



# Structural, Optical and Antibacterial Activity Studies on CMC/PVA Blend Filled with Three Different Types of Green Synthesized ZnO Nanoparticles

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## Abstract

In this work, zinc oxide (ZnO) was produced using extracts of *Thymus* (Z), *Hibiscus rosa-sinensis* (K), and *Daucus carota* (G). Furthermore, sodium carboxymethyl cellulose (CMC) and polyvinyl alcohol (PVA) were combined with ZnO to form three novel nanocomposites. X-ray diffraction (XRD) was used for the structural analysis, where the semicrystalline nature of the (CMC/PVA)/ZnO nanocomposites was confirmed. The characteristics functional groups that arose inside the prepared samples were identified by Fourier transform infrared spectroscopy (FTIR). Evidence for the successful preparation of the pure ZnO particles and their nanocomposites was carried out using a transmission electron microscope (TEM). The ZnO nanoparticles are mostly spherical, irregularly distributed, and have radii ranging from 10 to 40 nm. Their anti-bacterial activity was studied against *B. subtilis*, *E. coli*, and *Candida albicans*. The inhibition zones of all the prepared samples against *E. coli* were 0, 19, 31, and 23 mm for PVA/CMC blend, PVA/CMC/ZnO (Z) (PCZ-Z), PVA/CMC/ZnO (K) (PCZ-K), and PVA/CMC/ZnO (G) (PCZ-G), respectively, compared to the streptomycin control Gram-positive standard with inhibition zone (34 mm). On the other hand, the inhibition zones of the prepared samples against *B. subtilis* were equal to 0, 26, 33, and 28 mm for CMC/PVA, PCZ-Z, PCZ-K, and PCZ-G, respectively. Based on these results, the PCZ-K sample is the most effective at resisting *E. coli* (91.17%) and *B. subtilis* (94.28%). These nanocomposites do not have harmful chemicals, making them strong candidates for use in biological applications.

**Keywords** CMC/PVA · ZnO nanoparticles · Plant extract · XRD · Antibacterial activity

## 1 Introduction

Nano-sized inorganic composites have exhibited extraordinary antibacterial activity at extremely low concentrations because of their exceptional physiochemical properties and high surface area-to-volume ratio [1, 2]. Moreover, these

compounds are thermally stable at high pressure. Some of them are considered non-toxic and even contain vital minerals for the human body. Metal and metal oxide nanoparticles (NPs), like gold, copper, silver, zinc oxide, manganese oxide, and titanium oxide, were identified as the most antibacterial inorganic materials [3–5].

As nanotechnology advances, more and more attention is being paid to how nanomaterials affect bacteria and fungi, with the hope that this may lead to improved hygiene in public places [6]. One of the non-toxic nanomaterials that might be employed in biomedical applications is zinc oxide nanoparticles (ZnO NPs). They are effective against cancer and microbes [7].

Among the many benefits of ZnO are its good electrochemical features, wide absorption spectrum, and strong chemical stability. In addition to being beneficial for electrical and optoelectronic applications like storage devices and solar cells [8], its large energy bandwidth and thermal stability also qualify it as a semiconductor. Likewise, their

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ability to absorb UV rays makes them an essential ingredient in enzymes, cosmetics, toothpaste, and sunscreens [9]. ZnO has piezoelectric and pyroelectric properties that make it useful for the development of sensors, generators, and photocatalysis. It has low toxicity and biocompatibility, facilitating its applications in biomedicine [10].

Several studies [11–13] have reported on the different techniques to synthesize ZnO nanoparticles and its modification over different blends. They also studied its biological activities, as reported by Nadeem et al. [11], where they studied the antimicrobial activity of iron (Fe) and cobalt (Co) co-doped ZnO nanoparticles and displayed an enhancement in antibacterial activity by co-doping with iron and cobalt. Moreover, Mukhtar et al. [12] reported the photocatalytic properties of titanium, vanadium, and yttrium oxides and showed an increase in photocatalytic activity by mixing them together, as well as displayed excellent antibacterial activities.

To avoid agglomeration and control excessive growth of nanoparticles, nanomaterials synthesized from organometallic precursors need the application of a capping agent that works as a stabilizer and makes colloids more stable [14]. The two most high-performance stabilizing agents are carboxymethyl cellulose (CMC) and polyvinyl alcohol (PVA) [15]. Hydrolysis of polyvinyl acetate yields PVA, a synthetic polymer with excellent biodegradability, biocompatibility, and hydrophilicity [16]. PVA is employed in different biological applications, where its hydrogels have significant potential for use in artificial grafts and tissue engineering. PVA is a low-cost, lightweight polymer with high mechanical and optical qualities that is also environmentally friendly. It can also be utilized in various industrial applications due to its superior chemical and physical qualities, such as textile sizing, solar cells, adhesives, soil stabilizers, surgical, optoelectronic, and biological devices [17, 18]. Additionally, one of the most important cellulose derivatives is carboxymethyl cellulose (CMC) which is treated with chloroacetic acid ( $\text{ClCH}_2\text{CO}_2\text{H}$ ). It's utilized in the cosmetic, paints, pharmaceuticals, mineral processing, food, textiles, ceramic foam, biodegradable films, and paper industries as a thickener, binder, suspension stabilizer (stabilizing agent), and water-retaining agent [19].

In the current study, zinc oxide (ZnO) nanoparticles were synthesized from *Thymus* (Z), *Hibiscus rosa-sinensis* (K), and *Daucus carota* (G) extracts. After that, ZnO NPs were combined with sodium carboxymethyl cellulose (CMC) and polyvinyl alcohol (PVA) to form three novel nanocomposites. The synthesized nanocomposites were investigated by different techniques, including TEM, XRD, UV–Vis, and FT-IR spectroscopy. The antibacterial activity of the prepared composites was studied against *B. subtilis*, *E. coli*, and *Candida albicans*.

