



Original Research

In vitro targeting and selective killing of mcf-7 and colo320dm cells by 5-fluorouracil anchored to carboxylated SWCNTs and MWCNTs

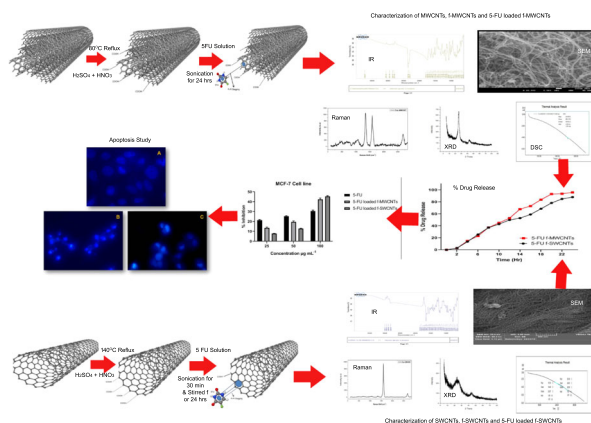
Rutuja V. Kamble¹ · Somnath D. Bhinge¹ · Shrinivas K. Mohite¹ · Dheeraj S. Randive² · Mangesh A. Bhutkar²

Received: 25 November 2020 / Accepted: 28 May 2021 / Published online: 14 June 2021
© The Author(s) 2021

Abstract

The intention of the present work was to synthesize the f-MWCNT and f-SWCNT terminated with proper functional group, loading of 5-Fluorouracil and to perform cytotoxic activity. Functionalization of MWCNTs and SWCNTs was achieved through the acid treatment ($H_2SO_4 + HNO_3$). 5-fluorouracil was loaded into the prepared functionalized CNTs, thereafter; in vitro drug loading capacity and % drug release were calculated. Also the prepared f-CNTs, 5-fluorouracil loaded CNTs were distinguished by using SEM, TGA, DSC, X-ray diffraction, Raman and FTIR spectroscopy. MCF-7 and COLO320DM cells were treated with selected concentrations of 5-FU loaded f-MWCNTs and f-SWCNTs to estimate the cytotoxic activity. It was observed that 5-FU loaded f-SWCNTs showed good activity against selected cell lines than others. Moreover, apoptosis percentage was reported to be 84.46 ± 4.3515 and 92.78 ± 2.6549 for 5-FU loaded f-SWCNTs against MCF-7 and COLO320DM cells respectively. It is evident from the results that the prepared drug loaded CNTs have comparable antitumor activity in cancer cell lines.

Graphical Abstract



1 Introduction

According to the report by WHO, breast, colon, liver, stomach, and lung cancer are the majority types of cancers, claiming lives of several cancer patients worldwide [1]. Breast cancer is the commonest type of cancer affecting around 21,00,000 women each year of which around 6,27,000 patients lose their life [2].

Localized tumors can be easily recovered with surgery; however, most of the aggressive cancers are metastasized and develop resistance to chemotherapy [3]. Therefore, the line of treatment is to use the fixed-dose combinations to inhibit the growth of cells and also to enhance the potential

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1007/s10856-021-06540-8>.

✉ Somnath D. Bhinge
somu1245@yahoo.co.in

¹ Department of Pharmaceutical Chemistry, RCP, Kasegaon, Maharashtra 415 404, India

² Department of Pharmaceutics, RCP, Kasegaon, Maharashtra 415 404, India

of their cytotoxic effect. Unfortunately, the toxicities of combination drug therapy is high to the patients owing to an increase in the drug numbers in the formulation [4]. Moreover, the cost of the anticancer agent(s) is very much high and ultimately, an increase of drug number, will lead to an increase in the cost of the formulation. Paclitaxel and 5-FU can be quoted as a classic example of fixed drug combination used in chemotherapy imparting good anticancer activity but unfortunately, they are interfering with each other [3, 5]. In addition, anticancer drugs pose a major issue with the solubility; often in aqueous medium owing to their hydrophobic nature [6]. To overcome the aforesaid disadvantages, several attempts have been made to develop formulations comprising of anticancer agents encapsulated in nanomaterials [6]. González-Lavado et al. [3] have reported that 5-FU loaded in nanomaterial (MWCNTs) exhibited good therapeutic effect against the human and murine cancer cells [3].

Interestingly, the ideal drug therapy involved with the targeted delivery and selective controlled release to optimize the therapeutic efficacy of the drug and to subsequently minimize its systemic toxicity [7]. In the entire world, Carbon is a dynamic element with their special arrangement and possessing a wide range of properties. For over 6000 years, carbon has been used for the reduction of metal oxides [8]. The different forms of carbon, namely graphite and diamond also belong to the family of chemical elements, and are in rolled layer(s) of graphite structure called carbon nanotubes (CNTs) [9, 10]. According to the layers the types of CNTs are classified as single-walled (SWCNTs) and multi-walled (MWCNTs) CNTs [11]. Moreover, CNTs exhibit wide range of biomedical applications owing to their extraordinary physio-chemical properties [12, 13]. Regrettably, CNTs have major technical impediment with their solubility which resulted in the lower biocompatibility, gastrointestinal absorption, blood transportation, and secretion in spite of being an excellent nanomaterial. Therefore, to overcome this disadvantage, Foldvari and Bagonluri [14], have projected two fundamental elements namely functionalization/modification and dispersion (biomolecular, surfactant-assisted and solvent dispersion) of CNTs [14]. Modification of the surface CNTs i.e., functionalization is a lucrative option for increasing the hydrophilic nature of CNTs. f-CNTs present excellent interaction properties between biological sites and act as target carriers in drug delivery systems, especially as anticancer treatments [15, 16]. A broad range of pharmacotherapeutic agents could be simply conjugated with the f-CNTs due to an easy surface modification [17]. As per the report of Son et al. [18] functionalized CNTs have been used as nanocarriers to transport anticancer drugs, genes, and proteins for chemotherapy [18]. Some researchers have loaded 5-FU in CNTs using numerous techniques, and also proved their anticancer potentials [3, 19–23]. However, none of the researcher has

attempted a study of synthesis, characterization and cytotoxic activity of 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs along with comparative data.

Thus, the present work was undertaken with an intention of synthesizing the f-MWCNT and f-SWCNT terminated with COOH groups, loading of 5-FU into the functionalized nanomaterial (MWCNTs and SWCNTs) and to characterize and perform a comparative study of their cytotoxic activity.

2 Materials and methods

2.1 Experimental

Highly pure (>99%) MWCNTs and (>85%) SWCNTs were procured from AD-nanotechnology Pvt. Ltd., Shimoga, Karnataka 577222. A gift sample of 5-fluorouracil was received from the Divi's Laboratory, Mumbai. All other AR grade chemicals were used for the entire study. The UV and IR spectra were recorded on a UV Spectrophotometer (Jasco V630, Japan) and FTIR spectroscopy (Jasco 4100, Japan) respectively. The solutions were stirred with magnetic stirrer (Remi Instruments, Mumbai). Dissolution test apparatus USP II (Lab India DS8000) was used for the analysis of drugs. The characterization of the prepared samples of CNTs was carried out using Scanning Electron Microscopy (SEM) (VEGA3 TESCAN), XRD (Bruker D2 X-Diffractometer), TGA (Shimadzu DTG-60) and Raman Spectroscopy (EZ Raman Model EZN B532). Breast cancer cell namely MCF-7 and COLO320DM was procured from the NCCS (National Center for Cell Sciences), Pune—411007.

2.2 Selection of the method for the functionalization of MWCNTs and SWCNTs

For COOH group functionalization of MWCNTs and SWCNTs, different types of techniques are available, amongst which base treatment, acid treatment and combined acid treatment are commonly used. The selection of method for the functionalization of MWCNTs and SWCNTs was based on the results of the dispersion stability study. f-MWCNTs and f-SWCNTs (5 mg) were treated with acid (HCl) and its combination (HCl + H₂SO₄). Thereafter, the treated CNTs were dispersed into 5 mL of phosphate buffer solution (pH 7.4) and sonicated for 15 min. Finally, the resultant solutions were kept in an air tight container for 15 days.

2.2.1 Functionalization of MWCNTs

100 mg pure MWCNTs were refluxed with 14M HNO₃ and conc. H₂SO₄ (3:1) at 80 °C for continuous 18 h. Subsequently, the resultant solution was filtered using 0.1 mm PTFE membrane filter paper, then, the residue was washed with deionized

water. Thereafter, the resultant solution was filtered under vacuum filtration, the f-MWCNT product was collected, and the f-MWCNTs were dried overnight at 80 °C under vacuum.

2.2.2 Functionalization of SWCNTs

100 mg pure SWCNTs was added to a solution containing H₂SO₄ and HNO₃ in the proportion 3:1 and 500 mL of distilled water, the entire resultant solution was then sonicated for 4.0 h. Subsequently, the resultant solution was filtered under vacuum filtration. The f-SWCNT product was collected, and the f-SWCNTs were dried overnight at 80 °C under vacuum [24].

