#### **DELIVERY SYSTEMS**

#### **Original Research**



# OXA-CuS@UiO-66-NH<sub>2</sub> as a drug delivery system for Oxaliplatin to colorectal cancer cells

Marjan Gholami<sup>1</sup> · Azadeh Hekmat<sup>1</sup> · Majid Khazaei<sup>2</sup> · Majid Darroudi 10<sup>3,4</sup>

Received: 16 December 2020 / Accepted: 14 June 2021 / Published online: 28 February 2022  $\ensuremath{\textcircled{}}$  The Author(s) 2022

#### Abstract

In this work, UiO-66-NH<sub>2</sub> was used to prepare a new delivery system by incorporating copper sulfide (CuS) into the pores. The CuS nanoparticles (NPs) were prepared to enhance the anticancer effects of Oxaliplatin (OXA) against colorectal cancer. The oxaliplatin was loaded into CuS@UiO-66-NH<sub>2</sub>. To characterize and investigate their cytotoxicity effects, powder X-ray diffraction (PXRD), Fourier transformation infrared spectroscopy (FT-IR), Brunauer–Emmett–Teller (BET) analysis, UV-Visible analysis, inductively coupled plasma mass spectrometry (ICP-MS), and MTT assay were considered to be performed. According to the observations, the cytotoxicity of OXA-CuS@UiO-66-NH<sub>2</sub> was greater than that of the OXA alone.

#### **Graphical Abstract**



Majid Darroudi darroudim@mums.ac.ir

- <sup>1</sup> Department of Biology, Science and Research Branch, Islamic Azad University, Tehran, Iran
- <sup>2</sup> Metabolic Syndrome Research Center, Mashhad University of Medical Sciences, Mashhad, Iran
- <sup>3</sup> Nuclear Medicine Research Center, Mashhad University of Medical Sciences, Mashhad, Iran
- <sup>4</sup> Department of Medical Biotechnology and Nanotechnology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

# **1** Introduction

The colorectal cancer (CRC) is one of the major causes of death in the world due to the lack of an effective drug that would cause fewer side effects [1, 2]. However, there have been significant improvements in the field of cancer by the exertion of new techniques in designing drug delivery systems. Novel pharmacological systems have proved to be effective in the targeted and specialized treatment of cancer and provided a new field that involves the usage of porous materials for delivery systems [3–5]. Oxaliplatin is a type of chemotherapy drug that is currently approved for the treatment of colorectal cancers. It is a hydrophilic compound of platinum that, like other platinum derivatives, including cisplatin and carboplatin, inhibits DNA synthesis [6]. The compounds that this drug creates to inhibit DNA synthesis are more toxic than cisplatin and carboplatin. In

addition, unlike cisplatin, this drug can inhibit RNA synthesis. However, the in vivo anti-tumor activity of Oxaliplatin has been reported to be low when used alone [7]. This low activity can be attributed to high partitioning to red blood cells and low drug accumulation in tumor tissues after intravenous injection [8]. On the other hand, the clinical efficacy of this drug is limited by its side effects such as neurotoxicity, which are also dose limiters. Side effects of Oxaliplatin and other chemotherapeutic drugs are the result of systemic administration and, as a result, the effect of these drugs on normal cells in addition to cancer cells [9]. Considering the above, it seems necessary to use a new drug delivery system that can reduce the side effects of the drug by entering the tumor site and increasing its antitumor activity by preventing it from entering normal tissues [10]. Therefore, one way to overcome these challenges is to use highly targeted chemical drugs using nanocarriers. In this work, the fabrication of CuS@UiO-66-NH<sub>2</sub> nanomaterials containing Oxaliplatin is considered for inactive targeting. With the remarkable nanoscience development that has been created over the past few decades, porous nanomaterials have been developed [11–13] that can conjointly alleviate various of the aforementioned side-effects when combined via OX into novel drug delivery systems (DDSs). Between the porous substances, metal-organic frameworks (MOFs) are the most flexible in terms of their design flexibility and applicability as novel nanomaterialsbased DDSs [13, 14]. The various toxicological shapes which recently introduced nano MOFs displayed negligible toxicity profiles for carboxylate-based nano MOFs [15]. They are sequestered by the liver and spleen, are biodegradable, degraded, and eliminated from the organism with urine or feces [16–18]. Though different MOF structures have been developed, UiO-66 and its functionalized forms (X-UiO-66, NH<sub>2</sub>, NO<sub>2</sub>, etc.) are of important interest as DDSs because of their intrinsic properties [19]. The MOFsconjugation via another material such as nanoparticles are interesting substitutes that could further increase their potential as the carrier in targeted drug delivery systems [11, 20]. These carriers are widely applied for enhancing the bioavailability, solubility, and controlled release of drugs [21–23]. The nanoporous carriers that are used for the preparation of new pharmaceutics include metalorganic frameworks, which have demonstrated high efficiency in the loading process and caused improvements in drug release [24]. Due to their unique properties such as large and malleable porosity, as well as the presence of modifiable organic groups, it is possible to adjust this framework for specific purposes such as regulating the size of the cavity [24–27]. It is also possible to synthesize metalbased nanoparticles into pores for obtaining new characteristics such as fluorescence to metal-organic frameworks [28–30]. By controlling the factors that affect the release rate of drugs and the amount of drug loading, metal-organic frameworks seem to be very idealistic and exhibit a promising horizon in drug delivery systems [30]. Zirconium-based Metal-organic frameworks, such as UiO-66 and UiO-66-NH<sub>2</sub>, can be considered as a practical alternative for designing drug delivery systems since they contain an adjustable cavity size and high compatibility with physiological systems [30-32]. Some of the advantages of using UiO-66-NH<sub>2</sub> include obtaining the benefits of biological adaptability and the ability to attract large amounts of drugs, as well as having control over the drug release [32-34]. According to the recent findings, copper sulfide nanoparticles can treat cancer by destroying cancerous tumors and stimulating the immune system against cancerous cells. In addition, the copper sulfide that is sized in a nanometer scale has been observed to display different features than its bulk form and upon being exerted in a controlled manner, they have proved to be safe and antineoplastic. It is also notable that next to directly annihilating the cancer cells, copper compounds cause an enhancement in the immune system to cope with cancer as well [34–40]. In this study, copper sulfide nanoparticles were integrated into the structure of UiO-66-NH<sub>2</sub> and applied in vitro to deliver Oxaliplatin to the colorectal cancer cells. The performance of this structure was investigated by the means of MTT assay and flow cytometry (FCM).