## 2 Materials and Methods

### 2.1 Materials

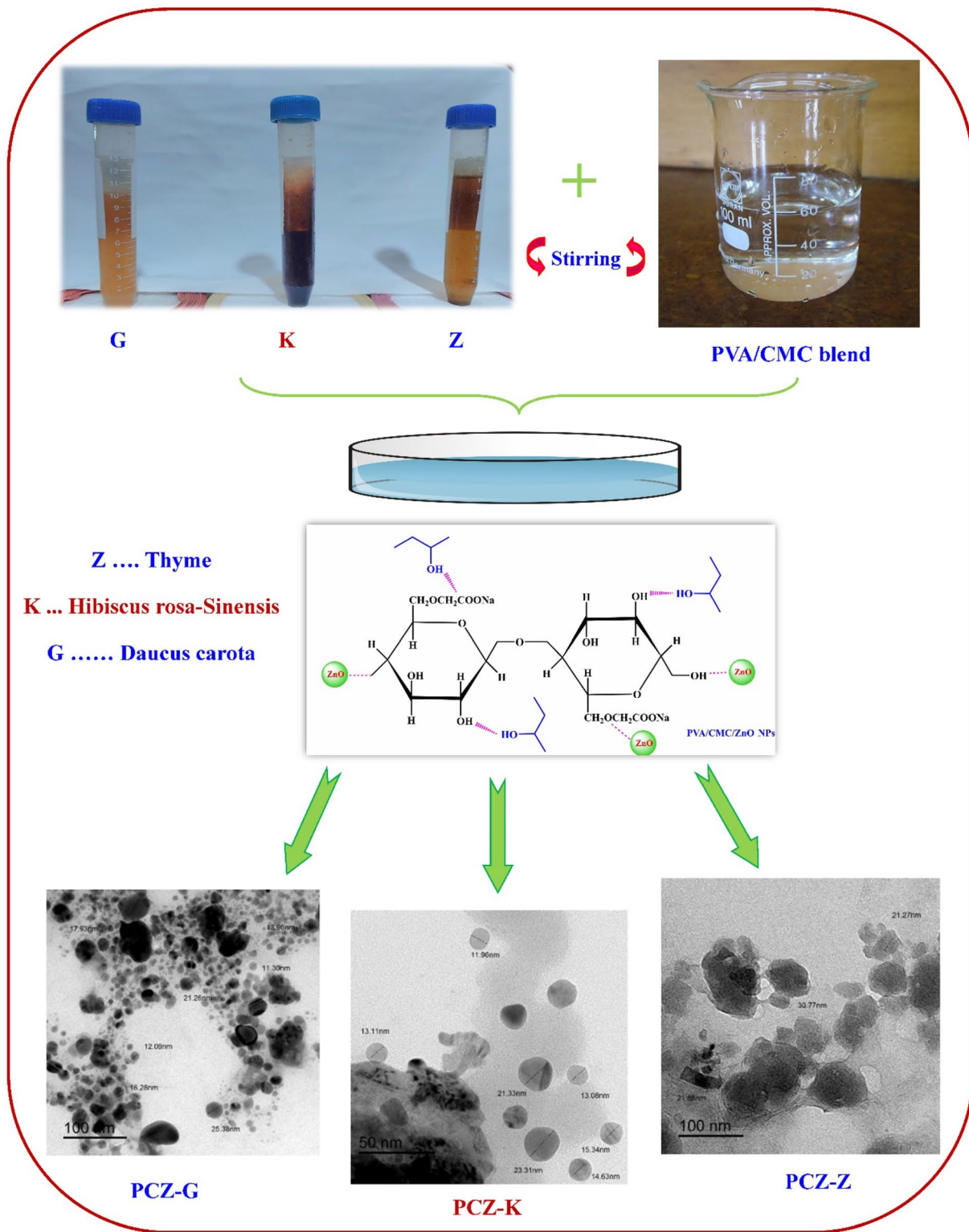
Polyvinyl alcohol (PVA) purchased from Acros (USA) has a molecular weight  $\approx 6000$  g/mol. Carboxymethyl cellulose (CMC) supplied by Lanxess (Germany), in combination with Zinc acetate dehydrate ( $\text{Zn}(\text{CH}_3\text{COO})_2 \cdot 2\text{H}_2\text{O}$ ) as a precursor supplied by the Sigma Aldrich company.

### 2.2 Synthesis of ZnO Nanoparticles

Thyme (Z), *Hibiscus rosa-sinensis* (K), and *Daucus carota* (G) were rinsed using tap water, followed by double-distilled water and ethanol to remove any trace of contamination. The plants were then dried at room temperature. About 10 g of each plant was ground in an agate mortar. The obtained powder was mixed separately with 250 mL of distilled water adjusted at about 75 °C for 1 h. The mixture was then filtered using Whatman filter paper No. 1. The obtained solutions were stored in dark bottles at 4 °C. A 0.01 M aqueous solution of zinc acetate dihydrate ( $\text{Zn}(\text{CH}_3\text{COO})_2 \cdot 2\text{H}_2\text{O}$ ) was prepared and stored as a stock solution. 95 mL of the stock solution was mixed with 5 mL of the plant extract, where each plant was in a separate flask. The resulting mixture was incubated for one hour at 75 °C while being continuously shaken at 150 rpm. As a result, bio-reduced salt ultimately settled in the flasks in the form of white precipitate. The supernatant was poured, and the precipitate was centrifuged and washed four times with deionized water to assure the elimination of contaminants [20].

