



# Stepwise Excited-state Double Proton Transfer and Fluorescence Decay Analysis

Tomasz Wróblewski<sup>1</sup> · Dzmitryi Ushakou<sup>1</sup>

Received: 8 July 2022 / Accepted: 12 October 2022 / Published online: 22 October 2022  
© The Author(s) 2022

## Abstract

This work considers excited state intramolecular proton transfers (ESIPT) occurred in multiple hydroxyl-containing compounds with one proton transfer site in the normal form. If several hydroxyl groups are located close to each other in a molecule, then the ESIPT process can lead to the next one. A proton donor site in the first ESIPT will be a proton acceptor during the second reaction. Therefore, a number of consecutive excited state proton transfers can occur. This work deals with the case of two successive proton transfers occurred in the molecular system. Such process is called as a stepwise excited state intramolecular double proton transfer (stepwise ESIDPT). It leads to the formation of two molecular tautomers. Therefore, fluorescence of such compounds can contain different emission bands correspond to emission of normal form and two tautomers. In this work, a rigorous analysis of fluorescence decay kinetics has been made using the model with three species, including a normal molecular form and two tautomers. The work presents theoretical framework of fluorescence decay analysis of ESIDPT process taking into account three species emission. Theoretically, the stepwise proton transfers can be consisted of more than two ESIPT reactions. It depends on molecular structure and number of involved hydroxyl groups. Here, a formal analysis of fluorescence decay kinetics has been made in the case of a stepwise process consisting of two proton transfers. Moreover, the quantum-chemical calculations have been performed in the case of scutellarein. It is a multiple hydroxyl-containing flavone and, therefore, it can be applied as a model molecule to study stepwise intramolecular proton transfers. The hypothetical scheme of ESIDPT has been proposed for this compound.

**Keywords** ESIPT · Stepwise ESIDPT · Fluorescence · Emission · Fluorescence decay kinetics

## Introduction

Intramolecular proton transfer processes can lead to significant changes in photophysical properties of compounds. For example, the occurrence of excited state intramolecular proton transfer (ESIPT) can cause dual fluorescence emission due to presence of two different forms of molecules – normal and tautomer [1–4]. One of the first observations of an extremely large Stokes shift caused by excited state intramolecular proton transfer was made for salicylic acid in 1955 by Weller [5]. A year later, Weller reported the dual fluorescence caused by ESIPT experimentally observed in methyl salicylate [6]. Nowadays, due to their properties,

compounds with ESIPT are used in a wide spectrum of applications, including production of luminescent materials [7], fluorescent chemosensors [8, 9], optoelectronic devices [10] and in particular solid state emitters [11], organic lasers [12] and white light-emitting diodes [13].

ESIPT occurs via an intramolecular hydrogen bonding way and it can be described by photophysical cycle with four energy levels. This model was proposed by Sengupta and Kasha in 1979 [14]. In this model, two levels correspond to the ground and the photoexcited states of the normal molecular form, and other two levels correspond to the ground and the excited states of the tautomer. The four energy level model is widely used in investigations of different aspects of ESIPT processes such as, for instance, effect of solvent on proton transfer dynamics [15] and impact of functional groups on ESIPT [16, 17]. An additional interesting issue is multiple ESIPT processes. There are works regarding double proton transfer in various organic compounds [18–22]. These studies consider molecules with two ESIPT sites. A

✉ Dzmitryi Ushakou  
UshakouD@yahoo.com

<sup>1</sup> Institute of Exact and Technical Sciences, Pomeranian University in Słupsk, str. Arciszewskiego 22b, Słupsk 76-200, Poland

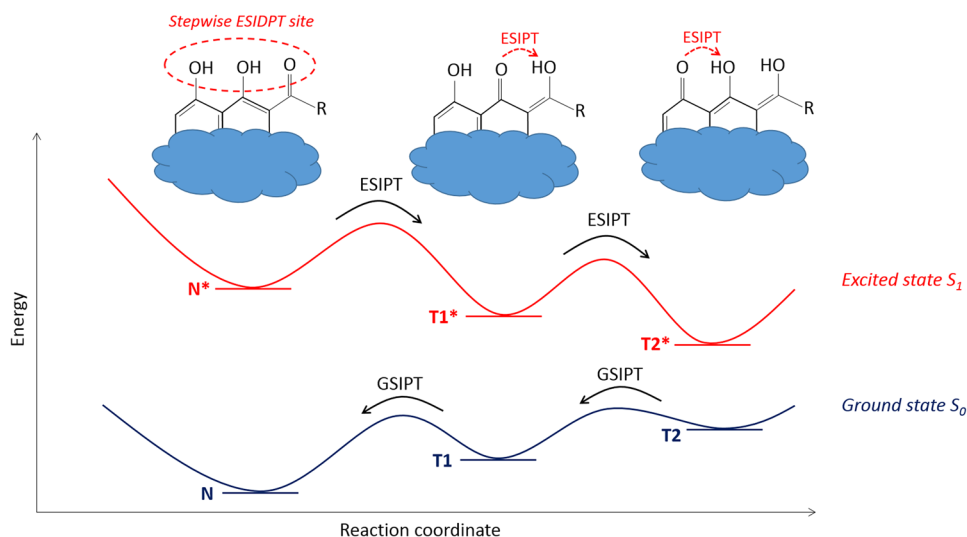
review on photochemical features of compounds with possible excited state double proton transfer was performed by Serdiuk and Roshal in 2017 [23]. In some cases, the presence of double proton transfer causes triple emission from different excited state species [24, 25].

Another interesting issue concerns to multiple hydroxyl-containing compounds with one proton transfer site in the normal form. In this case, if several hydroxyl groups are located close to each other in a molecule, then the ESIPT process can lead to the next one. A proton donor site in the first ESIPT will be a proton acceptor during the second reaction. Therefore, two consecutive excited state proton transfers can occur. Such process can be called as a stepwise excited state intramolecular double proton transfer (stepwise ESIDPT) because it consists of two successive proton transfers. It can occur due to ultrafast dynamics of ESIPT, which take a short time in comparison with excited state lifetimes. Numerous experimental studies have demonstrated that single ESIPT reaction is characterized by ultrafast speed and it can take from less than 1 ps [26–29] to several tens of picoseconds [30]. For instance, the excited state proton transfer is completed in about 0.5–0.6 ps in the case of N-(3-pyridinyl)-2-pyridinecarboxamide [31], and it is even more faster in the case of 3-hydroxyflavone where a time constant of 35–60 fs can be assigned to this process depending on solvent [32]. At the same time, an excited state lifetime of the normal and tautomer form is higher than 100 ps. The measurements have shown that typically the excited state lifetime is in the range from several hundred picoseconds [28, 29, 33, 34] to several nanoseconds [35]. For instance, the excited state tautomer decay time constant is equal to about 850 ps in the case of 3-hydroxyflavone [28]. Therefore, it allows to assume that proton transfer reactions can occur successively in the excited state of compounds containing multiple adjacent hydroxyl groups due to a high rate of ESIPT.