# 2 Materials and methods

#### 2.1 Chemicals and instruments

All of the chemicals were purchased from Sigma-Aldrich, Inc. and Merck GmbH (Darmstadt, Germany) unless stated otherwise and used without any further purification. Polyethylene glycol was procured from Pars Azma, Iran, while Methanol and ethanol were obtained from Pars alcohol, Iran. The following instruments were used for the experiments of this study: centrifuge (Froilabo Firlabo SW12R), ultrasonic bath (Bandelin Ultrasonic Bath DT 52 H SONOREX DIGITEC 240W with Heating), X-ray diffractometers equipped with Cu irradiation,  $\lambda = 1.5406 \text{ Å}$ (X'Pert Pro MPD), field emission scanning electron microscopy (TESCAN MIRA FESEM, Czech Republic), **BELSORP MINI X Surface Area and Pore Size Distribution** (from Japan), Fourier transform infrared spectrometer (Thermo Nicolet, Avatar 370 FTIR), the inductivity coupled plasma (OPTIMA 7300DV) and dynamic light scattering and zeta potential analyzer (Zetasizer Nano ZS, Malvern Instruments Ltd.).

#### 2.2 Nanoparticle synthesis

#### 2.2.1 Synthesis of UiO-66-NH<sub>2</sub>

Initially, 0.52 g of 2-aminoterephthalic acid was dissolved in 40 mL of DMF. Then, 0.50 g of  $ZrCl_4$  was dissolved in 20 mL of DMF and 4 mL of HCl by the usage of an ultrasonic bath for 20 min. Once the solutions were mixed, the process was continued by performing sonication for another 20 min. The mixture was poured inside a Teflonlined stainless steel autoclave and heated at 80 °C for 36 h. After being cooled off, the precipitate was separated through centrifugation (3000 rpm/min) to be washed three times with 20 mL of DMF and three times with 20 mL of HCl solutions. The obtained product was set aside at room temperature for two days and activated at 90 °C under vacuum before being used.

#### 2.2.2 Incorporation of copper sulfide nanoparticles into the pores CuS@UiO-66-NH<sub>2</sub>

To begin the process, 0.34 g of CuCl<sub>2</sub> was dissolved in ethanol (15 mL) and ethylene glycol (15 mL) at room temperature. Then, 0.10 g of activated UiO-66-NH<sub>2</sub> was added to the prepared solution and stirred for an hour at room temperature, which resulted in the formation of a green-blue solution (the first solution). In the following, 30 g of thioacetamide was dissolved in ethanol (10 mL) and ethylene glycol (10 mL) and stirred for 20 min at room temperature (the second solution). The second solution was appended to the first mixture at room temperature and allowed to rest for 1 h to achieve a pale yellow-colored solution. The mixture was heated at a temperature of 150 °C for 24 h in a Teflon-lined stainless steel autoclave. After being cooled off, the obtained precipitate was separated by centrifugation (4000 rpm/min) and washed three times with ethanol (20 mL) after being vortexed for 20 min. As the last step, the product was dried at 60 °C for 12 h and from this point, the resulting powder was labeled and called CuS@UiO-66-NH<sub>2</sub>.