#### 2.2.1 Preparation of the Polymer Blend and Its Nanocomposites

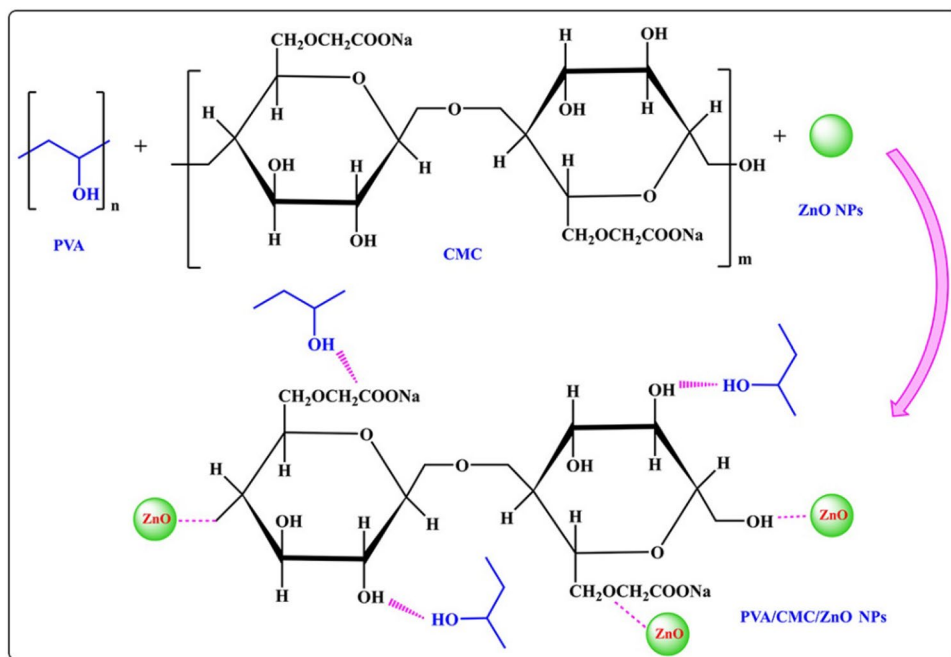
The preparation steps are shown in Scheme 1. In brief, 2 g of both polyvinyl alcohol (PVA) and carboxymethyl cellulose (CMC) were vigorously stirred individually in deionized water. The obtained solutions were then mixed for about 3 h until a clear, bubble-free mixture solution was obtained. The final mixture was then divided into four equal parts. The same quantity of ZnO nanoparticles synthesized from different plants, viz., thyme, rosella (*Hibiscus sabdariffa*), and carrots designated as shown in Table 1, was mixed with the blend sample using a sonicator homogenizer. Samples were then incubated at 50 °C for 2 days after being decanted into plastic Petri dishes to ensure evaporation of any solvent traces. The final product was in the form of thin films that were stored in a vacuum desiccator until use. Table 1 shows the symbols for the samples that were synthesized with different ZnO nanoparticles made from different plants.



Scheme 1 The preparation steps of the (CMC/PVA)/ZnO nanocomposites

**Table 1** Sample composition and designation

Sample	Symbol	PVA (wt%)	CMC (wt%)	ZnO	Source
PVA/CMC blend	Pure blend	50	50	0.0	
PVA/CMC/ZnO (Z)	PCZ-Z	49.95	49.95	0.1	Thymus
PVA/CMC/ZnO (K)	PCZ-K	49.95	49.95	0.1	<i>Hibiscus rosa-sinensis</i>
PVA/CMC/ZnO (G)	PCZ-G	49.95	49.95	0.1	<i>Daucus carota</i>

**Scheme 2** Molecular interaction of PVA and CMC

## 2.3 Measurements

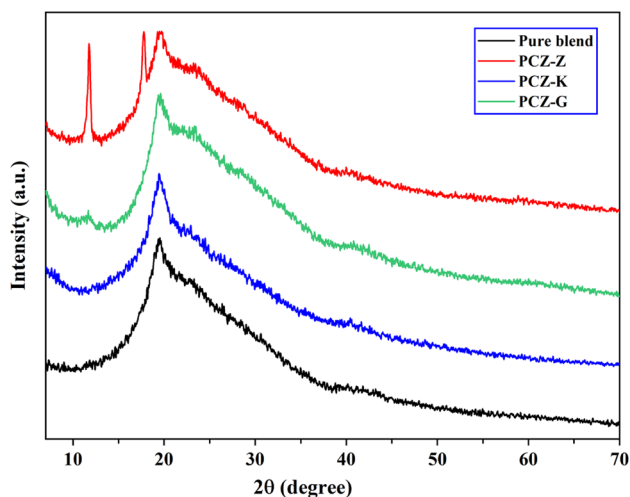
### 2.3.1 Characterization of Polymer Nanocomposites

The molecular interaction between the nanocomposite constituents is exhibited in Scheme 2. The crystallinity of the prepared nanocomposite samples was examined through an X-ray diffractometer (X'Pert PRO) with Cu  $K\alpha$  radiation at 30 kV and a wavelength of 0.15406 nm in room temperature. The structural features of all samples were investigated using a Fourier transform infrared (FTIR) spectrometer (Nicolet iS10) in the spectral region of  $4000\text{ cm}^{-1}$  to  $400\text{ cm}^{-1}$ . The prepared composites were coated on copper grids (200 mesh) and inspected by HRTEM (JEM-2100) at 200 kV for transmission electron microscopic examination to examine the particle size and morphology of the prepared nanocomposites. The optical properties of the prepared composites were tested using a spectrophotometer (V/570 UV/VIS–NIR, JASCO, Japan) in the range of 200–1100 nm.

### 2.3.2 Evaluation of Antibacterial Activity

Usually, the antibacterial activities of the present composites were estimated by the disc diffusion technique using inoculums consisting of  $10^6$  bacterial spreads on Mueller–Hinton agar plates [21]. The activities of *Bacillus subtilis* as Gram-positive bacteria, *Escherichia coli* as Gram-negative bacteria, and *Candida albicans* as fungi were examined for the synthesized samples by the qualitative technique. 20 mL of Mueller agar was placed into sterile petri dishes, allowed to harden, and then dried in the incubator. A total of roughly  $10^6$  cells were spread out on the agar plate using a sterilized glass rod, and the plate was then allowed to dry to a standard turbidity of 0.5 McFarland. The discs of samples were placed on top of agar plates that had been planted. The agar plates were incubated for one day at  $37\text{ }^\circ\text{C}$ . Each plate was tested after incubation. For each bacterium, positive control of streptomycin (100 mg/mL) is utilized. The disc's diameter as well as the diameter of the inhibition zones were measured. Zones are measured to the closest full millimeter





**Fig. 1** XRD spectra of CMC/PVA blend doped with ZnO-NPs extracted from three different plants

using calipers or a ruler placed on the back of the upright petri dish.

