

RESEARCH PAPER

Functionalized TiO₂ nanoparticles labelled with ²²⁵Ac for targeted alpha radionuclide therapy

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Abstract The ²²⁵Ac radioisotope exhibits very attractive nuclear properties for application in radionuclide therapy. Unfortunately, the major challenge for radioconjugates labelled with ²²⁵Ac is that traditional chelating moieties are unable to sequester the radioactive daughters in the bioconjugate which is critical to minimize toxicity to healthy, non-targeted tissues. In the present work, we propose to apply TiO₂ nanoparticles (NPs) as carrier for ²²⁵Ac and its decay products. The surface of TiO₂ nanoparticles with 25 nm diameter was modified with Substance P (5-11), a peptide fragment which targets NK1 receptors on the glioma cells, through the silan-PEG-NHS linker. Nanoparticles functionalized with Substance P (5-11) were synthesized with high yield in a two-step procedure, and the products were characterized by transmission electron microscopy (TEM), dynamic light scattering (DLS) and thermogravimetric analysis (TGA). The obtained results show that one TiO₂-bioconjugate nanoparticle contains in average 80 peptide molecules on its surface. The synthesized TiO₂-PEG-SP(5-11) conjugates were labelled with ²²⁵Ac by ion-exchange reaction on hydroxyl (OH) functional groups on the TiO₂ surface. The

labelled bioconjugates almost quantitatively retain ²²⁵Ac in phosphate-buffered saline (PBS), physiological salt and cerebrospinal fluid (CSF) for up to 10 days. The leaching of ²²¹Fr, a first decay daughter of ²²⁵Ac, in an amount of 30% was observed only in CSF after 10 days. The synthesized ²²⁵Ac-TiO₂-PEG-SP(5-11) has shown high cytotoxic effect in vitro in T98G glioma cells; therefore, it is a promising new radioconjugate for targeted radionuclide therapy of brain tumours.

Keywords Targeted radionuclide therapy · ²²⁵Ac · Titanium dioxide nanoparticles · Substance P · Treatment cancer cells · Nanomedicine

Introduction

Despite of the predominant role of β^- -particle emitters in radionuclide therapy trials (Iagaru et al. 2010), targeted radiotherapy based on α -particles is a promising alternative because they are highly cytotoxic agents, which deposit the whole of their energy within a few cell diameters (50–100 μ m). Cell survival studies have shown that α -particle-induced killing is independent of oxygenation state (Wulbrand et al. 2013) or cell-cycle during irradiation and that as few as 1–3 tracks across the nucleus may result in cell death (Macklis et al. 1988; Sgouros et al. 2010). Additionally, the short range of α -particles corresponds to the dimensions of tumour micrometastases, allowing for localized irradiation of target cells with limited toxicity to surrounding healthy cells.

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Recently, it has been found that in leukaemia, breast and brain cancer small subpopulations of tumour cells are able to self-renew and also to reconstitute the heterogeneous tumour-cell population (Al-Hajj et al. 2003; Singh et al. 2004). These, so called stem cancer cells, may be involved in the widespread metastatic dissemination of cancer (Wichal 2006). These findings have led to the suggestion that failure in tumour treatment may be associated with the failure to eradicate cancer stem cells (Al-Ejeh et al. 2011), especially that it has been found they are not sensitive to chemotherapy agents and to external and internal radiotherapy including β^- -emitters (Al-Hajj et al. 2004). Given the properties outlined above, tumour stem cells are ideal targets for targeted α -particle therapy (TAT) (Sgouros and Song 2008).

There are hundreds of α -particle emitting radionuclides, but only a few have properties suitable for developing therapeutic radiopharmaceuticals: ^{212}Bi ($t_{1/2} = 60$ min), ^{213}Bi ($t_{1/2} = 46$ min), ^{211}At ($t_{1/2} = 7.2$ h), ^{212}Pb ($t_{1/2} = 10.6$ h), ^{227}Th ($t_{1/2} = 18.7$ day), ^{223}Ra ($t_{1/2} = 11.4$ day) and ^{225}Ac ($t_{1/2} = 9.9$ day). However, most of the α -emitting radionuclides with optimal half-life for medical application, have one or more unstable daughter nuclides, which often emit α -particles as well. In principle such a cascade of α -particles can be very efficient in cancer treatment, but reliable delivery to the diseased site can be difficult to achieve. In the case of ^{225}Ac , the most perspective radionuclide for TAT (Sattiraju et al. 2017; Zhu et al. 2017), the four consecutive decay daughters (i.e. ^{221}Fr , ^{217}At , ^{213}Bi and ^{209}Pb) receive recoil energies ranging from 105 to 160 keV as a direct result of the law of conservation of momentum. The recoil energy of firstly produced ^{221}Fr is around 100 keV which is 1000 times greater than the energy of a chemical bond, implying that the recoiling daughter will always break free from the macrocyclic or acyclic chelating agents (Schwartz et al. 2011). Thus, produced daughter radionuclides are released from complexes, may accumulate in healthy organs and cause high toxicity which is a limiting factor in application of these radionuclides in targeted radiotherapy.

The new alternative strategy to sequester ^{225}Ac and its daughters at targeted side is based on the use of nanocarriers. Such encapsulation is expected to prevent, or at least considerably reduce, the release of the recoiling daughters and by this to avoid undesired accumulation in healthy organs and tissues. Moreover, the use of nanocarriers may improve delivery of larger amounts of activity to the diseased site. There has been

proposed several different types of nanostructures to retain ^{225}Ac as well as its daughter radionuclides. Single and multi-layer liposomes have been evaluated; however, experimental studies revealed that their retention properties were poor with less than 10% of ^{213}Bi retained (Sofou et al. 2004; Chang et al. 2008). Theoretical studies indicated that multi-walled polymersomes could achieve around 80% of ^{213}Bi retention, but their diameter has to be more than 800 nm (Thijssen et al. 2012). It would be very difficult for such large particles to extravasate/permeate/penetrate out of vasculature in vivo and also they can be clogged in microcapillary vessels. Woodward et al. (2011) developed 3–5 nm diameter monazite (LaPO_4) nanoparticles as carriers for ^{225}Ac , but these nanoparticles only partially retained daughters. More than ~50% of the ^{221}Fr and ^{213}Bi (from the decay of ^{221}Fr) were released from the nanoparticle lattice. Addition of two layers of LaPO_4 reduced leaching of ^{221}Fr to 20% (Rojas et al. 2015). Similar nanocarrier based on lanthanide phosphate nanoparticles additionally coated with gadolinium and gold layers was developed by McLaughlin et al. (2013). Although multilayered nanoparticles exhibited higher retention properties, their synthesis becomes a multi-step and time-consuming procedure.