Schematic diagram of the stepwise ESIDPT is depicted in Fig. 1. It presents two consecutive proton transfers within the molecule contained two adjacent hydroxyl groups. The stepwise double proton transfer occurs involving these hydroxyl groups and the adjacent carbonyl oxygen as a proton acceptor. Theoretically, the stepwise proton transfers can be consisted of more than two ESIPTs. It depends on molecular structure and number of involved hydroxyl groups. In this work, a formal analysis of fluorescence decay kinetics will be made in the case of a stepwise process consisting of two ESIPT reactions.

Experimental and theoretical study has shown that the stepwise ESIDPT occurs in 1,8-Dihydroxy-2-Naphthaldehyde [36, 37]. It is a naphthaldehyde derivative with two adjacent hydroxyl groups at positions 1 and 8. It was found that the first ESIPT is ultrafast and irreversible, and it takes less than 150 fs. The second ESIPT is reversible and the rates of forward and backward proton transfer are about  $(1.7 \text{ ps})^{-1}$  and  $(3.6 \text{ ps})^{-1}$ , respectively [36]. Due to ultrafast character of first ESIPT, the rate of this process was faster than the time response of the detection system used in the work [36]. For this reason, the standard model with two species has been applied to analyse the fluorescence decays. This model considers the population kinetics of both tautomeric forms without considering the population of the normal form, because the excited state of the normal form has been almost depopulated in an ultrashort period after excitation [36]. This assumption is fully justified and based on the obtained experimental data for 1,8-Dihydroxy-2-Naphthaldehyde. However, the applied model cannot be expanded on stepwise ESIDPT processes in general, because it does not take into account kinetic rate of the first proton transfer. Therefore, in this work, a rigorous analysis of fluorescence decay kinetics is made using the model with three species, including a normal molecular form and two tautomers. The analysis

**Fig. 1** Schematic diagram of the stepwise ESIDPT consisted of two consecutive proton transfers. The ground states are denoted as N, T1 and T2 for the normal form, tautomer 1 and tautomer 2, respectively. The excited first-singlet states are denoted as N\*, T1\* and T2\* for the normal form, tautomer 1 and tautomer 2, respectively. GSIPT means the ground state intramolecular proton transfer

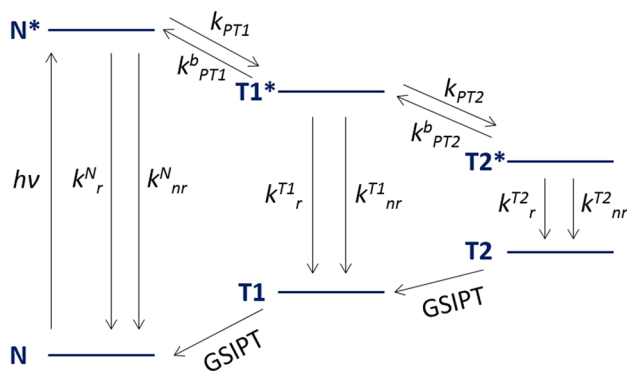


proposed here can be applied in further researches focused on time-resolved spectroscopy of the stepwise ESIDPT systems.

## Theoretical Description

The formal theoretical analysis and the experimental investigation of excited state proton transfer was made by Loken et al. in 1972 [38] and then elaborated by Laws and Brand in the late 1970s [39]. The excited state decay kinetics of two species (naphthol and naphtholate) has been studied, and the theoretical model has been developed in their works [38–40]. Although the proposed model was not initially used to investigate the intramolecular process, but it can be modified and expanded to describe the stepwise ESIDPT processes. This model is widely applied to study ES IPT reactions both in theoretical and practical works [30, 33, 41–43]. For instance, the four levels model of ES IPT was used for theoretical investigation of proton transfer and studying substituent effects on ES IPT mechanism in different derivatives of 2-(2-Hydroxyphenyl)benzothiazole [17].

To discuss the stepwise process consisting of two ES IPT reactions, a three molecular species model will be used below. This model consists of six states: the three ground states and the three excited states. A six energy levels diagram with kinetic rate constants is presented in Fig. 2. In this model, the excited normal form  $N^*$  is formed upon direct excitation ( $h\nu$  in the scheme) and can return to the ground state  $N$  by radiative relaxation (with the rate constant  $k_r^N$ )



**Fig. 2** Six energy levels diagram of the stepwise ESIDPT process consisting of two consecutive proton transfers. The ground states are denoted as  $N$ ,  $T1$  and  $T2$  for normal form, tautomer 1 and tautomer 2, respectively. The excited first-singlet states are denoted as  $N^*$ ,  $T1^*$  and  $T2^*$  for normal form, tautomer 1 and tautomer 2, respectively. ES IPT rate constants are denoted as  $k_{PT1}$  and  $k_{PT2}$ . Backward ES IPT rate constants are denoted as  $k_{PT1}^b$  and  $k_{PT2}^b$ . Rate constants of radiative and non-radiative relaxation are denoted by subscript  $r$  and  $nr$ , respectively. GS IPT means the ground state intramolecular proton transfer. Only the normal form  $N$  is directly excited by absorption of radiation  $h\nu$

or non-radiative relaxation ( $k_{nr}^N$ ). The excited normal form  $N^*$  can also transform into the excited tautomer  $T1^*$  due to forward ES IPT with the rate constant  $k_{PT1}$ . The excited tautomer  $T1^*$  can emit radiation ( $k_r^{T1}$ ) or move to the ground state by non-radiative relaxation ( $k_{nr}^{T1}$ ).  $T1^*$  state can also return to  $N^*$  state by reverse proton transfer. The rate constant of this backward ES IPT is denoted as  $k_{PT1}^b$  in the scheme. However, if an adjacent hydroxyl group will be involved in a next proton transfer reaction, then  $T1^*$  state can transform to the excited state of another tautomeric form  $T2^*$  with the kinetic rate constant  $k_{PT2}$ . The state  $T2^*$  can fluoresce ( $k_r^{T2}$ ) or move to the ground state by non-radiative relaxation ( $k_{nr}^{T2}$ ).  $T2^*$  state can also return to  $T1^*$  state if the backward ES IPT ( $k_{PT2}^b$ ) occurs.

