Hernandez LM, Xu EG, Farner JM, Tufenkji N (2019) Separation and analysis of microplastics and nanoplastics in complex environmental samples. *Acc Chem Res* 52(4):858–866
- Nizzetto L, Futter M, Langaas S (2016) Are agricultural soils dumps for microplastics of urban origin? *Environ Sci Technol* 50(20):10777–10779
- Nor NHM, Obbard JP (2014) Microplastics in Singapore's coastal mangrove ecosystems. *Mar Pollut Bull* 79(1–2):278–283
- O'Neill SM, Lawler J (2021) Knowledge gaps on Micro and Nanoplastics and human health: a critical review. *Case Studies in Chemical and Environmental Engineering*, p 100091
- Oberbeckmann S, Kreikemeyer B, Labrenz M (2018) Environmental factors support the formation of specific bacterial assemblages on microplastics. *Front Biol* 8:2709
- O'Connor D, Pan S, Shen Z, Song Y, Jin Y, Wu WM, Hou D (2019) Microplastics undergo accelerated vertical migration in sand soil due to small size and wet-dry cycles. *Environ Pollut* 249:527–534
- O'Donovan S, Mestre NC, Abel S, Fonseca TG, Carteny CC, Willems T, Prinsen E, Cormier B, Keiter SS, Bebianno MJ (2020) Effects of the UV filter, oxybenzone, adsorbed to microplastics in the clam *Scrobicularia plana*. *Sci Total Environ* 733:139102
- Omidi A, Naeemipoor H, Hosseini M (2012) Plastic debris in the digestive tract of sheep and goats: an increasing environmental contamination in Birjand, Iran. *Bull Environ Contam Toxicol* 88(5):691–694
- OSPAR Commission (2014) Marine litter regional action plan. OSPAR Secretariat, London
- Patricio Silva AL, Prata JC, Duarte AC, Barcelò D, Rocha-Santos T (2021) An urgent call to think globally and act locally on landfill disposable plastics under and after covid-19 pandemic: Pollution prevention and technological (Bio) remediation solutions. *Chem Eng J*, p 131201
- Peng J, Wang J, Cai L (2017) Current understanding of microplastics in the environment: occurrence, fate, risks, and what we should do. *Integr Environ Assess Manag* 13(3):476–482

- Pico Y, Alfarhan A, Barcelo D (2019) Nano-and microplastic analysis: Focus on their occurrence in freshwater ecosystems and remediation technologies. *TrAC Trends Anal Chem* 113:409–425
- Pitt JA, Kozal JS, Jayasundara N, Massarsky A, Trevisan R, Geitner N, Wiesner M, Levin ED, Di Giulio RT (2018) Uptake, tissue distribution, and toxicity of polystyrene nanoparticles in developing zebrafish (*Danio rerio*). *Aquat Toxicol* 194:185–194
- Plastics Europe (2017) *Plastics - the Facts 2017: an analysis of European Plastic Production, Demand and Waste Data*. https://www.plasticseurope.org/application/files/1715/2111/1527/Plastics_the_facts_2017_FINAL_for_website.pdf
- Prata JC (2018) Airborne microplastics: consequences to human health? *Environ Pollut* 234:115–126
- Prata JC, Silva ALP, Da Costa JP, Mouneyrac C, Walker TR, Duarte AC, Rocha-Santos T (2019) Solutions and integrated strategies for the control and mitigation of plastic and microplastic pollution. *Int J Environ Res Public Health* 16(13):2411
- Raju S, Carbery M, Kuttykattil A, Senathirajah K, Subash chandra bose SR, Evans G, Thavamani P, (2018) Transport and fate of microplastics in wastewater treatment plants: implications to environmental health. *Rev Environ Sci Biotechnol* 17(4):637–653
- Ramos JA, Barletta M, Costa MF (2012) Ingestion of nylon threads by Gerreidae while using a tropical estuary as foraging grounds. *Aquat Biol* 17(1):29–34