## 2.2.3 Preparation of OXA-CuS@UiO-66-NH<sub>2</sub> and OXA@UiO-66-NH<sub>2</sub>

As the first step, 1.0 g of CuS@UiO-66-NH<sub>2</sub> was dispersed in Oxaliplatin solution (5 mL, 5.0 mg/mL) and the suspension was stirred in the dark at room temperature for 48 h. Then, the resultant was centrifuged (4000 rpm/min) and rinsed once with distilled water to be dried later on at room temperature overnight. The obtained product was labeled as OXA-CuS@UiO-66-NH<sub>2</sub> and meanwhile, a similar procedure was performed to prepare OXA@UiO-66-NH<sub>2</sub>.

#### 2.2.4 Cell culture

For this experiment, we procured a CT26 cell line from the Iranian Pasteur Institute. An RPMI medium that contained FBS (10%) and penicillin/streptomycin antibiotics was exerted to culture the cells. The colorectal cancer cells were kept in an incubator chamber at 37 °C with a humidity of 5 and 95% of CO<sub>2</sub>.

#### 2.2.5 Growth inhibition studies

The colorectal cancer cells were treated with the prepared Nano drug and its constituents for 24 h; then, 20  $\mu$ L of MTT solution was added and incubated for 4 h. The MTT was reduced by succinate dehydrogenase during the incubation period, which resulted in the formation of violet formazan crystals. Thereafter, 150  $\mu$ L of DMSO was appended into each well to solubilize the existing crystal. Finally, the absorption was read and recorded by the application of a plate reader at  $\lambda_{max} = 570$  nm.

#### 2.2.6 Annexin V-FITC/PI staining

CT26 cancer cells were seeded in a 6-well plate (cell density =  $7 \times 10^4$  cells per well) to be treated with the composed Nano drug for 24 h and be compared to the control group. Then, the cells were suspended in a 200 µL X Annexin V binding buffer. After being centrifuged at  $400 \times g$  for 5 min, the pellet was separated and suspended in an Annexin V-FITC/PI staining solution. The mixture was incubated at ambient temperature for 15 min and analyzed afterward by the application of FACScalibur flow cytometer and FlowJo software.

# **3 Results**

#### 3.1 Powder X-ray diffraction

Diffraction is a phenomenon in which X-rays are scattered by atoms with a crystalline structure in specific spatial directions that reinforce each other and create stronger rays. This phenomenon is the only direct method of determining the crystalline structure and phase of varying materials. The main purpose of a diffraction experiment is to distinguish the angle of each peak and then, specify the distance of the atomic plane. The crystal structure of UiO-66-NH<sub>2</sub>, CuS-UiO-66-NH<sub>2</sub>, OXA-UiO-66-NH<sub>2</sub>, and OXA-CuS@UiO-66-NH<sub>2</sub> were compared with the reference patterns of CuS and UiO-66-NH<sub>2</sub> to monitor the formation of sample and changes that occur in the course of the functionalization process [36]. The UiO-66 structure with the Reference code of RUBTAK02 in Cambridge structural database <sup>31</sup> was



Fig. 1 PXRD of UiO-66-NH2, CuS, CuS@UiO-66-NH2, and OXA-CuS@UiO-66-NH2

used to analyze the compatibility of prepared UiO-66-NH<sub>2</sub> and the structural maintenance of CuS-UiO-66-NH<sub>2</sub> and OXA-UiO-66-NH<sub>2</sub> after being functionalized with the calculated pattern (Fig. 1). The characteristic peaks of UiO-66, including 7.3° and 8.5°, remained intact during the CuS synthesis and drug loading which was in correspondence to the calculated ones. Furthermore, the performed assessment on the synthesis of CuS contained a reference number of JCPDS # 01-079-2321 that was related to the hexagonal copper sulfide. According to the gathered data, the 2° values of 27.2° (100), 27.7° (101), 29.4° (102), 31.9° (103), 32.8° (006),  $48.0^{\circ}$  (110),  $52.7^{\circ}$  (108), and  $59.4^{\circ}$  (116) were associated with the synthesis of CuS. The presence of CuS in the CuS-UiO-66-NH2 and OXA-CuS@UiO-66-NH2 was proved by observing certain peaks at 29.4°, 32.8°, 48.0°, and 59.4°. The crystallite sizes of UiO-66-NH<sub>2</sub>, CuS-UiO-66-NH<sub>2</sub> and OXA-CuS@UiO-66-NH<sub>2</sub> were calculated by the Scherrer's equation and reported to be 17.9, 22.3, and 21.7 nm, respectively.