## 3 Results and Discussion

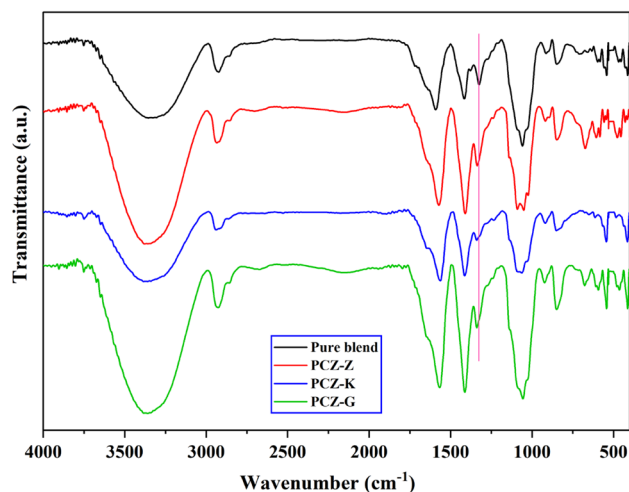
### 3.1 XRD Analysis

The XRD spectra of the CMC/PVA blend doped with ZnO-NPs extracted from the three different plants are displayed in Fig. 1. For the pure PVA/CMC blend, the protuberant broad peak at  $2\theta \approx 19.35^\circ$  was ascribed to the semicrystalline nature of PVA [22] and the crystalline cellulosic structure [18]. The other peak, centering at  $2\theta \approx 41.13^\circ$ , indicates the presence of a representative semi-crystalline structure, suggesting that CMC interacted strongly with PVA [23]. The first broad peak has shifted with the addition of ZnO nanoparticles, confirming the efficacy and compatibility of the ZnO-based filler in the polymer blend matrix. Some parameters, such as area under the main peaks ( $A_p$ ), d-spacing, and the full width at half maximum (FWHM), were calculated and listed in Table 2. The average crystalline size ( $D_{\text{cry}}$ ) was determined by using Sherrer's method that was previously discussed in detail in the literature [24–26].

From this table, it is observed that the broadness of the main peak had increased, which could be due to the lattice strain in the nanocomposites [27]. Further, the degree of crystallinity ( $X_{\text{cry}}\%$ ) was calculated using the Hermans–Weidinger approach [24]. The decrease in  $X_{\text{cry}}\%$  indicates that the amorphous areas were distributed throughout the host matrices, revealing that CMC/PVA/ZnO nanocomposites are semicrystalline in nature. The sharp peaks shown at  $2\theta \approx 11.71^\circ$  for the PCZ-G sample, and at  $\theta \approx 11.77^\circ$  and  $17.81^\circ$  for the PCZ-Z sample may be stemmed from the presence of ZnO-nanoparticles, confirming the above results and recommending them for use in advanced technological applications.

### 3.2 ATR-FTIR Spectroscopy

At room temperature, FT-IR spectroscopy was used to look at how the molecules in the current blend interacted with each other and with the nanoparticles. The FT-IR spectra of pure and modified CMC/PVA blends with ZnO-NPs extracted from three different plants were demonstrated in Fig. 2. The figure displayed that all the prepared samples have characteristic bands such as;  $3369\text{ cm}^{-1}$  (O–H stretching of CMC or physically adsorbed water) [28, 29],  $2928\text{ cm}^{-1}$  ( $\text{CH}_2$  asymmetrical str.),  $1644\text{ cm}^{-1}$  (C=O str.)



**Fig. 2** FT-IR spectra of the PVA/CMC polymer blend and the blend doped with ZnO-NPs extracted from three different plants

**Table 2** The Area under the peak ( $A_p$ ), FWHM,  $X_{\text{cry}}\%$ , d-spacing, average crystalline size ( $D_{\text{cry}}$ ), Urbach Energy ( $\Delta E$ ) and the indirect and direct optical energy gap values for the prepared samples

Sample	$A_p$	FWHM	$X_{\text{cry}}\%$	d-spacing	$D_{\text{cry}}$ (nm)	$E_{\text{igd}}$ (eV)	$E_{\text{gd}}$ (eV)	$\Delta E$ (eV)
Pure blend	13,620	12.44	39.08	4.49753	–	4.35	5.78	1.22
PCZ-Z	14,793	14.12	17.1	4.49787	10.8	4.25	5.50	1.00
PCZ-K	14,282	11.31	30.34	4.55624	13.58	3.52	4.80	1.16
PCZ-G	16,597	13.78	31.49	4.5153	11.15	4.00	5.22	0.97

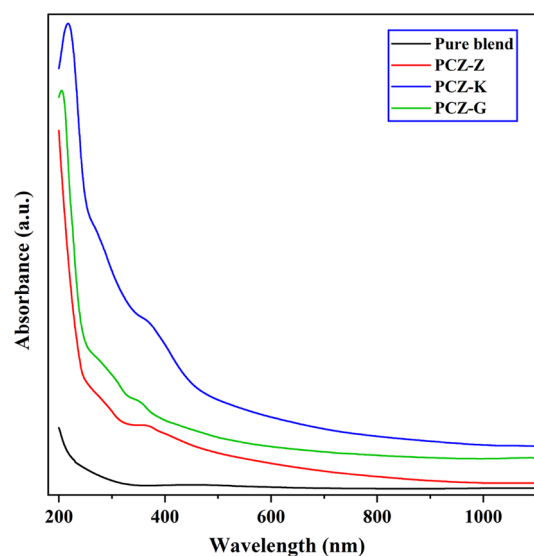
**Table 3** Bands assignments for the FT-IR spectra of the prepared films under study

Wavenumber (cm <sup>-1</sup> )	Band assignment	Source
3369	–OH stretching	PVA + CMC
2928	CH <sub>2</sub> asymmetrical stretching	CMC
1644	C=O stretching (shoulder)	CMC
1420	–CH <sub>2</sub> scissoring	CMC
1588	Asymmetric carboxylate (–COO <sup>-</sup> ) str	CMC
2857	CH <sub>2</sub> symmetrical stretching	PVA
1323	–OH bending	CMC + PVA
1057	C–O str	PVA
849	–CH rocking	PVA

[30], and 849 cm<sup>-1</sup> (C–H rocking of PVA) [31]. Table 3 summarizes additional band assignments for the FTIR spectra of the produced samples. The presence of oxygen in the ether bond allows hydrogen to interact with other molecules in the PVA and CMC polymers, leading to the development of a strong peak at 1092 cm<sup>-1</sup> [32]. This peak substantiates that the currently available nanocomposite samples have a semicrystalline structure. Furthermore, the intra-molecular interaction between the asymmetric carboxylate (–COO<sup>-</sup>) str. groups and the bending hydroxyl groups (–OH) is responsible for the two spectral bands seen at 1588 cm<sup>-1</sup> and about 1323 cm<sup>-1</sup> [33]. Having these two functional groups improves the structural characteristics of the nanocomposites formed from ZnO-NPs and the CMC/PVA blend [34]. Similar CMC characteristic bands at 1581, 1421, 1334, and 1056 cm<sup>-1</sup>, which were carbohydrate signature peaks, have been observed by previous reports [23, 35], indicating the existence of carboxymethyl substituents on the CMC backbone [36]. The observed shift in some peaks, such as –OH bending from 1323 to 1343 cm<sup>-1</sup> proves the constructive interaction of the current blend with ZnO NPs [37]. According to the available literature, the typical FT-IR bands of oxides are located between 600 and 400 cm<sup>-1</sup>. Thus, the vibrational bands observed in the range of 600–400 cm<sup>-1</sup> correspond to the characteristic ZnO-NPs stretching vibrations [26]. The enhanced structural characteristics of the nanocomposite samples are further supported by the increased –OH band intensity, which shows the formation of hydrogen bonds.