In the following discussion, concentration of  $N^*$ ,  $T1^*$  and  $T2^*$  species will be denoted as  $[N^*]$ ,  $[T1^*]$  and  $[T2^*]$ , respectively. Then, the differential rate expressions for the change of concentration of these species are given by

$$\frac{d[N^*]}{dt} = -(k_r^N + k_{nr}^N + k_{PT1})[N^*] + k_{PT1}^b[T1^*] \quad (1)$$

$$\frac{d[T1^*]}{dt} = -(k_r^{T1} + k_{nr}^{T1} + k_{PT1}^b + k_{PT2})[T1^*] + k_{PT1}[N^*] + k_{PT2}^b[T2^*] \quad (2)$$

$$\frac{d[T2^*]}{dt} = -(k_r^{T2} + k_{nr}^{T2} + k_{PT2}^b)[T2^*] + k_{PT2}[T1^*] \quad (3)$$

Considering all possible decay channels for each species, the following integral decay rate constants can be introduced:

$$K_N = k_r^N + k_{nr}^N + k_{PT1} \quad (4)$$

$$K_{T1} = k_r^{T1} + k_{nr}^{T1} + k_{PT1}^b + k_{PT2} \quad (5)$$

$$K_{T2} = k_r^{T2} + k_{nr}^{T2} + k_{PT2}^b \quad (6)$$

Then, the Eqs. (1)–(3) can be rewritten as

$$\frac{d[N^*]}{dt} = -K_N[N^*] + k_{PT1}^b[T1^*] \quad (7)$$

$$\frac{d[T1^*]}{dt} = -K_{T1}[T1^*] + k_{PT1}[N^*] + k_{PT2}^b[T2^*] \quad (8)$$

$$\frac{d[T2^*]}{dt} = -K_{T2}[T2^*] + k_{PT2}[T1^*] \quad (9)$$

The stepwise ESIDPT can be described by this system of the first order differential equations with the initial boundary condition that only the normal form is directly excited and

populated at  $t = 0$ . Therefore, the boundary condition can be written as follows

$$[N^*]_{at=0} = [N_0^*], [T1^*]_{at=0} = [T2^*]_{at=0} = 0 \quad (10)$$

After making some mathematical operations, the characteristic equation of the system (7)–(9) can be found as

$$x^3 + c_1x^2 + c_2x + c_3 = 0 \quad (11)$$

where

$$c_1 = K_N + K_{T1} + K_{T2} \quad (12)$$

$$c_2 = K_N K_{T1} + K_N K_{T2} + K_{T1} K_{T2} - k_{PT1} k_{PT1}^b - k_{PT2} k_{PT2}^b \quad (13)$$

$$\begin{aligned} c_3 &= K_N K_{T1} K_{T2} - K_{T2} k_{PT1} k_{PT1}^b - K_N k_{PT2} k_{PT2}^b = \\ &= K_N k_{PT2} (k_r^{T2} + k_{nr}^{T2}) + K_N K_{T2} (k_r^{T1} + k_{nr}^{T1}) \\ &\quad + K_{T2} k_{PT1}^b (k_r^N + k_{nr}^N) \end{aligned} \quad (14)$$

It is easily proven that  $c_1$ ,  $c_2$  and  $c_3$  are positive ( $c_1, c_2, c_3 > 0$ ). Therefore, all real roots of the Eq. (11) are negative, what corresponds to decay of the excited state population for all species. As it was mentioned before, forward ES IPT reactions are ultrafast in comparison with excited state relaxation processes. It means that

$$k_{PT1}, k_{PT2} \gg k_r^N, k_{nr}^N, k_r^{T1}, k_{nr}^{T1}, k_r^{T2}, k_{nr}^{T2} \quad (15)$$

The rate constants of the backward ES IPT ( $k_{PT1}^b$  and  $k_{PT2}^b$ ) can be either high (as the forward ES IPT rate) or low and even negligible [42]. It depends on compound properties and cannot be generalized. Then, taking into account the inequality (15), the expressions (12)–(14) can be rewritten as

$$c_1 \cong k_{PT1} + k_{PT1}^b + k_{PT2} + k_{PT2}^b \quad (16)$$

$$c_2 \cong k_{PT1} k_{PT2} + k_{PT1} k_{PT2}^b + k_{PT1}^b k_{PT2} \quad (17)$$

$$c_3 \cong k_{PT1} k_{PT2} (k_r^{T2} + k_{nr}^{T2}) + k_{PT1} k_{PT2}^b (k_r^{T1} + k_{nr}^{T1}) + k_{PT1}^b k_{PT2} (k_r^N + k_{nr}^N) \quad (18)$$

From algebra it is known that the characteristic cubic Eq. (11) has three distinct real roots if and only if its discriminant is positive [44]:

$$-4c_1^3c_3 + c_1^2c_2^2 + 18c_1c_2c_3 - 4c_2^3 - 27c_3^2 > 0 \quad (19)$$

Taking into account the inequality (15) and the expressions (16)–(18), it can be shown that  $c_1^2c_2^2, c_2^3 \gg c_1c_2c_3, c_1^3c_3$  and  $c_1c_2c_3, c_1^3c_3 \gg c_2^3$ , and it can also be proven that  $c_1^2c_2^2 - 4c_2^3 > 0$ . Therefore, in the following we will consider a case where the condition (19) is satisfied. In this case, all three roots of the characteristic Eq. (11) are real

negative numbers, which will be denoted further as  $x_1, x_2$  and  $x_3$  ( $x_1, x_2, x_3 < 0$ ).