In the present work, for sequestration of ^{225}Ac and its decay products, we propose to use titanium dioxide (TiO_2) nanoparticles (NPs), which are inorganic cation exchangers exhibiting high affinity for $^{225}\text{Ac}^{3+}$ and also $^{211}\text{Fr}^+$ and $^{213}\text{Bi}^{3+}$ cations. TiO_2 NPs were conjugated with a fragment of substance P (5-11) for targeting NK1 receptors overexpressed in brain carcinomas. We examined the ability of TiO_2 NPs to retain daughter products of ^{225}Ac and investigated the cytotoxicity of the radiobioconjugate on human *glioblastoma* multiforme cells (T98G cells).

Materials and methods

Reagents, radionuclide and cell line

The following chemical reagents were used directly without further purification: TiO_2 nanoparticles (anatase, ~25 nm, Sigma Aldrich), ethanol (96%, POCH), acetic acid (POCH), phosphate-buffered saline (PBS, Amresco), sodium chloride (POCH), anhydrous dimethylformamide (DMF) and triethylamine (TEA)

of 99% purity were from Sigma Aldrich, substance P (5-11) (Bachem), human serum (Sigma Aldrich, stored at -20°C), methoxyl silane and *N*-hydroxysuccinimide functionalized polyethylene glycol (silane-PEG-NHS, 2000 kDa, Nanocs) and deionized water (Millipore). ^{225}Ac was produced by radiochemical separation from ^{229}Th source as described before (Apostolidis et al. 2005; Zielinska et al. 2007). The activity of ^{225}Ac was quantified using a high-resolution γ -spectrometry when it was in secular equilibrium with its daughters, typically next day after sample collection.

T98G cells (human glioblastoma multiforme cells; CRL-1690; American Type Culture Collection) were cultured in DMEM/F12 K medium (Gibco) enriched with 10% foetal calf serum (FCS, Biological Industries, Israel) and streptomycin (100 $\mu\text{g}/\text{mL}$) and penicillin (100 IU/mL) (Sigma Aldrich). Cells were grown in humidified atmosphere with 5% CO_2 at 37°C . Prior to their in vitro use cells was detached using trypsin-EDTA (0.25%) (Biological Industries, Israel).

Instrumentation

The shape and diameter of titanium dioxide nanoparticles was determined by scanning electron microscopy (SEM, Zeiss) and transmission electron microscopy (TEM, LEO 912AB). The hydrodynamic diameter and zeta potential (ζ) were measured by dynamic light scattering (DLS, Malvern). The presence of surface coating of nanoparticles by PEG and substance P was determined by thermogravimetric analysis (TGA, Q500, TA Instruments). About 5 mg of dried powder was placed in a TGA furnace and the analysis was conducted from room temperature to 900°C at a heating rate of 10°C per min in the presence of inert (nitrogen) atmosphere.

Gamma-ray spectra of radionuclides were measured using coaxial HPGe detector (GX 1080) for gamma spectroscopy with multichannel analyzer DSA-1000 (Canberra Packard) with Genie 2000 software; detection range 10–5000 keV. High-performance liquid chromatography (HPLC) was performed using the ELITE LaChrom (VWR-Hitachi) system with L-2310 pump coupled to L-2455 diode array detector, radiometric (GabiStar, Raytest) detector and L-2350 column oven. Hamilton column (10 μm , 10×250 mm) eluted at the flow rate of 2 mL/min was used for the analytical and semi-preparative chromatography. The gradient elution system consisted of deionized water (A) and acetonitrile (B), both solvents contained 0.1% (v/v)

trifluoroacetic acid (TFA). The gradient conditions were as follows: 0 to 70% solvent B in 0–15 min, 70 to 95% solvent B in 15–20 min.

Conjugation of substance P (5-11) with PEG molecules

An equal molar amount of substance P (5-11) (SP(5-11)) was added to methoxyl silane and *N*-hydroxysuccinimide functionalized polyethylene glycol (silane-PEG-NHS) and dissolved in 250–300 μL DMF followed by 3 μL of TEA addition. Mixture was stirred for 72 h in an inert (nitrogen) gas atmosphere. After completion of the reaction (checked by HPLC), the solvent was removed under vacuum and the product (silan-PEG-SP(5-11)) was separated from unreacted substrates using above described HPLC system and conditions, and finally lyophilized.

Functionalization of TiO_2 surface with silane-PEG-SP(5-11)

The TiO_2 NPs were functionalized with earlier synthesized conjugate silane-PEG-SP(5-11) according to the procedure described in literature (Hermanson 2008). The acetic acid was added to 50 mL of ethanol (96%) to adjust pH to 4.5–5.5. Then about 5 mg of silan-PEG-SP(5-11) conjugate was dissolved and solution was stirred for 5 min. Next, 10 mg of TiO_2 NPs was added and the mixture was sonicated for 10 min followed by 2 h stirring at room temperature (RT). At the end, obtained product was washed several times with ethanol to remove unreacted silane-PEG-SP(5-11) and dried at 100°C for 30 min.

Labelling of TiO_2 -silane-PEG-SP(5-11) with ^{225}Ac

$^{225}\text{Ac}^{3+}$ cations are adsorbed on the surface of TiO_2 NPs through ion-exchange reaction on hydroxyl groups. For a typical labelling, ~ 1 mg of nanoparticles either bare TiO_2 NPs or TiO_2 -silane-PEG-SP(5-11) NPs was suspended in 2 mL of $^{225}\text{AcCl}_3$ (~ 100 kBq). The mixture was sonicated in an ultrasonic bath for 10 min and gently shaken on a circular stirrer for 1 h. After that time labelled NPs were centrifuged at 13000 rpm for 10 min, separated from supernatant and washed several times with deionized water. Finally, the product was suspended in 1 mL phosphate buffer (PB) and its activity measured on a γ -spectrometer to determine the labelling yield.