### 3.3 UV/Vis Study

Absorbance spectrum data of the prepared samples in the ultraviolet and visible regions were illustrated in Fig. 3. The  $\pi \rightarrow \pi^*$  transition is responsible for the 200 nm absorption edge in the UV region of the CMC/PVA mixture spectra [18]. After being filled with ZnO-NPs, the absorbance

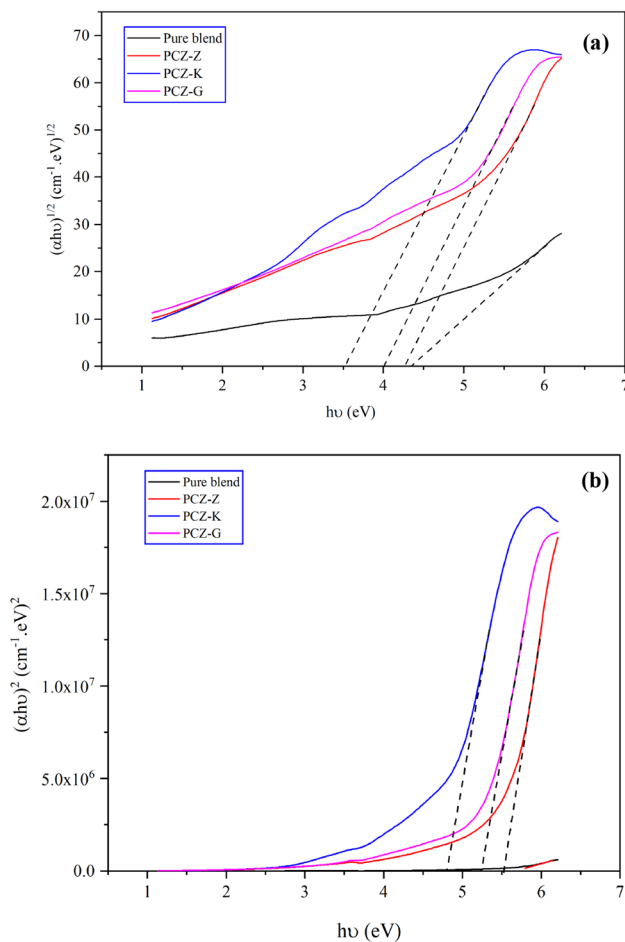
**Fig. 3** UV/VIS absorbance spectra for pure and filled CMC/PVA blend with ZnO-NPs extracted from the three different plants

values of the produced composites significantly increase, and a new absorption peak appears at about 350 nm. The presence of such a peak and its shift could be ascribed to the complexation and interaction behavior of the ZnO-NPs with polymeric matrices, affecting the optical band gap's value [38], which was associated with crystallinity variation in the nanocomposite, supporting the XRD findings [39]. The value of the optical energy gap ( $E_g$ ) could be determined by the equation of Mott and Davis [40], which analyzes the spectrum dependency of the absorption coefficient near the absorption edge.

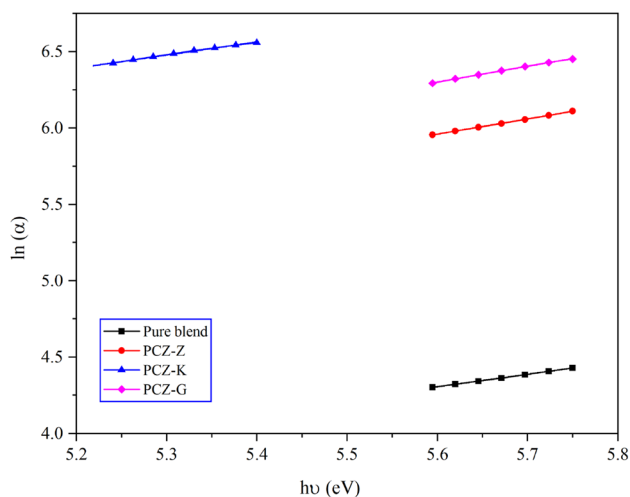
$$\alpha = \frac{B(h\nu - E_g)^m}{h\nu} \quad (1)$$

where B is an electronic transition probability constant and  $h\nu$  is the energy of the incident photons.  $m$  is equivalent to 2 or 1/2 for the allowed direct and indirect transitions, respectively.

Figure 4 displays  $(\alpha h\nu)^2$  and  $(\alpha h\nu)^{1/2}$  plots of versus photon energy ( $h\nu$ ) of the as-synthesized composites. Linear extrapolation of these data points along the  $h\nu$  axis yielded the optical band gap values shown in Table 2. The predicted optical energy gap reduces when ZnO-NPs are filled; this is ascribed to the function of ZnO-NPs in modifying the structural properties due to the development of varying polarons and defect contents, which are associated with the density of localized states  $N(E)$  [41]. Spectral measurements showed an extended tail that coincides with localised states in the valence band tail. The tail is longer because of ZnO-NPs defects, which allow it to reach into the conduction band at lower energies below the main edge. Therefore, the Urbach



**Fig. 4** The plots of **a**  $(\alpha h\nu)^{1/2}$ , and **b**  $(\alpha h\nu)^2$  versus  $(h\nu)$  for pure and filled CMC/PVA blend with ZnO-NPs extracted from the three different plants



**Fig. 5** Absorption efficiency ( $\alpha$ ) of a CMC/PVA blend including ZnO-NPs as a function of photon energy ( $h\nu$ )

formula [39] may be used to calculate the absorption coefficient ( $\alpha$ ) using just the energy tail width ( $\Delta E$ ) and the photon energy, both of which are correlated to thermal vibration in the lattice [22].

$$\alpha = \alpha_0 \exp\left(\frac{h\nu}{\Delta E}\right) \quad (2)$$

where  $\alpha_0$  is a constant

Urbach energy values ( $\Delta E$ ) were calculated using Fig. 5's absorption coefficient-photon energy relationship. The  $\Delta E$  values have been recorded in Table 2. The value of  $\Delta E$  for the pure blend is 1.22 eV, whereas the filled samples have values ranging from 1.16 and 0.97 eV.