- Ramos L, Berenstein G, Hughes EA, Zalts A, Montserrat JM (2015) Polyethylene film incorporation into the horticultural soil of small periurban production units in Argentina. *Sci Total Environ* 523:74–81
- Revel M, Châtel A, Mouneyrac C (2018) Micro (nano) plastics: A threat to human health? *Curr Opin Environ Sci Health* 1:17–23
- Rezaei M, Riksen MJ, Sirjani E, Sameni A, Geissen V (2019) Wind erosion as a driver for transport of light density microplastics. *Sci Total Environ* 669:273–281
- Riascos JM, Valencia N, Peña EJ, Cantera JR (2019) Inhabiting the technosphere: The encroachment of anthropogenic marine litter in Neotropical mangrove forests and its use as habitat by macrobenthic biota. *Mar Pollut Bull* 142:559–568
- Rist S, Baun A, Almeda R, Hartmann NB (2019) Ingestion and effects of micro-and nanoplastics in blue mussel (*Mytilus edulis*) larvae. *Mar Pollut Bull* 140:423–430
- Rochman CM, Brookson C, Bikker J, Djuric N, Earn A, Bucci K, Athey S, Huntington A, McIlwraith H, Munno K, De Frond H (2019) Rethinking microplastics as a diverse contaminant suite. *Environ Toxicol Chem* 38(4):703–711
- Rummel CD, Jahnke A, Gorokhova E, Kühnel D, Schmitt-Jansen M (2017) Impacts of biofilm formation on the fate and potential effects of microplastic in the aquatic environment. *Environ Sci Technol Lett* 4(7):258–267
- Sadeghi P, Sadeghi B, Marfavi Y, Kowsari E, Ramakrishna S, Chinnappan A (2021) Addressing the Challenge of Microfiber Plastics as the Marine Pollution Crisis Using Circular Economy Methods: a Review. *Mater Circ Econ* 3(1):1–23
- Sanchez W, Bender C, Porcher JM (2014) Wild gudgeons (*Gobiogobio*) from French rivers are contaminated by microplastics: preliminary study and first evidence. *Environ Res* 128:98–100
- Sandhiya R, Ramakrishna S (2020) Investigating the applicability of blockchain technology and ontology in plastics recycling by the adoption of ZERO plastic model. *Mater Circ Econ* 2:13
- Santillo D, Miller K, Johnston P (2017) Microplastics as contaminants in commercially important seafood species. *Integr Environ Assess Manage* 13:516–521
- Scherer C, Weber A, Lambert S, Wagner M (2018) Interactions of microplastics with freshwater biota. In: *Freshwater microplastics* (pp 153–180). Springer, Cham
- Schmaltz E, Melvin EC, Diana Z, Gunady EF, Rittschof D, Somarelli JA, Virdin J, Dunphy-Daly MM (2020) Plastic pollution solutions: emerging technologies to prevent and collect marine plastic pollution. *Environ Int* 144:106067
- Schneiderman ET (2015) Discharging microbeads to our waters: an examination of wastewater treatment plants in New York
- Schwaferts C, Niessner R, Elsner M, Ivleva NP (2019) Methods for the analysis of submicrometer-and nanoplastic particles in the environment. *TrAC Trend Anal Chem* 112:52–65
- Schwinghammer L, Krause S, Schaum C (2020) Determination of large microplastics: wet-sieving of dewatered digested sludge, co-substrates, and compost. *Water Sci Technol*
- Schymanski D, Goldbeck C, Humpf HU, Fürst P (2018) Analysis of microplastics in water by micro-Raman spectroscopy: release of plastic particles from different packaging into mineral water. *Water Res* 129:154–162
- Senathirajah K, Attwood S, Bhagwat G, Carbery M, Wilson S, Palanisami T (2021) Estimation of the mass of microplastics ingested—A pivotal first step towards human health risk assessment. *J Hazard Mater* 404:124004
- Seth CK, Shrivastav A (2018) Contamination of Indian sea salts with microplastics and a potential prevention strategy. *Environ Sci Pollut Res* 25(30):30122–30131