Stability studies of labelled TiO₂-silane-PEG-SP(5-11) NPs

In order to study leaching of ²²⁵Ac and its decay daughters, a portion of labelled TiO₂-silane-PEG-SP(5-11) NPs was dialyzed against 20 mL of 0.1 M PBS or saline (0.9% NaCl) for 10 days. Every day a 1-mL aliquot was withdrawn and the percentage of liberated activity from each daughter radionuclide was determined by its characteristic γ -ray peak. The stability study of radiolabelled nanoparticles in cerebrospinal fluid (CSF) was conducted using a centrifugal method. A 10 μ L sample of labelled TiO₂-silane-PEG-SP(5-11) NPs was added to 200 μ L of CSF, vortexed and incubated at 37 °C for 10 days. Every day, the probes with added nanoparticles were centrifuged and 100 μ L of CSF was tested for activity of ²²⁵Ac and its decay daughters.

Cytotoxicity evaluation

The impact of TiO₂ nanoparticles conjugated with substance P(5-11) fragment and labelled with ²²⁵Ac on metabolic activity and proliferation of T98G cells was measured with 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT, Sigma Aldrich) assay. The assay was performed as described by Lankoff et al. (2012). In brief, 1 day before the experiment, T98G cells were seeded in 96-well plates (TPP) at a density of 10³ cells/well in 100 μ L of culture medium. Cells were treated for 48, 72 and 96 h with increasing concentrations of non-radiolabelled TiO₂ nanoparticles (0.0625–1 mg/mL) or the nanoparticles labelled with ²²⁵Ac (1.25–20 kBq/mL). At least three independent experiments in six replicate wells were conducted. After the described treatment, cell culture medium was removed and 100 μ L of 3 mg/mL MTT solution was added to each well. After 3 h incubation at 37 °C the MTT solution was removed. Remaining insoluble formazan crystals were dissolved in 100 μ L dimethylsulfoxide (DMSO, Sigma Aldrich) and absorbance of the solution was measured at 570 nm in plate reader spectrophotometer Infinite M200 (Tecan).

Statistical analysis

At least three independent experiments in six replicate wells were conducted for each toxicity point. Difference between samples and control were evaluated using GraphPad Prism 5.0 software (GraphPad Software

Inc., USA). Toxicological data were evaluated by Kruskal-Wallis One Way Analysis of Variance on Ranks (ANOVA) followed by post hoc Dunnett's method. Differences were considered statistically significant when the *p* value was less than < 0.05.

Results and discussion

Currently ²²⁵Ac is the most perspective radionuclide for targeted α -therapy in nuclear medicine. However, one significant factor limiting its use is the difficulty in sequestration of its decay products ²²¹Fr and ²¹³Bi at the targeted site. In our approach, we propose to use TiO₂ nanoparticles as carriers for ²²⁵Ac and its decay daughters. Previous studies on TiO₂ NPs properties indicated their high sorption affinity for 3+ metal cations (Metwally and Rizk 2014) and heavy alkali metal cations (Filipowicz et al. 2014) and facile functionalization and modification of their surface through hydroxyl groups (Bogdan et al. 2017). Therefore, these reports provided a compelling rationale to evaluate TiO₂ NPs as potential carriers for stable retention of ²²⁵Ac and its decay daughters.

The morphology, shape and diameter of bare TiO₂ NPs were characterized by SEM and TEM microscopy. The TiO₂ NPs exhibit nearly spherical morphology and their diameter ranges from 12 to 25 nm, as depicted on Fig. 1.

In order to synthesize a potential radiopharmaceutical exhibiting affinity to NK1 receptors overexpressed on glioma cells, attachment of an active biomolecule is required, in our case, an undecapeptide regulatory neuropeptide, substance P (SP). However, the native SP is metabolized in vivo due to enzymatic cleavages. The fragment SP(5-11) is the most abundant metabolite of native SP and it is still biologically active as the binding site with NK1 receptors is localized in the region of 7–11 amino acids. We have previously shown that this fragment radiolabelled with different radionuclides exhibit high affinity (IC₅₀ = 38 nM) and interaction with NK1 receptors on T98G cells (Piotrowska et al. 2017; Lyczko et al. 2017). Additionally, fragment SP(5-11) contains L-glutamine in position 5 what allows coupling of silane-PEG-NHS via NHS-active ester to form bio-conjugate that further is used to functionalize the TiO₂ NPs surface.

Figure 2 shows the procedure for TiO₂ NPs surface functionalization. In a first step, the silane-PEG-NHS

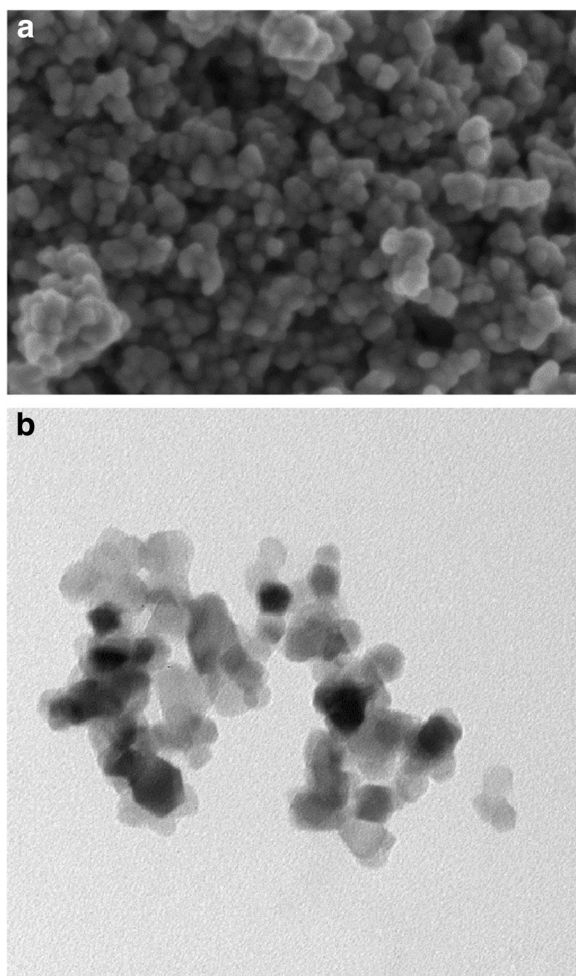


Fig. 1 SEM (a) and TEM (b) images of bare TiO₂ nanoparticles

linker was coupled to the SP(5-11) fragment and the obtained bioconjugate silane-PEG-SP(5-11) was attached to the surface of TiO₂ NPs by siloxane bond formation. Since the OH groups on TiO₂ surface are active sites for both ²²⁵Ac adsorption and also for conjugation through silanization process, the number of silane-PEG-SP(5-11) molecules must be low enough to remain sufficient number of OH groups for adsorption of ²²⁵Ac and its decay products.