Hence, the solution of the system (7)–(9) is given by

$$[N^*](t) = [N_0^*] \left( \alpha_1 e^{-\frac{t}{\tau_1}} + \alpha_2 e^{-\frac{t}{\tau_2}} + \alpha_3 e^{-\frac{t}{\tau_3}} \right) \quad (20)$$

$$[T1^*](t) = [N_0^*] \left( \beta_1 e^{-\frac{t}{\tau_1}} + \beta_2 e^{-\frac{t}{\tau_2}} + \beta_3 e^{-\frac{t}{\tau_3}} \right) \quad (21)$$

$$[T2^*](t) = [N_0^*] \left( \gamma_1 e^{-\frac{t}{\tau_1}} + \gamma_2 e^{-\frac{t}{\tau_2}} + \gamma_3 e^{-\frac{t}{\tau_3}} \right) \quad (22)$$

where  $\tau_1 = -\frac{1}{x_1}$ ,  $\tau_2 = -\frac{1}{x_2}$  and  $\tau_3 = -\frac{1}{x_3}$ ;  $x_1, x_2$  and  $x_3$  ( $x_1, x_2, x_3 < 0$ ) are the roots of the characteristic Eq. (11). The pre-exponential factors are related by the following expressions:

$$\beta_i = \frac{K_N - \frac{1}{\tau_i}}{k_{PT1}^b} \alpha_i, \gamma_i = \frac{k_{PT2}}{K_{T2} - \frac{1}{\tau_i}} \beta_i = \frac{k_{PT2}}{k_{PT1}^b} \frac{K_N - \frac{1}{\tau_i}}{K_{T2} - \frac{1}{\tau_i}} \alpha_i \quad (i = \overline{1, 3}) \quad (23)$$

To satisfy the initial boundary condition (10), the pre-exponential factors must also fulfil the following criteria:

$$\sum_{i=1}^3 \alpha_i = 1 \quad (24)$$

$$\sum_{i=1}^3 \beta_i = 0 \Rightarrow \sum_{i=1}^3 \frac{\alpha_i}{\tau_i} = K_N \quad (25)$$

$$\sum_{i=1}^3 \gamma_i = 0 \Rightarrow \sum_{i=1}^3 \frac{K_N - \frac{1}{\tau_i}}{K_{T2} - \frac{1}{\tau_i}} \alpha_i = 0 \quad (26)$$

Using Vieta's formulas [45] for cubic equations, the following expressions can be derived:

$$\frac{1}{\tau_1} + \frac{1}{\tau_2} + \frac{1}{\tau_3} = c_1 \quad (27)$$

$$\frac{1}{\tau_1\tau_2} + \frac{1}{\tau_1\tau_3} + \frac{1}{\tau_2\tau_3} = c_2 \quad (28)$$

$$\frac{1}{\tau_1\tau_2\tau_3} = c_3 \quad (29)$$

In turn, from (28) and (29), the expression for the sum of the three decay times can be found:

$$\tau_1 + \tau_2 + \tau_3 = \frac{c_2}{c_3} \quad (30)$$

Based on the Eqs. (20)–(22), the measured fluorescence intensity decays can be derived as

$$I_N(t) = s_N k_r^N [N^*](t) = s_N k_r^N \left[ N_0^* \left( \alpha_1 e^{-\frac{t}{\tau_1}} + \alpha_2 e^{-\frac{t}{\tau_2}} + \alpha_3 e^{-\frac{t}{\tau_3}} \right) \right] \quad (31)$$

$$I_{T1}(t) = s_{T1} k_r^{T1} [T1^*](t) = s_{T1} k_r^{T1} \left[ N_0^* \left( \beta_1 e^{-\frac{t}{\tau_1}} + \beta_2 e^{-\frac{t}{\tau_2}} + \beta_3 e^{-\frac{t}{\tau_3}} \right) \right] \quad (32)$$

$$I_{T2}(t) = s_{T2} k_r^{T2} [T2^*](t) = s_{T2} k_r^{T2} \left[ N_0^* \left( \gamma_1 e^{-\frac{t}{\tau_1}} + \gamma_2 e^{-\frac{t}{\tau_2}} + \gamma_3 e^{-\frac{t}{\tau_3}} \right) \right] \quad (33)$$

where  $I_N$ ,  $I_{T1}$  and  $I_{T2}$  are the measured intensities of the  $N^*$ ,  $T1^*$  and  $T2^*$  form emission, respectively;  $s_N$ ,  $s_{T1}$  and  $s_{T2}$  are the spectral sensitivities of the detection channels at wavelengths of the  $N^*$ ,  $T1^*$  and  $T2^*$  form emission, respectively.

Therefore, time-resolved fluorescence measurements and determining components of the emission decay kinetics allows to find parameters  $c_1$ ,  $c_2$  and  $c_3$ . In turn, they can be used to estimate the rate constants of relaxation and ESIPT processes from the Eqs. (12)–(14) or (16)–(18). If not the only normal form is directly excited and there are molecular tautomers before excitation, then the constraints (25) and (26) imposed by the boundary conditions should be changed.

It can be noted that, if the inequality (19) is violated, then the kinetics of the populations will be expressed by a single exponential decay with harmonic oscillations, and the characteristic time of exponential decay will be the same for all species.

## Examples of Compounds with Multiple Adjacent Hydroxyl Groups

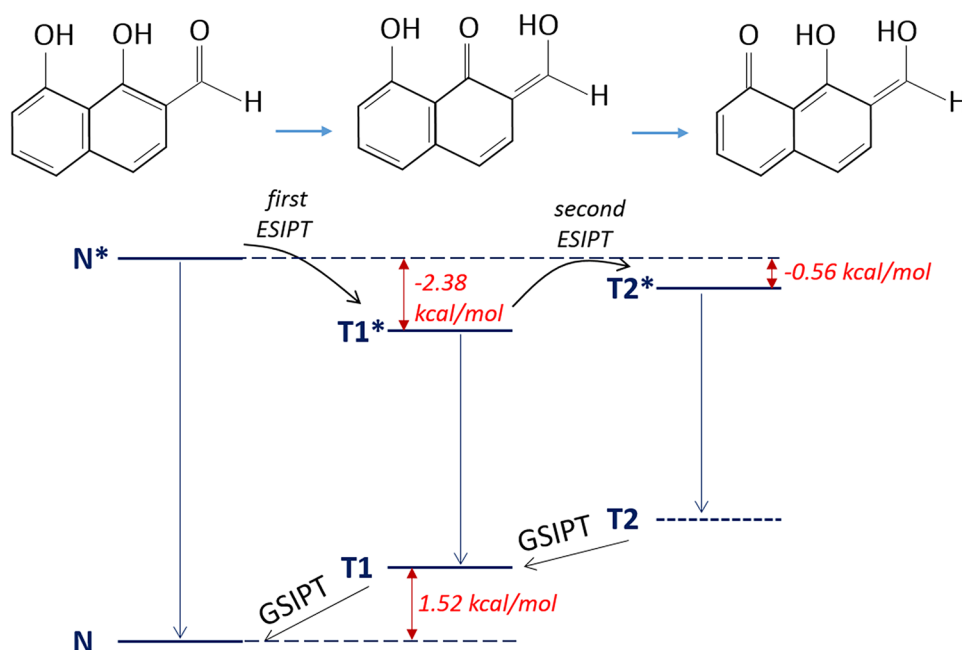
In this section, two compounds containing multiple adjacent hydroxyl groups will be considered from the point of view of the possibility of ESIDPT process.