#### 3.2 Fourier transform infrared spectroscopy

Fourier transform infrared spectroscopy (FTIR) is a wellknown technique applied for the identification of different materials (Fig. 2). The most important application of FTIR is the performance of a qualitative analysis that includes identifying the functional groups and determining the structure of organic, inorganic, or hybrid species [21]. The FTIR spectrum of UiO-66-NH<sub>2</sub> was consistent with the results that had been reported [39, 41] and used for monitoring the changes that occurred during the synthesizing process and encapsulation. The existence of OXA in UiO-



Fig. 2 FTIR Spectra of CuS, UiO-66-NH<sub>2</sub>, Cus@-UiO-66-NH<sub>2</sub>, and OXA-CuS@UiO-66-NH<sub>2</sub>

66-NH<sub>2</sub> has been confirmed since the OXA loaded UiO-66-NH<sub>2</sub> had exhibited aliphatic C-H stretchings at 2850–2990 cm<sup>-1</sup> after OXA encapsulation. The successful synthesis of CuS has been also approved by the appearance of peaks at  $621 \text{ cm}^{-1}$ , which had been related to CuS vibrations that appeared as a shoulder on the OXA-CuS@UiO-66-NH<sub>2</sub> curve at 597 cm<sup>-1</sup>.

#### 3.3 Brunauer-Emmett-Teller surface area analysis

Brunauer-Emmett-Teller (BET) analysis is one of the methods exerted for measuring and evaluating surface porosity since the accurate measurement of surface area and cavities in many applications, such as catalysts, Nano adsorbents, compounds and additives, pharmaceutical, and food industries as well as nanostructures including metal nanoparticles, nanotubes, nanofibers, etc. is considered of great importance. Among the different methods that are available for determining the porosity, the BET method is known to be based on gas absorption. Herein, the results of this particular analysis exhibited the differences in surface area and porosity before and after functionalization (Fig. 3 and Table 1). Accordingly, it can be perceived that the surface area has faced a significant decrease after functionalization. The most visible changes have occurred after the incorporation of CuS into the UiO-66-NH<sub>2</sub> pores. A decrease in pore size and total pore volume after CuS incorporation is indicative of CuS penetration into the pores while the internal surface area was involved as well.



Fig. 3 The BET analysis of samples

Table 1 Surface area, pore size, and total pore of the UiO-66-NH<sub>2</sub>, CuS@UiO-66-NH<sub>2</sub>, and OXA-CuS@UiO-66-NH<sub>2</sub>

Sample	Surface area (m <sup>2</sup> /g)	Pore size (Å)	Total pore volume $(p/p_0 = 0.990)$
UiO-66-NH <sub>2</sub>	845.9	27.8	0.5874
CuS@UiO-66-NH <sub>2</sub>	357.7	18.5	0.2406
OXA-CuS@UiO-66-NH <sub>2</sub>	69.0	18.5	0.2527

However, there have not been signs of alterations in the pore volume and pore size after the process of drug loading. The attachment of OXA could be highlighted as the most plausible conclusion of this work, which was achieved by exerting the hydrogen bond that exists on the surface of OXA-CuS@UiO-66-NH<sub>2</sub>, leading to a significant decrease in the surface area after the loading since the windows had been blocked by OXA. It is also notable that the presence of CuS can prevent drug penetration into the pores. We have detected a lower loading percentage in comparison with that of the UiO-66-NH<sub>2</sub>, which could be due to the inability of CuS@UiO-66-NH<sub>2</sub> in utilizing its internal surface area after having CuS synthesized into the pores.

#### 3.4 Field emission scanning electron microscopy

Field emission scanning electron microscopy (FESEM) analysis is one of the most widely used methods for studying the surface and morphology of materials. The spherical morphology, which can be observed in the FESEM images, was not altered even after functionalization (Fig. 4). The mean particle size distributions of UiO-66-NH<sub>2</sub>, CuS@UiO-66-NH<sub>2</sub>, and OXA-CuS@UiO-66-NH<sub>2</sub> were measured to be  $150.9 \pm 40.46$ ,  $149.7 \pm 53.19$ , and  $122.5 \pm 48.94$  nm, respectively; however, the particles

appeared in lower sizes after the process of drug loading. Mechanical stirring resulted in decreasing the grain sizes obtained from the solid phase surface analysis. The aggregation of UiO-66-NH<sub>2</sub> and other nanoparticles can be indicated by observing the comparison between the crystallite size and the grain size obtained from FESEM images.

#### 3.5 Dynamic light scattering (DLS)

After measuring the results of DLS analyses, we discovered signs of particle aggregation throughout the solution (Fig. 5). In addition, we observed higher amounts of aggregation in sizes that were over 10 times larger than the crystallite sizes, while the dynamic particle sizes were larger than the solid form as well. The probable reasons for this phenomenon include the interaction of solvent molecules with the surface of nanoparticles and the presence of amine functionals on the MOF. Therefore, it could be assumed that hydrogen bonds stand as the most important interaction in the dynamic sizes of prepared nanoparticles. According to Fig. 5, there was no evident increase in the particle sizes after the modification process, and consequently, any conclusion associated with a tendency to increase or decrease in sizes after modifications is denied.