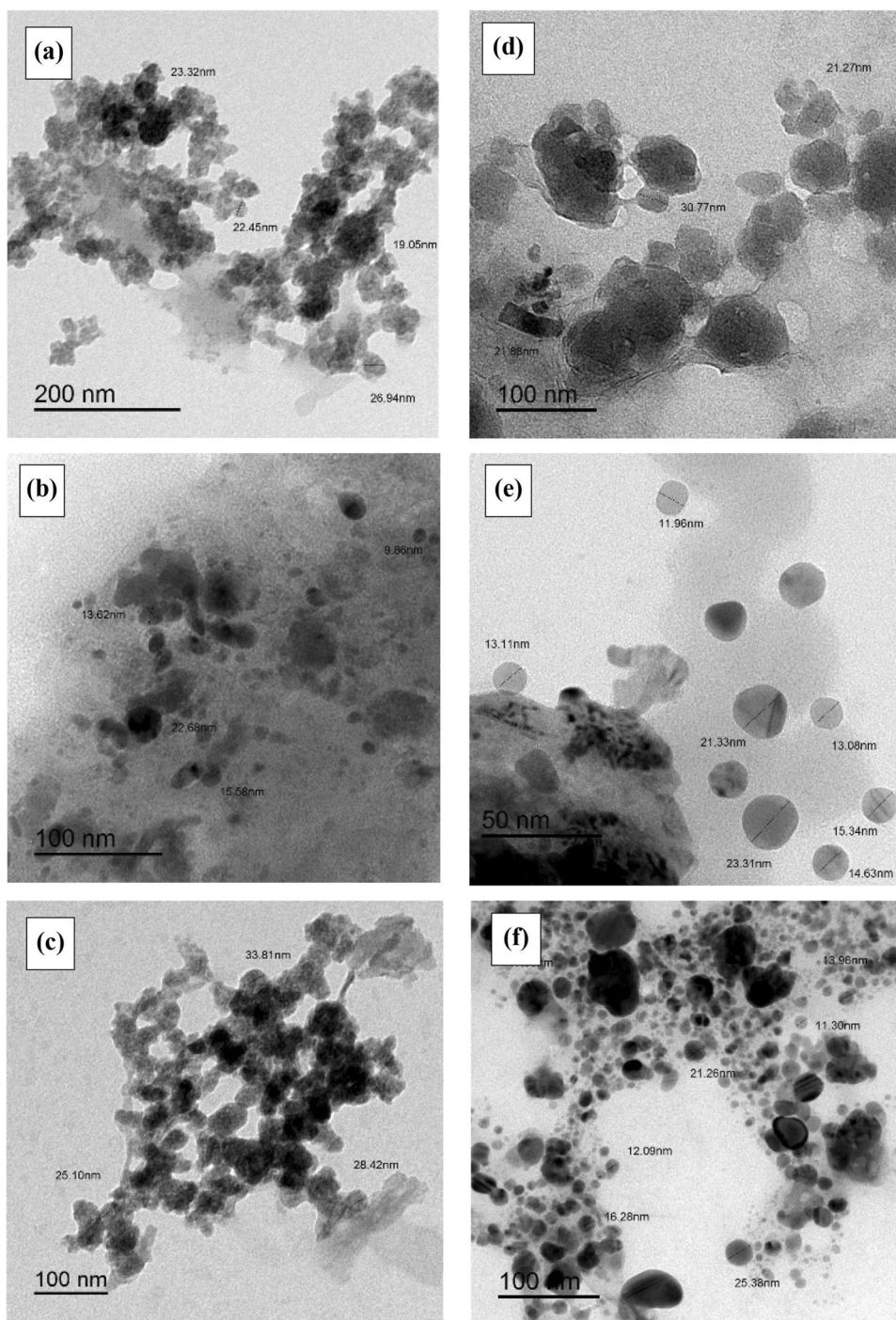
### 3.4 Transmission Electron Microscopy (TEM)

The morphology, nanoparticle size, and shape of the produced ZnO-NPs were checked using TEM images. Figure 6 displays TEM images for the purely obtained ZnO-NPs extracted from the three different plants and also shows the ZnO-NPs distributed in the aqueous solution of the CMC/PVA blend. The nanomaterials are mostly spherical, irregularly distributed, and their radii ranged from 10 to 40 nm. As well as, the images revealing that after addition of the CMC/PVA blend, the ZnO NPs were well dispersed over the blend, especially PCZ-K, which could have an excellent feature in the microbial activity. According to the nanoparticle size distribution histograms shown in Fig. 7, the average sizes of ZnO extracted from the three different plants were found to be 25.4, 15.2, and 17.9 nm for blended PCZ-G, PCZ-K, and PCZ-Z, respectively. So, the PCZ-K sample has the smallest particle size, which could affect the catalytic activity toward the different microbial organisms.

### 3.5 Antibacterial Activity

Figure 8 displays the antibacterial activity of CMC/PVA, PCZ-Z, PCZ-K and PCZ-G nanocomposites against *B. subtilis*, *E. coli*, and *Candida albicans*. The inhibition zone diameter was given in millimeters on the agar plate and calculated as an average value after each sample was repeated three times. The results are listed in Table 4. CMC/PVA did not display any antibacterial or antifungal activities. On the other hand, PCZ-Z, PCZ-K and PCZ-G displayed good antibacterial activity. The inhibition zone was bigger in *Hibiscus sabdariffa* than in *Daucus carota* and the thymus, which was interesting. These antibacterial activities for ZnO samples may result from numerous recommended mechanisms; the first mechanism is due to the size of nanoparticles of ZnO; when the particle size was decreased, the antibacterial activity was increased because of the concentration was increased [42]. The second process might include the introduction of  $Zn^{2+}$  ions