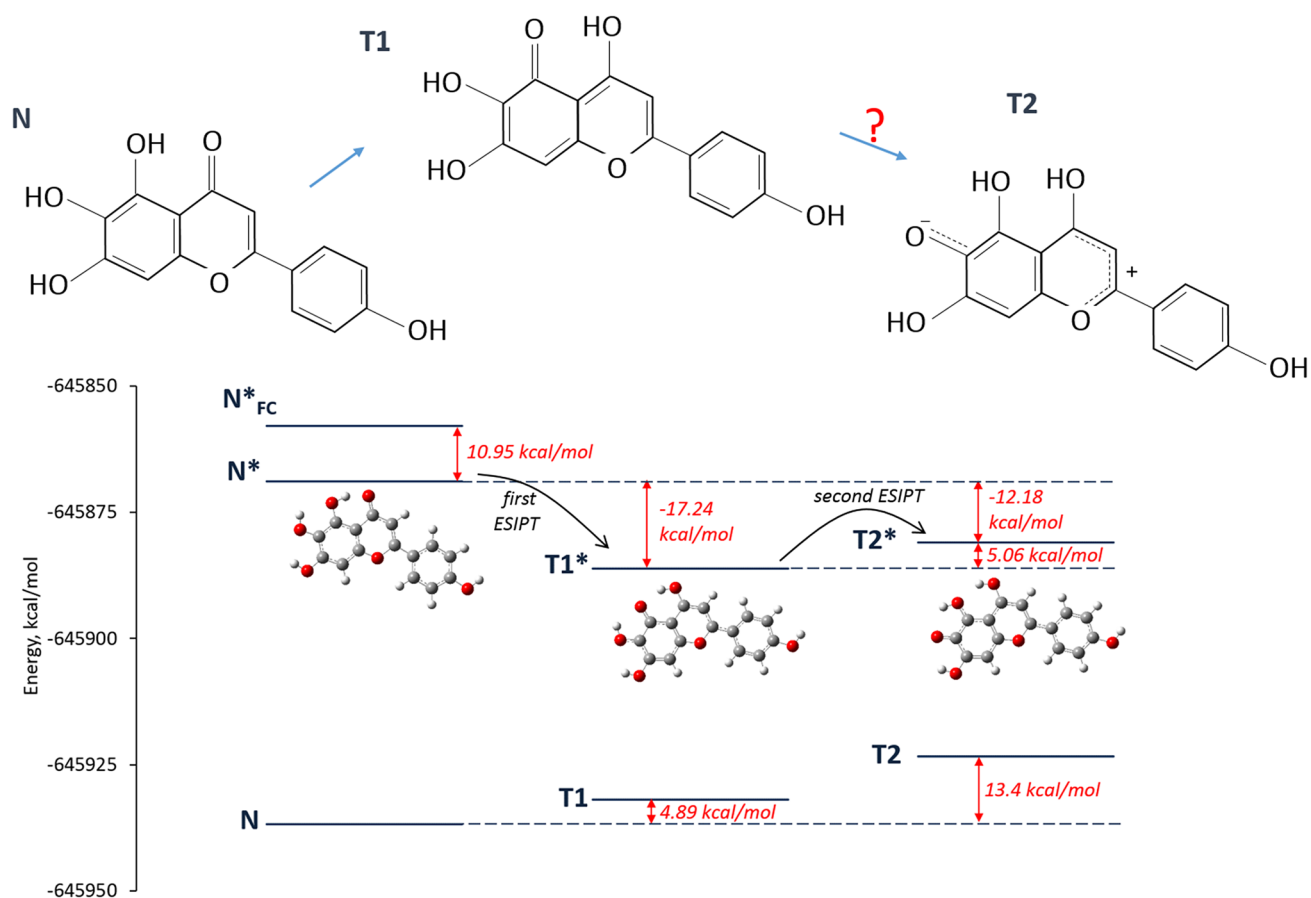
Theoretical and experimental studies have shown that ESIDPT occurs in 1,8-Dihydroxy-2-Naphthaldehyde [36, 37]. The schematic diagram of the proton transfer reactions is depicted in Fig. 3. It was found that the first ESIPT is ultrafast and irreversible, and it takes less than 150 fs. The second ESIPT is fast and reversible, and the rates of forward and backward proton transfer are about  $(1.7 \text{ ps})^{-1}$  and  $(3.6 \text{ ps})^{-1}$ , respectively [36].

It is known, that the ESIPT reaction leads to strong changes in the molecular charge distribution [46, 47]. Therefore, in the case of stepwise ESIDPT, the charge redistribution should occur twice. For this reason, in some compounds with multiple adjacent hydroxyl groups just one ESIPT process occurs while a second proton transfer is blocked because of the impossibility of further charge redistribution in molecule.

The following example is tautomerization in scutellarein. The recent study has shown the possibility of ESIPT in scutellarein [48]. The structure of this flavone also contains three adjacent hydroxyl groups, that allows to consider a hypothetical ESIDPT process. As above mentioned, the first and second proton transfers are associated with charge redistribution within molecule. In the case of such benzopyran

**Fig. 3** Energy diagram of the stepwise ESIDPT in 1,8-Dihydroxy-2-Naphthaldehyde in cyclohexane. Calculated relative energies are taken from the work of Peng et al. [36]. The calculations have been performed at the B3LYP/6–31 + G(d,p) and TD-B3LYP/6–31 + G(d,p) levels for the ground and excited states, respectively [36]





**Fig. 4** Energy diagram of the three different form of scutellarein in acetonitrile. N, T1 and T2 are denoted the ground state of normal form, tautomer 1 and tautomer 2, respectively.  $N^*_{FC}$ ,  $N^*$ ,  $T1^*$  and  $T2^*$  are denoted the Franck–Condon state, excited relaxed normal form and excited tau-

omer 1 and tautomer 2, respectively. The detailed results of the quantum-chemical calculations for each form are given in the Supplementary Information

ring system as scutellarein, ESIDPT process causes a significant change in the electronic distribution. It will lead to a drastic growth of the dipole moment of a tautomeric form. It determines a high activation energy of tautomerization (from T1 to T2). At the same time, the calculations show that the difference between energies of T1 and T2 forms is equal to

about 5 kcal/mol. Therefore, it could be assumed that second ESIPT can occur in scutellarein with low kinetic rate.