TGA provided experimental evidence for the presence of silane-PEG-SP(5-11) on the TiO₂ NPs surface. Figure 3 shows comparison of TGA thermograms of bare TiO₂ and TiO₂ functionalized with silane-PEG-SP(5-11). Both thermograms indicate a loss of mass ~ 1% in the range 25–100 °C and additional 0.8% in the range 100–300 °C, which can be attributed to desorption

of physically adsorbed and intercrystalline water, respectively. The two additional mass losses can be noticed on the thermogram of functionalized TiO₂. The first mass loss observed between 270 and 400 °C (1.5%) is due to peptide molecule degradation and the second between 375 and 450 °C (1%) is related to the silane bond degradation.

On the basis of obtained TGA results the number of silane-PEG-SP(5-11) coating molecules per one TiO₂ nanoparticle can be estimated. The calculation was performed under assumption that the nanoparticle is spherical with medium diameter of 20 nm, as measured by TEM, and that the density of material is 3.8 g cm⁻³. The obtained results show that in average 80 silane-PEG-SP(5-11) molecules are bound to the surface of one TiO₂ nanoparticle. Taking into account that the ion-exchange capacity of TiO₂ nanoparticles is about 1 meq g⁻¹ (Filipowicz et al. 2014), the number of available OH groups on one TiO₂ nanoparticle should exceed 3.8×10^4 . This indicates that even after functionalization of the TiO₂ nanoparticle surface with silane-PEG-SP(5-11) molecules, still the great majority of hydroxyl groups remain capable for binding of ²²⁵Ac³⁺ and formed in the decay process ²²¹Fr⁺ and ²¹³Bi³⁺ cations.

Bare TiO₂ NPs and TiO₂-silane-PEG-SP(5-11) were also characterized by dynamic light scattering (DLS). The obtained values of hydrodynamic diameters and zeta (ζ) potentials are presented in Table 1. Due to the hydration layer formed on bare nanoparticles, the hydrodynamic diameter measured by DLS is usually greater than that measured by TEM. The observed increase of the hydrodynamic diameter for TiO₂ NPs after functionalization process is another confirmation that silanization reaction proceeded and silane-PEG-SP(5-11) molecules were indeed coupled to the surface of TiO₂ NPs. The measured negative zeta potential value of -28.5 mV for TiO₂-silane-PEG-SP(5-11) indicates that particles repel each other and do not aggregate what was confirmed by monitoring changes in hydrodynamic diameter over many days.

Bare TiO₂ and TiO₂-silane-PEG-SP(5-11) NPs were radiolabelled with ²²⁵Ac (100 kBq) at pH = 6.0 with a high yield of $99.8 \pm 2.1\%$ ($n = 13$) and $98.2 \pm 1.3\%$ ($n = 10$), respectively. The obtained results indicate that there is no difference in labelling yield between both nanoparticles and that surface functionalization did not have any negative impact on the radiolabelling

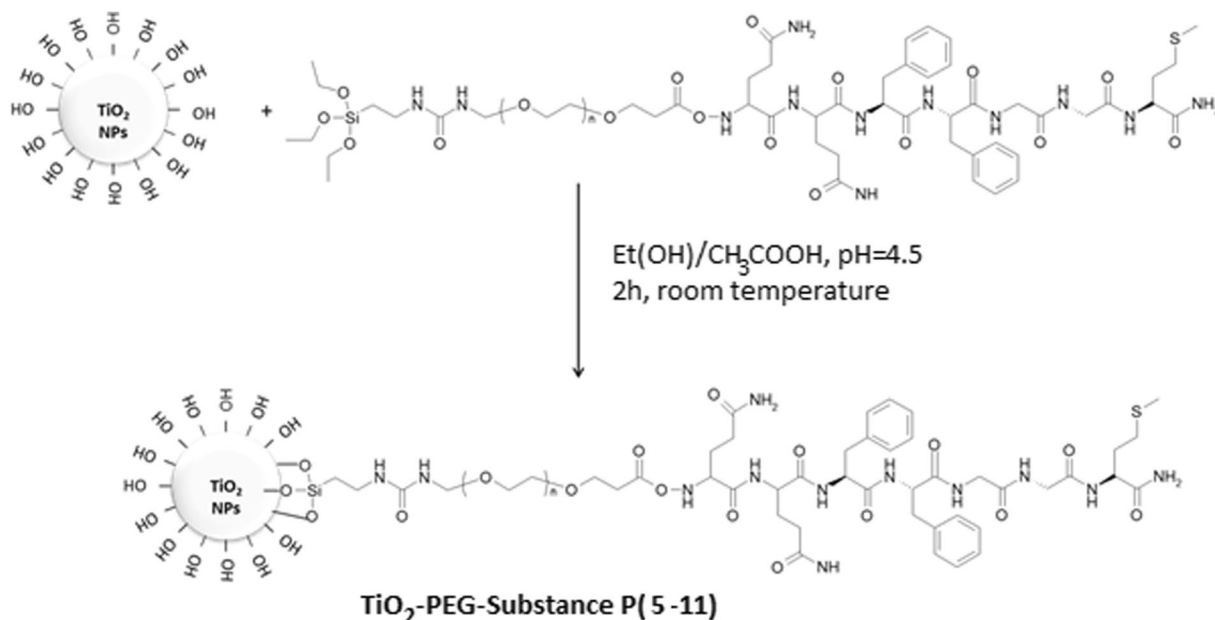


Fig. 2 Functionalization of titanium dioxide nanoparticles with PEG-SP(5-11)

process. It was expected, as previous calculations confirmed that the number of attached silane-PEG-SP(5-11) molecules is significantly much lower than the number of available OH functional groups responsible for ion-exchange approach with ^{225}Ac .