2.3 Preparation of 5-fluorouracil drug loaded f-MWCNTs and f-SWCNTs

2.3.1 MWCNTs

100 mg of 5-fluorouracil was dissolved in 10 mL solution containing ethanol:water (1:9) [25]. Subsequently, 800 mg f-MWCNTs were added to this solution with continuous agitation using an ultra sonicator. Thereafter, the resultant mixture was stirred for 3 h, and the dispersion obtained was filtered using membrane filter paper (0.5 µm, Sigma Aldrich) equipped with vacuum filtration assembly. Finally, the residue was washed using ultra pure water. The final drug loaded f-MWCNTs were dried at 40 °C for 24 h [25]. Dry solid product was then stored until further use, whereas the collected filtrate containing unbound 5-fluorouracil was used for the estimation of 5-fluorouracil loading capacity of MWCNTs by using UV–Vis spectrophotometer. The collected filtrate containing unbound 5-fluorouracil was scanned at 280 nm, is the characteristic absorbance wavelength of 5-fluorouracil. Also calibration curve of 5-fluorouracil was accomplished with the parameter. Finally, obtained results were recorded to estimate the drug loading capacity as per Eq. 1; Tarawneh et al. [26].

$$\text{Drug loading capacity} = \frac{W_{\text{initial 5-fluorouracil}} - W_{\text{unbound 5-fluorouracil}}}{W_{\text{initial 5-fluorouracil}}} \times 100 \quad (1)$$

2.3.2 SWCNTs

Briefly, 100 mg of 5-fluorouracil was completely soluble in 800 mL of deionized water to get concentration at 0.125 mg mL⁻¹. Then, 800 mg of f-SWCNTs were dispersed in a 5-fluorouracil solution. The resultant dispersion was sonicated for continuous 30 min (30 °C). Furthermore, the resultant dispersion was stirred continuously for 24 h in the dark with a stirrer. Consequently, the pH of the resultant dispersion was maintained at 4.0 to achieve optimum adsorption of 5-fluorouracil onto the f-SWCNTs. Furthermore, the resultant

dispersion was centrifuged for next 15 min with 5000 rpm. It was then filtrated and rinsed with water. The rinsing procedure was followed with repeated four cycles, finally residue was oven dried at 65 °C and the resultant solid product was then stored in a air tight container till further use, whereas, collected supernatant having unbound 5-fluorouracil was used for the estimation of 5-fluorouracil loading capacity of SWCNTs by using UV–Vis spectrophotometer. The collected supernatant residue containing unbound 5-fluorouracil was scanned at 280 nm which is the characteristic absorbance wavelength of 5-fluorouracil. Also the calibration curve of 5-fluorouracil was plotted. Finally, obtained results were taken to estimate the drug loading capacity as per Eq. 1; Tarawneh et al. [26].

2.4 Characterization of f-MWCNTs, f-SWCNTs, 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs

2.4.1 Fourier transforms infrared spectroscopy (FTIR)

The FTIR spectra of the MWCNTs, SWCNTs, f-MWCNTs, f-SWCNTs, 5FU loaded f-MWCNTs and 5FU loaded f-SWCNTs were recorded in the scale of 4200–400 cm⁻¹.

2.4.2 Scanning electron microscopy (SEM)

Structure of the MWCNTs, SWCNTs, f-MWCNTs, f-SWCNTs, 5FU loaded f-MWCNT, and 5FU loaded f-SWCNT were confirmed by SEM and the images were captured by VEGA3 TESCAN Scanning microscope (Japan) and Jeol SEM (USA).

2.4.3 Raman spectroscopy

The radial breathing mode of CNTs is commonly used to evaluate their diameter by Raman spectroscopy. Degree of disorder of MWCNTs, f-MWCNTs, 5-FU loaded f-MWCNTs, SWCNTs, f-SWCNTs, and 5-FU loaded f-SWCNTs were determined. Side wall of the CNTs were structurally modified with an introduction of defects (sonication) or with the attachment of different chemical groups (functionalization) were also confirmed [26].

2.4.4 Differential scanning calorimeter (DSC)

DSC is employed to check the relative specific heat associated with transitions in CNTs, which help to provide qualitative and quantitative data about physical and chemical changes, involve exothermic and endothermic processes, or changes in heat capacity. Thermal properties of the MWCNTs, SWCNTs, f-MWCNTs, f-SWCNTs, 5FU loaded f-MWCNT, and 5FU loaded f-SWCNT were characterized by the DSC analysis [27].

2.4.5 X-Ray diffraction (XRD)

XRD analysis is analytical technique which is used for the phase characterization of crystalline materials [28]. Samples were scanned in the angular range of 100–600 in a Brucker D2 Phaser X-Diffractometer instrument. The f-MWCNTs, f-SWCNTs, 5-FU loaded f-MWCNTs, and 5-FU loaded f-SWCNTs were placed into instrument, X-ray passed through sample, and results were noted.

2.4.6 Thermo-gravimetric analysis (TGA)

TGA provides the details about chemical and physical properties namely sublimation, absorption, vaporization, adsorption, desorption, chemisorptions, dehydration, decomposition, and redox reactions [29]. The report of TGA represents the change in mass of CNTs substance is recorded as a function of time or temp. or upon heating a material, its weight increases or decreases. The plot of change in weight verses temperature known is as thermogram. Physical and chemical properties of the MWCNTs, SWCNTs, f-MWCNTs, f-SWCNTs, 5FU loaded f-MWCNT, and 5FU loaded f-SWCNT were characterized by the TGA analysis.

2.5 In vitro drug release

Dialysis tubing method was followed for the in vitro drug release analysis of 5-FU from f-SWCNTs and f-MWCNTs containing drug in simulated gastric or an intestinal dissolution fluid [30], the required simulated fluid (pH 7.4, intestinal) was prepared in accordance with the procedure mentioned by Sobh et al. [23].

Before starting the experiments, dialysis bag/tubing having MW cutoff 12,000 Da and and 76 mm in size (Sigma Aldrich) was placed in the selected buffer solution for continuous 3.0 h [31]. Then, 1 mg of the samples was added into 3.0 mL of the dissolution media (buffer saline) containing dialysis tubing. Subsequently, sample containing tubing was kept in a closed receptor compartment containing dissolution media (50 mL, Buffer Saline) with stirring (100 rpm) at 37.0 ± 1 °C [31]. The buffer saline volume level was maintained during the experiment. Finally, 3 mL of drug loaded CNTs sample was withdrawn at regular time interval for UV analysis at the maximum absorption wavelength using the standard calibration curve method.

2.6 Cytotoxic activity

MCF-7 and COLO320DM cells were separately treated with 5-FU loaded f-MWCNT and f-SWCNTs with selected concentrations at 25.00, 50.00, and 100.00 $\mu\text{g mL}^{-1}$. Then the cells were CO₂ incubated at a selected strength of 1×10^4 cells

mL^{-1} in the culture medium for continuous 1 day with 37.00 ± 2.00 °C. Consequently, 100 μL of 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs were dropped into the 96 well micro plates. Then the cells were again incubated for 24 h at 37.00 ± 1.00 °C with CO₂ incubator. Further 20 μL MTT dye was added in a well, and then incubation was continued for continuous 4hr at aforesaid conditions. Lastly, 200 μL Dimethyl sulfoxide solution was added in the each well and kept for further 10 min. Elisa microplate plate reader was used to measure the absorbance at 550–570 nm in triplicate [18, 32].

2.7 Cell morphology studies of 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs by DAPI staining

Microplate covered with cover slip (24 well) having a flat bottom was selected to study the cell morphology. MCF-7 and COLO320DM cells were separately seeded in a plate, further; the plate was incubated in a CO₂ incubator with a 37.00 ± 1.00 °C temperature for an overnight. Subsequently, 100 $\mu\text{g mL}^{-1}$ of 5-FU loaded CNTs (MWCNTs and SWCNTs) solution was added into the labeled well. After the addition of CNTs, plate was again incubated for continuous 1 day (24 h) at 37.00 ± 1.00 °C. Then, the PBS solution was used for washing purpose and the resultant solution was fixed with 4% formaldehyde for half an hour. Further, 20 μL of DAPI staining solution was added in well containing CNTs and incubated for continuous 5 min at ambient temperature in the dark condition. DAPI treated samples of control, 5-FU loaded MWCNTs and 5-FU loaded SWCNTs were examined under fluorescent microscopy.

2.8 Statistical analysis

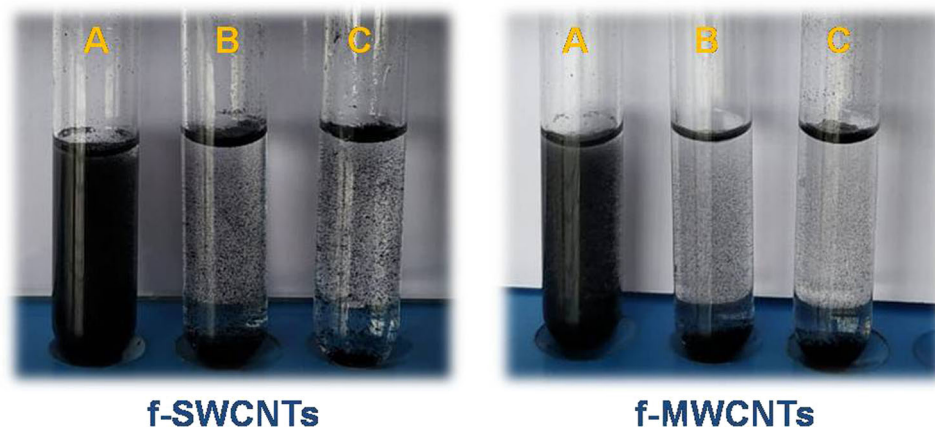
Statistical content of the drug release and cytotoxicity data was examined on Version 5.0, GraphPad Prism. All the cytotoxicity study experiments were conducted in triplicate. The obtained results were explored by using one-way ANOVA with standard error mean (SEM) differences were considered significant with a value as $p < 0.05$.