This prediction has been verified by quantum-chemical calculations of total energies for normal and tautomeric forms. The obtained results and hypothetical scheme of ESIDPT for scutellarein is presented in Fig. 4. The

**Table 1** Calculated total and relative energies and dipole moments of different forms of scutellarein in acetonitrile

	Ground state			Excited state		
	Total energy, kcal/mol	Relative energy (to the N ground state), kcal/mol	Dipole moment, D	Total energy, kcal/mol	Relative energy (to the N ground state), kcal/mol	Dipole moment, D
Normal form, N	-645936.76	0	4.85	-645868.83	67.93	6.53
Normal form (the Franck–Condon state), $N^*_{FC}$	-	-	-	-645857.88	78.88	4.85
Tautomer 1, T1	-645931.87	4.89	8.84	-645886.07	50.69	11.26
Tautomer 2, T2	-645923.36	13.40	12.72	-645881.01	55.75	12.93

The calculations have been performed at the B3LYP/cc-pVTZ and TD-B3LYP/cc-pVTZ levels for the ground and excited states, respectively

optimization of structures and energy calculations have been performed using Gaussian 09 program [49] based on Density Functional Theory (DFT) at the B3LYP/cc-pVTZ and TD-B3LYP/cc-pVTZ levels [50] for the ground and excited states, respectively. The Polarizable Continuum Model (PCM) together with the Integral Equation Formalism variant (IEF-PCM) [51] has been applied for modelling of solvent effects on molecular structure in acetonitrile.

The calculated total and relative energies and dipole moments are presented in Table 1. The detailed results of the quantum-chemical calculations and obtained optimized structures are given in the Supplementary Information.

However, it should be noted that, three-dimensional potential energy surfaces can provide additional information about transition states and about a way of proton-transfer process. Here, the quantum-chemical calculations have been limited to only studying the stable states (normal and tautomer forms) without transition states. This issue can be considered in detail in the further researches focused on this problem.

## Conclusions

The formal analysis of fluorescence decay kinetics has been made for compounds with stepwise excited state intramolecular double proton transfer (stepwise ESIDPT). Such processes can be observed in molecules with multiple adjacent hydroxyl groups. A three molecular species model has been used instead of model with two molecular forms in the case of single ES IPT. It considers normal form and two different tautomers. This model consists of six energy levels: the three ground states and the three excited states of molecular forms. The analytical solution shows that the fluorescence intensity kinetics are expressed by three-component exponential decays. It was shown that time-resolved fluorescence measurements will allow to estimate decay parameters that are directly related to kinetic rates of intramolecular proton transfers and relaxation processes of normal and tautomeric molecular forms. Moreover, the analytical relations between them and the emission decay parameters have been obtained in the work.

The obtained analytical results could be used in further experimental investigations of compounds with ESIDPT. Time-resolved emission spectroscopy together with the reported results could be used to estimate kinetic rates of proton transfers and, as a result, study of mechanism and reversibility of these intramolecular processes. However, due to ultrafast character of these processes, resolution of time-resolved measurements should be better than in the case of compounds with single ES IPT process.

The quantum-chemical calculations have been performed in the case of scutellarein. The calculated energies of excited states of normal form and tautomers have allowed to propose the hypothetical scheme of stepwise intramolecular proton transfers in this molecule.

**Supplementary Information** The online version contains supplementary material available at <https://doi.org/10.1007/s10895-022-03042-w>.

**Acknowledgements** The cost of Open Access publication was covered under the Transformative Agreement between Springer Nature and ICM Poland (Program publikowania otwartego Springer 2022-2024).

**Authors' Contributions** Tomasz Wróblewski: Conceptualization; Methodology; Quantum-chemical calculations; Writing–Review & Editing. Dzmitryi Ushakou: Conceptualization; Methodology; Formal and mathematical analysis; Quantum-chemical calculations; Writing–Original Draft, Review & Editing. All authors reviewed the manuscript.

**Availability of Data and Materials** The data that supports the findings of this study are available in the Supporting Information, and any additional data regarding the research are available from the corresponding author, D. Ushakou, upon a reasonable request.

## Declarations

**Ethical Approval** Not applicable.

**Competing interests** The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

## References

1. Lakowicz JR (2006) Principles of Fluorescence Spectroscopy, 3rd edn. Springer Science+Business Media, LLC, Baltimore, MD, USA
2. Yang W, Chen X (2014) Dual fluorescence of excited state intra-molecular proton transfer of HBFO: Mechanistic understanding, substituent and solvent effects. *Phys Chem Chem Phys* 16(9):4242–4250. <https://doi.org/10.1039/c3cp54462a>
3. Ushakou DV, Tomin VI (2018) Spectroscopic methods for the study of energetic characteristics of the normal and photoproduct forms of 3-hydroxyflavones. *Spectrochim Acta A Mol Biomol Spectrosc* 204:40–47. <https://doi.org/10.1016/j.saa.2018.06.022>
4. Chrayteh A, Ewels C, Jacquemin D (2020) Dual fluorescence in strap ES IPT systems: A theoretical study. *Phys Chem Chem Phys* 22(2):854–863. <https://doi.org/10.1039/c9cp06261k>