However, during our studies, it was found that due to low acidic character of the OH functional groups, the radiolabelling process is pH dependent and in acidic environment (pH = 3) the yield decreases even to only 27% for bare TiO₂ NPs. Therefore, the

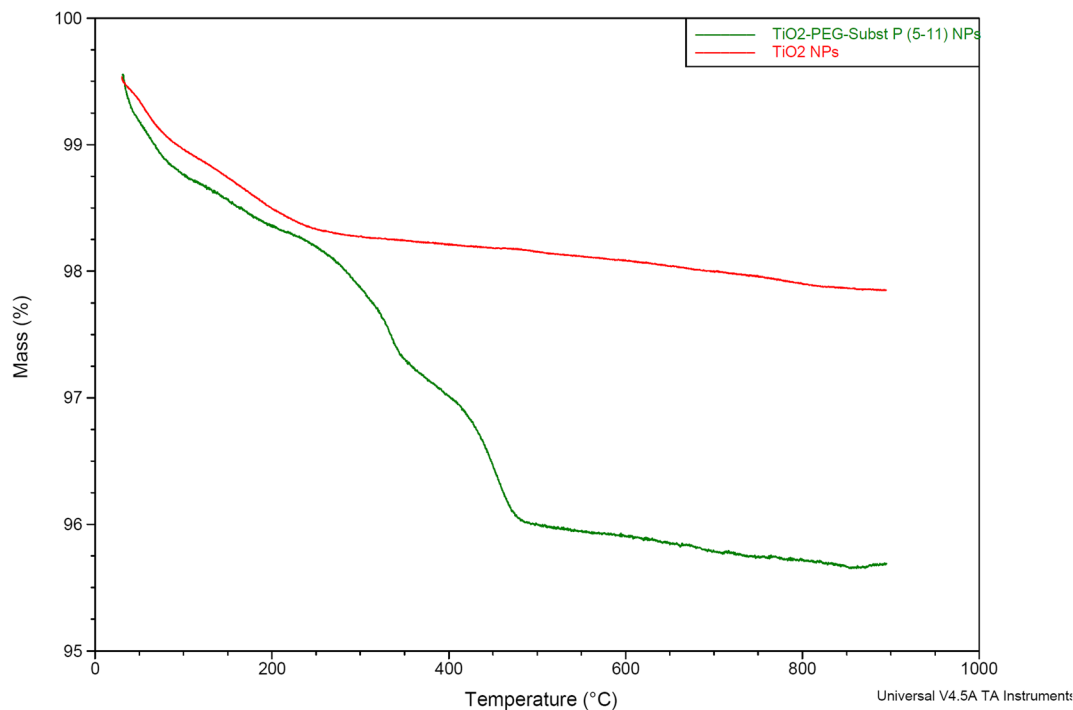


Fig. 3 Thermograms of bare TiO₂ NPs and TiO₂ NPs functionalized with silane-PEG-SP(5-11)

Table 1 Hydrodynamic diameter and zeta (ζ) potential of bare and functionalized TiO₂ NPs (pH = 7.4, PB buffer)

Nanoparticles	Hydrodynamic diameter	Zeta (ζ) potential (mV)
TiO ₂	76.02 ± 5.54	− 23.2
TiO ₂ -silane-PEG-SP(5-11)	98.78 ± 4.50	− 28.5

optimal pH for radiolabelling reaction is 6.0 and it was used in further studies.

The stability of TiO₂-silane-PEG-SP(5-11) NPs labelled with ²²⁵Ac was examined in 0.02 M PBS (pH = 7.4), physiological salt (0.9% NaCl) and cerebrospinal fluid (CSF) for 4 days (Table 2). The leaching of mother radionuclide ²²⁵Ac and its first decay product ²²¹Fr was measured on γ -spectrometer. In PBS and physiological salt NPs retained more than 95% of ²²⁵Ac and ²²¹Fr over 10 days. In CSF, the fraction of ²²⁵Ac released from NPs was still less than 5%, but the retention of ²²¹Fr was decreasing for the first 2 days and later stabilized and remained constant up to 4 days. The high stability in CSF is very important, as treatment of glioblastoma multiforme brain tumour is mostly done through intratumoral or intracavitary injection of radiopharmaceutical, which than can readily penetrate brain parenchyma and target widely disseminated cancerous cells.

The obtained retention results might be interpreted on the base of recoil energy in the ²²⁵Ac \rightarrow ²²¹Fr reaction and subsequent α decays. Calculated, using the SRIM program (SRIM 2010), recoil range of ²²¹Fr in water exceeds 84 nm, but in the material with density close to TiO₂-anatase shows only about 44 nm. When the decay series is taken into consideration, we can assume that the radionuclides ²¹⁷At, ²¹³Bi and ²⁰⁹Pb (daughters of ²²¹Fr) receive similar recoil energy. The escape of the radioactive isotopes from recoiled daughter and targeting agents allows them to freely migrate in the body, causing toxicity to healthy tissues and decreasing the