3 Result

3.1 Selection of the best method for the Functionalization of MWCNTs and SWCNTs

The results of the dispersion study (after 15 days) revealed that combined acid (H₂SO₄ + HNO₃) treatment produce better dispersion (Fig. 1). Therefore, in the present study the acid treatment (H₂SO₄ + HNO₃) was adopted for functionalization of MWCNTs and SWCNTs.

Fig. 1 Dispersion of MWCNTs and SWCNTs after 15 days treatment with (A) combined acid ($\text{H}_2\text{SO}_4 + \text{HNO}_3$), (B) plain acid (HCl), and (C) base



3.2 Characterization of f-MWCNTs, f-SWCNTs, 5-FU loaded f-MWCNTs, and 5-FU loaded f-SWCNTs

3.2.1 FTIR analysis

The functional group and band pattern of the MWCNTs, SWCNTs, f-MWCNTs, f-SWCNTs, 5-FU, 5-FU loaded MWCNTs, and 5-FU loaded SWCNTs was confirmed using FT-IR (Fig. 2).

The IR spectra of MWCNT exhibited weak peaks at 818.22 and 1639.34 cm^{-1} corresponding to the bond stretching of $\text{C}=\text{C}$ and $\text{C}-\text{C}$ present in alkene group respectively. After the functionalization of MWCNTs and SWCNTs; $\text{C}=\text{O}$ (carbonyl), $\text{C}-\text{O}$ and $\text{O}-\text{H}$ (hydroxyl) bond stretching vibration of COOH group were expected in the IR spectra. However, the IR spectra of f-MWCNTs showed the peaks for carbonyl, hydroxyl and $\text{C}-\text{O}$ groups at 1745.41 , 3061.09 , and 1070.93 cm^{-1} respectively, which confirmed the functionalization of MWCNTs. Whereas, the IR spectra of 5FU loaded f-MWCNTs showed the peaks for carbonyl group present in the drug at 1746.35 and 1534.96 cm^{-1} . In addition, hydroxyl, $\text{C}-\text{O}$, $\text{C}-\text{C}$, $\text{C}-\text{N}$, and $\text{N}-\text{H}$ groups present in 5-FU were noted at 3010.77 , 1089.85 , 1351.26 , 986.38 , and 3319.35 cm^{-1} respectively.

The IR spectra of SWCNT exhibited weak peaks at 834.99 and 1638.98 cm^{-1} which is characteristic to the bond stretching of the $\text{C}=\text{C}$ bending and $\text{C}-\text{C}$ in alkene group respectively (Fig. 2). Whereas, the IR spectra of f-SWCNTs showed the peaks for carbonyl, hydroxyl and $\text{C}-\text{O}$ groups at 1650.61 , 3000.11 , and 1193.42 cm^{-1} respectively. Thus, the graph of hydroxyl, carbonyl and $\text{C}-\text{O}$ confirmed functionalization of MWCNTs. The IR spectra of 5-FU loaded f-SWCNTs showed the peaks for carbonyl group at 1728.13 and 1504.08 cm^{-1} . Hydroxyl, $\text{C}-\text{O}$, $\text{C}-\text{C}$, $\text{C}-\text{N}$, and $\text{N}-\text{H}$ groups were noted at 3146.74 , 1113.42 , 1374.81 , 1018.35 , and 3551.11 cm^{-1} respectively. Thus, graph of hydroxyl, carbonyl, and $\text{C}-\text{O}$ thereby confirmed the functionalization of MWCNTs.

3.2.2 SEM analysis

The results of SEM exhibited the morphology of MWCNTs, SWCNTs, f-MWCNTs, f-SWCNTs, 5-FU loaded f-MWCNTs, and 5-FU loaded f-SWCNTs. As revealed in Fig. 3, craggy surface and rounded nanosheets of MWCNTs and SWCNTs were observed. Figure 3A, B, C revealed that MWCNTs, f-MWCNTs, and 5-FU loaded f-MWCNTs with an average size in the range of 18.53 ± 4.2790 , 19.86 ± 4.5550 , and $26.89 \pm 6.1540\text{ nm}$ respectively, whereas, SWCNTs, f-SWCNTs, and 5-FU loaded f-SWCNTs were noted to be 12.08 ± 3.9720 , 17.47 ± 3.5040 , and $18.70 \pm 3.0980\text{ nm}$ (Fig. 3D, E, F). Moreover, histograms of all CNTs are depicted in Fig. 4.

3.2.3 Raman spectroscopy

Degree of disorder of MWCNTs, SWCNTs, f-MWCNTs, f-SWCNTs, 5-FU loaded f-MWCNTs, and 5-FU loaded f-SWCNTs were confirmed by the Raman spectroscopy (Fig. 5). A Raman spectrum D and G band of MWCNTs and f-MWCNTs were noted at 1350 and 1595 cm^{-1} . Disorder in the C (carbon) system and radial breathing form was identified with a D band at 1350 cm^{-1} and graphite form of graphene i.e., G band was noted at 1620 cm^{-1} in the Raman spectra of MWCNTs, f-MWCNTs, and 5-FU loaded f-MWCNTs. Zhao et al. [33]. have confirmed D band peak at 1341 cm^{-1} for disordered form of graphene and 1581 cm^{-1} for graphite form of graphene [33]. A Raman spectrum D and G band of SWCNTs and f-SWCNTs were noted at 1330 and 1585 cm^{-1} respectively. Disorder in the carbon system was observed with a band D at 1330 cm^{-1} (weak peak) and graphite structure of graphene band G was noted at 1585 cm^{-1} (strong peak) in the Raman spectra of MWCNTs, f-MWCNTs, 5-FU loaded f-MWCNTs. The radial breathing mode (RBM) in MWCNTs and SWCNTs was observed at below the 300 cm^{-1} wave number [32, 34].

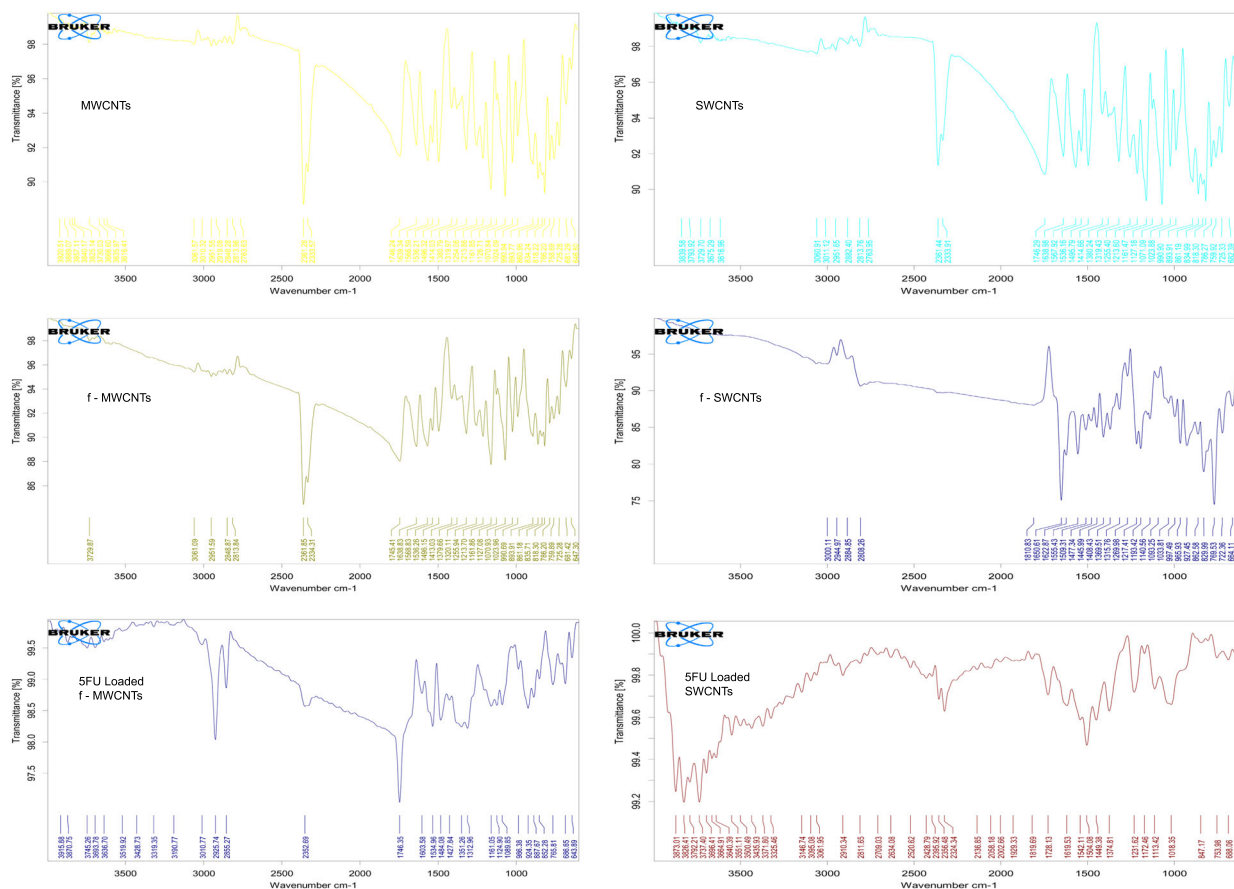


Fig. 2 IR studies of MWCNTs, f-MWCNTs, 5-FU loaded f-MWCNTs, SWCNTs, f-SWCNTs, and 5-FU loaded f-SWCNTs

3.2.4 Differential scanning calorimeter (DSC)

DSC curves for f-MWCNTs and 5-FU loaded f-MWCNTs are depicted in Fig. 6A, C. The f-MWCNTs and 5-FU loaded f-MWCNTs noted sharp exothermic peak at 245.56 °C, corresponding to the melting transitions temperature and decomposition of MWCNTs. Sharp endothermic DSC peak signifies that MWCNTs were in pure crystalline state. In case of f-MWCNTs the DSC pattern of 5-FU loaded f-MWCNTs, indicated that most of the 5-FU was uniformly dispersed in CNTs at molecular level.