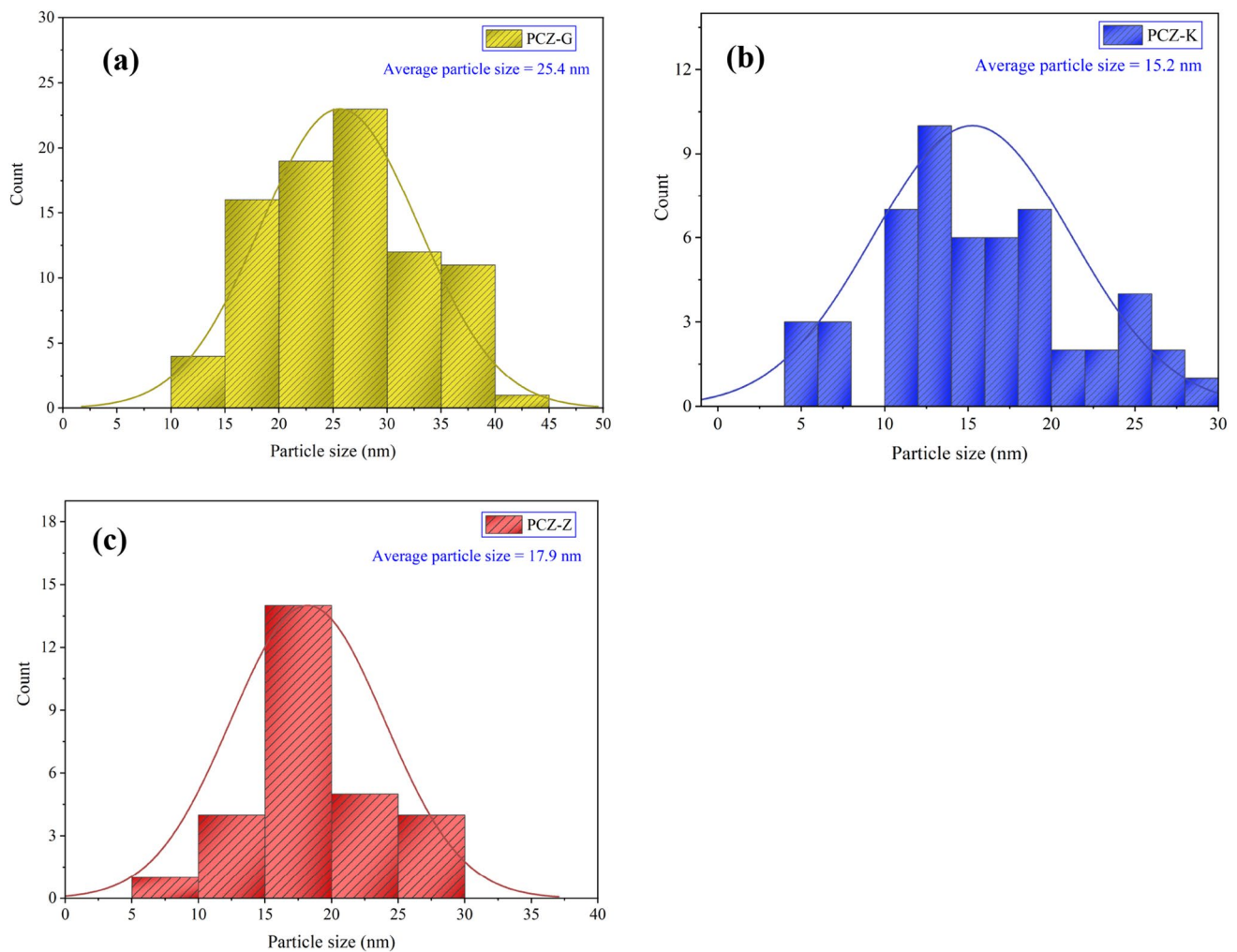
**Fig. 6** TEM Images of **a** pure synthesized ZnO-NPs (G), **b** pure synthesized ZnO-NPs (K), **c** pure synthesized ZnO-NPs (Z), **d** blended PCZ-G, **e** PCZ-K, and **f** PCZ-Z



into the bacterial growth medium [43, 44]. So, all the samples containing ZnO-NPs displayed excellent antibacterial activity and showed a deadly effect on the tested bacteria. The inhibition zones of all the prepared samples against *E. coli* was 0, 19, 31, and 23 mm for CMC/PVA, PCZ-Z, PCZ-K and PCZ-G, respectively, compared to the streptomycin control gram-positive standard with inhibition zone (34 mm). On the other hand, the inhibition zones of the prepared samples against *B. subtilis* were equal to 0, 26,

33, and 28 for CMC/PVA, PCZ-Z, PCZ-K and PCZ-G, respectively. These results reveal that PCZ-K sample has the maximum antibacterial efficiency for *E. coli* (91.17%) and *B. subtilis* (94.28%). Unfortunately, all the prepared samples have no activity toward *Candida albicans*, which means these samples do not have any antifungal activities. The improved antibacterial activity may be related to the CMC/PVA support that prevents the accumulation of ZnO nanoparticles and surface defects in ZnO [45].





**Fig. 7** Nanoparticle size distribution histograms of **a** PCZ-G, **b** PCZ-K, and **c** PCZ-Z

The higher antibacterial activity of ZnO NPs could be related to the production of reactive oxygen species and the accumulation on the surface or deposition in the cytoplasm of the cells, as detected in the previous results for *S. aureus* [43]. The smallest particle size of 15.2 nm in the case of PCZ-K has been found to strongly inhibit the survival of pathogenic microorganisms tested compared with other results. So, the inhibitory efficacy of ZnO NPs is very much dependent on their size, which is similar to other findings [46].