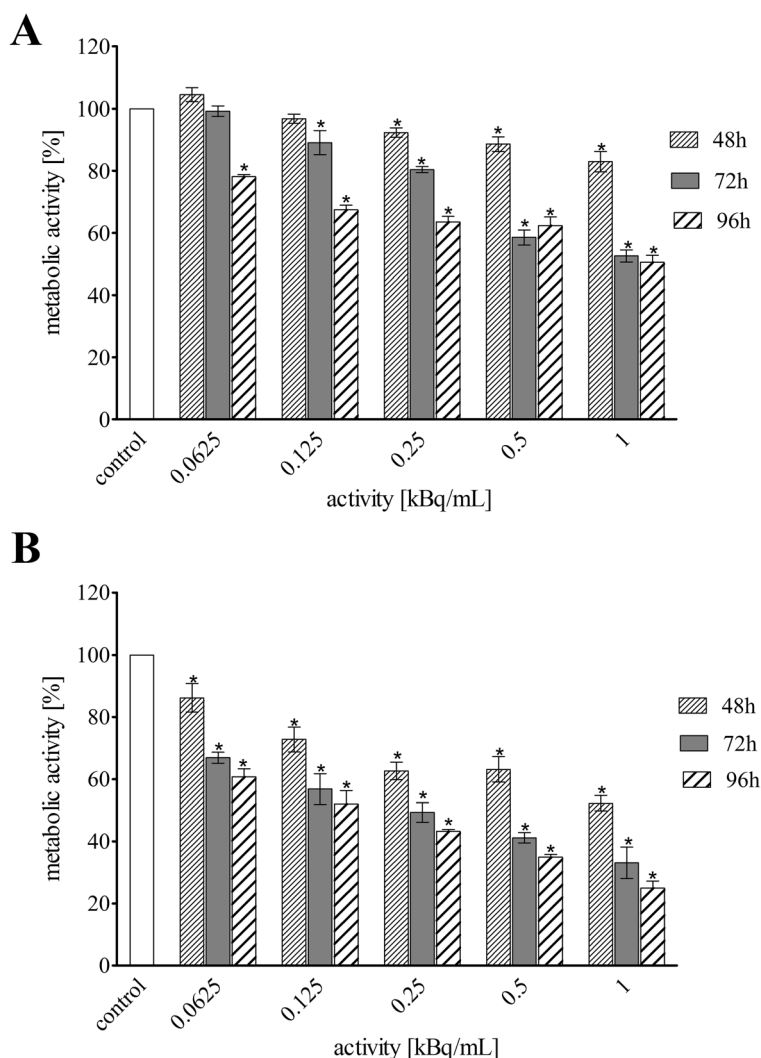
therapeutic dose delivered to the diseased site. The renal toxicity induced by ²¹³Bi is considered to be the major constraint to apply ²²⁵Ac in a large number of clinical trials (Jaggi et al. 2005; Schwartz et al. 2011). Based on the literature data, one would expect that higher amount of ²²¹Fr and ²¹³Bi nuclides would be ejected from a 20-nm-diameter TiO₂ nanoparticle than observed in our studies. The observed smaller release of the daughter radionuclide ²²¹Fr (< 5% in PBS and saline solution) suggests that a fraction of the recoil energy takes part in translation of the whole NP rather than being entirely dissipated through atomic displacements resulting, as it occurs in the bulk, in shorter recoil range. Additionally, in the case of TiO₂ which is an effective cation exchanger, we observe subsequent reloading of escaped recoils ²²¹Fr⁺ and ²¹³Bi³⁺ as it contains hydroxyl surface groups with high affinity for these cations (Filipowicz et al. 2014; Bogdan et al. 2017; Perekhozheva et al. 1985; Bilewicz et al. 1991). This process can also explain slightly lower stability of radiolabelled NPs in CSF, as in this medium hydroxyl surface groups of NPs are blocked by the proteins and peptides present in CSF what prevents rebinding of released daughter radionuclides.

Cytotoxicity studies were performed on human glioma T98G cells with NK1 receptor overexpression. Cells were exposed to different radioactivities of ²²⁵Ac-silane-TiO₂-PEG and ²²⁵Ac-silane-TiO₂-PEG-SP(5-11) as well as different concentrations of non-radioactive TiO₂-silane-PEG and TiO₂-silane-PEG-SP(5-11) for up to 96 h. The effect of all studied compounds was assessed by the MTT assay. The metabolic activity of cells exposed to ²²⁵Ac-silane-TiO₂-PEG-SP(5-11) decreased rapidly with increasing radioactive doses and incubation time (Fig. 4b). On the contrary, ²²⁵Ac-silane-TiO₂-PEG NPs control, without targeting biomolecule, reduced cells viability, although the degree was not so much significant. Thus, the obtained data clearly indicate that observed cytotoxicity was specific to NK1 receptors and directly correlated with its expression level on tumour cells. The non-radiolabelled TiO₂-silane-PEG and TiO₂-silane-PEG-SP(5-11) NPs were relatively non-toxic as decrease in metabolic activity was observed only for the highest concentration (1 mg/mL) (Fig. 5). This concentration reduced the metabolic activity of T98G cells after 96 h incubation to 75 and 73% of control value for TiO₂-silane-PEG and TiO₂-silane-PEG-SP(5-11) NPs, respectively. These results reveal that indeed ²²⁵Ac attached to NPs is a source of toxicity.

Table 2 Retention of ²²⁵Ac and ²²¹Fr on TiO₂-silane-PEG-SP(5-11) NPs radiolabelled with ²²⁵Ac

Solution	²²¹ Fr retention (%)			
	24 h	48 h	72 h	96 h
PBS (pH = 7.4)	97.7	98.9	96.8	98.8
Physiological salt (0.9% NaCl)	98.5	98.4	93.3	94.8
Cerebrospinal fluid (CSF)	78.2	70.6	68.5	–

Fig. 4 Metabolic activity (MTT assay) of T98G cells treated with different radioactivities of ^{225}Ac -TiO₂-silane-PEG (a) and ^{225}Ac -TiO₂-silane-PEG-SP(5-11) (b) NPs for 48, 72 and 96 h. Data are expressed as percent of control and the mean \pm SD from three independent experiments