# 3.6 OXA-UiO-66-NH<sub>2</sub> and OXA-CuS@UiO-66-NH<sub>2</sub> MTT assay

To analyze the results of survival measurement tests such as the MTT assay, we exerted Elisa as a plate reader device (Fig. 6). In this method, the mean absorption of three wells for each dosage is evaluated. Furthermore, the mean absorption of each dosage minus the amount of blank was also used for further calculations (generally done by Elisa reader). To complete this measuring process, the mean absorption of each dosage was divided by the mean absorption of control wells to be multiplied by 100. The calculated numbers represent a percentage of survival rate or Viability. The lack of any significant cytotoxicity from UiO-66-NH<sub>2</sub> was indicated by the results of MTT. The  $IC_{50}$ of OXA-CuS@UiO-66-NH<sub>2</sub> was reported to be 7.97 ppm, which is lower than that of the OXA-UiO-66-NH<sub>2</sub> (37.58 ppm), suggesting that the presence of CuS led to a higher percentage of cellular death. After the loading of OXA obtained by ICP (10.8 and 26.1 %), the concentrations of OXA based IC<sub>50</sub> were 0.8 ppm and 9.8 ppm for the cases of OXA-CuS@UiO-66-NH2 and OXA-UiO-66-NH2, respectively; as it can be observed, OXA-CuS@UiO-66-NH<sub>2</sub> has been more effective than OXA (IC<sub>50</sub>  $\pm$  1.05 ppm). Regarding how the IC<sub>50</sub> of CuS and CuS@UiO-66-NH<sub>2</sub> were 40.7 and 85.1 ppm, respectively, the cytotoxicity of CuS against



Fig. 4 FESEM images and size distributions of the UiO-66-NH<sub>2</sub>, CuS@UiO-66-NH<sub>2</sub>, and OXA-CuS@UiO-66-NH<sub>2</sub>







colorectal cancer cells is quite evident, which also displayed signs of synergism throughout the treatment process at lower doses.

#### 3.7 Flow cytometry

The effects of OXA-CuS@UiO-66-NH<sub>2</sub> on the apoptosis process of the CT26 cell line were investigated by the means of flow cytometry (Fig. 7). After treating the cell for 48 h with a dosage of 100 nM, the amount of apoptosis was calculated through the application of Flow Jo software. According to the results, OXA-CuS@UiO-66-NH<sub>2</sub> is capable of stimulating the apoptosis process of the CT26 cell line [42].

# **4** Discussion

To design drug delivery systems using MOFs, the MOF and guest interactions are needed to be optimized to control spontaneous drug release [14, 17, 23, 43, 44]. The MOF nanocomposites appear to be effective for this purpose. Using metal oxide/MOF nanocomposites such as incorporated magnetic NPs [45] can be used to facilitate drug release by a stimulant. For example, nimesulide-loaded Fe<sub>3</sub>O<sub>4</sub>@HKUST-1 has been used in the treatment of pancreatic cancer, where the release has lasted over 11 days [46].  $\gamma$ -Fe<sub>2</sub>O<sub>3</sub>@MIL-53(Al) was also investigated for controlled ibuprofen release which was over 7 days [47]. The other nanomaterials were gold nanorods that were incorporated into [Al(OH)(1,4-ndc)]<sub>n</sub>,



Fig. 6 MTT assay of CuS, OXA, CuS@UiO-66, OXA@UiO-66-NH<sub>2</sub>, and OXA-CuS@UiO-66-NH<sub>2</sub>

which was also controlled anthracene release [48]. The hybrid nanomaterials including metal oxide/MOFs nanocomposites appear to be promising and effective in inducing drug release by a stimulus. However, Few reports presented them for drug delivery and further investigation is needed to fully understand their potential. Copper sulfide (CuS) is a theranostic agent [49] that can be conjugated with drugs and used for imaging of the tumor microenvironment which can complete the lackings of the MOFs for tracking and monitoring purposes. CuS NPs can be used to design drug delivery systems based on metal oxide/ MOF nanocomposites for theranostic purposes, which could also lead to a better release profile. Herein, the CuS incorporated UiO-66 was used for the first time to investigate the characteristic of the OXA-CuS@UiO-66-NH2 in the killing of the CT-26 cells. PXRD analyses have shown the successful synthesis of the MOF and also CuS nanoparticles. To make sure the CuS NPs were formed into the pores of the MOF, the