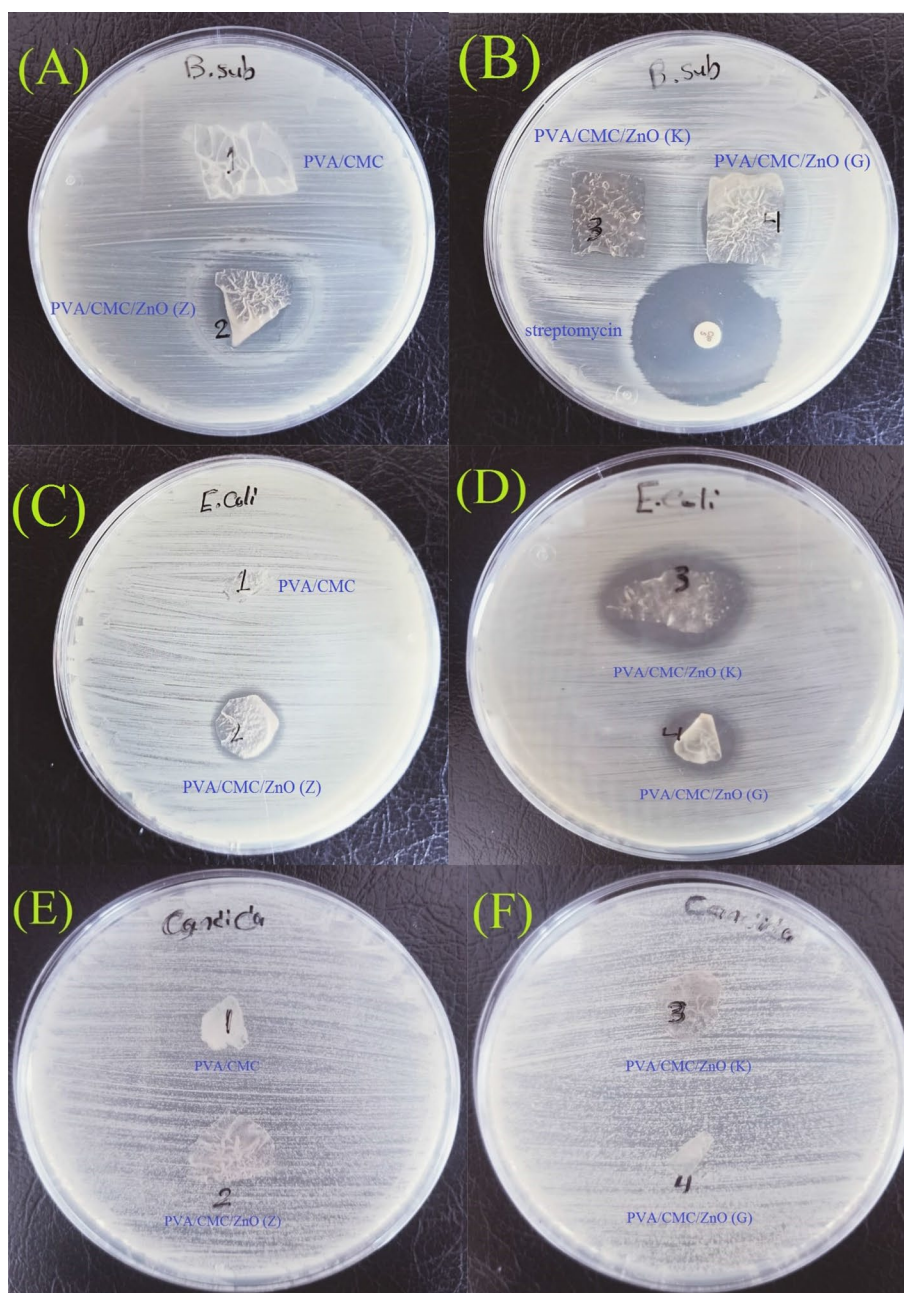
The mechanisms of antibacterial activity of ZnO particles are not well understood, although some statements were proposed, such as the binding of ZnO particles on bacterial surfaces due to the electrostatic forces could be a mechanism [47], the generation of hydrogen peroxide could be the main factor of antibacterial activity [48], or the production of reactive oxygen species, that elevate membrane lipid peroxidation, which causes membrane leakage of reducing sugars, DNA, proteins, and reduces cell viability [49]. Herein,

Zn<sup>2+</sup> ions collide with the cell membrane of bacteria having a negative charge. These electrostatic collision of positive Zn<sup>2+</sup> ions and negative cell membrane may prevent bacterial evolution. So, microbes are feeble that can weaken successively with time. Also, the creation of reactive oxygen species (ROS), including H<sub>2</sub>O<sub>2</sub>, OH<sup>\*</sup>, and O<sup>2\*-</sup> played a crucial character in antibacterial sensation. The O<sup>2\*-</sup>, OH<sup>-</sup> being negatively charged, may stick to the material's surface and can damage DNA [50].

## 4 Conclusions

Pure and doped CMC/PVA blends with ZnO-NPs films were synthesized by the casting method using water as a solvent. The prepared composites were characterized by XRD, FTIR, TEM, and UV/Vis spectroscopy. UV/Vis and FTIR spectroscopy showed the complexation and interaction of both polymers and/or ZnO-NPs appeared in the growth and red-shift

**Fig. 8** Comparison of inhibition zone test between **A, B** *Bacillus subtilis*, **C, D** *Escherichia coli* and **E, F** *Candida albicans* for different samples



**Table 4** Average inhibition zone and approximated efficiency for all prepared samples compared with streptomycin as a positive control

Microbe	Inhibition zone (mm)					Approximated efficiency (%)
	A	B	C	D	E	
<i>Bacillus subtilis</i>	35	–	26	33	28	0, 74, 94 and 80%
<i>Escherichia coli</i>	34	–	19	31	23	0, 56, 91 and 68%
<i>Candida albicans</i>	27	–	–	–	–	0, 0, 0, 0%

(A) Streptomycin as a positive control, (B) CMC/PVA, (C) PCZ-Z, (D) PCZ-K, (E) PCZ-G

of the UV region, which is ascribed to the intermolecular interaction between the hydrogen bonds. The XRD analysis revealed the semi-crystalline nature of the CMC/PVA blend,

which decreased after filling with ZnO-NPs. TEM images confirm the presence of ZnO-NPs with a spherical shape and a diameter between 10 and 40 nm. TEM images also

revealed that the CMC/PVA blend is an effective host matrix for the encapsulation of ZnO-NPs, acting as a good capping agent and giving it environmental and chemical stability. All these data support the idea of complexation between ZnO-NPs and polymer matrix. Antibacterial tests revealed that all samples containing ZnO-NPs had exceptional antibacterial activity and were lethal to the bacteria tested. The PCZ-K sample, in particular, displays a high antibacterial efficiency against *E. coli* (91.17%) and *B. subtilis* (94.28%).

**Author Contributions** AYY: resources, data curation, methodology, validation, formal analysis, investigation, conceptualization, supervision, software, visualization, writing—original draft, writing—review & editing. AMA: resources, methodology, formal analysis. RSS: data curation, methodology, validation, formal analysis, investigation, conceptualization, writing—original draft, writing—review & editing. AET: data curation, methodology, validation, formal analysis, investigation, conceptualization, software, visualization, writing—original draft.

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## Declarations

**Conflict of interest** The authors declare that we have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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