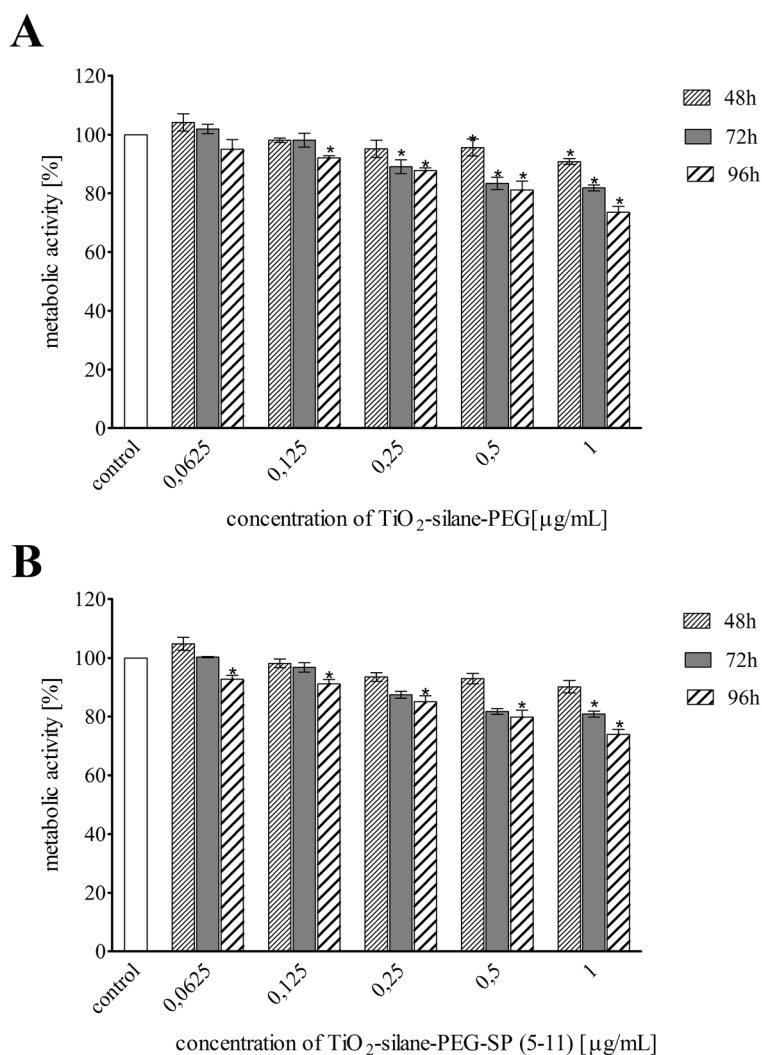
The cytotoxicity of ^{225}Ac -silane-TiO₂-PEG-SP(5-11) is very high in comparison to other radiopharmaceuticals including conjugates radiolabelled with α -emitters like ^{213}Bi or ^{211}At (Gadbois et al. 1996; Dziawer et al. 2017), however, it is comparable with other radiobioconjugates of ^{223}Ra and ^{225}Ac (Borchardt et al. 2003; Piotrowska et al. 2017). The decay processes of ^{225}Ac and ^{223}Ra include four α and two β^- emissions to a stable ^{209}Bi and ^{207}Pb daughters. Because a very large amount of energy (~ 28 MeV) is released during these decay processes, much smaller amount of radionuclides activity is required to produce the desired effect and ^{225}Ac and ^{223}Ra radionuclides demonstrate extreme cytotoxicity. Comparison of obtained cytotoxicity results of ^{225}Ac -silane-TiO₂-PEG-SP(5-11) NPs with literature data for radiopharmaceuticals labelled with ^{213}Bi (Friesen

et al. 2007) demonstrate that several logs less ^{225}Ac radioactivity is required to reach similar LD₅₀ value, because of the longer half-life and multiple α -emissions.

Conclusion

We have shown that TiO₂ NPs functionalized with substance P (5-11) can be used to deliver and retain ^{225}Ac and its daughter radioisotopes at a target site; thereby reducing the absorbed dose to non-targeted organs. The TiO₂ NPs successfully retain a large fraction of the ^{225}Ac decay products without compromising the tumoricidal properties of the α radiation. Because of the reloading of the ^{225}Ac decay products the retention efficiencies of TiO₂ NPs are comparable to the radiolabelled LnPO₄-Au core shell

Fig. 5 Metabolic activity (MTT assay) of T98G cells treated with different concentration of non-radiolabelled TiO_2 (a) and TiO_2 -silane-PEG-SP(5-11) (b) NPs for 48, 72 and 96 h. Data are expressed as percent of control and the mean \pm SD from three independent experiments



nanostructures; however, the synthesis process is much simpler and less time-consuming. Furthermore, it has been shown that ^{225}Ac - TiO_2 -silane-PEG-SP(5-11) exhibits satisfactory stability in CSF what is very important as it is intended for direct intratumoral or post-resection application. The intravenous injection of the ^{225}Ac - TiO_2 -silane-PEG-SP(5-11) is excluded due to the relatively large size and high hydrophilicity which prevents crossing the blood-brain barrier.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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References

- Al-Ejeh F, Smart CE, Morrison BJ, Chenevix-Trench G, Lopez JA, Lakhani SR, Brown MP, Khanna KK (2011) Breast cancer stem cells: treatment resistance and therapeutic opportunities. *Carcinogenesis* 32:650–658
- Al-Hajj M, Becker MW, Wicha M, Weissman I, Clarke MF (2004) Therapeutic implications of cancer stem cells. *Curr Opin Genet Dev* 14:43–47