Fig. 7 Flow cytometry charts of UiO-66-NH2 and OXA-CuS@UiO-66-NH2 (a and b) and statistics (c)

bet analyses were taken. It was revealed that surface area and the pore size were decreased after the synthesis of CuS NPs. It can be deduced the metal oxide occupied the pore and even closed some windows. After drug loading, FTIR analyses confirmed the presence of C-H stretching of the OXA ligands. PXRD also confirmed that the structure was remained intact after drug loading. Considering the crystallite, FESEM, and hydrodynamic sizes, it appears aggregation is much more intense in the aqueous media. Despite all these obstacles, the samples were meticulously prepared and the powders were thoroughly suspended in the solution for biological tests. The cytotoxicity assay has displayed the effectiveness of the prepared drug delivery system in the killing of the cancer cells concerning OXA IC<sub>50</sub>. It appears the presence of CuS into the pores could work synergistically with the loaded drug. The flow cytometry also confirmed the aforementioned results.

# 5 Conclusion

In conformity to the results of analyzing the synthesized OXA-CuS@UiO-66-NH<sub>2</sub> by the BET method, the formation of CuS within the pores was successfully achieved, however, it led to the lack of using the internal surface for OXA loading. Furthermore, the presence of crystallites aggregation caused an increase in the solid phase and dynamic sizes. In addition, the existence of hydrogen bonds resulted in intensifying the dynamic sizes and caused the 10 percent adsorption of OXA, which was indicated through the ICP analysis. It was also notable that apparently, the higher loading of OXA in UiO-66-NH<sub>2</sub> was due to the internal surface area. Despite all of these facts, it is interesting that CuS was able to act as a cytotoxic agent with more efficacy as a result of being exerted in combination with OXA. This work suggests that the prepared Nano capsule could function more efficiently if the fluorescence of nanoparticles or their photothermal properties were also investigated.

**Acknowledgements** We thankfully acknowledge the technical support for this article provided by Mashhad University of Medical Sciences and the Islamic Azad University of Tehran. The authors also would like to appreciate everyone who helps us in this work.

Author contributions MG: Writing original draft, synthesized and characterized the compounds using XRD, UV-Vis, TRM, FTIR techniques. AH: Data acquisition, analysis and interpretation, review, and editing, Funding, and resources acquisition. MK: Data acquisition, analysis and interpretation, review, and editing. MD: Supervision, Project administration, review & editing, Funding, and resources acquisition.

#### **Compliance with ethical standards**

Conflict of interest The authors declare no competing interests.

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons. org/licenses/by/4.0/.

# References

- Fardsadegh B, Jafarizadeh-Malmiri H. Aloe vera leaf extract mediated green synthesis of selenium nanoparticles and assessment of their in vitro antimicrobial activity against spoilage fungi and pathogenic bacteria strains. Green Process Synth. 2019;8:399–407.
- Ren L, Wu Z, Ma Y, Jian W, Xiong H, Zhou L. Preparation and growth-promoting effect of selenium nanoparticles capped by polysaccharide-protein complexes on tilapia. J Sci Food Agric. 2021;101:476–85.
- Rahdar A, Aliahmad M, Azizi Y. NiO nanoparticles: synthesis and characterization. 2015.
- Xinyi L, Zhiyong W, Yongtai Z, Yongming C. Nanoscale metal organic frameworks for drug delivery. Prog Chem. 2019;31:783.
- Abdul Rahman I, Ayob MTM, Radiman S. Enhanced photocatalytic performance of NiO-decorated ZnO nanowhiskers for methylene blue degradation. J Nanotechnol. 2014;2014:212694 https://doi.org/10.1155/2014/212694.
- Mehmood RK. Review of Cisplatin and oxaliplatin in current immunogenic and monoclonal antibody treatments. Oncology Rev. 2014;8.
- Raymond E, Faivre S, Woynarowski JM, Chaney SG, editors. Oxaliplatin: mechanism of action and antineoplastic activity. Semin Oncol. 1998.
- Gustavsson B, Carlsson G, Machover D, Petrelli N, Roth A, Schmoll H-J, et al. A review of the evolution of systemic chemotherapy in the management of colorectal cancer. Clin Colorectal Cancer. 2015;14:1–10.
- Cassidy J, Misset J-L, editors. Oxaliplatin-related side effects: characteristics and management. Seminars in oncology; 2002: WB Saunders.
- Hoff PM, Saad ED, Costa F, Coutinho AK, Caponero R, Prolla G, et al. Literature review and practical aspects on the management of oxaliplatin-associated toxicity. Clin Colorectal Cancer. 2012;11:93–100.
- Cai W, Wang J, Chu C, Chen W, Wu C, Liu G. Metal–organic framework-based stimuli-responsive systems for drug delivery. Adv Sci. 2019;6:1801526.
- Lin Q, Li W, Liu D, Zhao M, Zhu X, Li W, et al. Porous silicon carrier delivery system for curcumin: preparation, characterization, and cytotoxicity in vitro. ACS Appl Bio Mater. 2019;2:1041–9.
- He C, Lu K, Liu D, Lin W. Nanoscale metal–organic frameworks for the co-delivery of cisplatin and pooled siRNAs to enhance therapeutic efficacy in drug-resistant ovarian cancer cells. J Am Chem Soc. 2014;136:5181–4.

