



Bioprocess Optimization of Nutritional Parameters for Enhanced Anti-leukemic L-Asparaginase Production by *Aspergillus candidus* UCCM 00117: A Sequential Statistical Approach

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Abstract

Sequential optimization of bioprocess nutritional conditions for production of glutaminase-near-free L-asparaginase by *Aspergillus candidus* UCCM 00117 was conducted under shake flask laboratory conditions. Catalytic and anti-cancer activities of the poly-peptide were evaluated using standard in vitro biochemical methods. Medium nutrients were selected by one-factor-at-a-time (OFAT) approach while Plackett–Burman design (PBD) screened potential factors for optimization. Path of steepest ascent (PSA) and response surface methodology (RSM) of a Min-Run-Res V fractional factorial of a central composite rotatable design (CCRD) were employed to optimize factor levels towards improved enzyme activity. A multi-objective approach using desirability function generated through predictor importance and weighted coefficient methodology was adopted for optimization. The approach set optimum bioprocess conditions as 49.55 g/L molasses, 64.98% corn steep liquor, 44.23 g/L asparagine, 1.73 g/L potassium, 0.055 g/L manganese and 0.043 g/L chromium (III) ions, at a composite desirability of 0.943 and an L-asparaginase activity of 5216.95U. The Sephadex-200 partially-purified polypeptide had a specific activity of 476.84 U/mg; 0.087U glutaminase activity, 36.46% yield and 20-fold protein purification. Anti-cancer activity potentials of the catalytic poly-peptide were dose-dependent with IC₅₀ (µg/mL): 4.063 (HL-60), 13.75 (HCT-116), 15.83 (HeLa), 11.68 (MCF-7), 7.61 (HepG-2). The therapeutic enzyme exhibited 15-fold more cytotoxicity to myeloid leukemia cell line than to normal (HEK 238 T) cell. Optimum temperature and pH for activity were within physiological range. However, significant interactions between exposure time and levels of each of temperature and pH made interpretations of residual enzyme activities difficult. The manganese-dependent L-asparaginase from *Aspergillus s candidus* UCCM 00117 is recommended for further anticancer drug investigations.

Keywords Sequential optimization · *Aspergillus candidus* · L-Asparaginase activity · Renewable substrates · Anti-cancer activity · Selectivity index

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Introduction

Cancers are among the topmost causes of global death and arise from uncontrolled proliferation of certain cells, tissues and organs including breast, lung, blood, liver, pancreas, stomach, colon and cervix, after exposure to certain environmental, nutritional or habitual agents (Sultana et al. 2019; Ervina et al. 2020). Certain types of cancers can also develop genetically and are heritable within genetically-related populations (Sultana et al. 2019). Cancerous cells characteristically have a requirement for asparagine having lost their ability to express the gene that encodes asparagine synthetase (Sarquis et al. 2004). In human carcinomas therefore, the primary target of anti-cancer compounds is the central sensor for nutrient and energy availability which regulates cell cycle, growth and proliferation (Luengo et al. 2017; Zhang et al. 2018). Cancer treatments have largely proceeded through surgery, radiotherapy, immunotherapy and chemotherapy employing diverse agents including inorganic (Jaros et al. 2019; Chen et al. 2020) and organic compounds or a combination of both (Bokas et al. 2020). Peptide molecules from different biological sources including plants (de Oliveira et al. 2015; Eswaraiah et al. 2020), animals (Elrayess et al. 2020) and microorganisms (Sereena and Sebastian 2020; Harish et al. 2020) have demonstrated precision in target identification and activity, and have therefore received tremendous attention as key components of anti-cancer chemotherapy in recent years.

L-asparaginases (EC 3.5.1.1) are amidohydrolases which primarily catalyze the hydrolytic cleavage of L-asparagine into L-aspartic acid and ammonia often-times with secondary catalytic cleavage of L-glutamine to L-glutamic acid and ammonia using glutaminase activity (Avramis 2012; Cachumba et al. 2016). The enzyme is frequently included in modern treatment regime for lymphoblastic leukemia and other lymphomas, and in acrylamide reduction protocol in the food industry (Kukurova et al. 2009; Pieters et al. 2011; Shakambari et al. 2019). L-asparaginase is frequently sourced from plants (Oza et al. 2010; Mohamed et al. 2015) and microorganisms (Sahu et al. 2007; Hassan et al. 2018). Foremost among microorganisms that produce L-asparaginases include species of *Escherichia* and *Erwinia* however; other bacteria including *Bacillus velezensis* (Mostafa et al. 2019), *Enterobacter cloacae* (Husain et al. 2016), *Streptomyces rochei* (El-Naggar and El-Shweihy 2020) and *Geobacillus thermodenitrificans* (Nadeem et al. 2019) have also been reported. Species of the genera *Aspergillus*, *Penicillium*, *Fusarium* (Sarquis et al. 2004) and *Trichosporon* (Ashok et al. 2019) dominate literature as fungal L-asparaginase producers. *Aspergillus* species most reported include

Aspergillus niger (Vala et al. 2018), *Aspergillus terreus* and *Aspergillus tamari* (Baskar and Sahadevan 2012; Sarquis et al. 2004).

Applications of bacterial and fungal L-asparaginases, especially from species of *Bacillus* (Alrumman et al. 2019) and *Aspergillus* (Prabavathy and Phoebe 2020), in anti-cancer research have been documented. However, the difficulty in obtaining sufficient plant L-asparaginases to meet its high demand and the allergic reactions that attend microbial L-asparaginases during clinical applications owing to secondary glutaminase co-activity are significant drawbacks in anti-cancer chemotherapy (Cecconello et al. 2020). Remedial attempts at toxicity reduction during chemotherapy have included the use of pegylated forms of the product (Appel et al. 2008) as well as sourcing for producing strains with low glutaminase activity (Mahajan et al. 2012). Currently, there is an upsurge in reports on glutaminase free L-asparaginase (El-Naggar et al. 2018) which presupposes that anti-cancer activity of L-asparaginases may not require the associated glutaminase activity (Chan et al. 2014). However, complete elimination of glutaminase activity from L-asparaginase may render the medication therapeutically ineffective (Mahajan et al. 2012). A workable remedy may well be the production of L-asparaginases with low glutaminase activity and optimizing fermentation conditions that enhance their production.

Microbial fermentations have been at the forefront of production of L-asparaginases by solid-state and submerged methods (Shakambari et al. 2019). However, yields of the product have been low and with poor production economics (Farag et al. 2015). Sourcing for high-yielding strains, improving them through genetic and metabolic manipulations; optimization of their bioprocess conditions of media, operations and downstream processing; remain reliable approaches towards yield improvement (Baskar and Sahadevan 2012; Yingling and Zhengfang 2013). Nutritional conditions for microbial fermentations have been reported to account for 30–40% of production economics of bioprocesses; sources of carbon and nitrogen being the most critical. Selection of significant nutrients for inclusion in microbial fermentation media and optimizing their levels are subjects that are vigorously pursued by industries (Shakambari et al. 2019). A sequence of reported techniques in that regard include one-factor-at-a-time (OFAT) approach that does the primary selection (Long et al. 2018) followed by a two-level factorial design like Plackett–Burman design (PBD) that establishes significance of selected factors (Mechmeche et al. 2019; Gururaj et al. 2020). A third procedure in the optimization process is path of steepest ascent (PSA) which moves significant coefficients from the first-order model of PBD close to their optimum with subsequent resetting of center points for response surface methodology (RSM) (Huang 2019). The RSM is a mathematical

approximation method that employs special experimental design methods in sequential fashion to vary and combine levels for significant interaction effects of variables to bring about an improved condition of target response (Kowalski et al. 2005).

The present study reports the exploration of the sequence of optimization experiments conducted to enhance the fermentative production of a glutaminase near-free poly-peptide with L-asparaginase activity by a new addition to *Aspergillus* species in the quest for low-toxicity L-asparaginases. We are not aware of a prior report on L-asparaginase production by a strain of *Aspergillus candidus*. A description of the spectrum of anti-cancer activity of the therapeutic catalytic poly-peptide and its prospects for safety and stability are also reported.

Materials and Methods

Microorganism

The producing mold, *Aspergillus candidus* UCCM 00117, earlier isolated by Asitok and Ekpenyong (2019) and sharing 100% sequence homology with *Aspergillus candidus* JN-YG-3-2 (MG554228.1), was retrieved from the University of Calabar Collection of Microorganisms, UCCM (www.wfcc.info/ccinfo/collection/by_id/652). The mold was reactivated in glucose-peptone broth for 72 h at room temperature (28 ± 2 °C). Next, the culture was diluted 1000-fold and then plated on Czapek-Dox agar for 72 h at room temperature. Viability and L-asparaginase-producing potential were checked by transferring one agar plug (6 mm) of culture from Czapek-Dox agar onto freshly prepared L-asparagine agar medium composed (g/L) of Na_2HPO_4 6; KH_2PO_4 3; $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.2; $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ 0.05, NaCl 0.5; CaCl_2 0.5; KCl 0.5, 1% asparagine and supplemented with 0.25% phenol red. Development of pink coloration between 36 and 60 h was indicative of retained viability and activity for L-asparagine hydrolysis. The reactivated culture was transferred onto potato dextrose agar (PDA) on Petri dish and maintained in a refrigerator until required for further studies.

One-Factor-at-a-Time (OFAT) Optimization of Variables for Maximum L-Asparaginase Activity

Carbon Source

The preferred extraneous carbon source to drive maximum L-asparaginase activity by the study mold was selected by the traditional one-factor-at-a-time (OFAT) approach. The method involved varying the source of carbon and holding every other parameter constant (Ekpenyong et al. 2017a).

Glucose (GLU), mannose (MAN) and fructose (FRU) as monosaccharides, sucrose (SUC), maltose (MAL) and lactose (LAC) as disaccharides and agro-industrial waste products including molasses (MOL), cassava processing effluent (CPE) and rice processing effluent (RPE) were the carbon substrates screened. Sugars were purchased from Sigma-Aldrich (Sigma-Aldrich Inc., St. Louis, MO, USA) while effluents were obtained from local industries in Lagos and Calabar, Nigeria and stored at -20 °C until required.

The minimal medium for screening contained (g/L) Na_2HPO_4 6; KH_2PO_4 3; $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.2; $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ 0.05, NaCl 0.5; CaCl_2 0.5; KCl 0.5 (Asitok and Ekpenyong 2019) and was supplemented with 1 mL trace mineral solution composed (g/L) of $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ 0.005; $\text{MnSO}_4 \cdot 4\text{H}_2\text{O}$ 0.005; H_3BO_3 0.005; $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ 0.005; $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$ 0.005; MoCl_5 0.005, BaCl_2 0.005; $\text{CrCl}_3 \cdot 6\text{H}_2\text{O}$ 0.005; FeCl_3 0.005 and $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$ 0.005 (Ekpenyong et al. 2017b). The fully-constituted medium was supplemented with 1% (w/v) L-asparagine (Merck) and pH adjusted to 5.8 using 1 N HCl. Medium was dispensed into 250 mL Erlenmeyer flasks at 20% (v/v) and carbon sources (except the effluents) incorporated at 1% (w/v). The effluents were retrieved from the freezer and allowed to thaw under ambient laboratory conditions and subsequently boiled to facilitate removal of un-dissolved solids. Upon cooling, effluent was centrifuged at 10,000 rpm for 20 min and supernatant dispensed as 20% (v/v) into 250 mL Erlenmeyer flasks without adjusting pH (Nitschke et al. 2004). A control flask without extraneous carbon where asparagine served as carbon source was also set up. All arrangements were made in triplicates and flasks sterilized by autoclaving at 121 °C for 15 min.

Upon cooling, one agar plug (6 mm) of reactivated and equilibrated mold culture was aseptically added to each flask. Flasks were plugged with sterile cotton wool and incubated at 30 °C on a rotary shaker agitating at 150 rpm for 72 h. Fermentation broth was first filtered with Whatman No.1 filter paper to remove mycelia and then centrifuged at 8,000 rpm for 15 min. Supernatant was twice filtered with 0.45 μm and 0.22 μm membrane filter (Millipore, USA) and L-asparaginase activity quantified for each flask by the Nesslerization protocol of Imada et al. (1973) using Tris–HCl buffer at pH 7.2. One unit of L-asparaginase activity (U) was defined as the amount of enzyme required to liberate one micromole of ammonia by L-asparagine hydrolysis in one minute under the study conditions. A one-way analysis of variance (one-way ANOVA) in GraphPad Prism 8 software (GraphPad, San Diego, CA, USA) was used to compare performances of the carbon sources and significant mean differences were separated by Tukey HSD test using 95% confidence interval.

Nitrogen Source

Extraneous nitrogen sources were also screened by the OFAT approach. Nitrogen sources included ammonium chloride (NH_4Cl), potassium nitrate (KNO_3), ammonium nitrate (NH_4NO_3), bean processing effluent (BPE), corn steep liquor (CSL), peptone (PEP) and tryptone (TYP). Organic and inorganic nitrogen sources were added at 1% (w/v) to minimal medium supplemented with 1% (w/v) L-asparagine and selected carbon substrate. Effluent substrates were prepared as described in the section for carbon screening and used as also described. A control flask without extraneous nitrogen was also prepared. Experimental set-up, sterilization, inoculation, incubation, harvest, L-asparaginase activity assay and statistical analyses were as described under carbon screening.

Inoculum Size

To select most suitable inoculum size (spore density) for the fermentation, inoculum sizes were screened by the OFAT approach by inoculating 3% (v/v) spore suspension of different inoculum sizes ranging from 10^3 to 10^9 spore-forming units per milliliter (sfu/mL) into minimal medium containing selected extraneous carbon and nitrogen substrates. Working spore concentrations were prepared by the spectrophotometric method as described in Ekpenyong et al. (2020a) and experimental set up, incubations, harvest, determinations of enzyme activity and statistical analyses were as described under carbon screening section.