- Weller A (1955) Über die fluoreszenz der salizylsäure und verwandter verbindungen. *Naturwissenschaften* 42(7):175–176. <https://doi.org/10.1007/BF00595299>
- Weller A (1956) Innermolekularer Protonenübergang im angeregten Zustand. *Zeitschrift für Elektrochemie, Ber Bunsenges Phys Chem* 60(9–10):1144–1147. <https://doi.org/10.1002/bbpc.19560600938>
- Zhao J, Ji S, Chen Y, Guo H, Yang P (2012) Excited state intramolecular proton transfer (ESIPT): From principal photophysics to the development of new chromophores and applications in fluorescent molecular probes and luminescent materials. *Phys Chem Chem Phys* 14(25):8803–8817. <https://doi.org/10.1039/C2CP23144A>
- Wu J, Liu W, Ge J, Zhang H, Wang P (2011) New sensing mechanisms for design of fluorescent chemosensors emerging in recent years. *Chem Soc Rev* 40(7):3483–3495. <https://doi.org/10.1039/c0cs00224k>
- Yang Y, Yang L, Ma F, Li Y, Qiu Y (2022) Theoretical investigation on the fluorescent sensing mechanism for recognizing formaldehyde: the TDDFT calculation and excited-state nonadiabatic dynamics. *Chin Phys B*. <https://doi.org/10.1088/1674-1056/ac80af>
- Kwon JE, Park SY (2011) Advanced organic optoelectronic materials: Harnessing excited-state intramolecular proton transfer (ESIPT) process. *Adv Mater* 23(32):3615–3642. <https://doi.org/10.1002/adma.201102046>
- Padalkar VS, Seki S (2016) Excited-state intramolecular proton-transfer (ESIPT)-inspired solid state emitters. *Chem Soc Rev* 45(1):169–202. <https://doi.org/10.1039/c5cs00543d>
- Yan C, Wang X, Liao L (2020) Organic lasers harnessing excited state intramolecular proton transfer process. *ACS Photonics* 7(6):1355–1366. <https://doi.org/10.1021/acsp Photonics.0c00407>
- Peng H, Kang C, Liang M, Chen C, Demchenko A, Chen C, Chou P (2011) En route to white-light generation utilizing nanocomposites composed of ultrasmall CdSe nanodots and excited-state intramolecular proton transfer dyes. *ACS Appl Mater Interfaces* 3(5):1713–1720. <https://doi.org/10.1021/am200229t>
- Sengupta PK, Kasha M (1979) Excited state proton-transfer spectroscopy of 3-hydroxyflavone and quercetin. *Chem Phys Lett* 68(2–3):382–385. [https://doi.org/10.1016/0009-2614\(79\)87221-8](https://doi.org/10.1016/0009-2614(79)87221-8)
- Yang Y, Chen Y, Zhao Y, Shi W, Ma F, Li Y (2019) Under different solvents excited-state intramolecular proton transfer mechanism and solvatochromic effect of 2-(2-hydroxyphenyl) benzothiazole molecule. *J Lumin* 206:326–334. <https://doi.org/10.1016/j.jlumin.2018.10.077>
- Yang Y, Ding Y, Shi W, Ma F, Li Y (2020) The effects of amino group meta- and para-substitution on ESIPT mechanisms of amino 2-(2'-hydroxyphenyl) benzazole derivatives. *J Lumin* 218:116836. <https://doi.org/10.1016/j.jlumin.2019.116836>
- Yang Y, Luo X, Ma F, Li Y (2021) Substituent effect on ESIPT mechanisms and photophysical properties of HBT derivatives. *Spectrochim Acta A Mol Biomol Spectrosc* 250:119375. <https://doi.org/10.1016/j.saa.2020.119375>
- Zhao J, Chen J, Liu J, Hoffmann MR (2015) Competitive excited-state single or double proton transfer mechanisms for bis-2,5-(2-benzoxazolyl)-hydroquinone and its derivatives. *Phys Chem Chem Phys* 17(18):11990–11999. <https://doi.org/10.1039/c4cp05651e>
- Zhao J, Dong H, Zheng Y (2018) Theoretical insights into the excited state double proton transfer mechanism of deep red pigment alkanin. *J Phys Chem A* 122(5):1200–1208. <https://doi.org/10.1021/acs.jpca.7b10492>
- Vérité PM, Guido CA, Jacquemin D (2019) First-principles investigation of the double ESIPT process in a thiophene-based dye. *Phys Chem Chem Phys* 21(5):2307–2317. <https://doi.org/10.1039/c8cp06969g>
- Sun C, Zhang X, Diao L, Cao B, Yin H, Shi Y (2020) How the atomic electron-accepting ability affect the double ESIPT process of 2,5-bis(benzoxazol-2-yl)thiophene-3,4-diol? *J Lumin* 225:117329. <https://doi.org/10.1016/j.jlumin.2020.117329>
- Wang J, Liu Q, Yang D (2020) Theoretical insights into excited-state hydrogen bonding effects and intramolecular proton transfer (ESIPT) mechanism for BTS system. *Sci Rep* 10:5119. <https://doi.org/10.1038/s41598-020-61804-7>
- Serdiuk IE, Roshal AD (2017) Exploring double proton transfer: A review on photochemical features of compounds with two proton-transfer sites. *Dyes Pigm* 138:223–244. <https://doi.org/10.1016/j.dyepig.2016.11.028>
- Sun W, Li S, Hu R, Qian Y, Wang S, Yang G (2009) Understanding solvent effects on luminescent properties of a triple fluorescent ESIPT compound and application for white light emission. *J Phys Chem A* 113(20):5888–5895. <https://doi.org/10.1021/jp900688h>
- Sahu S, Ila, Shankar B, Sathiyendiran M, Krishnamoorthy G (2018) Molecular aggregation to obtain conformer specific enhanced emissions from a triple emissive ESIPT dye. *J Photochem Photobiol A Chem* 353:416–423. <https://doi.org/10.1016/j.jphotochem.2017.11.044>
- Douhal A, Lahmani F, Zewail AH (1996) Proton-transfer reaction dynamics. *Chem Phys* 207:477–498. [https://doi.org/10.1016/0301-0104\(96\)00067-5](https://doi.org/10.1016/0301-0104(96)00067-5)
- Lochbrunner S, Wurzer AJ, Riedle E (2000) Ultrafast excited-state proton transfer and subsequent coherent skeletal motion of 2-(2'-hydroxyphenyl)benzothiazole. *J Chem Phys* 112(24):10699–10702. <https://doi.org/10.1063/1.481711>
- Chevalier K, Wolf MMN, Funk A, Andres M, Gerhards M, Diller R (2012) Transient IR spectroscopy and ab initio calculations on ESIPT in 3-hydroxyflavone solvated in acetonitrile. *Phys Chem Chem Phys* 14(43):15007–15020. <https://doi.org/10.1039/c2cp41077j>
- Verma P, Steinbacher A, Schmiedel A, Nuernberger P, Brixner T (2016) Excited-state intramolecular proton transfer of 2-acetylandan-1,3-dione studied by ultrafast absorption and fluorescence spectroscopy. *Struct Dyn* 3:023606. <https://doi.org/10.1063/1.4937363>
- Tang K-C, Chang M-J, Lin T-Y, Pan H-A, Fang T-C, Chen K-Y, Hung W-Y, Hsu Y-H, Chou P-T (2011) Fine tuning the energetics of excited-state intramolecular proton transfer (ESIPT): White light generation in a single ESIPT system. *J Am Chem Soc* 133(44):17738–17745. <https://doi.org/10.1021/ja2062693>
- Böhnke H, Bahrenburg J, Ma X, Röttger K, Näther C, Rode MF, Sobolewski A, Temps F (2018) Ultrafast dynamics of the ESIPT photoswitch: N-(3-pyridinyl)-2-pyridinecarboxamide. *Phys Chem Chem Phys* 20(4):2646–2655. <https://doi.org/10.1039/c7cp06145e>
- Ameer-Beg S, Ormson SM, Brown RG, Matousek P, Towrie M, Nibbering ETJ, Fogg P, Neuwahl FVR (2001) Ultrafast measurements of excited state intramolecular proton transfer (ESIPT) in room temperature solutions of 3-hydroxyflavone and derivatives. *J Phys Chem A* 105(15):3709–3718. <https://doi.org/10.1021/jp0031101>
- Shynkar VV, Mély Y, Duportail G, Piémont E, Klymchenko AS, Demchenko AP (2003) Picosecond time-resolved fluorescence studies are consistent with reversible excited-state intramolecular proton transfer in 4'-(dialkylamino)-3-hydroxyflavones. *J Phys Chem A* 107(45):9522–9529. <https://doi.org/10.1021/jp035855n>
- Zhang Z, Chen Y-A, Hung W-Y, Tang W-F, Hsu Y-H, Chen C-L, Meng F-Y, Chou P-T (2016) Control of the reversibility of excited-state intramolecular proton transfer (ESIPT) reaction: Host-polarity tuning white organic light emitting diode on a new thiazolo[5,4-d]thiazole ESIPT system. *Chem Mater* 28(23):8815–8824. <https://doi.org/10.1021/acs.chemmater.6b04707>
- Skonieczny K, Yoo J, Larsen JM, Espinoza EM, Barbasiewicz M, Vullev VI, Lee C, Gryko DT (2016) How to reach intense luminescence for compounds capable of excited-state intramolecular proton transfer? *Chem Eur J* 22:7485–7496. <https://doi.org/10.1002/chem.201504944>
- Peng C-Y, Shen J-Y, Chen Y-T, Wu P-J, Hung W-Y, Hu W-P, Chou P-T (2015) Optically triggered stepwise double-proton transfer in an intramolecular proton relay: A case study of 1,8-dihydroxy-2-naphthaldehyde. *J Am Chem Soc* 137(45):14349–14357. <https://doi.org/10.1021/jacs.5b08562>