- Al-Hajj MA, Wichal MS, Benito-Hernandez A, Morrison SJ, Clarke MF (2003) Prospective identification of tumorigenic breast cancer cells. *Proc Nat Acad Sci USA* 100:3983–3988
- Apostolidis C, Molinet R, Rasmussen G, Morgenstern A (2005) Production of ac-225 from Th-229 for targeted alpha therapy. *Anal Chem* 77:6288–6291
- Bilewicz A, Dybczynski R, Narbutt J (1991) Ion exchange of alkali metals on hydrous titanium dioxide in neutral and alkaline solutions. *J Radioanal Nucl Chem* 148:359–371
- Bogdan J, Plawinska-Czarnak J, Zarzynska J (2017) Nanoparticles of titanium and zinc oxides as novel agents in tumor treatment: a review. *Nanoscale Res Lett* 12:225
- Borchardt PE, Yuan RR, Miederer M, McDevitt MR, Scheinberg DA (2003) Targeted actinium-225 in vivo generators for therapy of ovarian cancer. *Cancer Res* 63:5084–5090
- Chang M, Seideman J, Sofou S (2008) Enhanced loading efficiency and retention of ^{225}Ac in rigid liposomes for potential targeted therapy of micrometastases. *Bioconjug Chem* 19:1274–1282
- Dziawer L, Koźmiński P, Męczynska-Wielgosz S, Pruszyński M, Łyczko M, Was B, Celichowski G, GrobelnyJastrzebski J, Bilewicz A (2017) Gold nanoparticle bioconjugates labelled with ^{211}At for targeted alpha therapy. *RSC Adv* 7:41024–41032
- Filipowicz B, Pruszyński M, Krajewski S, Bilewicz A (2014) Adsorption of ^{137}Cs on titanate nanostructures. *J Radioanal Nucl Chem* 301:889–895
- Friesen C, Glatting G, Koop B, Schwarz K, Morgenstern A, Apostolidis C, Debatin K, Reske SN (2007) Breaking chemoresistance and radioresistance with ^{213}Bi anti-CD45 antibodies in leukemia cells. *Cancer Res* 67:1950–1958
- Gadbois DM, Crissman HA, Nastasi A, Habberset R, Wang SK, Chen D, Lehnert BE (1996) Alterations in the progression of cells through the cell cycle after exposure to alpha particles or gamma rays. *Radiat Res* 146:414–424
- Hermanson GT (2008) *Bioconjugate techniques*, 3rd. edn. Academic Press, Amsterdam
- Iagaru A, Mitra ES, Ganjoo K, Knox SJ, Goris ML (2010) ^{131}I -Tositumomab (Bexxar®) vs. ^{90}Y -Ibritumomab (Zevalin®) therapy of low-grade refractory/relapsed non-Hodgkin lymphoma. *Mol Imag Biol* 12:198–203
- Jaggi JS, Kappel BJ, McDevitt MR, Sgouros G, Flombaum CD, Cabassa C, Scheinberg DA (2005) Efforts to control the errant products of a targeted in vivo generator. *Cancer Res* 65:4888–4895
- Lankoff A, Sandberg WJ, Wegierek-Ciuk A, Lisowska H, Refsnes M, Sartowska B (2012) The effect of agglomeration state of silver and titanium dioxide nanoparticles on cellular response of HepG2, A549 and THP-1 cells. *Toxicol Lett* 208:197–213
- Łyczko M, Pruszyński M, Majkowska-Pilip A, Łyczko K, Was B, Meczynska-Wielgosz S, Kruszewski M, Szkliniarz K, Jastrzebski J, Stolarz A, Bilewicz A (2017) ^{211}At labeled substance P (5–11) as potential radiopharmaceutical for glioma treatment. *Nucl Med Biol* 53:1–8
- Macklis RM, Kinsey BM, Kassis AI, Ferrara JL, Archer RW, Hines JJ, Coleman CN, Adelstein SJ, Burakoff SJ (1988) Radioimmunotherapy with alpha particle-emitting immunoconjugates. *Science* 240:1024–1026
- McLaughlin MF, Woodward J, Boll RA, Wall JS, Rondinone AJ, Kennel SJ, Mirzadeh S, Robertson JD (2013) Gold coated lanthanide phosphate nanoparticles for targeted alpha generator radiotherapy. *PLoS One* 8:e54531
- Metwally SS, Rizk HE (2014) Preparation and characterization of nano-sized iron-titanium mixed oxide for removal of some lanthanides from aqueous solution. *Sep Sci Technol* 49:2426–2436
- Perekhozheva TN, Sharygin LM, Albantova GP (1985) Cation-exchange properties of an adsorbent based on hydrated titanium-dioxide. *Inorg Mat* 21:364–367
- Piotrowska A, Męczynska-Wielgosz S, Majkowska-Pilip A, Koźmiński P, Wójciuk G, Cędrowska E, Bruchertseifer F, Morgenstern A, Kruszewski M, Bilewicz A (2017) Nanozeolite bioconjugates labeled with ^{223}Ra for targeted alpha therapy. *Nucl Med Biol* 47:10–18
- Rojas JV, Woodward JD, Chen N, Rondinone AJ, Castano CH, Mirzadeh S (2015) Synthesis and characterization of lanthanum phosphate nanoparticles as carriers for ^{223}Ra and ^{225}Ra for targeted alpha therapy. *Nucl Med Biol* 42:614–620
- Sattiraju A, Solingapuram K, Sai K, Xuan A, Pandya DN, Almaguel FG, Wadas TJ, Herpai DM, Debinski W, Mintz A (2017) IL13RA2 targeted alpha particle therapy against glioblastomas. *Oncotarget* 8:42997–43007
- Schwartz J, Jaggi JS, O'Donoghue JA, Ruan S, McDevitt M, Larson SM, Scheinberg DA, Humm JL (2011) Renal uptake of bismuth-213 and its contribution to kidney radiation dose following administration of actinium-225-labeled antibody. *Phys Med Biol* 56:721–733
- Sgouros G, Song H (2008) Cancer stem cell targeting using the alpha-particle emitter, ^{213}Bi : mathematical modeling and feasibility analysis. *Cancer Biother Radiopharm* 23:74–81
- Sgouros G, Roeske JC, McDevitt MR, Palm S, Allen BJ, Fisher DR, Brill AB, Song H, Howell RW, Akabani G (2010) MIRD pamphlet no. 22 (abridged): radiobiology and dosimetry of α -particle emitters for targeted radionuclide therapy. *J Nucl Med* 51:311–328
- Singh SK, Hawkins C, Clarke ID, Squire JA, Bayani J, Hide T, Henkelman RM, Cusimano MD, Dirks PB (2004) Identification of human brain tumour initiating cells. *Nature* 432:396–401
- Sofou S, Thomas JL, Lin H, McDevitt MR, Scheinberg DA (2004) Sgouros G (2004) engineered liposomes for potential alpha particle therapy of metastatic cancer. *J Nucl Med* 45:253
- SRIM. Particle Interactions with Matter. <http://www.srim.org/2010>
- Thijssen L, Schaart DR, de Vries D, Morgenstern A, Bruchertseifer F, Denkova AG (2012) Polymersomes as nano-carriers to retain harmful recoil nuclides in alpha radionuclide therapy: a feasibility study. *Radiochim Acta* 100:473–481
- Wichal MS (2006) Cancer stem cells and metastasis: lethal seeds—commentary. *Clin Cancer Res* 12:5606–5607
- Woodward J, Kennel SJ, Stuckey A, Osborne D, Wall J, Rondinone AJ, Standaert RF, Mirzadeh S (2011) LaPO4 nanoparticles doped with actinium-225 that partially sequester daughter radionuclides. *Bioconjug Chem* 22:766–776
- Wulbrand C, Seidl C, Gaertner FC, Bruchertseifer F, Morgenstern A, Essler M, Senekowitsch-Schmidtke R (2013) Alpha-particle emitting ^{213}Bi -anti-EGFR immunoconjugates eradicate tumor cells independent of oxygenation. *PLoS One* 8:e64730
- Zhu C, Sempkowski M, Holleran T, Linz T, Bertalan T, Josefsson A, Bruchertseifer F, Morgenstern A, Sofou S (2017) Alpha-particle radiotherapy: for large solid tumors diffusion trumps targeting. *Biomaterials* 130:67–75
- Zielinska B, Apostolidis C, Bruchertseifer F, Morgenstern A (2007) An improved method for the production of Ac-225/Bi-213 from Th-229 for targeted alpha therapy. *Solv Extr Ion Exch* 25:339–349