Metal Ions

Metal ions including Na^+ , K^+ , Mg^{2+} , Ca^{2+} , Zn^{2+} , Mn^{2+} , Ni^{2+} , Cu^{2+} , Co^{2+} , Fe^{2+} , Fe^{3+} , Pb^{2+} , Cr^{3+} and Mo^{5+} were screened for their effects on L-asparaginase production. All metal salts were purchased as chlorides from Sigma-Aldrich except ferrous and magnesium salts which were purchased as hydrated sulfates from Fisher Scientific (Fisher Scientific UK Ltd, Loughborough, UK). A complete minimal medium with all salts in place as detailed under carbon screening section plus selected carbon, nitrogen and inoculum size served as control. Effect of metal ion on L-asparaginase activity was scored on the basis of L-asparaginase activity quantified in the absence of the respective metal. Higher L-asparaginase activity indicated inhibition in the presence of the metal while lower values indicated enhancement of activity. All metal ions were tested at 1 mM concentration and experimental set-up, incubations, harvest and determinations were as described for carbon screening. Effects of metals on L-asparaginase activity were compared by one-way ANOVA and significant means separated by Dunnett test comparing metal performance to the control at 95% confidence interval.

Significant reduction in L-asparaginase activity selected respective metal for further studies.

Plackett–Burman Design Screening of Selected Variables

The Plackett–Burman design (PBD) was employed to screen a combination of 12 major and minor nutrients obtained from OFAT experiments to develop an appropriate fermentation medium for production of L-asparaginase with maximum enzyme activity, Y . This 2^2 factorial design completely ignores possible two-way factor interactions but focuses on their main effects on the response(s). The variables included in this study design were molasses (MOL), corn steep liquor (CSL), inoculum volume (INV), asparagine (ASP), Na^+ , K^+ , Mg^{2+} , Zn^{2+} , Mn^{2+} , Fe^{2+} , Co^{2+} and Cr^{3+} . All 12 factors were tested at 2 levels; high (+1) and low (−1) and the design made with MINITAB 17 (MiniTab Inc, Pennsylvania, USA). The design comprised 25 experimental runs having been augmented with 5 center points to enable estimation of experimental error and test the fit of the first-order model. The factor names and their levels as employed in PBD are presented in Table DT5 of Mendeley Data (Ekpenyong et al. 2020b). Only predictors with significant ($p < 0.05$) main effects selected by backward elimination were employed to build the first-order regression model for further studies. The general fitted first-order model developed by least squares method is given as Eq. 1:

$$y = \beta_0 + \sum_{i=1}^k \beta_i x_i + \varepsilon \quad (1)$$

where y is the predicted response, β_0 the intercept, β_i the coefficients of predictors (significant) x_i , $i = 1, 2, \dots, k$.

Path of Steepest Ascent (PSA) Experimentation

The path of steepest ascent (PSA) is an intermediary gradient protocol in RSM optimization experiment employed to move identified significant predictors of a process or system close to the optimum through sequential experimentation (Kowalski et al. 2005). The goal of the PSA is to identify a new operating region for significant process improvement. It finds a suitable approximation in the first-order model from the PBD which provides the coefficients of significant factors to direct operating region towards PSA subject to a given distance ρ , from the center point.

Since a one unit (using coded units) steepest ascent point away from the center point is desired to maximize the response, y , the coordinates of the significant factor levels were determined as follows:

- i. The slope of the model was calculated using the largest absolute regression coefficient, β_j in the model and denoted Δx_j as step size 1
- ii. Other step sizes were then calculated using Eq. 2 as follows:

$$\Delta x_i = \frac{\beta_i}{(\beta_j/\Delta x_j)} \quad i = 1, 2, \dots \quad ki \neq j \quad (2)$$

- iii. Then, the coded step sizes were converted to natural levels using the relationship between coded and natural levels as given in Eq. 3

$$dX_i = S_i(\rho) \quad i = 1, 2, \dots \quad ki \neq j \quad (3)$$

where dX_i is the change in natural level of predictor variables, Δx_i change in coded level of predictor variables, S_i the slope with respect to each predictor and ρ the natural level change (proportion) in the predictor, j , with largest absolute coefficient, β .

Therefore, using the fitted first-order model, the PSA approach moved levels of significant predictors away from the original design center along the path of steepest ascent towards maximum L-asparaginase activity. The levels of significant variables in the PSA experiment that yielded the highest L-asparaginase activity were employed as center points in RSM experiment.

Response Surface Modeling of Significant Predictors for Maximum L-Asparaginase Activity

The matrix for the response surface method (RSM) of optimization was obtained through central composite rotatable design (CCRD) using Design Expert 12 software (Stat Ease Inc., Minnesota, USA), with each numerical factor evaluated at 5 levels. Based on the number of significant factors selected by PBD, a special kind of fractional factorial design called Min-Run-Res V type factorial design was adopted for RSM. The design comprised 6 center points for lack-of-fit (LoF) test and 34 non-center points made up of 12 axial (-1.56508 and $+1.56508$) and 22 factorial (-1 and $+1$) points; making a total of 40 experimental runs. The predictors were coded as x_1, x_2, x_3, x_4, x_5 and x_6 corresponding to molasses, corn steep liquor, asparagine, potassium, manganese and chromium. Molasses was composed (%) of dry solids 82.1; Sucrose 50.7; Invert sugar 0.4, Raffinose 1.1; Ash 12.0; Total nitrogen 1.7; Purity quotient of 63.1 and pH of 7.5. Corn steep liquor was composed, in percent weight (wt%), of crude protein 47.2; ash (oxide) 15.3; lactic acid 24.8, phytic acid 7.8, fat 0.33; reducing sugar 2.8, water 32, pH 4.1. The actual levels of the factors were varied based on the steepest ascent steps earlier developed in the PSA

section as follows: $X_1 = (44.61)/4.935$, $X_2 = (67.97)/2.995$, $X_3 = (40.05)/4.175$, $X_4 = (1.88)/0.146$, $X_5 = (50)/5.000$, $X_6 = (47.42)/4.57$. Details are as presented in Table DT9 of the Mendeley Dataset (Ekpenyong et al. 2020b).

The requirement to report specific activity of the enzyme and the yield coefficient of the bioprocess made the inclusion of biomass concentration (g/L) and total protein (mg) as additional response variables necessary, at this stage of the study. Therefore, three response variables were determined in the RSM experiments namely biomass concentration (Y_1), total protein (Y_2) and L-asparaginase activity (Y_3). Biomass concentration was determined by the conventional oven dry weight technique (Banerjee et al. 1993) while total protein and L-asparaginase activity were determined by the methods of Bradford (1976) and Imada et al. (1973) respectively. All 40 completely randomized experiments were conducted in triplicates and results reported as mean responses. Data was subjected to multiple regression analysis using least squares to build/fit models. The scalar notation of the general quadratic function for RSM is given below;

$$Y = \beta_0 + \sum_{i=1}^k \beta_i x_i + \sum_{i=1}^k \beta_{ii} x_i^2 + \beta_{ij} x_i x_j + \epsilon \quad (4)$$

where y is the response, $\beta_0, \beta_i, \beta_{ii}$ and β_{ij} are the regression coefficients for the constant, linear (x_i), quadratic (x_i^2) and interaction terms ($x_i x_j$) of the k factors respectively and ϵ the error term of response variable computation. Only significant ($p < 0.05$) predictors of each response was included in the final model. Adequacy of models was evaluated by examining model diagnostic plots, adjusted and predicted r^2 values, LoF F and p -values and percent coefficient of variation (%CV). Significant model predictors were evaluated by t -test of their coefficients and model curvatures presented as contour (2-D), surface (3-D) plots.

Multi-Objective Optimization of Response Variables

Since the three responses of biomass concentration (Y_1), total protein (Y_2) and L-asparaginase (Y_3) were measured during the same batch fermentation set-up under the same conditions; mono-objective optimization of responses became impracticable. This research therefore undertook a multi-objective optimization approach of the responses using the desirability function provided by response optimizer in Design Expert. The major thrusts of the research were to maximize specific activity of L-asparaginase (U/mg); calculated as the ratio of L-asparaginase activity to the amount of total protein in the culture supernatant, as well as its yield coefficient ($Y_{p/x}$) calculated as ratio of L-asparaginase concentration to biomass concentration and expressed

as g/g. To achieve this therefore, a medium formulated to maximize L-asparaginase activity while minimizing biomass concentration and total protein would be ideal. The desirability function is a mathematical protocol that recommends a set of operational conditions that meet all set goals with the best desirability (Sperb et al. 2018). The composite desirability of a multi-response function was defined as the geometric mean of the desirability of individual responses and is given by the expression;

$$D = (d_1 \times d_2 \times d_3 \times \dots \times d_n)^{\frac{1}{n}} = \left(\prod_{i=1}^n d_i \right)^{\frac{1}{n}} \quad (5)$$

where D is the composite desirability, n is the number of responses, d_1 , d_2 , d_3 and d_n are desirability for individual responses. The equation could be weighted to reflect the importance of the responses in the desirability measure where the weight, w_i satisfies the function $0 < 1 < w_i$, with $w_1 + w_2 + w_3 + \dots + w_n = 1$, as follows:

$$D = (d_1^{w_1} \times d_2^{w_2} \times d_3^{w_3} \times \dots \times d_n^{w_n})^{\frac{1}{n}} \quad (6)$$

Confirmation Experiments for RSM Multi-objective Optimization

Triplicate experiments were set up in 1 L Erlenmeyer flasks containing media (20% v/v) formulated according to significant factor settings recommended by the multi-objective numerical optimization. The three responses were measured as earlier described and specific L-asparaginase activity and yield coefficient calculated. Results from confirmation experiments were compared with those obtained in the numerical optimization and a less than 5% difference in L-asparaginase activities was accepted as confirmatory for optimal settings.

Fractionation of L-Asparaginase by Ammonium Sulfate

The sterile crude L-asparaginase broth was gradually brought to 40% saturation with ammonium sulfate (enzyme grade) and the precipitate removed by centrifugation at 12,000 rpm for 10 min. The resulting supernatant was then brought to 80% saturation with the salt and further purification conducted as described in Asitok and Ekpenyong (2019). The resulting fraction was dialyzed against 0.05 M Tris–HCl buffer (pH 7.2) at 4 °C for 24 h with 12 changes. Total protein (Bradford 1976), L-asparaginase and glutaminase activities (Imada et al. 1973) were determined from the concentrated dialyzed supernatant.

DEAE-Cellulose Column Purification

The dialyzed enzyme fraction obtained from the fractionation experiment was loaded into a di-ethyl-amino-ethyl (DEAE)-cellulose 10 cm column (Sigma-Aldrich) and further purification conducted as described by Sahu et al. (2007). Total protein and activities of L-asparaginase and glutaminase were determined from the pooled concentrated dialyzed fraction as described earlier.

Gel Filtration with Sephadex G-200

The concentrated fraction of the crude enzyme from the cellulose column purification step was applied onto Sephadex G-200 gel and further purification performed as described in Husain et al. (2016). The eluted fractions were pooled into a composite sample, dialyzed as earlier described and concentrated with 0.05 M sucrose and protein concentration, L-asparaginase and L-glutaminase activities of the partially-purified enzyme determined as described earlier.

In-Vitro Anti-cancer Activities of *Aspergillus candidus* UCCM-00117 L-Asparaginase

The in vitro anti-cancer activity of the partially-purified L-asparaginase in this study was investigated using the MTT-based cytotoxicity assay of Skehan et al. (1990). Human myeloid leukemia (HL-60), colorectal carcinoma (HCT-116), cervical epithelial carcinoma (HeLa), breast cancer (MCF-7) and hepatocellular carcinoma (HepG-2) cell lines were employed to evaluate the therapeutic potentials of the enzyme. Toxicity to normal cells was tested against non-tumor human embryonic cell line (HEK 283 T) to evaluate the selective toxicity of the enzyme. Enzyme dosages were two-fold dilutions from enzyme stock solution ranging from 1.148 to 1174.898 µg/mL. *Mycoplasma* sterility tests, viability and subsequent cytotoxicity studies using the sulforhodamine B (SRB) assay were as described in Asitok and Ekpenyong (2019) and Ekpenyong et al. (2020a). Experiments were conducted in triplicates and results, expressed as % cell viability, were presented as means of triplicate determinations. Cell viability was calculated using the relationship below:

$$\%Cellviability = \left(\frac{A_{570T}}{A_{570C}} \right) \times 100 \quad (7)$$

where A_{570T} is the absorbance at 570 nm of cells treated with L-asparaginase and A_{570C} is the absorbance at 570 nm

of untreated (control) cells. Data was analyzed by non-linear regression using the sigmoidal four-parameter logistic (4PL) model presented as Eq. 8.

$$y = d + \frac{a - d}{1 + \left(\frac{x}{c}\right)^b} \quad (8)$$

where x is the independent variable and y , the dependent variable; the 4 estimated parameters include a ; the minimum value that can be obtained (i.e. what happens at 0 dose), d ; the maximum value that can be obtained (i.e. what happens at infinite dose), c ; the point of inflection (i.e. the point on the sigmoidal curve half-way between a and d) called IC_{50} or EC_{50} and b ; the Hill's slope of the curve.

Selectivity index (SI) of the enzyme was calculated as a ratio of IC_{50} of non-tumor cell line to those of cancer cell lines and data was analyzed by two-way ANOVA.

Evaluation of Temperature, pH and Metal Ion Effects on L-Asparaginase Activity and Stability

The temperature and pH effects on L-asparaginase activity earlier reported in Asitok and Ekpenyong (2019) were repeated to confirm the optimum temperature (5–70 °C) and pH (3–12) for maximum activity of the enzyme. Reported values were means of triplicate determinations \pm standard deviation. One way analysis of variance (one-way ANOVA) was used to compare data and significant mean differences were separated by Tukey HSD at $p < 0.05$. Furthermore, the thermal stability of the enzyme was evaluated by first exposing the enzyme in 50 mM Tris–HCl buffer (pH 7.5) to different temperatures including 40, 50, 60, 70 and 80 °C for 0, 15, 30, 45, 60, 75, 90 min. The enzyme was allowed to refold on ice for 15 min and then residual activity determined. Similarly, stability of the enzyme to increasing pH levels was evaluated by incubating enzyme in different buffers of varying pH values (Iboyo et al. 2017) ranging from 4.5 to 10.5 at 4 °C for 0, 6, 12, 18, 24 and 30 h. Residual activity of the enzyme was again determined after incubation as described for thermal stability. Data for enzyme stability studies were analyzed by two-way ANOVA in GraphPad Prism 8 to compare significant means and identify possible interactions at $p < 0.05$.