- Shamsuyeva M, Endres HJ (2021) Plastics in the context of the circular economy and sustainable plastics recycling: comprehensive review on research development, standardisation and market. *Composites Part C: Open Access*, p 100168
- Shen M, Zhang Y, Zhu Y, Song B, Zeng G, Hu D, Wen X, Ren X (2019) Recent advances in toxicological research of nanoplastics in the environment: A review. *Environ Pollut* 252:511–521
- Shim WJ, Song YK, Hong SH, Jang M (2016) Identification and quantification of microplastics using Nile Red staining. *Mar Pollut Bull* 113(1–2):469–476
- Simon M, van Alst N, Vollertsen J (2018) Quantification of microplastic mass and removal rates at wastewater treatment plants applying Focal Plane Array (FPA)-based Fourier Transform Infrared (FT-IR) imaging. *Water Res* 142:1–9
- Simon M, Vianello A, Vollertsen J (2019) Removal of >10 µm Microplastic Particles from Treated Wastewater by a Disc Filter. *Water* 11(9):1935
- Song YK, Hong SH, Eo S, Jang M, Han GM, Isobe A, Shim WJ (2018) Horizontal and vertical distribution of microplastics

- in Korean coastal waters. *Environ Sci Technol* 52(21):12188–12197
- Stock F, Kochleus C, Bänisch-Baltruschat B, Brennholt N, Reifferscheid G (2019) Sampling techniques and preparation methods for microplastic analyses in the aquatic environment—A review. *TrAC Trends Anal Chem*
- Stockholm convention, UNEP, Marine Pollution Litter and Microplastics, Stockholm convention on Persistent Organic Pollutants (2018)
- Sun J, Dai X, Wang Q, van Loosdrecht MC, Ni BJ (2019) Microplastics in wastewater treatment plants: Detection, occurrence and removal. *Water Res* 152:21–37
- Sun H, Lei C, Xu J, Li R (2021) Foliar uptake and leaf-to-root translocation of nanoplastics with different coating charge in maize plants. *J Hazard Mater* 416:125854
- Sun J, Zhu ZR, Li WH, Yan X, Wang LK, Zhang L, Jin J, Dai X, Ni B J (2021) Revisiting Microplastics in Landfill Leachate: Unnoticed Tiny Microplastics and Their Fate in Treatment Works. *Water Res* 190:116784
- Sutton R, Mason SA, Stanek SK, Willis-Norton E, Wren IF, Box C (2016) Microplastic contamination in the San Francisco Bay, California, USA. *Mar Pollut Bull* 109(1):230–235
- Tagg AS, Harrison JP, Ju-Nam Y, Sapp M, Bradley EL, Sinclair CJ, Ojeda JJ (2017) Fenton's reagent for the rapid and efficient isolation of microplastics from wastewater. *Chem Comm* 53(2):372–375
- Talvitie J, Heinonen M, Pääkkönen JP, Vahtera E, Mikola A, Setälä O, Vahala R (2015) Do wastewater treatment plants act as a potential point source of microplastics? Preliminary study in the coastal Gulf of Finland. *Baltic Sea Water Sci Technol* 72(9):1495–1504
- Talvitie J, Mikola A, Koistinen A, Setälä O (2017) Solutions to microplastic pollution—Removal of microplastics from wastewater effluent with advanced wastewater treatment technologies. *Water Res* 123:401–407
- Tchounwou PB, Yedjou CG, Patlolla AK, Sutton DJ (2012) Heavy metal toxicity and the environment. *Molecular, clinical and environmental toxicology*, pp 133–164
- Thompson RC (2015) Microplastics in the marine environment: sources, consequences and solutions. In: *Marine anthropogenic litter* (pp 185–200). Springer, Cham
- UNEP (2014) Plastic debris in the ocean. *UNEP Year Book 2014 Emerging issues update*
- Uurasjärvi E, Hartikainen S, Setälä O, Lehtiniemi M, Koistinen A (2020) Microplastic concentrations, size distribution, and polymer types in the surface waters of a northern European lake. *Water Environ Res* 92(1):149–156