- 14. Huxford RC, Della Rocca J, Lin W. Metal–organic frameworks as potential drug carriers. Curr Opin Chem Biol. 2010;14:262–8.
- Ruyra À, Yazdi A, Espín J, Carné-Sánchez A, Roher N, Lorenzo J, et al. Synthesis, culture medium stability, and in vitro and in vivo zebrafish embryo toxicity of metal–organic framework nanoparticles. Chem–A Eur J. 2015;21:2508–18.
- Li X, Lachmanski L, Safi S, Sene S, Serre C, Greneche J-M, et al. New insights into the degradation mechanism of metal-organic frameworks drug carriers. Sci Rep. 2017;7:1–11.
- 17. Horcajada P, Gref R, Baati T, Allan PK, Maurin G, Couvreur P, et al. Metal–organic frameworks in biomedicine. Chem Rev. 2012;112:1232–68.
- Baati T, Njim L, Neffati F, Kerkeni A, Bouttemi M, Gref R, et al. In depth analysis of the in vivo toxicity of nanoparticles of porous iron (III) metal–organic frameworks. Chem Sci. 2013;4:1597–607.
- Lazaro IA, Forgan RS. Application of zirconium MOFs in drug delivery and biomedicine. Coord Chem Rev. 2019;380:230–59.
- 20. Filippousi M, Turner S, Leus K, Siafaka PI, Tseligka ED, Vandichel M, et al. Biocompatible Zr-based nanoscale MOFs coated with modified poly (ε-caprolactone) as anticancer drug carriers. Int J pharmaceutics. 2016;509:208–18.
- Sabouri Z, Sabouri M, Amiri MS, Khatami M, Darroudi M. Plantbased synthesis of cerium oxide nanoparticles using Rheum turkestanicum extract and evaluation of their cytotoxicity and photocatalytic properties. Mater Technol. 2020:1-14. https://doi.org/ 10.1080/10667857.2020.1863573.
- 22. Rane AV, Kanny K, Abitha VK, Thomas S. Chapter 5 Methods for Synthesis of Nanoparticles and Fabrication of Nanocomposites. In: Mohan Bhagyaraj S, Oluwafemi OS, Kalarikkal N, Thomas S, eds. Synthesis of Inorganic Nanomaterials. Woodhead Publishing; 2018. pp. 121-39.
- Horcajada P, Chalati T, Serre C, Gillet B, Sebrie C, Baati T, et al. Porous metal–organic-framework nanoscale carriers as a potential platform for drug delivery and imaging. Nat Mater. 2010;9:172–8.
- 24. Lin J, Wu X, Zheng C, Zhang P, Huang B, Guo N, et al. Synthesis and properties of epoxy-polyurethane/silica nanocomposites by a novel sol method and in-situ solution polymerization route. Appl Surf Sci. 2014;303:67–75.
- 25. Nilash MM, Hashemzadeh A, Fakhari AR, Amini MM. Novel Schiff base-functionalized metal–organic framework nanoparticles for dispersive solid phase extraction of copper ions from vegetable and water samples. Analytical Methods 2019;11:2683–91.
- 26. Shaabani A, Mohammadian R, Hashemzadeh A, Afshari R, Amini MM. Amine-functionalized MIL-101 (Cr) embedded with Co (ii) phthalocyanine as a durable catalyst for one-pot tandem oxidative A 3 coupling reactions of alcohols. N J Chem. 2018;42:4167–74.
- Noghabi MP, Parizadeh MR, Ghayour-Mobarhan M, Taherzadeh D, Hosseini HA, Darroudi M. Green synthesis of silver nanoparticles and investigation of their colorimetric sensing and cytotoxicity effects. J Mol Struct. 2017;1146:499–503.
- Leszczyński MK, Kornowicz A, Prochowicz D, Justyniak I, Noworyta K, Lewiński J. Straightforward synthesis of singlecrystalline and redox-active Cr (II)-carboxylate MOFs. Inorg Chem. 2018;57:4803–6.
- Inyinbor Adejumoke A, Adebesin Babatunde O, Oluyori Abimbola P, Adelani Akande Tabitha A, Dada Adewumi O, Oreofe Toyin A. Water pollution: effects, prevention, and climatic impact. Water Challenges of an Urbanizing World. 2018;33.
- Haghighi E, Zeinali S. Nanoporous MIL-101 (Cr) as a sensing layer coated on a quartz crystal microbalance (QCM) nanosensor to detect volatile organic compounds (VOCs). RSC Adv. 2019;9:24460–70.
- ÇAyir TaŞDemİRcİ T. Synthesis of copper-doped nickel oxide thin films: Structural and optical studies. Chem Phys Lett. 2020;738:136884.https://doi.org/10.1016/j.cplett.2019.136884.