DSC curves for f-SWCNTs and 5-FU loaded f-SWCNTs are depicted in Fig. 6B, D. The f-SWCNTs and 5-FU loaded f-SWCNTs noted sharp exothermic peak at 254.98 °C, corresponding to the melting transitions temperature and decomposition of SWCNTs. Sharp endothermic DSC peak signifies that MWCNTs used was in pure crystalline state. In case of f-SWCNTs the DSC pattern of 5-FU loaded f-SWCNTs, which indicate the most of drug was uniformly dispersed in CNTs at the molecular level.

3.2.5 X-Ray diffraction (XRD)

XRD analysis of f-MWCNTs, f-SWCNTs exhibited two broad diffraction patterns with 2θ values of 25.0° and 45.0°. For 5-FU loaded f-MWCNTs, the leading peak at $2\theta = 43.00^\circ$ which is typical peak with an intense and sharp form was observed for f-MWCNTs, which indicated good crystallinity of 5-FU loaded MWCNTs composite. It was also observed that after loading of drug in f-MWCNTs and f-SWCNTs composite, crystallinity was noted with evidence of new varietal and low-intensity peaks (Fig. 7A, B, C, D).

3.2.6 Thermo-gravimetric analysis (TGA)

TGA/DTA was executed at a scanning range from 30 to 600 °C under N_2 gas with heating rate of $20^\circ C min^{-1}$. Inflection temperatures of COOH functionalized SWCNTs were 480, 260, and 60 °C, and the corresponding mass changes occurred at -1.5%, -1.1%, and -1.9%, respectively, as shown in the DTG curves.

Whereas the inflection temperatures of 5-FU loaded f-SWCNTs were observed to be 470, 260, and 50 °C and the

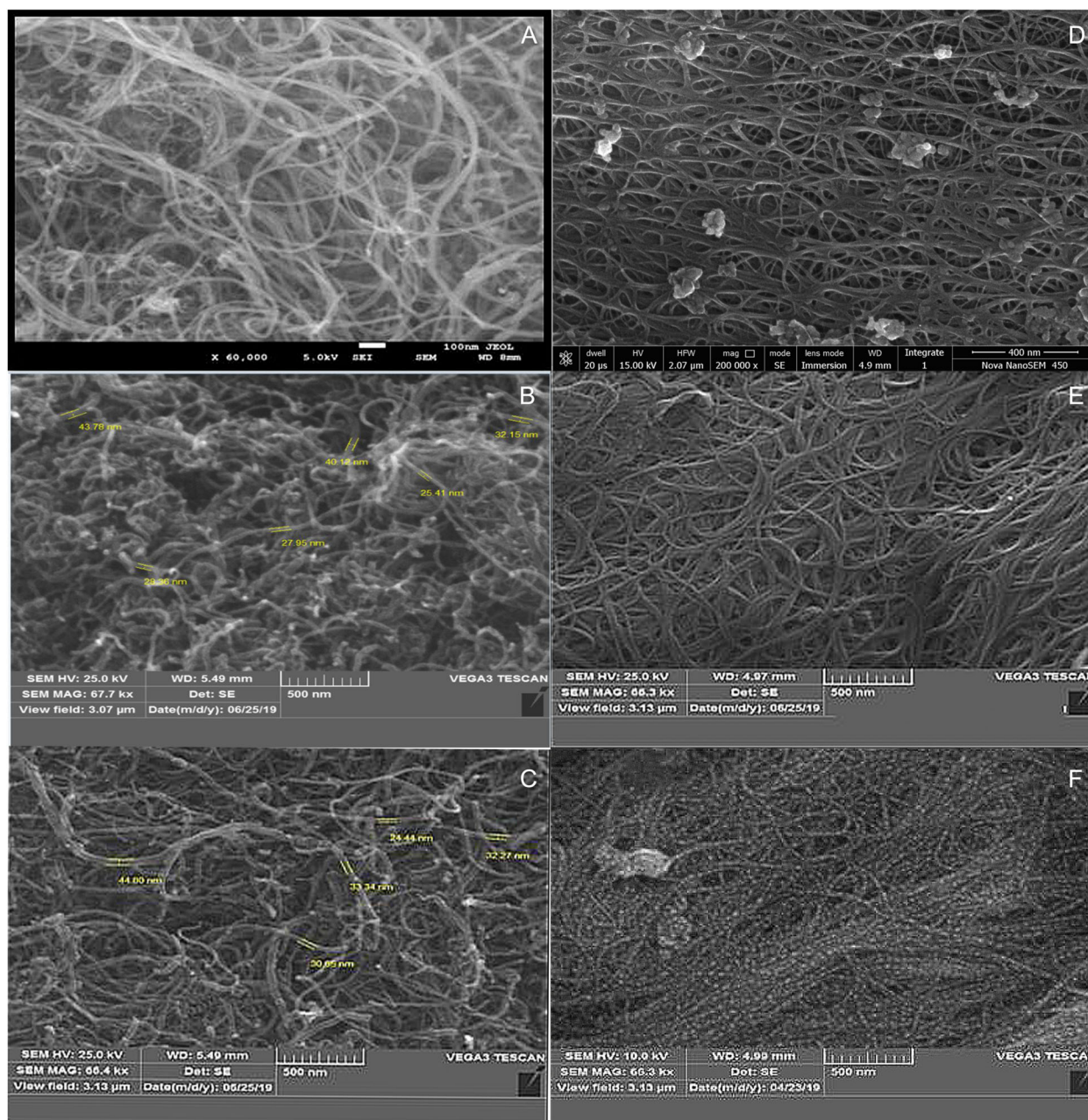


Fig. 3 SEM analysis of (A) MWCNTs, (B) f-MWCNTs, and (C) 5-FU loaded f-MWCNTs (D) SWCNTs, (E) f-SWCNTs, and (F) 5-FU

corresponding mass changes occurred at -0.7% , -1.1% , and -0.8% , respectively, as shown in the DTG curves. Figure 8 showed the differences in the total mass of the f-SWCNTs and 5-FU-SWCNT samples corresponding to the drug.

3.3 The loading rate of 5-FU onto f-MWCNTs and f-SWCNTs

The drug loading rate is the vital part of the targeted drug delivery approach. 5-FU entrapment efficiency was found to be significantly higher for f-MWCNTs and f-SWCNTs than

MWCNTs and SWCNTs at different ratios of the CNTs:5-FU. The 5-FU entrapment efficiency was studied at different ratio of the f-MWCNTs:5-Fu; f-SWCNTs:5-FU. The starting ratio of CNTs (f-SWCNTs/f-MWCNTs):5-FU were 1:0.5(w/w), which increased the concentration of 5-FU only upto 1:4 (w/w). The 5-FU entrapments in CNTs were quantitatively noted at 280 nm by UV-Vis spectroscopy. 5-FU entrapments were observed to be $95.83 \pm 2.57\%$ and 93.43 ± 1.65 in f-MWCNTs and f-SWCNTs respectively. 5-FU is the aromatic heterocyclic compound, due to unique property π - π staging and hydrophobic interactions possible