- Van Cauwenberghe L, Janssen CR (2014) Microplastics in bivalves cultured for human consumption. *Environ Pollut* 193:65–70
- Van den Berg P, Huerta-Lwanga E, Corradini F, Geissen V (2020) Sewage sludge application as a vehicle for microplastics in eastern Spanish agricultural soils. *Environ Pollut* 261:114198
- Veerasingam S, Ranjani M, Venkatachalapathy R, Bagaev A, Mukhanov V, Litvinyuk D, Verzhvetskaia L, Gunganathan L, Vethamony P (2020) Microplastics in different environmental compartments in India: Analytical methods, distribution, associated contaminants and research needs. *TrAC Trends Anal Chem* 133:116071
- Venkata Mohan S and Katakowala R (2021) The Circular Chemistry Conceptual Framework: a way forward to Sustainability in Industry 4.0. *Curr Opin Green Sustain Chem* 28:100434
- Venkata Mohan S, Dahiya S, Amulya K, Ranaprathap K, Vanitha TK (2019) Can circular bioeconomy be fueled by waste biorefineries—a closer look. *Bioresour Technol Rep* 7:100277
- Verla AW, Enyoh CE, Verla EN, Nwarnorh KO (2019) Microplastic–toxic chemical interaction: a review study on quantified levels, mechanism and implication. *SN Appl Sci* 1(11):1–30
- Viršek M K, Lovšin M N, Koren Š, Kržan A, Peterlin M (2017) Microplastics as a vector for the transport of the bacterial fish pathogen species *Aeromonas salmonicida*. *Mar Pollut Bull* 125(1–2):301–309
- Wahl A, Le Juge C, Davranche M, El Hadri H, Grassl B, Reynaud S, Gigault J (2021) Nanoplastic occurrence in a soil amended with plastic debris. *Chemosphere* 262:127784
- Walker TR (2021) (Micro) plastics and the UN sustainable development goals. *Curr Opin Green Sustain Chem* 100497
- Wang J, Lv S, Zhang M, Chen G, Zhu T, Zhang S, Teng Y, Christie P, Luo Y (2016) Effects of plastic film residues on occurrence of phthalates and microbial activity in soils. *Chemosphere* 151:171–177
- Wang W, Ndungu AW, Li Z, Wang J (2017) Microplastics pollution in inland freshwaters of China: a case study in urban surface waters of Wuhan, China. *Sci Total Environ* 575:1369–1374
- Wang W, Ge J, Yu X (2020a) Bioavailability and toxicity of microplastics to fish species: a review. *Ecotoxicol Environ Saf* 189:109913
- Wang W, Ge J, Yu X, Li H (2020b) Environmental fate and impacts of microplastics in soil ecosystems: progress and perspective. *Sci Total Environ* 708:134841
- Wang Q, Adams CA, Wang F, Sun Y, Zhang S (2021) Interactions between microplastics and soil fauna: a critical review. *Crit Rev Environ Sci Technol*, pp 1–33
- Wei W, Huang QS, Sun J, Wang JY, Wu SL, Ni BJ (2019) Polyvinyl chloride microplastics affect methane production from the anaerobic digestion of waste activated sludge through leaching toxic bisphenol-A. *Environ Sci Technol* 53(5):2509–2517
- Weinstein JE, Crocker BK, Gray AD (2016) From macroplastic to microplastic: Degradation of high-density polyethylene, polypropylene, and polystyrene in a salt marsh habitat. *Environ Toxicol Chem* 35(7):1632–1640
- Wong KHJ, Lee KK, Kuok Ho DT, Yap PS (2020) Microplastics in freshwater and terrestrial environments: prevalence, facts, impacts and sustainable solutions. *Sci Total Environ*. <https://doi.org/10.1016/j.scitotenv.2020.137512>
- Wright SL, Ulke J, Font A, Chan KL A, Kelly FJ (2020) Atmospheric microplastic deposition in an urban environment and an evaluation of transport. *Environ Int* 136:105411
- Wu WM, Yang J, Criddle CS (2017) Microplastics pollution and reduction strategies. *Front Environ Sci Eng* 11(1):6