Inhibition or enhancement of enzyme activity by metal ions was investigated by incubating the enzyme reaction mixture with 1 mM concentration of each metal ion (Na^+ , K^+ , Mg^{2+} , Ca^{2+} , Ba^{2+} , Zn^{2+} , Ni^{2+} , Fe^{2+} , Fe^{3+} , Cr^{3+} , Mn^{2+} , Co^{2+} , Cu^{2+} and Mo^{5+}). Relative activities were calculated using assay without metal as control. One-way analysis of variance was used to compare data for significant mean

differences at $p < 0.05$. Range finding experiments for significant activity enhancing metal ions were conducted using 0.5, 1.0, 1.5, 2.0, 2.5 and 3.0 mM concentration. All experiments were conducted in triplicates and relative activities calculated and reported as mean value \pm standard deviation.

Results and Discussion

One-Factor-at-a-Time (OFAT) Experiments

Frequently in microbial fermentations, the carbon substrate that drives cellular metabolism differs from the one that enhances target metabolite synthesis (Abalos et al. 2002). Among the 9 carbon substrates screened by OFAT, only sucrose, cassava processing effluent and molasses significantly ($p < 0.05$) enhanced L-asparaginase activity above control (Fig. 1a). The Tukey's multiple comparisons test and its 95% confidence interval plot (Fig. DF1 in Mendeley Data: Ekpenyong et al. 2020b) showed that molasses was the best ($p < 0.0001$) of all substrates tested with L-asparaginase activity of 1281 ± 32.71 U against 933.0 ± 20.02 U in the control. Selection of molasses as most suitable carbon substrate for microbial production of value-added metabolites is not new. Asitok and Ekpenyong (2019) reported selection of sucrose, the dominant carbohydrate in molasses, as best carbon substrate for the mold. Ebeed and Aboutaleb (2014) had earlier reported the preference of black strap molasses as carbon source for L-asparaginase production by *Acinetobacter radioresistance* PS14. Production of L-asparaginase on agro-industrial wastes have been suggested as a sustainable approach towards improved production economics of biotechnological products.

The OFAT experiment to select appropriate extraneous nitrogen source for improved L-asparaginase activity using molasses as extraneous carbon substrate revealed that tryptone (1387 ± 62.18 U) and corn steep liquor (1602 ± 93.34 U) were the only nitrogen sources which significantly improved the activity of the enzyme (Fig. 1b). Corn steep liquor was selected with a slight edge over tryptone ($p = 0.0188 < 0.05$) as best enhancing nitrogen source while urea significantly ($p < 0.0001$) inhibited L-asparaginase activity, allowing only an activity of 873.8 ± 90.87 U. Inhibition of L-asparaginase activity by urea had earlier been reported by Li et al. (2018) to proceed through sustained denaturation of thermo-stable L-asparaginase from *Pyrococcus yanosii* CH1 expressed in *Bacillus subtilis* 168. Production of L-asparaginase by *Aspergillus niger* on corn steep liquor had earlier been reported by Zia et al. (2013). One of the remarkable things about using agro-industrial wastes as substrates for fermentation is the multiple roles they can play. In the present study it served as nitrogen source in combination with asparagine

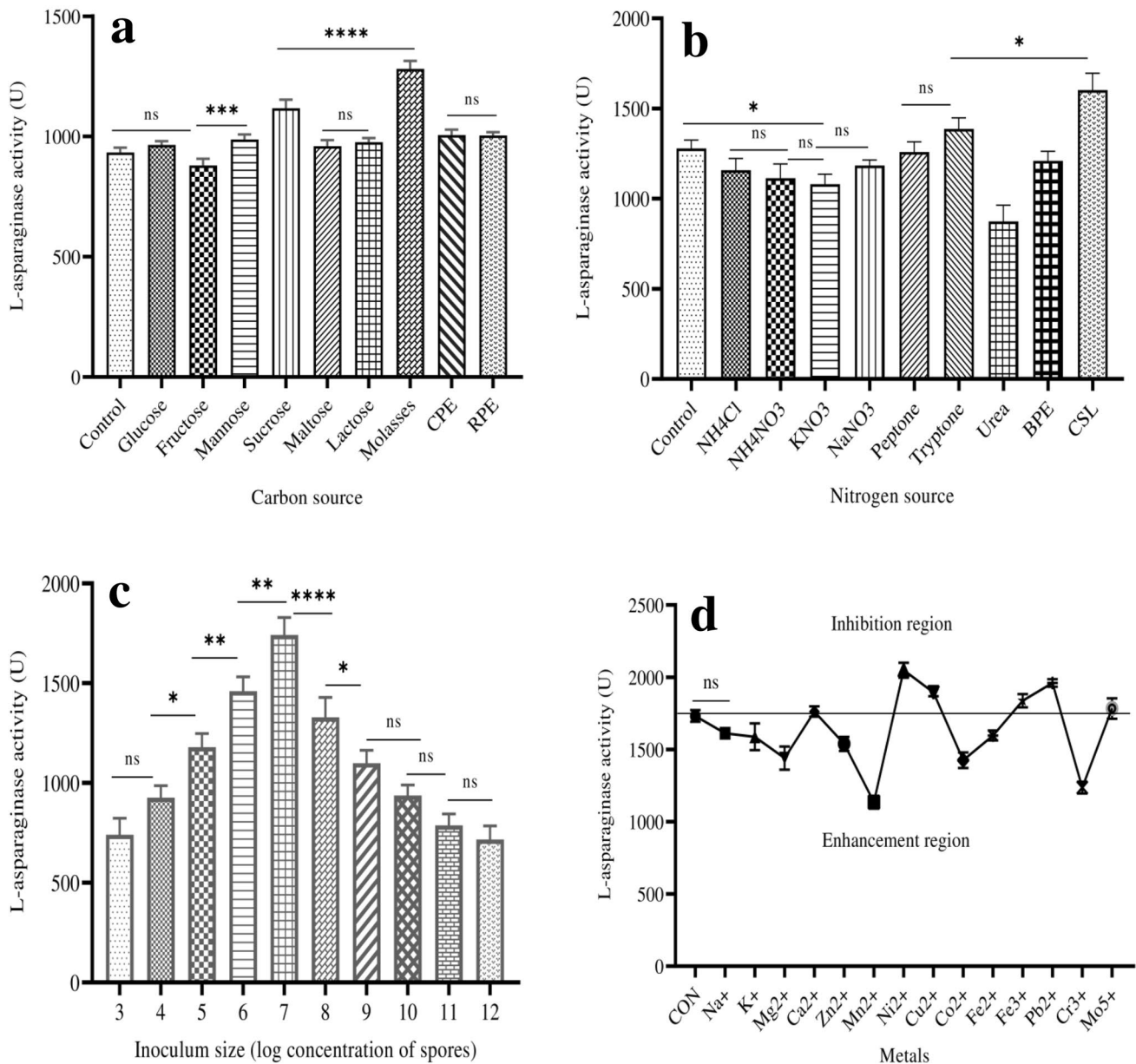


Fig. 1 One-factor-at-a-time (OFAT) plots to select most enhancing **a** carbon substrate **b** nitrogen source **c** spore density **d** metal ions for enhanced L-asparaginase production

and carbon source in combination with beet molasses and asparagine.

The results presented as Fig. 1c show that L-asparaginase activity increased with increasing spore density from 740.4 ± 82.35 U at 10^3 spore forming units per milliliter (sfu/mL) to 1742 ± 87.16 U when spore density was 10^7 sfu/mL. Tukey multiple comparisons test of a one-way ANOVA revealed that L-asparaginase activity reported at 10^7 sfu/mL was significantly higher than those at 10^6 sfu/mL (1460 ± 72.01 U) below it and 10^8 sfu/mL (1329 ± 99.98 U) above it. A similar range of inoculum size was reported by

Zia et al. (2013) for L-asparaginase production by *Aspergillus niger*.

Finally, out of 14 metal ions screened by OFAT for their effects on L-asparaginase activity, results, presented as Fig. 1d, show that L-asparaginase activity reduced from 1732 ± 39.76 U in the control to 1135 ± 44.45 U in the absence of Mn^{2+} and 1237 ± 41.00 U in the absence of Cr^{3+} suggesting significant ($p < 0.0001$) requirement for these ions in the fermentation medium. However, L-asparaginase activity significantly increased to 2049 ± 51.83 U in the absence of Ni^{2+} and a little less in the absence of

Pb²⁺, Cu²⁺ and Fe³⁺ suggesting significant inhibition of L-asparaginase production or activity in their presence. This is the first report on L-asparaginase activity enhancement by trivalent chromium. However, among all metals studied by El-Naggar et al. (2018) on their effects on *Streptomyces brolosae* NEAE-115 L-asparaginase, maximal enzyme activity was observed in the presence of manganese. Dunnett's multiple comparisons test showed that incorporation or non-incorporation of Na⁺, K⁺ and Mo⁵⁺ ions did not significantly influence L-asparaginase activity. The dataset for these experiments is available at Mendeley Data (Ekpenyong et al. 2020b). The report by El-Naggar et al. (2018) also supports the inhibitory effects of nickel and trivalent iron on L-asparaginase production.

Plackett–Burman Designed Experiments

In this study, PBD was applied to screen 12 factors from the OFAT experiments and the design matrix with the experimental and predicted responses of all combinations of factors is presented in Table 1. Highest experimental and predicted L-asparaginase activities of 1948.86 and 1974.0U respectively, were obtained with the factor combination in run 22 highlighted in yellow. The full regression model is presented as Eq. 9 and incorporates all 12 predictors; significant and non-significant. Their corresponding coefficient table with *t*-test result has been published as dataset in Mendeley Data (Ekpenyong et al. 2020b).

Table 1 Design matrix of coded levels of factors in Plackett–Burman design screening for L-asparaginase production by *Aspergillus candidus* UCCM 00117 and their experimental and predicted responses

Run	MOL	CSL	INV	ASP	Na+	K+	Mg2+	Zn2+	Mn2+	Co2+	Fe2+	Cr3+	eASPase act	pASPase act
1	-1	1	1	-1	-1	-1	-1	1	-1	1	-1	1	1555.31	1577.51
2	-1	1	-1	1	-1	1	1	1	1	-1	-1	1	1887.96	1919.39
3	1	-1	1	1	-1	-1	-1	-1	1	-1	1	-1	1784.72	1762.58
4	1	1	-1	-1	-1	-1	1	-1	1	-1	1	1	1883.89	1858.46
5	-1	1	1	-1	1	1	-1	-1	-1	-1	1	-1	1563.58	1530.53
6	-1	-1	1	1	-1	1	1	-1	-1	-1	-1	1	1701.55	1693.25
7	1	1	-1	-1	1	1	-1	1	1	-1	-1	-1	1836.29	1811.48
8	1	-1	-1	1	1	-1	1	1	-1	-1	-1	-1	1639.74	1621.16
9	1	1	1	-1	-1	1	1	-1	1	1	-1	-1	1798.46	1811.48
10	0	0	0	0	0	0	0	0	0	0	0	0	1707.28	1711.21
11	1	-1	1	-1	1	1	1	1	-1	-1	1	1	1694.84	1714.64
12	0	0	0	0	0	0	0	0	0	0	0	0	1693.29	1711.21
13	0	0	0	0	0	0	0	0	0	0	0	0	1652.21	1711.21
14	-1	1	1	1	1	-1	-1	1	1	-1	1	1	1838.88	1837.08
15	1	1	-1	1	1	-1	-1	-1	-1	1	-1	1	1887.85	1835.18
16	1	-1	-1	-1	-1	1	-1	1	-1	1	1	1	1733.84	1714.64
17	-1	-1	1	-1	1	-1	1	1	1	1	-1	-1	1484.75	1504.92
18	1	1	1	1	-1	-1	1	1	-1	1	1	-1	1654.95	1705.88
19	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	1344.81	1363.49
20	0	0	0	0	0	0	0	0	0	0	0	0	1678.49	1711.21
21	-1	-1	-1	-1	1	-1	1	-1	1	1	1	1	1679.33	1634.21
22	1	-1	1	1	1	1	-1	-1	1	1	-1	1	1948.86	1974.2
23	-1	-1	-1	1	-1	1	-1	1	1	1	1	-1	1729.78	1705.38
24	-1	1	-1	1	1	1	1	-1	-1	1	1	-1	1682.24	1648.67
25	0	0	0	0	0	0	0	0	0	0	0	0	1717.25	1711.21

MOL Molasses, CSL Corn steep liquor, INV inoculum volume, ASP Asparagine; Na⁺ = NaCl; K⁺ = KCl; Mg²⁺ = MgSO₄·7H₂O; Zn²⁺ = ZnCl₂; Mn²⁺ = MnCl₂; Co²⁺ = CoCl₂; Fe²⁺ = FeSO₄·7H₂O; Cr³⁺ = CrCl₃·6H₂O. Coded values are low (-), high (+) and center points (0) to facilitate determination of lack-of-fit of the model; eASPase act = experimental L-asparaginase activity; pASPase act = predicted L-asparaginase activity. Values of L-asparaginase activity are means of triplicate determinations

$$\hat{Y} = 1711.21 + 69.76MOL + 42.36CSL - 13.99INV + 59.07ASP + 9.05Na^+ + 41.16K^+ - 5.81Mg^{2+} - 10.95Zn^{2+} + 70.71Mn^{2+} - 1.04Co^{2+} + 8.02Fe^{2+} + 64.65Cr^{3+} \quad (9)$$

where, \hat{Y} = predicted L-asparaginase activity, MOL = molasses, CSL = corn steep liquor, INV = inoculum volume, ASP = asparagine, Na^+ = sodium ions, K^+ = potassium ions, Mg^{2+} = magnesium ions, Zn^{2+} = zinc ions, Mn^{2+} = manganese (II) ions, Co^{2+} = cobalt (II) ions, Fe^{2+} = Iron (II) ions and Cr^{3+} = chromium (III) ions. The model was significant with an r^2 of 0.9747, adjusted r^2 of 0.9493, predicted r^2 of 0.8785, non-significant lack-of-fit test of $F(8, 12) = 1.64$, $p = 0.333 > 0.05$. However, to achieve the primary objective of PBD viz screening, the backward elimination method was adopted to reduce the model to include only significant ($p < 0.05$) predictors as in Eq. 10 below:

$$\hat{Y} = 1711.21 + 69.76MOL + 42.36CSL + 59.07ASP + 41.16K^+ + 70.71Mn^{2+} + 64.65Cr^{3+} \quad (10)$$

The reduced model was also significant ($p < 0.0001$) at $F = 59.73$, r^2 of 0.9522, adjusted r^2 of 0.9362, predicted r^2 of 0.9037, a non-significant lack-of-fit test of $F(14, 18) = 2.02$, $p = 0.260 > 0.05$. The ANOVA table for the modified model is presented as Table 2 and shows that Mn^{2+} and molasses made the most contributions towards enhanced L-asparaginase activity as indicated by their large F -values of 85.22 and 82.95 respectively. A careful examination of the predicted r^2 values reveal that the reduced model gives better prediction of L-asparaginase activity (pred. $r^2 = 90.37\%$) than the full model (pred. $r^2 = 87.85\%$). The diagnostic plots for the second model are given as Fig. 2. The Pareto chart (Fig. 2a) confirms the selection of the six predictors in

Eq. 9 as important variables for L-asparaginase production while Fig. 2b reveals their levels of importance through main effects plot. The residual plots (Fig. 2c) confirm the adequacy of the reduced model to explain the events leading to the activity of L-asparaginase observed during fermentation.