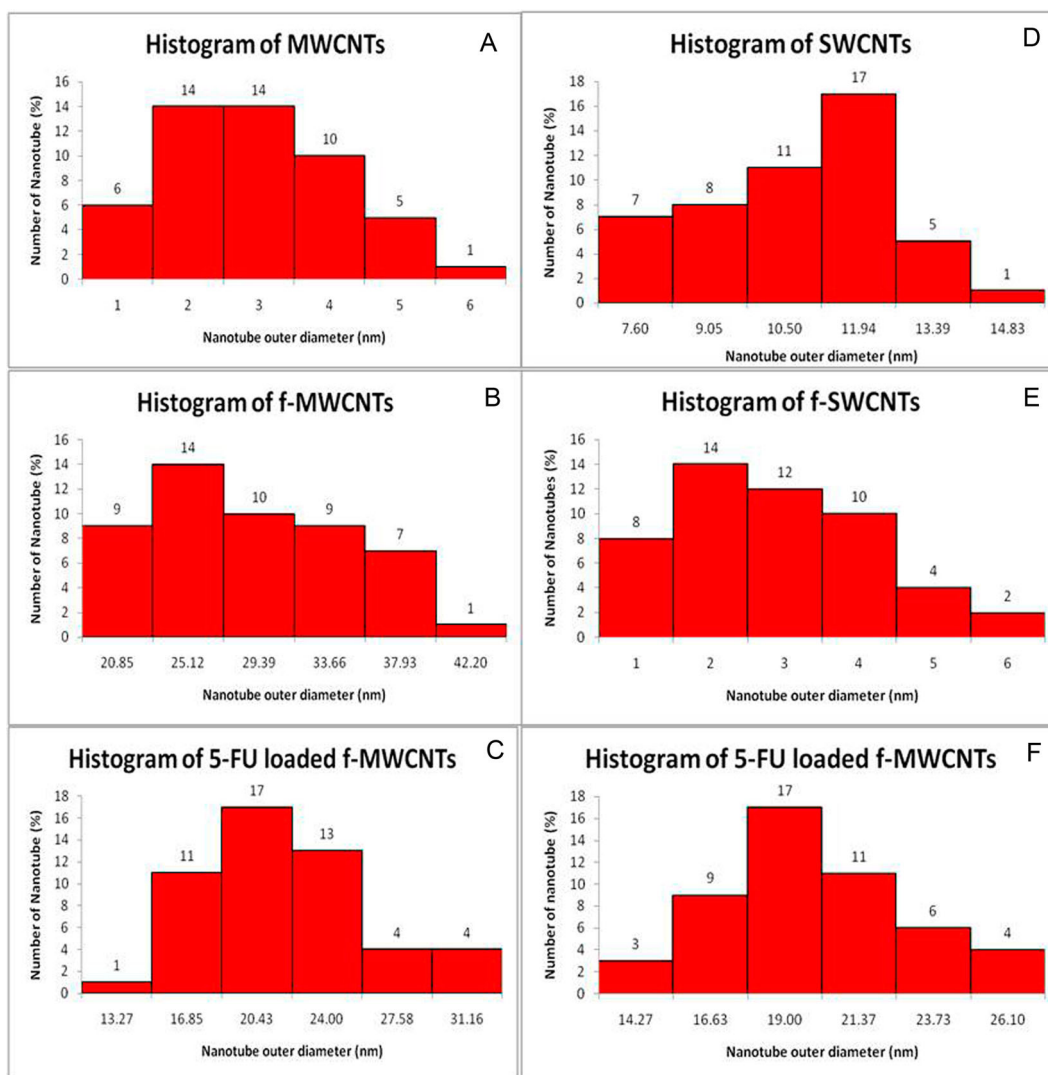


Fig. 4 Histogram of (A) MWCNTs, (B) f-MWCNTs, and (C) 5-FU loaded f-MWCNTs (D) SWCNTs, (E) f-SWCNTs, and (F) 5-FU loaded

in side wall of CNTs and the chances of increasing the entrapment of 5-FU in CNTs. 5-FU is pyrimidine analog containing C=O and N-H group. Thus, the present COOH and OH group in the f-MWCNTs and f-SWCNTs form strong hydrogen bond with C=O and NH group.

3.4 In vitro drug release of 5-FU loaded MWCNTs

Phosphate buffer solution with pH 7.4 was used as dissolution media for conducting in vitro drug release. The drug release profiles of 5-FU loaded f-MWCNTs and f-SWCNTs are shown in Fig. 8. For 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs, the drug release rate was observed to be 44.98 ± 0.7038 and 42.89 ± 0.4904 (at pH 7.4) respectively after 10 h, whereas, 95.90 ± 0.4425 and $88.00 \pm 0.7471\%$ (at pH 7.4) was noted for 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs after 24 h, respectively (Fig. 8). 5-FU loaded f-MWCNTs and f-SWCNTs illustrated an initial burst

release, attributed to the 5-FU loosely anchored to the walls of CNTs or held within the CNTs.

3.5 Cytotoxicity study

Cytotoxicity study was carried out with MTT proliferation assay as previously described by Bhinge et al. 2020 [35], to confirm the sensitivity of MCF-7 and COLO320DM cells against 5-FU (bulk form), 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs. MTT assay setup was executed for the inhibition of MCF-7 and COLO320DM cell proliferation. The selected cell line was treated with various strength solutions of 5-FU (bulk), 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs.

In this study, cell viability was confirmed after addition of 25.00, 50.00, and $100.00 \mu\text{g mL}^{-1}$ solution of 5-FU, 5-FU loaded f-MWCNTs, and 5-FU loaded f-SWCNT after exact 50 h. The inhibition rates were noted to be

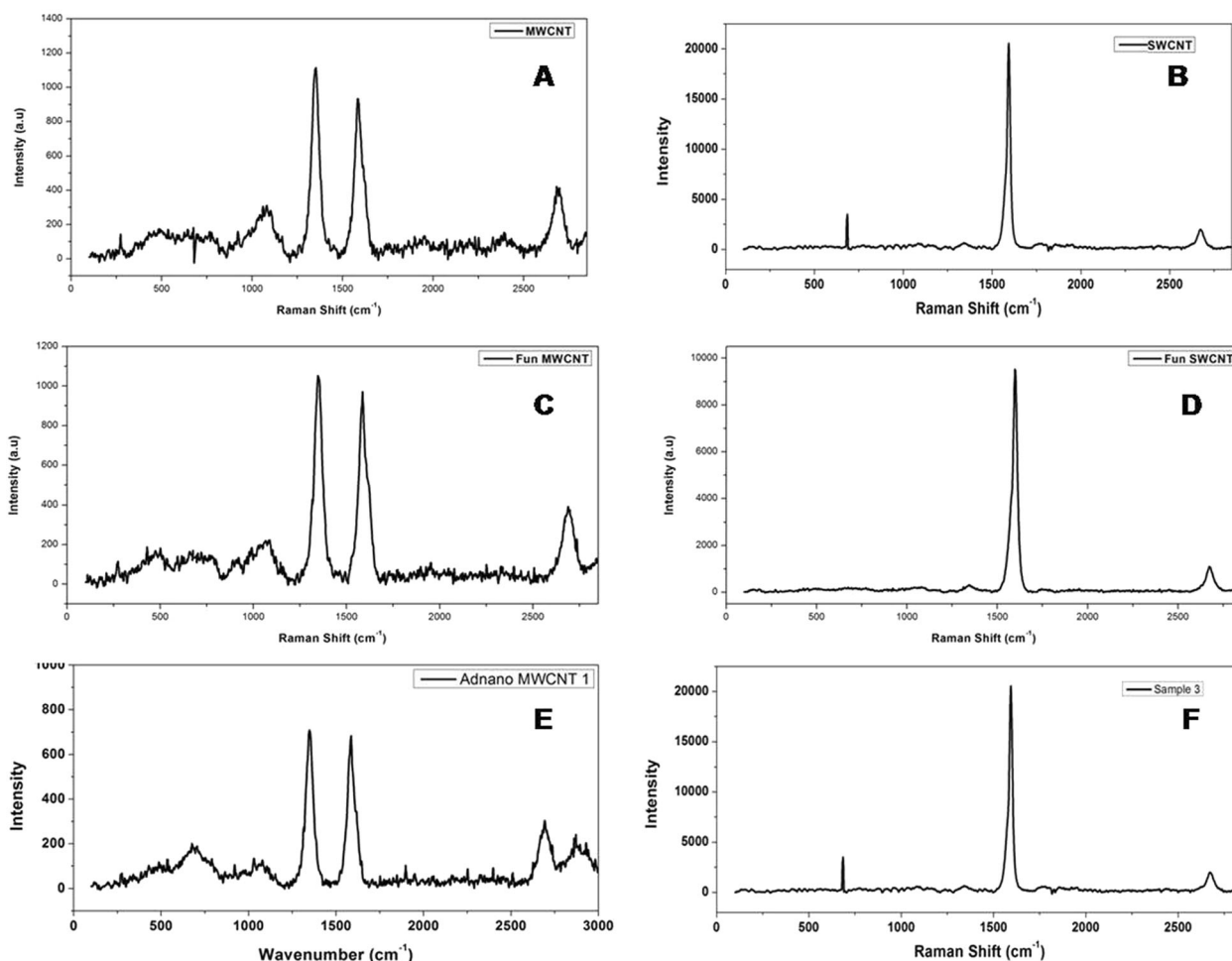


Fig. 5 Raman spectroscopy of (A) MWCNTs, (B) SWCNTs, (C) f-MWCNTs, (D) f-SWCNTs, (E) 5-FU loaded f-MWCNTs, and (F) 5-FU loaded f-SWCNTs

30.59 ± 1.0037 , 45.28 ± 0.9184 , and $42.25 \pm 1.5297\%$ for $100.00 \mu\text{g mL}^{-1}$ of 5-FU, 5-FU loaded f-SWCNTs, and 5-FU loaded f-MWCNTs respectively against MCF-7 cell lines. While, % inhibition was to be noted at 56.72 ± 0.9455 , 67.12 ± 0.7259 , and $58.87 \pm 0.7305\%$ at a concentration of $100 \mu\text{g mL}^{-1}$ for 5-FU, 5-FU loaded f-SWCNTs, and 5-FU loaded f-MWCNTs respectively against the COLO320DM cells (Fig. 9).

3.6 Cell morphology studies of 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs by DAPI staining for fluorescence microscopy

Morphology changes in MCF-7 and COLO320DM cells were identified with DAPI staining approach with the treatment of 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs.

An image of the fluorescence microscopy for control treated and cell treated with 5-FU loaded f-MWCNTs,

and 5-FU loaded f-SWCNTs are depicted in Fig. 10. The control exhibited normal intact nuclei of the cells with weak homogenous blue staining pattern. On the contrary, the cells treated with 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs were noted as smaller nuclei with bright chromatin condensation, by blebbing, nuclear fragmentation, and apoptotic bodies means smaller spherical fragments formation. Apoptosis percentage was reported to be 4.22 ± 1.86 , 41.72 ± 3.56 , and 84.46 ± 4.35 for control, 5-FU loaded f-MWCNTs, and 5-FU loaded f-SWCNTs, respectively, against the MCF-7 cells. Whereas, in the COLO320DM cells the % inhibition was observed to be 2.65 ± 2.0123 , 45.58 ± 2.0925 , and 92.78 ± 2.6549 for control, 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs respectively (Table 1). The results clearly indicated that 5-FU loaded MWCNTs and 5-FU loaded SWCNTs induced apoptosis in MCF-7 and COLO320DM cells when compared with control (Fig. 10).