- Wu P, Huang J, Zheng Y, Yang Y, Zhang Y, He F, Chen H, Quan G, Yan J, Li T, Gao B (2019) Environmental

- occurrences, fate, and impacts of microplastics. *Ecotoxicol Environ Saf* 184:109612
- Xu M, Halimu G, Zhang Q, Song Y, Fu X, Li Y, Li Y, Zhang H (2019) Internalization and toxicity: A preliminary study of effects of nanoplastic particles on human lung epithelial cell. *Sci Total Environ* 694:133794
- Xu S, Ma J, Ji R, Pan K, Miao A J (2019) Microplastics in aquatic environments: occurrence, accumulation, and biological effects. *Sci Total Environ*, p 134699
- Xu C, Zhang B, Gu C, Shen C, Yin S, Aamir M, Li F (2020) Are we underestimating the sources of microplastic pollution in terrestrial environment? *J Hazard Mater* 400:123228
- Yang X, Lwanga EH, Bemani A, Gertsen H, Salanki T, Guo X, Fu H, Xue S, Ritsema C, Geissen V (2019a) Biogenic transport of glyphosate in the presence of LDPE microplastics: A mesocosm experiment. *Environ Poll* 245:829–835
- Yang L, Li K, Cui S, Kang Y, An L, Lei K (2019b) Removal of microplastics in municipal sewage from China's largest water reclamation plant. *Water Res* 155:175–181
- Yee MSL, Hii LW, Looi CK, Lim WM, Wong SF, Kok YY, Tan BK, Wong CY, Leong CO (2021) Impact of microplastics and nanoplastics on human health. *Nanomaterials* 11(2):496
- Yuan W, Zhou Y, Liu X, Wang J (2019) New perspective on the nanoplastics disrupting the reproduction of an endangered fern in artificial freshwater. *Environ Sci Technol* 53(21):12715–12724
- Zada L, Leslie HA, Vethaak AD, Tinnevelt GH, Jansen JJ, de Boer JF, Ariese F (2018) Fast microplastics identification with stimulated Raman scattering microscopy. *J Raman Spectrosc* 49(7):1136–1144
- Zhang GS, Liu YF (2018) The distribution of microplastics in soil aggregate fractions in south-western China. *Sci Tot Environ* 642:12–20
- Zhang D, Liu HB, Hu WL, Qjn XH, Yan CR, Wang HY (2016) The status and distribution characteristics of residual mulching film in Xinjiang. *China J Integr Agric* 15(11):2639–2646
- Zhang Q, Xu EG, Li J, Chen Q, Ma L, Zeng EY, Shi H (2020) A review of microplastics in table salt, drinking water, and air: direct human exposure. *Environ Sci Technol* 54(7):3740–3751
- Zhang Z, Chen Y (2020). Effects of microplastics on wastewater and sewage sludge treatment and their removal: a review. *Chem Eng Sci* 382:122955
- Zhao S, Danley M, Ward JE, Li D, Mincer TJ (2017) An approach for extraction, characterization and quantitation of microplastic in natural marine snow using Raman microscopy. *Anal Methods* 9(9):1470–1478
- Zheng J, Suh S (2019) Strategies to reduce the global carbon footprint of plastics. *Nat Clim Chang* 9(5):374–378
- Zhou G, Wang Q, Zhang J, Li Q, Wang Y, Wang M, Huang X (2020) Distribution and characteristics of microplastics in urban waters of seven cities in the Tuojiang River basin, China. *Environ Res* 189:109893
- Zhu H, Fu SF, Su Y, Zhang Y (2021) Effects of nanoplastics on microalgae and their trophic transfer along food chain: Recent advances and perspectives. *Environ Sci Processes Impacts*. <https://doi.org/10.1039/D1EM00438G>
- Ziajahromi S, Neale PA, Rintoul L, Leusch FD (2017) Wastewater treatment plants as a pathway for microplastics: development of a new approach to sample wastewater-based microplastics. *Water Res* 112:93–99
- Zuccarello P, Ferrante M, Cristaldi A, Copat C, Grasso A, Sangregorio D, Fiore M, Conti G O (2019) Exposure to microplastics (< 10 µm) associated to plastic bottles

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.