- Kim KH, Takahashi C, Abe Y, Kawamura M. Effects of Cu doping on nickel oxide thin film prepared by sol–gel solution process. Optik 2014;125:2899–901.
- 33. Amita, Deepak, Arun, Rana PS, editors. Synthesis, characterization and sunlight catalytic performance of Cu doped NiO nanoparticles. AIP Conference Proceedings; 2019: AIP Publishing LLC.
- Ahmad Z, Afzal AM, Khan MF, Manzoor A, Khalil HMW, Aftab S. Copper-doped nickle-oxide nanoparticles for photocatalytic degradation of erichrome black-T and methylene blue and its solar cell applications. J Nanoelectronics Optoelectron. 2019;14:1304–12.
- 35. Hameeda B, Mushtaq A, Saeed M, Munir A, Jabeen U, Waseem A. Development of Cu-doped NiO nanoscale material as efficient photocatalyst for visible light dye degradation. Toxin Rev. 2020:1-11. https://doi.org/10.1080/15569543.2020.1725578.
- 36. Mohamed EF. Removal of organic compounds from water by adsorption and photocatalytic oxidation 2011.
- 37. Zhang Y, Mahdavi B, Mohammadhosseini M, Rezaei-Seresht E, Paydarfard S, Qorbani M, et al. Green synthesis of NiO nanoparticles using calendula officinalis extract: chemical charactrization, antioxidant, cytotoxicity, and anti-esophageal carcinoma properties. Arab J Chem. 2021;14:103105.
- Khan SA, Shahid S, Hanif S, Almoallim HS, Alharbi SA, Sellami H. Green synthesis of chromium oxide nanoparticles for antibacterial, antioxidant anticancer, and biocompatibility activities. Int J Mol Sci. 2021;22:502.
- Khandel P, Shahi SK, Soni DK, Yadaw RK, Kanwar L. Alpinia calcarata: potential source for the fabrication of bioactive silver nanoparticles. Nano Convergence. 2018;5:1–17.
- Somasundaram G, Rajan J. Effectual role of Abelmoschus esculentus (okra) extract on morphology, microbial and Photocatalytic activities of CdO tetrahedral clogs. J Inorg Organomet Polym Mater. 2018;28:152–67.
- Khatri A, Rana PS. Visible light photocatalysis of methylene blue using cobalt substituted cubic NiO nanoparticles. Bull Mater Sci. 2019;42:1–11.
- 42. Sankar S, Sharma SK, An N, Lee H, Kim DY, Im YB, et al. Photocatalytic properties of Mn-doped NiO spherical nanoparticles synthesized from sol-gel method. Optik. 2016;127:10727–34.
- Cunha D, Ben Yahia M, Hall S, Miller SR, Chevreau H, Elkaïm E, et al. Rationale of drug encapsulation and release from biocompatible porous metal–organic frameworks. Chem Mater. 2013;25:2767–76.
- 44. Hashemzadeh A, Drummen GP, Avan A, Darroudi M, Khazaei M, Khajavian R, et al. When a metal-organic framework mediated smart drug delivery meets gastrointestinal cancers. J Mater Chem B. 2021;50.
- 45. Abney CW, Taylor-Pashow KM, Russell SR, Chen Y, Samantaray R, Lockard JV, et al. Topotactic transformations of metal–organic frameworks to highly porous and stable inorganic sorbents for efficient radionuclide sequestration. Chem Mater. 2014;26:5231–43.
- Deshpande RK, Waterhouse GI, Jameson GB, Telfer SG. Photolabile protecting groups in metal–organic frameworks: preventing interpenetration and masking functional groups. Chem Commun. 2012;48:1574–6.
- 47. Küsgens P, Rose M, Senkovska I, Fröde H, Henschel A, Siegle S, et al. Characterization of metal-organic frameworks by water adsorption. Microporous Mesoporous Mater. 2009;120:325–30.
- Khaletskaya K, Reboul J, Meilikhov M, Nakahama M, Diring SP, Tsujimoto M, et al. Integration of porous coordination polymers and gold nanorods into core–shell mesoscopic composites toward light-induced molecular release. J Am Chem Soc. 2013;135:10998–1005.
- 49. Funkhouser J. Reinventing pharma: the theranostic revolution. Curr Drug Disco. 2002;2:17–9.