Path of Steepest Ascent Experiments

Results of the path of steepest ascent (PSA) experiments are reported in Table 3 and Fig. 3. The table summarizes the steps taken to arrive at the steepest ascent step sizes used to develop the steepest ascent experiments. The primary objective of the PSA was to identify plausible center points for the predictors suggested by the PBD first-order model for use in RSM. Table 3 assigned codes to the six predictors as X_1 , X_2 , X_3 , X_4 , X_5 and X_6 to correspond to molasses, corn steep

liquor, asparagine, K^+ , Mn^{2+} and Cr^{3+} respectively. In consideration of the predictor coefficients in Eq. 9, the predictors were varied by factors of 4.935 g/L, 2.995%, 4.175 g/L, 0.146 g/L, 5 mg/L and 4.57 mg/L respectively. With these step changes, experiment 7 yielded the highest L-asparaginase activity of 3272 ± 42.64 U beyond which the activity took a descent (Fig. 3). The path of steepest ascent clearly identified the values of predictors which should form the center points in RSM as $(X_1, X_2, X_3, X_4, X_5, X_6) = (44.61, 67.97, 40.05, 1.876, 50, 47.42) = (0, 0, 0, 0, 0, 0)$.

Table 2 Analysis of variance of significant predictors of the modified first-order model for L-asparaginase activity from Plackett–Burman design (PBD) screening

Source	Degrees of Freedom	Adjusted sum of Squares	Adjusted mean square	F-value	p-value
Regression	6	420,483	70,080	59.73	0.000
Molasses	1	97,336	97,336	82.95	0.000
Corn steep liquor	1	35,887	35,887	30.58	0.000
Asparagine	1	69,789	69,789	59.48	0.000
Potassium ions	1	33,880	33,880	28.87	0.000
Manganese ions	1	99,999	99,999	85.22	0.000
Chromium (III) ions	1	83,891	83,891	71.24	0.000
Error	18	21,121	1173		
Lack-of-Fit	14	18,509	1322	2.02	0.26
Pure Error	4	2612	653		
Total	24	441,603			

$S = 34.25$; $r^2 = 95.22\%$; Adjusted $r^2 = 93.62\%$; Predicted $r^2 = 90.37\%$

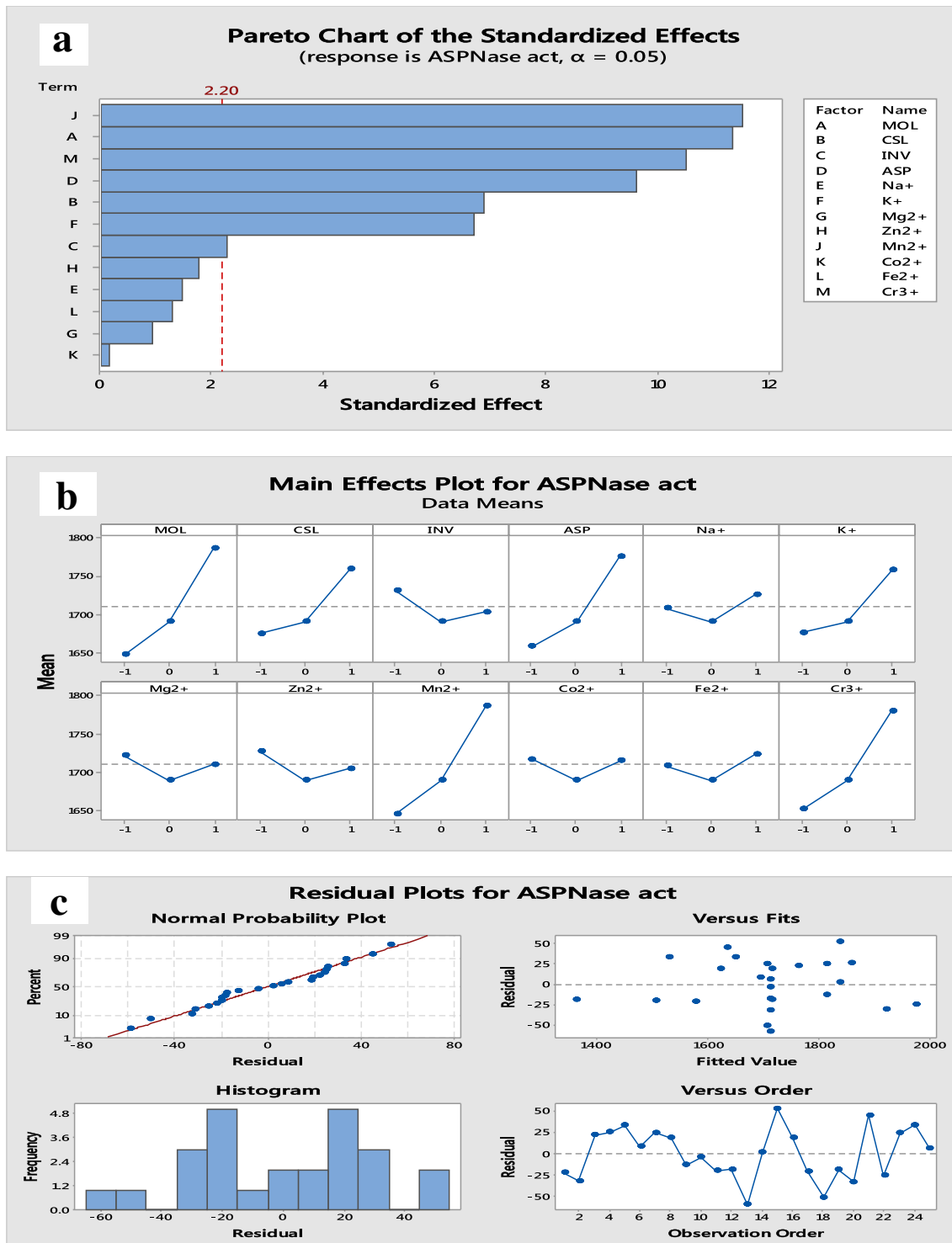


Fig. 2 Plackett–Burman design **a** Pareto plot showing significant factor selection. **b** Main effects plot of each factor contribution. **c** Diagnostic plots to test adequacy of the first-order regression model for significant predictors towards L-asparaginase production

Table 3 Path of steepest ascent (PSA) optimization of significant predictors for enhanced L-asparaginase activity during fermentation by *Aspergillus candidus* UCCM 00117

	x_1	x_2	x_3	x_4	x_5	x_6	MOL ^a	CSL ^b	ASP ^c	K ⁺ ^d	Mn ²⁺ ^e	Cr ³⁺ ^f	Mean L-ASPase act ± SE ^l
Base point (origin) ^g	0	0	0	0	0	0	15	50	15	1	20	20	
Original step unit ^h							5	5	5	0.25	5	5	
Coefficients, β ⁱ	69.76	42.36	59.07	41.16	70.71	64.65	X ₁	X ₂	X ₃	X ₄	X ₅	X ₆	
Steepest ascent step distance, Δ	0.987 ^j	0.599 ^j	0.835 ^j	0.582 ^j	1 ^j	0.914 ^j	4.935 ^k	2.995 ^k	4.175 ^k	0.146 ^k	5 ^k	4.57 ^k	
Exp.1 (Origin)	0	0	0	0	0	0	15	50	15	1	20	20	1683 ± 34.46
Exp. 2 (Origin + Δ^m)	0.987	0.599	0.835	0.582	1	0.914	19.935	52.995	19.175	1.146	25	24.57	1987 ± 55.15
Exp. 3 (Origin + 2 Δ)	1.974	1.198	1.67	1.164	2	1.828	24.87	55.99	23.35	1.292	30	29.14	2262 ± 41.14
Exp. 4 (Origin + 3 Δ)	2.961	1.797	2.505	1.746	3	2.742	29.805	58.985	27.525	1.438	35	33.71	2495 ± 51.16
Exp. 5 (Origin + 4 Δ)	3.948	2.396	3.34	2.328	4	3.656	34.74	61.98	31.7	1.584	40	38.28	2659 ± 36.98
Exp. 6 (Origin + 5 Δ)	4.935	2.995	4.175	2.91	5	4.57	39.675	64.975	35.875	1.73	45	42.85	2919 ± 40.50
Exp. 7 (Origin + 6 Δ)	5.922	3.594	5.01	3.492	6	5.484	44.61	67.97	40.05	1.876	50	47.42	3272 ± 42.64
Exp. 8 (Origin + 7 Δ)	6.909	4.193	5.845	4.074	7	6.398	49.545	70.965	44.225	2.022	55	51.99	2769 ± 55.10
Exp. 9 (Origin + 8 Δ)	7.896	4.792	6.68	4.656	8	7.312	54.48	73.96	48.4	2.168	60	56.56	2362 ± 47.69

Exp. Experiment, L-ASPase act L-asparaginase activity, $x_1 - x_6$ model codes for predictors, X₁ – X₆ Actual predictor codes

N/B: ^aMolasses; ^bCorn steep liquor; ^cAsparagine; ^dPotassium ions; ^eManganese (II) ions; ^fChromium (III) ions; ^gPredictor levels at center points in PBD; ^hOriginal step units of predictors in the PBD matrix; ⁱCoefficients of significant predictors in the first-order model from PBD; ^jSteepest ascent coded unit step distance; ^kcoded ascent natural unit step distance; ^lStandard error of means of triplicate determinations of L-asparaginase activity; ^msteepest ascent step change

Fig. 3 Path of steepest ascent (PSA) experiments plot to maximize L-asparaginase activity

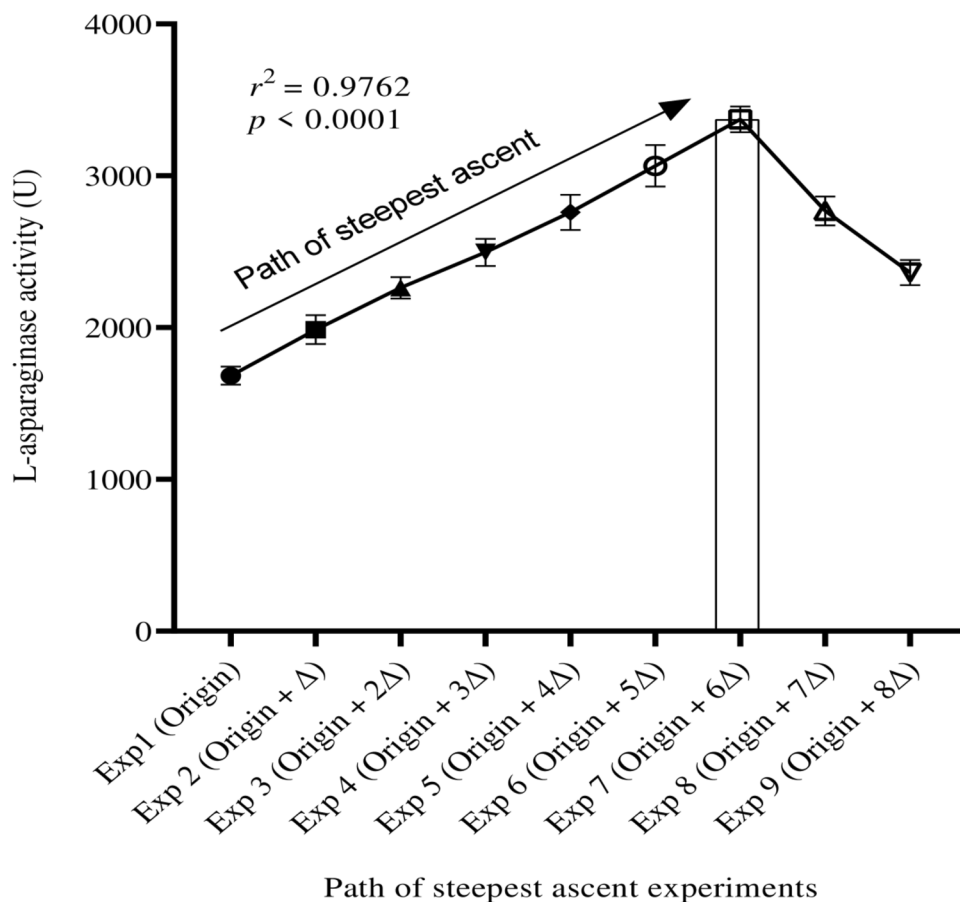


Table 4 Design codes, experimental and predicted values of responses of a surface methodology