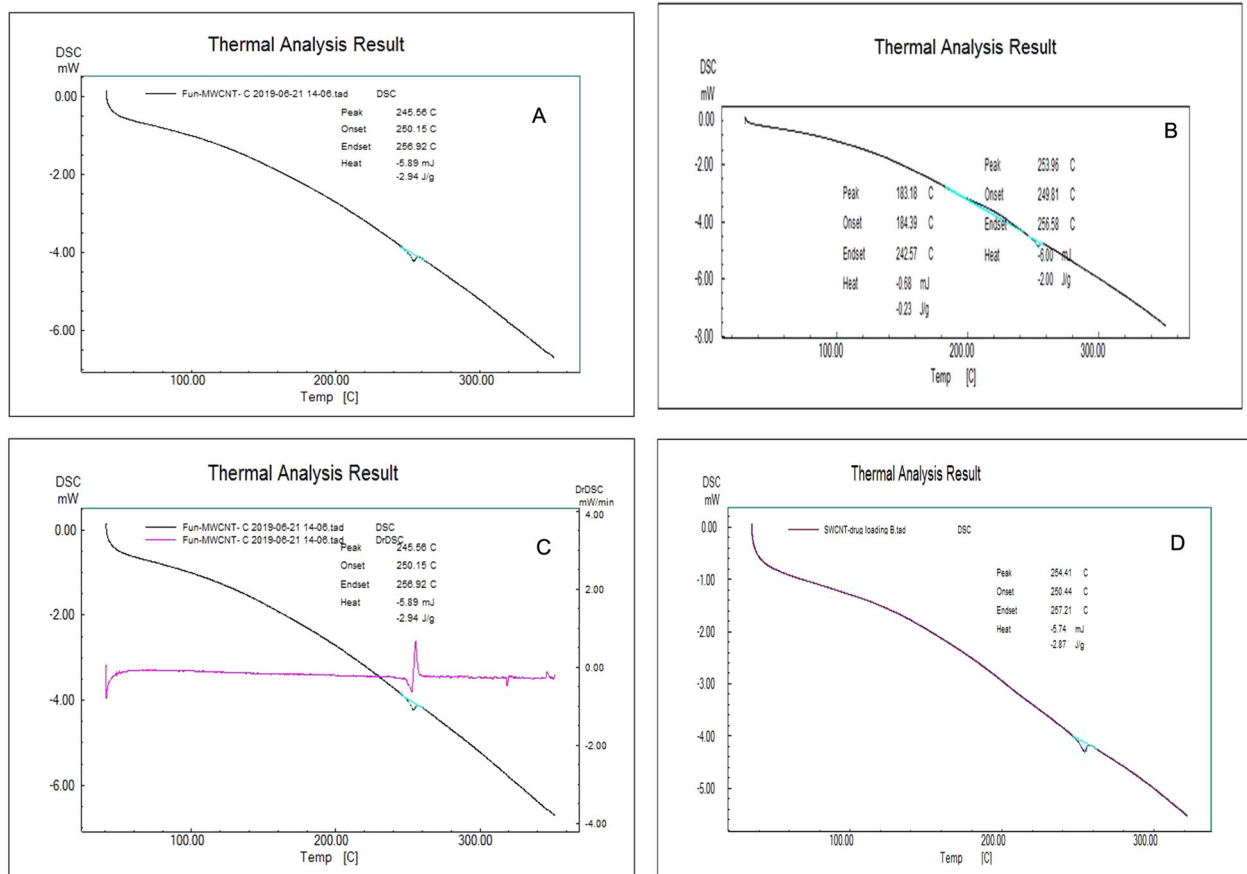


Fig. 6 Differential scanning calorimeter (DSC) images of (A) f-MWCNTs, (B) f-SWCNTs, (C) 5-FU loaded f-MWCNTs, and (D) 5-FU loaded f-SWCNTs

4 Discussion

Conventional drug delivery system is often confronted with the problem of poor site specificity which may lead to unwanted effects of the drugs; especially in the case of anticancer therapies. In the effective treatment of tumors it is vital that the drug should target the tumor site in optimum concentration, which can be achieved by use of specific carriers. Therefore, it's an urgent need to find out a suitable carrier for the anticancer drug.

CNTs are considered as a promising tool for the delivery of various therapeutic molecules owing to its small size and high specific surface areas which facilitates their adsorption on biological substrates [18]. CNTs are known to increase the cell permeability in cancer cells [18].

Moreover, due to effective interaction properties between biological site(s), CNT's are considered to be excellent target carriers in drug delivery systems, especially for cancer treatments [15, 16]. MWCNTs and SWCNTs exhibit wide ranges of physical properties. Plain CNTs are hydrophobic in nature and exhibit less interaction toward the targeted site, however, modification of the surface CNTs

i.e., functionalization is a lucrative option for increasing the hydrophilic nature of CNTs which will contribute to maximum chemical binding of drug moieties thereby increasing the drug concentration in the CNTs. Thus, in the present study the CNTs have been functionalized with acids to anchor the $-\text{COOH}$ group at the side wall of the CNTs and thereafter, the drug under study was anchored with the modified CNTs- COOH .

Pyrimidine analogs have been commonly used in cancer treatments and the most common derivative namely; 5-FU is an example of this class which could treat several solid tumors like liver, rectal, colon, pancreatic, ovarian, breast, gastric, and bladder cancer etc. However, 5-FU poses the problem of poor absorption and has been reported with severe side effects. Therefore, it is necessary to increase its site specific action which will facilitate its effective absorption along with fewer side effects. An attempt was thus made in the direction to incorporate 5-FU into the CNTs as discussed earlier.

González-Lavado et al. [3]. clearly stated that 5-FU was loaded into plain CNTs via π -stacking onto the side wall of MWCNTs [3]. However, plain CNTs possess less disperse

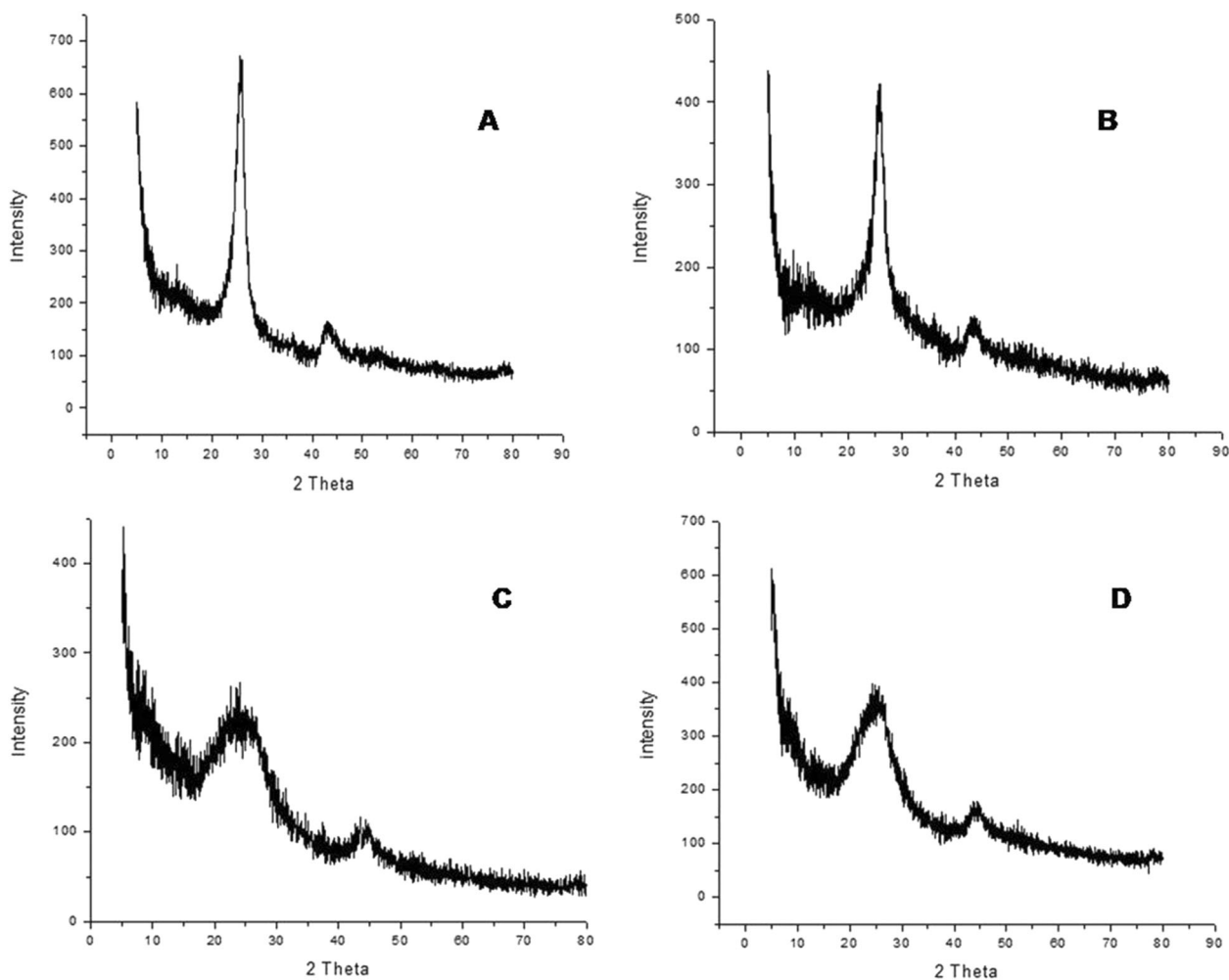


Fig. 7 X-ray diffraction images of (A) f-MWCNTs, (B) f-SWCNTs, (C) 5-FU loaded f-MWCNTs, and (D) 5-FU loaded f-SWCNTs

properties and less stability for loading of 5-FU. Therefore, in the present study we have employed f-CNTs instead of plain CNTs. The probable mechanism involved may be in accordance with the mechanism stated by González-Lavado et al. [3]. Also, another possibility is an interaction of NH group of 5-FU with COOH group of f-CNTs with liberation of water and the said fact is depicted in IR spectrum of 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs wherein, the peak of C–O group is not observed, however in IR spectra of f-MWCNTs and f-SWCNTs the group peaks are clearly seen (Fig. 2). Moreover, Fig. 8 depicts the TGA study where about 8% of the total mass of f-SWCNT sample corresponds to 5-FU loaded f-SWCNT.