37. Xie B, Wang K, Jia P, Liu X, Cui G (2022) Excited-State Double Proton Transfer of 1,8-Dihydroxy-2-Naphthaldehyde: A MS-CASPT2//CASSCF Study. *Chin J Chem Phys* 35:422. <https://doi.org/10.1063/1674-0068/cjcp2109163>
38. Loken M, Hayes J, Gohlke J, Brand L (1972) Excited-state proton transfer as a biological probe. Determination of rate constants by means of nanosecond fluorometry. *Biochemistry* 11(25):4779–4786. <https://doi.org/10.1021/bi00775a022>
39. Laws WR, Brand L (1979) Analysis of two-state excited-state reactions. the fluorescence decay of 2-naphthol. *J Phys Chem* 83(7):795–802. <https://doi.org/10.1021/j100470a007>
40. Brand L, Laws W (1983) Excited-State Proton Transfer. *Time-Resolved Fluorescence Spectroscopy in Biochemistry and Biology*. Springer Science+Business Media, LLC, New York, USA, pp 319–340
41. Tomin VI, Ushakou DV (2016) Characterization of ESIPT reactions with instant spectra of fluorescence and complexation processes. *J Lumin* 178:94–105. <https://doi.org/10.1016/j.jlumin.2016.05.042>
42. Tomin VI, Demchenko AP, Chou P-T (2015) Thermodynamic vs. kinetic control of excited-state proton transfer reactions. *J Photochem Photobiol, C* 22:1–18. <https://doi.org/10.1016/j.jphotochemrev.2014.09.005>
43. Yang Y, Zhao J, Li Y (2016) Theoretical study of the ESIPT process for a new natural product quercetin. *Sci Rep* 6:32152. <https://doi.org/10.1038/srep32152>
44. Jacobson N (1985) *Basic Algebra I*, 2nd edn. W.H. Freeman and Company, New York, USA
45. Vinberg EB (2003) *A course in algebra*. American Mathematical Society, Providence, USA
46. Demchenko AP, Tang K-C, Chou P-T (2013) Excited-state proton coupled charge transfer modulated by molecular structure and media polarization. *Chem Soc Rev* 42(3):1379–1408. <https://doi.org/10.1039/c2cs35195a>
47. Yang D, Yang G, Jia M, Song X, Zhang Q, Zhang T, Gao H (2018) Theoretical research on excited-state intramolecular proton coupled charge transfer modulated by molecular structure. *RSC Adv* 8(52):29662–29669. <https://doi.org/10.1039/c8ra05945d>
48. Ushakou D, Wróblewski T (2022) Scutellarein in organic solvents: Changes in spectroscopic properties caused by solute-solvent interactions. *Spectrochim Acta A Mol Biomol Spectrosc* 273:121027. <https://doi.org/10.1016/j.saa.2022.121027>
49. Frisch MJ, Trucks G, Schlegel H, Scuseria G, Robb M, Cheeseman J, Scalmani G, Barone V, Mennucci B, Petersson G, Nakatsuji H, Caricato M, Li X, Hratchian H, Izmaylov A, Bloino J, Zheng G, Sonnenberg J, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Vreven T, Montgomery J Jr, Peralta J, Ogliaro F, Bearpark M, Heyd J, Brothers E, Kudin K, Staroverov V, Keith T, Kobayashi R, Normand J, Raghavachari K, Rendell A, Burant J, Iyengar S, Tomasi J, Cossi M, Rega N, Millam J, Klene M, Knox J, Cross J, Bakken V, Adamo C, Jaramillo J, Gomperts R, Stratmann R, Yazyev O, Austin A, Cammi R, Pomelli C, Ochterski J, Martin R, Morokuma K, Zakrzewski V, Voth G, Salvador P, Dannenberg J, Dapprich S, Daniels A, Farkas O, Foresman J, Ortiz J, Cioslowski J, Fox D (2010) Gaussian 09, Revision C.01. Gaussian Inc, Wallingford CT
50. Becke AD (1993) Density-functional thermochemistry. III. the role of exact exchange. *J Chem Phys* 98(7):5648–5652. <https://doi.org/10.1063/1.464913>
51. Scalmani G, Frisch MJ (2010) Continuous surface charge polarizable continuum models of solvation. I. General formalism. *J Chem Phys* 132(11):114110. <https://doi.org/10.1063/1.3359469>

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.