Run	x_1	x_2	x_3	x_4	x_5	x_6	eY_1	pY_1	eY_2	pY_2	eY_3	pY_3
1	1	-1	-1	-1	1	1	355.63	355.47	224.56	224.52	4924.78	4927.74
2	-1.565	0	0	0	0	0	351.92	352.46	236.26	237.25	3198.57	3194.52
3	-1	1	1	1	1	1	350.24	350.13	227.87	227.82	3974.55	3978.16
4	-1	-1	1	1	-1	1	345.23	345.14	222.13	222.01	3327.49	3331.28
5	1	1	1	-1	-1	1	387.91	387.34	265.39	265.11	3815.84	3825.71
6	0	0	0	0	0	0	361.73	363.88	255.23	255.32	3294.84	3290.22
7	1	1	1	1	-1	-1	358.64	358.11	227.45	227.17	3724.18	3727.87
8	1	-1	1	-1	-1	-1	349.29	348.7	221.86	221.38	4235.27	4239.41
9	0	0	0	0	0	0	365.34	363.88	255.2	255.32	3263.64	3290.22
10	-1	-1	-1	1	-1	-1	347.23	347.15	218.27	218.07	3086.24	3084.91
11	1.565	0	0	0	0	0	375.22	376.38	259.22	259.55	3698.59	3685.13
12	1	1	1	-1	-1	-1	358.23	359.24	239.77	240.5	3628.46	3622.72
13	0	0	0	0	0	0	363.38	363.88	256.48	255.32	3264.89	3290.22
14	0	0	0	0	-1.565	0	356.27	357.19	235.06	236.01	3258.37	3249.51
15	-1	1	-1	1	1	-1	338.22	338.12	223.55	223.42	3175.44	3173.92
16	1	1	1	-1	1	-1	352.38	351.74	230.67	230.34	4235.47	4239.25
17	0	0	0	0	0	0	365.37	363.88	257.54	255.32	3284.46	3290.22
18	0	0	1.565	0	0	0	348.77	349.87	234.06	234.8	3958.84	3942
19	-1	1	1	-1	-1	-1	348.23	347.72	220.92	220.26	3799.05	3800.42
20	0	0	0	0	1.565	0	350.05	350.83	235.82	236.19	3945.74	3937.08
21	0	0	0	0	0	1.565	371.27	371.91	260.21	260.48	3523.57	3503.22
22	0	-1.565	0	0	0	0	358.37	359.06	245.01	245.66	3684.53	3675.18
23	-1	-1	-1	-1	1	-1	339.12	338.99	213.99	213.76	3351.78	3350.49
24	0	0	0	-1.565	0	0	360.28	361.4	245.25	246.3	3874.73	3865.7
25	1	1	-1	1	1	1	380.83	380.7	257.32	257.38	3452.85	3455.58
26	1	-1	-1	1	-1	1	368.34	368.23	246.92	246.9	3174.18	3177.1
27	1	1	-1	-1	-1	-1	348.39	347.85	233.55	233.11	3012.75	3012.37
28	-1	-1	-1	-1	-1	1	346.37	346.27	225.5	225.3	3148.66	3150.42
29	-1	-1	1	1	1	-1	335.24	335.11	218.37	218.22	3975.12	3975.87
30	0	0	0	0	0	-1.565	346.36	347.42	233.43	234.48	3296.38	3299.21
31	-1	-1	1	-1	1	1	337.83	337.69	225.45	225.3	4869.77	4873.61
32	1	-1	-1	1	1	-1	362.98	362.84	244.43	244.39	3429.48	3429.35
33	-1	1	-1	-1	1	1	358.52	358.4	225.74	225.61	3321.11	3322.69
34	0	0	-1.565	0	0	0	351.29	351.89	239.46	240.03	2735.98	2735.3
35	0	0	0	1.565	0	0	366.25	366.83	253.11	253.38	3346.24	3337.75
36	-1	1	-1	1	-1	1	362.54	362.47	235.13	235.03	3242.57	3244.1
37	0	0	0	0	0	0	366.28	363.88	256.45	255.32	3326.45	3290.22
38	0	1.565	0	0	0	0	371.37	372.37	255.22	255.89	3305.22	3297.06
39	1	-1	1	1	1	1	361.45	361.3	240.1	240.14	4632.14	4637.14
40	0	0	0	0	0	0	367.21	363.88	255.71	255.32	3244.83	3290.22

x_1, x_2, x_3, x_4, x_5 and x_6 = model codes for molasses, corn steep liquor, asparagine, potassium ions, manganese (II) ions and chromium (III) ions respectively; eY_1 = Experimental biomass concentration (g/L); pY_1 = Predicted biomass concentration (g/L); eY_2 = experimental total protein (mg); pY_2 = Predicted total protein (mg); eY_3 = experimental L-asparaginase activity (U); pY_3 = Predicted L-asparaginase activity (U); Values of responses are means of triplicate determinations.

RSM Modeling for Improved L-Asparaginase Activity in *Aspergillus candidus* UCCM 00117

In addition to L-asparaginase activity (Y_3), two more responses were measured during the RSM experimentation

namely biomass concentration (Y_1) and total protein (Y_2) to enable the determination of specific activity of the enzyme and yield coefficient of the process. The design matrix, experimental and predicted responses are presented in Table 4. The table shows that maximum biomass

Table 5 Analysis of variance (ANOVA) for reduced quadratic regression model of biomass concentration (Y_1)

Source	Sum of squares	df	Mean square	F-value	p-value
Model	5335.32	19	280.81	82.17	< 0.0001
X ₁ -molasses	1290.71	1	1290.71	377.7	< 0.0001
X ₂ -corn steep liquor	384.91	1	384.91	112.64	< 0.0001
X ₃ -asparagine	16.99	1	16.99	4.97	0.0374
X ₄ -K ⁺	84.87	1	84.87	24.83	< 0.0001
X ₅ -Mn ²⁺	77.09	1	77.09	22.56	0.0001
X ₆ -Cr ³⁺	1365.27	1	1365.27	399.52	< 0.0001
X ₁ X ₂	16.23	1	16.23	4.75	0.0415
X ₁ X ₃	27.36	1	27.36	8.01	0.0104
X ₁ X ₄	52.61	1	52.61	15.4	0.0008
X ₁ X ₆	24.38	1	24.38	7.13	0.0147
X ₂ X ₃	52.17	1	52.17	15.27	0.0009
X ₂ X ₄	94.67	1	94.67	27.7	< 0.0001
X ₂ X ₅	17.62	1	17.62	5.16	0.0344
X ₂ X ₆	306.78	1	306.78	89.77	< 0.0001
X ₃ X ₄	89.57	1	89.57	26.21	< 0.0001
X ₃ X ₅	56.62	1	56.62	16.57	0.0006
X ₃ ²	384.93	1	384.93	112.64	< 0.0001
X ₅ ²	219.01	1	219.01	64.09	< 0.0001
X ₆ ²	37.08	1	37.08	10.85	0.0036
Residual	68.35	20	3.42		
Lack-of-fit	48.33	15	3.22	0.805	0.6617
Pure error	20.01	5	4		
Cor total	5403.67	39			

$r^2=0.9874$; Adjusted $r^2=0.9753$; Predicted $r^2=0.9269$; Standard deviation=1.85; Mean=357.24; C.V. %=0.5175; Adequate precision=40.37; PRESS=395.21; BIC=208.72; AICc=219.15

concentration, Y_1 of 87.91 g/L (predicted as 87.34 g/L) was obtained in run 5 where molasses, corn steep liquor, asparagine and Cr³⁺ were at their high levels (+ 1), with K⁺ and Mn²⁺ at their low levels (− 1). This run also accounted for highest total protein, Y_2 of 65.39 mg (predicted as 65.11 mg) indicating strong significant positive correlation ($r=0.8747$, $p<0.0001$) between the two responses. Highest L-asparaginase activity (Y_3) of 4924.78 U (predicted as 4927.74 U) was recorded for run 1 where corn steep liquor, asparagine and K⁺ were at their low levels while molasses, Mn²⁺ and Cr³⁺ were at their high levels. Conversely, lowest L-asparaginase activity of 2735.98 U (predicted as 2735.3 U) was observed in run 34 where all predictors were held at their center points (0) except asparagine which was supplied at its lowest or axial level (− 1.56508).

Analysis of variance (ANOVA) for biomass concentration (Y_1) model is presented as a reduced model containing only significant predictors of the response, Y_1 obtained by backward elimination method (Table 5). From the table, it is observed that the modified model was significant ($F=82.17$, adjusted $r^2=0.9753$, predicted $r^2=0.9269$, $p<0.0001$) and adequate to explain variations about the data within the experimental region as evaluated

by its non-significant lack-of-fit (LoF) test value of $F(15, 20)=0.805$, $p=0.6617>0.05$. The diagnostic plot of actual versus predicted Y_1 plot (Fig. 4a) further confirms the adequacy of the model. Coefficients of the predictors were employed to build the reduced second-order regression function for biomass concentration as follows:

$$Y_1 = 63.88 + 7.64X_1 + 4.25X_2 + 1.73X_4 - 2.03X_5 + 7.82X_6 + 1.11X_1X_3 + 1.83X_1X_4 + 1.20X_1X_6 + 1.46X_2X_3 - 2.05X_2X_4 + 4.24X_2X_6 - 1.98X_3X_4 - 1.67X_3X_5 - 5.31X_3^2 - 4.03X_5^2 - 1.72X_6^2 \quad (11)$$

Analysis of variance (ANOVA) for total protein (Y_2) model is also presented as a reduced quadratic function that discriminated against non-significant predictor terms through backward elimination (Table 6). Out of the 27 terms required to build the full model, 5 terms with non-significant ($p>0.05$) F -value were eliminated. The reduced model was significant at $F=269.35$, adjusted $r^2=0.9934$, predicted $r^2=0.9803$, $p<0.0001$. The non-significant LoF test value of $F(12, 17)=1.97$, $p=0.2352>0.05$ indicated adequacy of the model. The goodness-of-fit, r^2 of experimental versus predicted Y_2 plot (Fig. 4b) was also in reasonable agreement

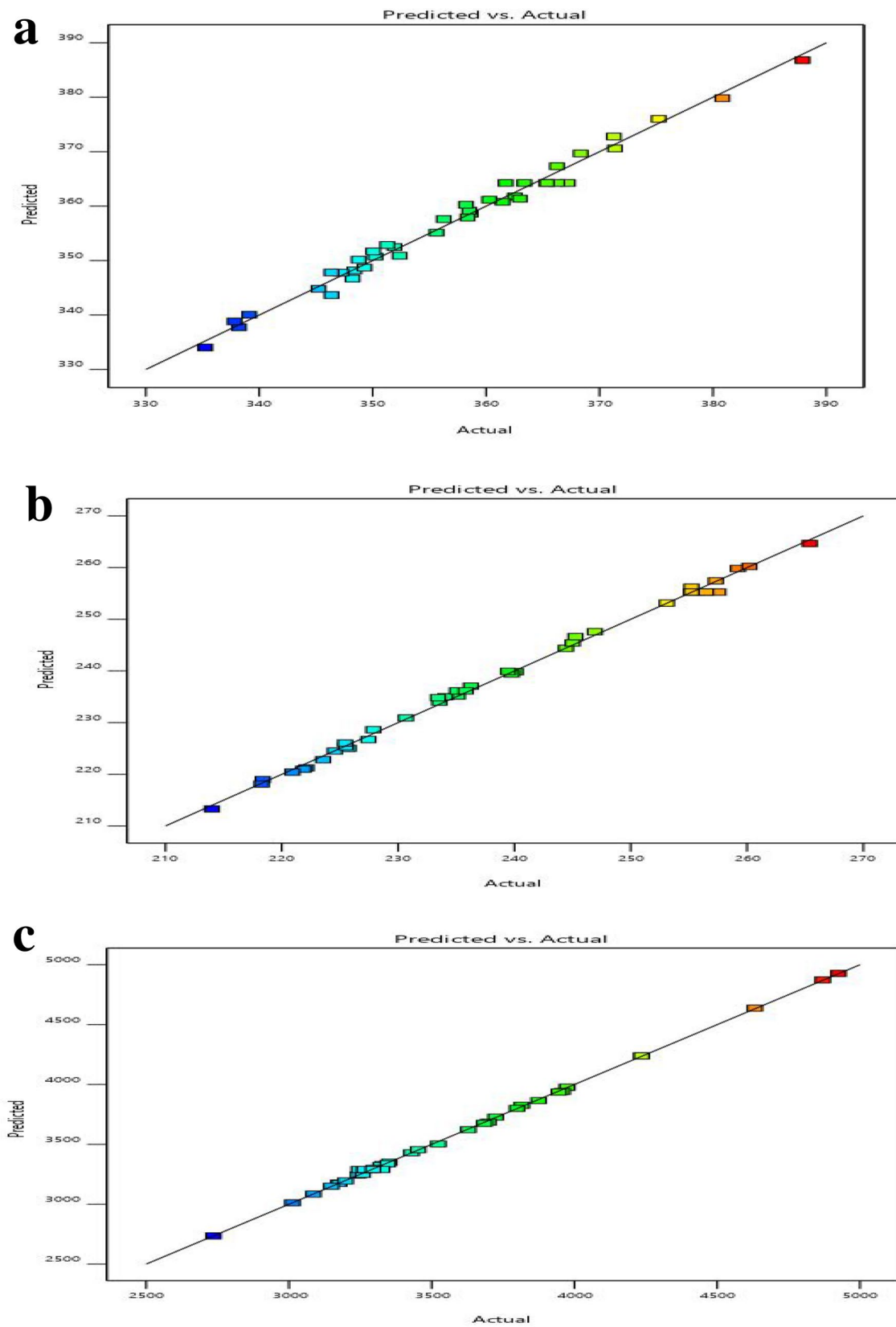


Fig. 4 Experimental versus predicted response surface methodology plots to evaluate the adequacy of **a** biomass concentration **b** total protein **c** L-asparaginase activity second-order regression models