In the present study the CNTs were functionalized with the mixture of $\text{H}_2\text{SO}_4 + \text{HCl}$, which was selected based on the revealed results of the dispersion study for 15 days, which have been earlier reported. Moreover, all the synthesized CNTs were characterized using hyphenated tools viz; FTIR, SEM, Raman, DSC, XRD, and TGA analysis. The aforesaid techniques revealed the morphological

properties of CNTs and drug moiety. 5-FU entrapment was observed to >90% for both CNTs due to may be π - π staging and hydrophobic interactions on side wall of MWCNTs and SWCNTs as discussed earlier. Also, the % drug release of 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs exhibited control release pattern over a period of 24 h as compared to pure 5-FU, therefore, the system will be beneficial to achieve targeted as well as controlled drug delivery. Importantly, the prepared CNTs were evaluated for the in vitro anticancer activity. It was observed that the drug loaded f-SWCNTs exhibited better % inhibition rate than the 5-FU loaded f-MWCNTs and plain 5-FU against MCF-7 and COLO320DM cell line. 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs showed better percent inhibition than the 5-FU may be due to cell permeability nature of the CNTs [18]. Furthermore, probable mechanism was verified with apoptosis studies, our study revealed that all studied parameters exhibited almost same results for both CNTs, however in apoptosis study 5-FU loaded f-SWCNTs showed the maximum apoptotic

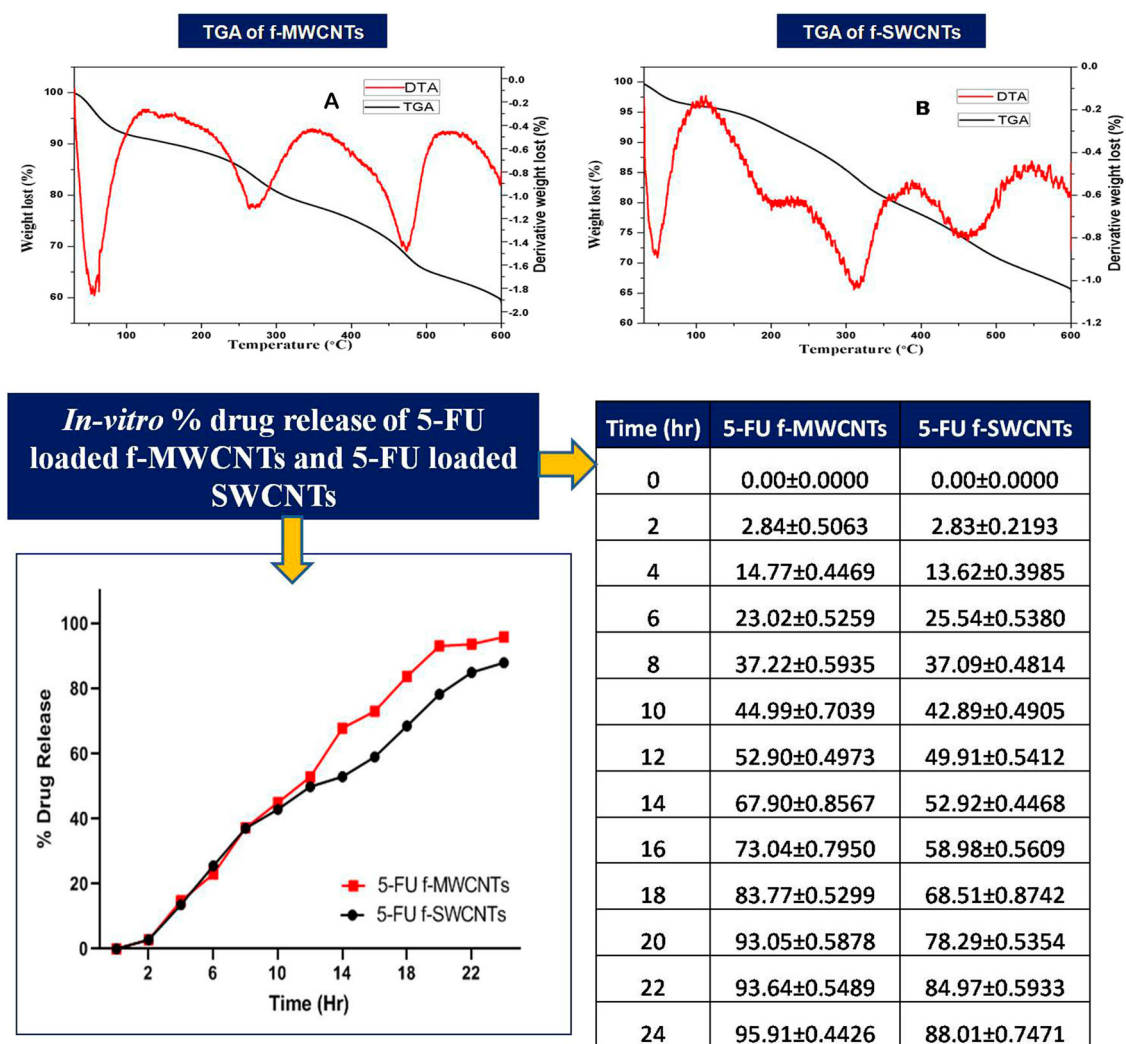


Fig. 8 Thermo-gravimetric analysis (TGA) images of (A) f-SWCNTs and (B) 5-FU loaded f-SWCNTs; and In vitro % Drug release of 5-FU loaded f-MWCNTs and 5-FU

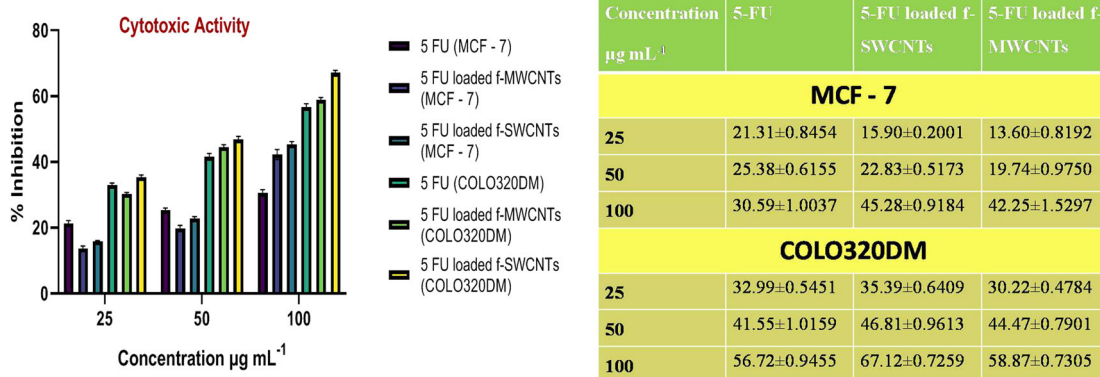


Fig. 9 Cytotoxic activity of 5-FU, 5-FU loaded MWCNTs, and 5-FU SWCNTs against the MCF-7 and COLO320DM cell line

cell ($84.46 \pm 4.35\%$), as compared to 5-FU loaded f-MWCNTs (41.72 ± 3.56). Based upon the said facts, the 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs can

enhance the uptake of 5-FU in MCF-7 and COLO320DM cells. This is presumably due to the high aspect ratio and the high surface area of the CNTs. In this research, we have