Table 6 Analysis of variance (ANOVA) for reduced quadratic regression model of total protein (Y_2)

Source	Sum of squares	df	Mean square	F-value	p-value
Model	8092.77	22	367.85	269.35	< 0.0001
X_1 -molasses	1118.34	1	1118.34	818.87	< 0.0001
X_2 -corn steep liquor	266.48	1	266.48	195.13	< 0.0001
X_3 -asparagine	52.27	1	52.27	38.28	< 0.0001
X_4 - K^+	93.92	1	93.92	68.77	< 0.0001
X_6 - Cr^{3+}	1400.88	1	1400.88	1025.76	< 0.0001
X_1X_2	126.78	1	126.78	92.83	< 0.0001
X_1X_4	42.32	1	42.32	30.98	< 0.0001
X_1X_5	22.64	1	22.64	16.58	0.0008
X_2X_3	14.1	1	14.1	10.32	0.0051
X_2X_4	98.41	1	98.41	72.06	< 0.0001
X_2X_5	53.02	1	53.02	38.82	< 0.0001
X_2X_6	54.33	1	54.33	39.78	< 0.0001
X_3X_4	338.45	1	338.45	247.82	< 0.0001
X_3X_6	14.19	1	14.19	10.39	0.005
X_4X_5	201.44	1	201.44	147.5	< 0.0001
X_5X_6	64.25	1	64.25	47.04	< 0.0001
X_1^2	104.21	1	104.21	76.31	< 0.0001
X_2^2	44.3	1	44.3	32.44	< 0.0001
X_3^2	709.25	1	709.25	519.33	< 0.0001
X_4^2	64.88	1	64.88	47.5	< 0.0001
X_5^2	818.31	1	818.31	599.19	< 0.0001
X_6^2	134.2	1	134.2	98.27	< 0.0001
Residual	23.22	17	1.37		
Lack-of-fit	19.16	12	1.6	1.97	0.2352
Pure error	4.06	5	0.8119		
Cor total	8115.99	39			

$r^2=0.9971$; Adjusted $r^2=0.9934$; Predicted $r^2=0.9803$; Standard deviation=1.17; Mean=238.94; C.V. %=0.4891; Adequate precision=57.98; PRESS=160.23; BIC=176.60; AICc=206.76

with the model r^2 value of 0.9971, thus validating the adequacy of the model. Model coefficient terms were used to build the second-order polynomial equation for Y_2 as follows:

$$\begin{aligned}
 Y_2 = & 55.27 + 7.27X_1 + 3.45X_2 - 1.57X_3 + 2.07X_4 \\
 & + 8.17X_6 + 2.62X_1X_2 + 1.52X_1X_4 - 1.13X_1X_5 \\
 & + 0.8729X_2X_3 - 2.28X_2X_4 - 1.70X_2X_5 \\
 & + 1.71X_2X_6 - 4.32X_3X_4 + 0.8929X_3X_6 \\
 & + 3.34X_4X_5 - 1.91X_5X_6 - 2.79X_1^2 - 1.82X_2^2 \\
 & - 7.27X_3^2 - 2.20X_4^2 - 7.81X_5^2 - 3.16X_6^2 \quad (12)
 \end{aligned}$$

The ANOVA for L-asparaginase activity regression model (Y_3) is presented as a full quadratic function in Table 7. All 27 predictors of the second-order function of 6 factors were significant for Y_3 and the model's F -value of 649.96 was also significant ($p < 0.0001$). The difference between adjusted and predicted goodness-of-fit, r^2 of 0.9978 and 0.9909 respectively, was not more than 0.2 suggesting that the model was adequate for predictions (Fig. 4c). The LoF

Fischer-ratio statistic of $F(7, 12) = 0.3965$ was not significant ($p = 0.8701 > 0.05$) suggesting that whatever component of the proposed quadratic model that may have lacked fit was not important enough to alter the predicted L-asparaginase activity value. The model is presented as Eq. 13 below.

$$\begin{aligned}
 Y_3 = & 3290.22 + 156.74X_1 - 120.80X_2 + 385.51X_3 - 168.67X_4 \\
 & + 219.66X_5 + 65.18X_6 - 123.68X_1X_2 - 27.84X_1X_3 - 82.78X_1X_4 \\
 & + 94.40X_1X_5 + 82.44X_1X_6 - 24.80X_2X_3 + 150.70X_2X_4 \\
 & - 90.14X_2X_5 - 21.49X_2X_6 - 37.49X_3X_4 + 87.61X_3X_5 \\
 & - 22.43X_3X_6 - 94.86X_4X_5 - 95.95X_4X_6 + 98.14X_5X_6 \\
 & + 61.08X_1^2 + 79.98X_2^2 + 19.77X_3^2 + 127.17X_4^2 + 123.73X_5^2 + 45.32X_6^2 \quad (13)
 \end{aligned}$$

The L-asparaginase activity model predictor coefficients inform about the magnitude of contributions of the predictors to the model while the signs indicate the direction of the contribution or effect of the predictor. Equation 13 reveals that asparagine (X_3) and Mn^{2+} (X_5) made the largest linear contribution to predicted L-asparaginase

Table 7 Analysis of variance (ANOVA) for the full quadratic model of L-asparaginase activity (Y_3)

Source	Sum of squares	df	Mean square	F-value	p-value
Model	9.39E+06	27	3.48E+05	649.96	< 0.0001
X_1 -molasses	4.56E+05	1	4.56E+05	852.66	< 0.0001
X_2 -corn steep liquor	2.71E+05	1	2.71E+05	506.48	< 0.0001
X_3 -asparagine	2.76E+06	1	2.76E+06	5158.31	< 0.0001
X_4 - K^+	5.29E+05	1	5.29E+05	987.39	< 0.0001
X_5 - Mn^{2+}	8.97E+05	1	8.97E+05	1674.7	< 0.0001
X_6 - Cr^{3+}	78,926.2	1	78,926.2	147.44	< 0.0001
X_1X_2	2.56E+05	1	2.56E+05	477.76	< 0.0001
X_1X_3	12,958.3	1	12,958.3	24.21	0.0004
X_1X_4	1.15E+05	1	1.15E+05	214	< 0.0001
X_1X_5	1.49E+05	1	1.49E+05	278.33	< 0.0001
X_1X_6	1.14E+05	1	1.14E+05	212.26	< 0.0001
X_2X_3	10,278.95	1	10,278.95	19.2	0.0009
X_2X_4	3.80E+05	1	3.80E+05	709.25	< 0.0001
X_2X_5	1.36E+05	1	1.36E+05	253.74	< 0.0001
X_2X_6	7722.39	1	7722.39	14.43	0.0025
X_3X_4	23,493.29	1	23,493.29	43.89	< 0.0001
X_3X_5	1.28E+05	1	1.28E+05	239.74	< 0.0001
X_3X_6	8412.16	1	8412.16	15.71	0.0019
X_4X_5	1.50E+05	1	1.50E+05	281.05	< 0.0001
X_4X_6	1.54E+05	1	1.54E+05	287.49	< 0.0001
X_5X_6	1.61E+05	1	1.61E+05	300.8	< 0.0001
X_1^2	49,744.54	1	49,744.54	92.93	< 0.0001
X_2^2	85,295.11	1	85,295.11	159.34	< 0.0001
X_3^2	5213.79	1	5213.79	9.74	0.0088
X_4^2	2.16E+05	1	2.16E+05	402.91	< 0.0001
X_5^2	2.04E+05	1	2.04E+05	381.4	< 0.0001
X_6^2	27,383.95	1	27,383.95	51.16	< 0.0001
Residual	6423.58	12	535.3		
Lack-of-fit	2293.1	7	327.59	0.3965	0.8701
Pure error	4130.48	5	826.1		
Cor Total	9.40E+06	39			

$r^2=0.9993$; Adjusted $r^2=0.9978$; Predicted $r^2=0.9909$; Standard deviation=23.14; Meana=3576.08; C.V. %=0.6470; Adequate precision=113.26; PRESS=85,742.41; BIC=419.96; AICc=520.31

activity. Their contributions were positive suggesting that they enhanced the activity of the enzyme. Corn steep liquor and K^+ , on the other hand, had the largest negative coefficients in the model suggesting they largely inhibited L-asparaginase activity. The largest positive interaction coefficient is that for corn steep liquor and asparagine (X_2X_3) which enhanced L-asparaginase activity. Contrariwise, the interaction between molasses and corn steep liquor (X_1X_2) and that between K^+ and Mn^{2+} (X_4X_5) had the largest negative coefficients and so largely reduced the activity of the enzyme. All the quadratic terms of the model contributed towards improved L-asparaginase activity with K^+ and Mn^{2+} making the highest contributions to the improvement. This position is the complete opposite

for Y_1 and Y_2 models where quadratic terms of their models only reduced the respective responses.

Interaction model predictors for all three responses are presented as surface (3-D) plots in Fig. 5. Figure 5a reveals that highest predicted concentration of fungal biomass would be 378.69 g/L if levels of corn steep liquor and Cr^{3+} were adjusted to 0.995 (70.96%) and 0.992 (51.95 mg/L) in natural units respectively, with the remaining 4 predictors held at their center points. Highest interaction term contribution to total protein was also by corn steep liquor and Cr^{3+} when their levels were adjusted to 0.984 and 0.996 respectively (Fig. 5b) corresponding to 70.92% and 51.97 mg/L respectively, in natural units. This lends credence to the strong significant positive relationship between biomass

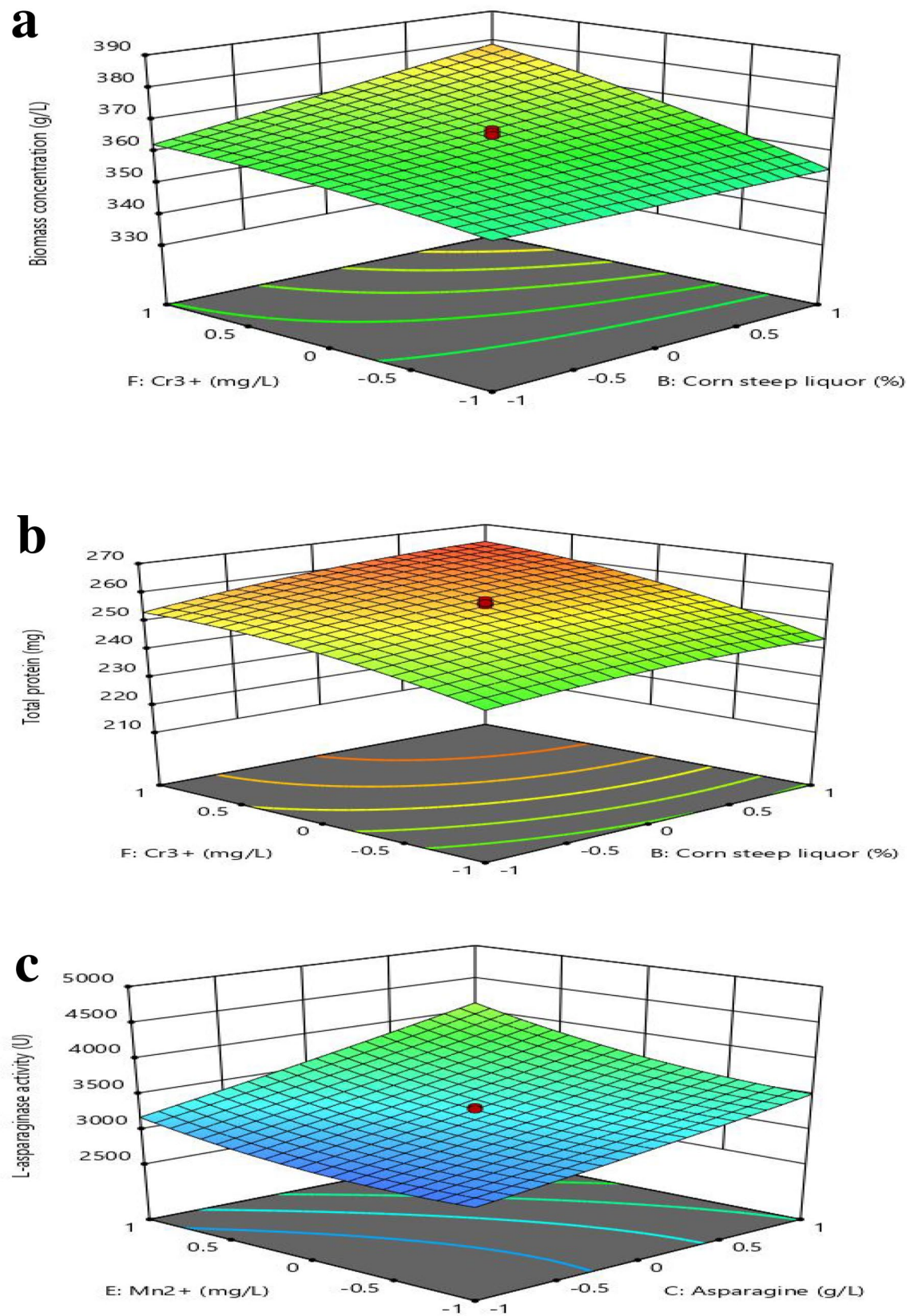


Fig. 5 Surface plots of most significant two-way factor interactions towards maximum **a** biomass concentration **b** total protein and **c** L-asparaginase activity

concentration and total protein ($r=0.8747$). Figure 5c is the surface plot for most significant two-way interaction of predictors towards enhanced L-asparaginase activity. The figure shows that L-asparaginase activity will be as high as 4124.6 U if levels of both asparagine and Mn^{2+} would be adjusted to 0.988 which would correspond to 44.22 g/L and 54.99 mg/L respectively, in natural units while all other factors are held at their center points.

Multi-objective Optimization of Responses Using the Desirability Function

Biomass concentration (Y_1) and total protein (Y_2) are by themselves significant predictors of L-asparaginase concentration, and by extension its activity (Y_3). The total activity of an enzyme is not nearly as important as its specific activity; calculated as a ratio of total enzyme activity to the amount of total protein. Similarly, the yield coefficient (yield of product on biomass), Y_p/x , of a microbial fermentation

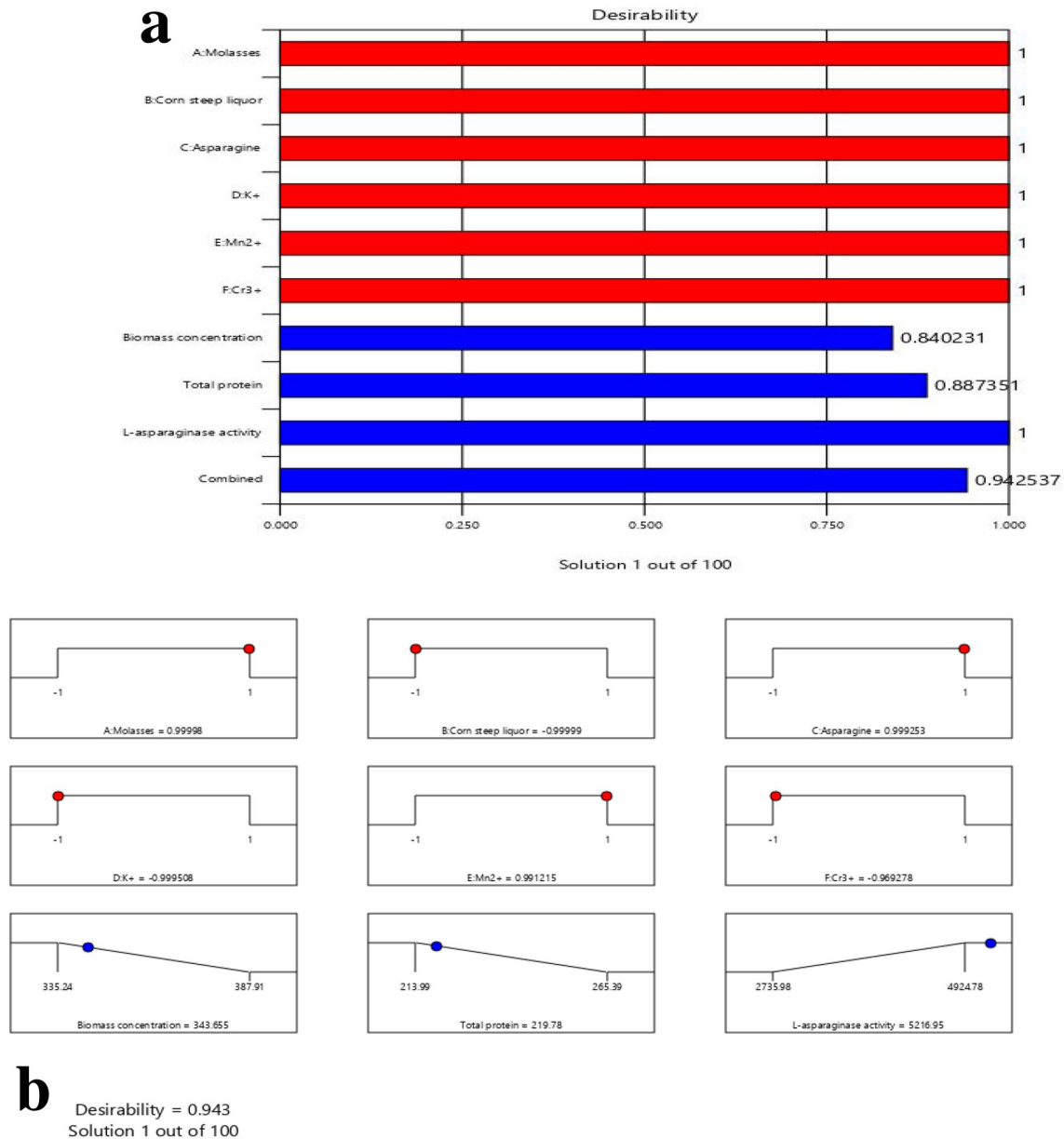


Fig. 6 Multi-objective plots for optimization of bioprocess nutrient levels towards enhanced L-asparaginase activity. **a** The bar graph of desirability plot showing individual response desirability and recommended composite (combined) desirability when all factors were

given maximum desirability of 1. **b** The ramp plot showing final factor settings and the responses obtained at the composite desirability recommended

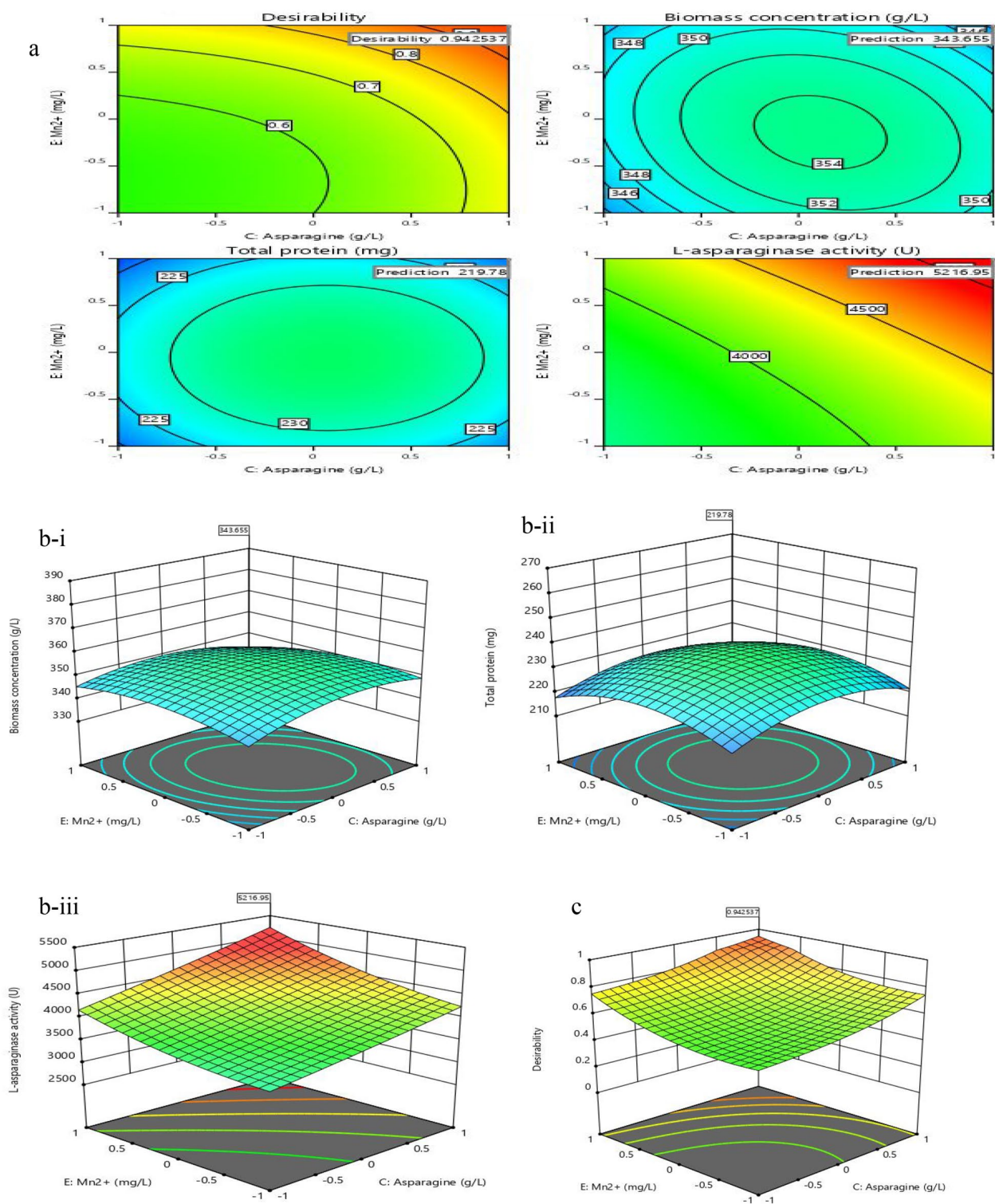


Fig. 7 Contour and surface plots of multi-objective optimal solutions for fermentation responses **a** A 4-in-1 contour plot of asparagine/ Mn^{2+} interaction towards minimum biomass concentration, minimum total protein and maximum L-asparaginase. **b-i** Surface plot for mini-

imum biomass concentration. **b-ii** Surface plot for minimum total protein. **b-iii** Surface plot for maximum L-asparaginase activity. **c** Surface plot for the composite desirability for maximum L-asparaginase activity

product is calculated in relation to the concentration of biomass that synthesized it. Since the two goals of specific activity and yield coefficient would only be enhanced at high L-asparaginase activity and concentration respectively, it became imperative to search for predictor levels that would maximize L-asparaginase concentration and its activity, while keeping the denominators at minimum levels. The goals of the multi-objective optimization were therefore set to minimize both Y_1 and Y_2 while maximizing Y_3 . Secondly, all the factors and responses were held at equal weight of 1 but with variable scaling of the importance of the three responses. A scale of 1 indicated least importance while 5 indicated most importance. Results showed that a scaling of importance of 1, 3 and 5 for Y_1 , Y_2 and Y_3 respectively was most appropriate since it produced the highest composite desirability for the study. The desirability of each response was calculated as 0.840 for Y_1 , 0.887 for Y_2 and 1.000 for Y_3 (Fig. 6a). A composite desirability, D , solved from Eq. 6 was calculated as 0.943 by adjusting levels of predictors to minimize Y_1 and Y_2 while maximizing Y_3 . The optimal solution given by the multi-objective optimization was therefore 0.99998, -0.99999, 0.999253, -0.999508, 0.991215, -0.969278 for X_1 , X_2 , X_3 , X_4 , X_5 , X_6 respectively (Fig. 6b). These levels corresponded to 49.55 g/L molasses, 64.98% corn steep liquor, 44.23 g/L asparagine, 1.73 g/L K^+ , 0.055 g/L Mn^{2+} and 0.043 g/L Cr^{3+} in natural units. These conditions resulted in a biomass concentration of 343.655 g/L, total protein of 219.78 mg and L-asparaginase activity of 5216.945 U. The contour, surface and desirability plots in Fig. 7 are graphical representations of the optimized solutions for the responses described.

Validation Experiments for Multi-objective Response Surface Optimization

The triplicate confirmation experiments set up to verify the optimal settings by the multi-objective function towards maximum L-asparaginase activity produced 344.76 ± 23.74 g/L fungal biomass, 218.53 ± 19.59 mg total protein and an L-asparaginase activity of 5187.65 ± 184.38 U.

These results gave a yield coefficient, Y_p/x , of 15.05 U/(g/L) and a specific activity of 23.74 U/mg. When these results were compared to the suggestions by the multi-objective function, it was found that none of the corresponding differences was greater than 5% indicating that the optimal conditions suggested by the optimizer were in reasonable agreement with real life situations.

Purification, Activity and Fold Determinations of *Aspergillus candidus* UCCM 00117 L-Asparaginase

Results of purification, activity and fold determinations of the L-asparaginase obtained by fermentation of *Aspergillus candidus* UCCM 00117 in the optimized medium is summarized in Table 8. The table shows that enzyme purity, expressed in terms of fold, increased with each purification step until a final enzyme with ~20-fold purity was obtained. On the contrary, L-asparaginase yield (%) or recovery decreased with every step of the purification process until a final yield of 36.46% of the enzyme, corresponding to 1.45 mg of a final total protein of 3.97 mg was obtained. L-asparaginase yield by *Streptomyces rochei* subsp. *chromatogens* NEAE-K reported by El-Naggar and El-Shweihy (2020) from a similar optimization study was 30%, although with lower fold of purification. The L-glutaminase activity of the enzyme also decreased from an initial activity of 82.131U in the sterile broth to a final activity of 0.087U, when L-asparaginase activity was 1891.64U with a specific activity of 476.48 U/mg. Since potent L-asparaginases in anti-cancer drug formulations possess a range of untoward reactions due to glutaminase activity, *Aspergillus candidus* UCCM 00117L-asparaginase holds promise for very minimal toxicity during therapy.

Literature is replete with reports on microbial production and anti-cancer activities of glutaminase-free L-asparaginase (Hussain et al. 2016; El-Naggar et al. 2018; Ashok et al. 2019). However, research is not very clear on the role of glutaminase activity during L-asparaginase treatment of human cancers. Glutaminase activity of L-asparaginase may not be required for anti-cancer activity in asparaginase

Table 8 Effects of purification steps on total protein, activities, yield and fold of *Aspergillus candidus* UCCM 00117 L-asparaginase

Purification step	Total protein (mg)	L-asparaginase		GUNase activity (U)	ASNase yield (%)	Fold
		Total activity (U)	Specific activity (U/mg)			
Sterile broth	218.53	5187.65	23.739	82.131	100.00	1.000
(NH ₄) ₂ SO ₄ Fractionation + dialysis	31.51	3843.74	121.985	5.482	74.09	5.139
DEAE-cellulose column	11.24	2997.29	266.663	0.531	57.78	11.233
Sephadex G-200 + dialysis	3.97	1891.64	476.484	0.087	36.46	20.072

ASNase L-asparaginase, GUNase L-glutaminase, DEAE-cellulose column Diethylaminoethyl-cellulose column chromatography

Fig. 8 Dose–response curves of L-asparaginase cytotoxic activity against human cancer cell lines

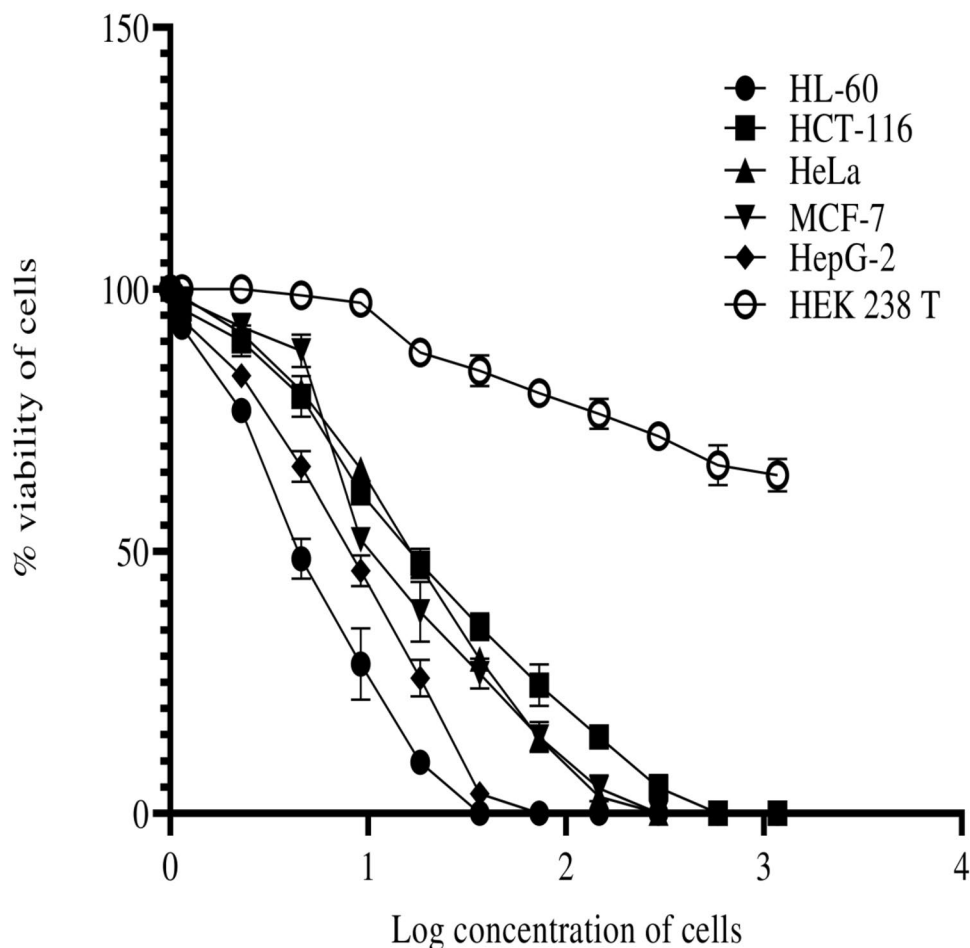


Table 9 Therapeutic potential indicators of *Aspergillus candidus* UCCM 00117 L-asparaginase

Cell lines	IC ₅₀ (μg/mL)	MCC (μg/mL)	SI	r ²	Adjusted r ²	RMSE
HL-60	4.063	36.73	15.15	0.9978	0.9969	1.873
HCT-116	13.75	587.49	4.48	0.9982	0.9976	1.611
HeLa	15.83	293.77	3.89	0.9989	0.9985	1.358
MCF-7	11.68	293.77	5.27	0.9903	0.9866	4.103
HepG2	7.610	73.45	8.09	0.9956	0.9939	2.715
HEK 238 T	61.54	ND	-	0.9884	0.9840	1.475

HL-60 Human myeloid leukemia, *HCT-116* Human colorectal carcinoma, *HeLa* Cervical epitheloid carcinoma, *MCF-7* Breast carcinoma, *HepG-2* Hepatocellular carcinoma, *HEK 238T* Human embryonic kidney cell line, *IC₅₀* Half-maximal inhibitory concentration, *MCC* Maximum cytotoxic concentration, *SI* Selectivity index, *r²* Coefficient of determination, *Adjusted r²* Adjusted coefficient of determination, *RMSE* Root mean squared error; *ND* Not determined

synthetase-negative cancers (Chan et al. 2014) but may well be for durable pre-clinical activity against T- and B-cell lymphoblastic leukemia (Nguyen et al. 2018; Chan et al. 2019).

Anti-cancer Activity of the Partially-Purified Enzyme

Results of the anti-cancer activity of the partially-purified L-asparaginase presented in Fig. 8 reveal a dose-dependent therapeutic effect where % cell viability decreased with increasing enzyme concentration. The human myeloid leukemia (HL-60) cell line was most

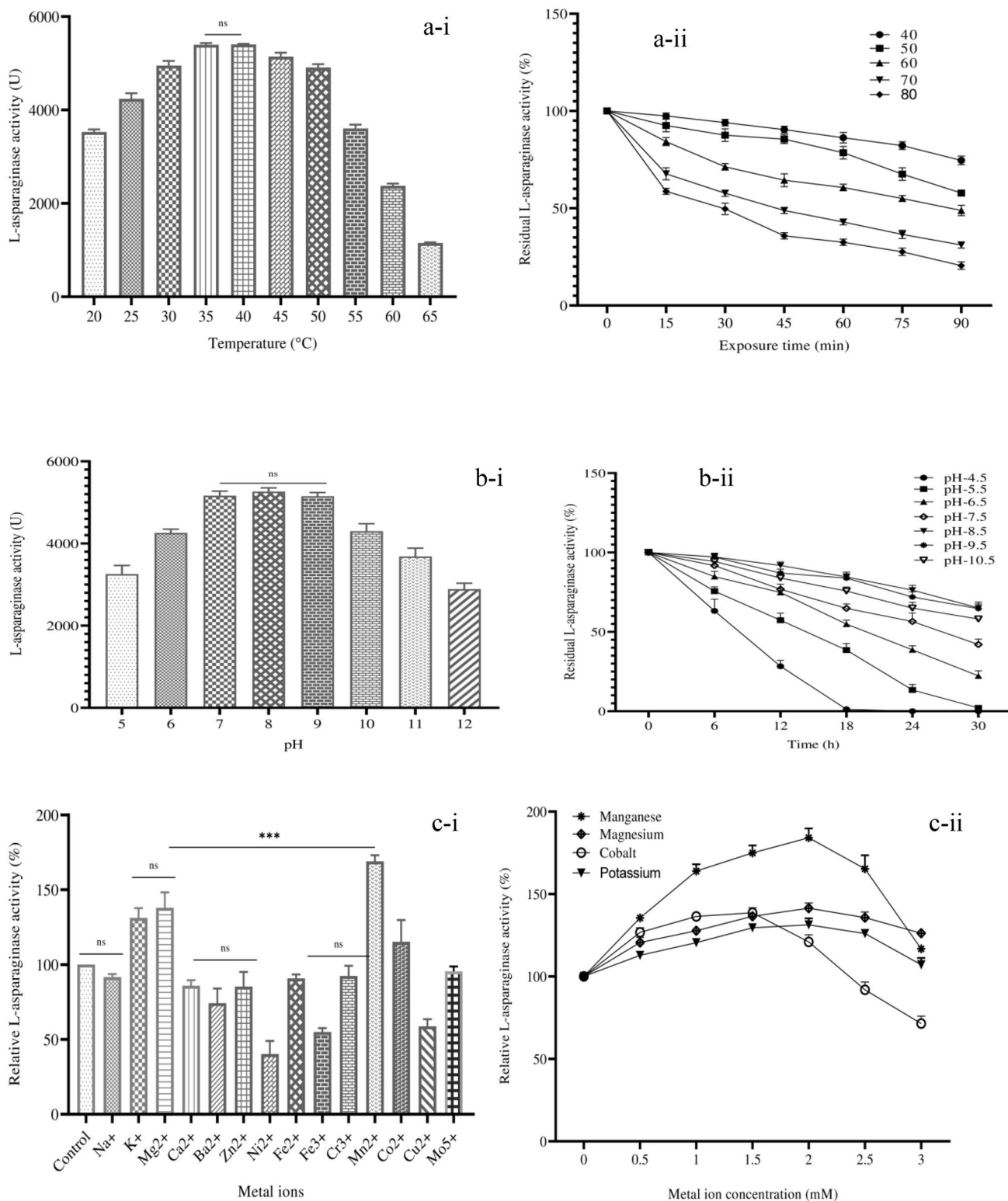


Fig. 9 Activity and stability plots of temperature, pH and metal ion effects on L-asparaginase activity. **a-i** Effect of temperature on L-asparaginase activity. **a-ii** Effect of temperature on stability of L-asparaginase activity. **b-i** Effect of pH on L-asparaginase activity.

b-ii Effect of pH on stability of L-asparaginase activity. **c-i** Effect of metal ions on L-asparaginase activity. **c-ii** Concentration range finding plot of L-asparaginase activity enhancing metal ions

sensitive to the enzyme with 100% cytotoxicity occurring at maximum cytotoxic concentration (MCC) between 18.37 and 36.73 $\mu\text{g}/\text{mL}$ after 48 h exposure (Table 9). Half-maximal inhibitory concentration (IC_{50}) of the metabolite against HL-60 cell line was 4.063 $\mu\text{g}/\text{mL}$ while those for HCT-116, HeLa, MCF-7 and HepG2 cell lines were 13.75, 15.83, 11.68 and 7.610 $\mu\text{g}/\text{mL}$ respectively. Alrumman et al. (2019) reported IC_{50} of 17.02, 14.55 and 11.66 $\mu\text{g}/\text{mL}$ for their purified L-asparaginase against HCT-116, MCF-7 and HepG-2 cell lines respectively. Their results are in reasonable agreement with the findings of this study considering the profile likelihood of 9.214–17.99 $\mu\text{g}/\text{mL}$ and 7.062–16.54 $\mu\text{g}/\text{mL}$ for HCT-116 and MCF-7 respectively (Ekpenyong et al. 2020b). The four parameter logistic models for all test cell lines were significant (Table 9) including that for non-tumor human embryonic kidney cell line (HEK-238 T). The selectivity index (SI); a measure of discrimination of anti-cancer agent against normal cells; determined for HL-60 cell was ~ 15 suggesting that the therapeutic enzyme was 15-fold more cytotoxic against the myeloid leukemia cell than to normal cell (Table 9). The index was greater than 2 for all tested cell lines indicating low toxicity of the compound (Suffness and Pezzuto 1990). El-Naggar and El-Shweihy (2020) reported similar success ($\text{SI} > 2$) with *Streptomyces rochei* L-asparaginase against HepG-2 and MCF-7 cell lines. Anti-cancer substances with IC_{50} less than 30 $\mu\text{g}/\text{mL}$ have been reported to hold promise for cancer drug development (de Oliveira et al. 2016). *Aspergillus candidus* UCCM 00117 L-asparaginase appears to be a potent anti-cancer molecule for the treatment of a number of human carcinomas especially blood and liver cancers.

Effects of Temperature, pH and Metal Ions on L-Asparaginase Activity and Stability

Results of investigation into optimum temperature and pH for L-asparaginase activity are presented in Figs. 9a and b respectively. Figure 9a-i reveals that optimum temperature of activity for the study enzyme was 40 $^{\circ}\text{C}$ (5402 ± 13.29 U). However, Tukey HSD multiple comparisons analysis of the one-way ANOVA informed that there was no statistically significant difference between activity of the enzyme at 35 $^{\circ}\text{C}$ and at 40 $^{\circ}\text{C}$. Since the human physiological temperature is 37 $^{\circ}\text{C}$ which lies between 35 and 40 $^{\circ}\text{C}$, the L-asparaginase in this study is considered suitable for treatment of different human carcinomas. This finding agrees with those of El-Naggar et al. (2018) who reported a temperature optimum of 37 $^{\circ}\text{C}$ for *Streptomyces brolllosae* NEAE-115 L-asparaginase. Results of a two-way ANOVA of the thermal stability of the enzyme indicated significant interaction between duration of exposure and temperature thus making

interpretation of residual activity results difficult as shown in Fig. 9a-ii. This implies that the residual activity is a function of both the temperature and duration of exposure. The pH optimum for the enzyme ranged from 7 to 9, accommodating the typical tight range of the human physiological pH of 7.35–7.45, and suggesting once more the suitability of the enzyme for in-vivo therapeutic applications (Fig. 9b-i). Once again, pH stability result was difficult to interpret in isolation from exposure time since the interaction effect was significant (Fig. 9b-ii). Strangely, this is the only report that considers the statistical implication of physicochemical parameters on stability of enzyme activity. Relative activity of the enzyme with regard to metal inhibition or enhancement was also studied and the result is presented in Fig. 9c. Figure 9c-i shows that Ba^{2+} , Ni^{2+} , Fe^{3+} and Cu^{2+} were potent inhibitors of L-asparaginase activity while K^{+} , Mg^{2+} , Mn^{2+} and Co^{2+} enhanced it. No significant difference existed between relative activities of the remaining cations and that of the control. Highest enhancement (168.94%) was mediated by Mn^{2+} at 1 mM which is in agreement with the findings of El-Naggar et al. (2018) who reported a relative activity of 145.15% and Li et al. (2018) who reported mediation of 190% relative L-asparaginase activity of *Pyrococcus yayanosii* CH1 L-asparaginase expressed in *Bacillus subtilis* 168 by Mn^{2+} . Maximum enhancement of our L-asparaginase activity by the metal occurred at 2 mM with relative activity of 184.17% (Fig. 9c-ii) suggesting that the assay medium for this L-asparaginase may require some re-constitution by manganese boosting.

Conclusions

Sequential statistical optimization involving OFAT, PBD, PSA and CCRD-RSM coupled with multi-objective optimization selected and optimized levels of molasses, corn steep liquor, asparagine, K^{+} , Mn^{2+} and Cr^{3+} as significant medium nutrients for enhanced L-asparaginase production by *Aspergillus candidus* UCCM 00117. Utilization of two renewable substrates (molasses and corn steep liquor) for fermentation may contribute significantly towards a sustainable bioprocess with attractive production economics. A combination of ammonium sulfate fractionation, column chromatography and dialysis recovered substantial amount of the therapeutic enzyme (36.46%) with 20-fold purity. The high specific activity (476.48 U/mg) of the Mn^{2+} -dependent L-asparaginase, coupled with low glutaminase activity (0.087 U), low half-maximal inhibitory concentration (< 20 $\mu\text{g}/\text{mL}$) and high selectivity index (> 3) offers potential for further anti-cancer drug investigation and development.

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Compliance with Ethical Standards

Conflict of interest The authors have no conflicts of interest to declare that are relevant to the content of this article.

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