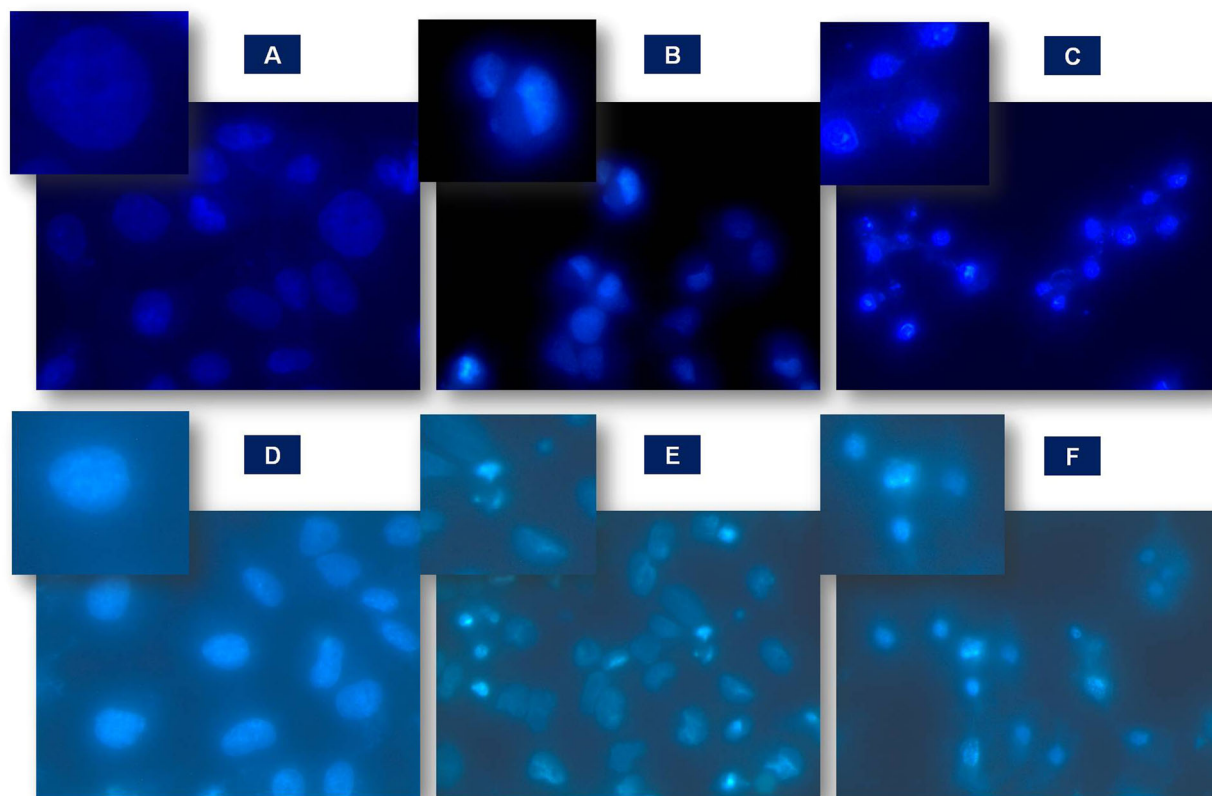


Fig. 10 Apoptosis activity of (A) Control of MWCNTs, (B) 5-FU loaded MWCNTs, and (C) 5-FU SWCNTs against the MCF-7 cell line; and (D) control of SWCNTs, (E) 5-FU loaded MWCNTs and (F) 5-FU SWCNTs against the COLO320DM cell line

Table 1 Apoptosis study of control, 5-FU loaded f-MWCNTs, and 5-FU loaded f-SWCNTs

Concentration $\mu\text{g mL}^{-1}$	Control	5-FU loaded f-SWCNTs	5-FU loaded f-MWCNTs
MCF-7			
100	4.22 ± 1.8617	84.46 ± 4.3515	41.72 ± 3.5603
COLO320DM			
100	2.65 ± 2.0123	92.78 ± 2.6549	45.58 ± 2.0925

tried to explore the comparison between 5-FU loaded f-MWCNTs and 5-FU loaded f-SWCNTs anticancer activity along with % of apoptosis, which was not focused in earlier studies. Also, there is further scope to explore more research on CNTs coated exteriorly with biocompatible layers, e.g., PEG, polysaccharide, protein, and DNA to study their effect on the efficacy of coated CNTs which have not been a scope of our study. The results and findings of the present investigation will be helpful and provide an insight on coating of CNTs exteriorly with suitable biocompatible polymers.

5 Conclusion

MWCNTs and SWCNTs have been considered as lucrative options for the targeted and controlled drug delivery owing to their intrinsic properties. As per our intention, we have functionalized both CNTs (single and multi walled) and loaded 5-FU drug into the f-CNTs. Thereafter, the drug loaded CNTs were characterized using advanced tools and studied for their anticancer potential against MCF-7 and COLO320DM cell lines. The drug loaded f-SWCNTs exhibited excellent % cell inhibition than the 5-FU. Moreover, apoptosis study proved that the MCF-7 and COLO320DM cells treated with 5-FU loaded MWCNTs and 5-FU loaded SWCNTs exhibited smaller nuclei size with a bright chromatin condensation, blebbing, nuclear fragmentation, and formation of apoptotic bodies. With these obtained results, it can be concluded that the prepared drug loaded CNTs possess significant comparable antitumor activity in breast cancer cell lines thus, providing a hopeful technique to enhance the efficacy of traditional chemotherapies. Furthermore, CNT's may prove to be immensely beneficial owing to its diverse properties in effective management of cancer therapy.

Acknowledgements The authors are thankful to the Secretary KES society Kasegaon, and Principal, KE society's, Rajarambapu College of Pharmacy, Kasegaon, Sangli, MS for providing research facilities.

Author contributions Conceptualization: SDB, DSR; data curation: SDB, DSR, MAB, and RK; formal analysis: SDB, DSR, SKM, RK, and MAB; investigation: SDB, DSR; methodology: RK: SDB, DSR; project administration: SDB; software: RK, SDB, DSR; supervision: SDB: DSR, MAB; validation: SDB; DSR, SKM, MAB; roles/writing—original draft: SDB, DSR, RK; writing—review and editing: SDB, DSR, MAB, and RK.

Compliance with ethical standards

Conflict of interest The authors declare no competing interests.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- World Health Organization. Cancer. Fact Sheet #297. 2015. <http://www.who.int/mediacentre/factsheets/fs297/en/>. Accessed 15 Sep 2019.
- World Health Organization. Cancer. 2018. <https://www.who.int/cancer/prevention/diagnosis-screening/breast-cancer/en/>. Accessed 15 Sep 2019.
- González-Lavado E, Valdivia L, García-Castaño A, González F, Pesquera C, Valiente R, et al. Multi-walled carbon nanotubes complement the anti-tumoral effect of 5-Fluorouracil. *Oncotarget*. 2019;10:2022–9. <https://doi.org/10.18632/oncotarget.26770>.
- Da-Yong L. Drug combinations. Personalized Cancer Chemotherapy. 1st ed. Woodhead Publishing. 2015. Pp. 37–41.
- Johnson KR, Wang L III, Miller MC, Willingham MC, Fan W. 5-Fluorouracil interferes with paclitaxel cytotoxicity against human solid tumor cells. *Clin Cancer Res*. 1997;3:1739–45.
- Motevalli SM, Eltahan AS, Liu L, Magrini A, Rosato N, Guo W, et al. Co-encapsulation of curcumin and doxorubicin in albumin nanoparticles blocks the adaptive treatment tolerance of cancer cells. *Biophys Rep*. 2019;5:19–30. <https://doi.org/10.1007/s41048-018-0079-6>.
- Debbage P. Targeted drugs and nanomedicine: present and future. *Curr Pharm Des*. 2009;15:153–72.
- Prato M, Kostarelos KAB. Functionalized carbon nanotubes in drug design and discovery. *Acc Chem Res*. 2008;41:16–27.
- Iijima. Helical microtubules of graphitic carbon. *Nature*. 1991;354:56–8.
- Aqel A, El-Nour KMMA, Ammar RAA, Al-Warthan A. Carbon nanotubes, science and echnology part (I) structure, synthesis and characterization. *Arab J Chem*. 2012;5:1–23.
- Sinha N, Yeow JTW. Carbon nanotubes for biomedical applications. *IEEE Trans Nanobioscience*. 2005;4:180–95.
- Cui D, Zhang H, Sheng J, Wang Z, Toru A, He R, et al. Effects of CdSe/ZnS quantum dots covered multi-walled carbon nanotubes on murine embryonic stem cells. *Nano Biomed Eng*. 2010;2:236–44.
- Huang P, Zhang C, Xu C, Bao L, Li Z. Preparation and characterization of near-infrared region absorption enhancer carbon nanotubes hybridmaterials. *Nano Biomed Eng*. 2010;2:225–30.
- Foldvari M, Bagonluri M. Carbon nanotubes as functional excipients for nanomedicines: I pharmaceutical properties. *Nanomedicine*. 2008;3:173–82.
- Bosi S, Da Ros T, Spalluto G, Prato G. Fullerene derivatives: an attractive tool for biological applications. *Eur J Med Chem*. 2003;38:913–23.
- Oberdörster E. Manufactured nanomaterials (fullerenes, C60) induce oxidative stress in the brain of juvenile largemouth bass. *Environ Health Perspect*. 2004;112:1058–62.
- Pagona G, Tagmatarchis N. Carbon nanotubes: materials for medicinal chemistry and biotechnological applications. *Curr Med Chem*. 2006;13:1789–98.
- Son KH, Hong JH, Lee JW. Carbon nanotubes as cancer therapeutic carriers and mediators. *Int J Nanomed*. 2016;11:5163–85.
- Boncel S, Herman AP, Budniok S, Jędrysiak RG, Jakóbi-Kolon A, Skepper JN, et al. In vitro targeting and selective killing of T47D breast cancer cells by purpurin and 5-fluorouracil anchored to magnetic CNTs: nitrene-based functionalization versus uptake, cytotoxicity and intracellular fate. *ACS Biomater Sci Eng*. 2016;2:1273–85.
- Fan X, Jiao G, Gao L, Jin P, Li X. Preparation and drug delivery of graphene-carbon nanotubes-Fe3O4 nanoparticles hybrid. *J Mater Chem B*. 2013;1:2658–64.
- Kaur S, Mehra NK, Jain K, Jain NK. Development and evaluation of targeting ligand anchored CNTs as prospective targeted drug delivery system. *Artif Cells Nanomed Biotechnol*. 2015;45:242–50.
- Khalili Z, Ganji MD, Mehdizadeh M. Fluorouracil functionalized Pt-doped carbon nanotube as drug delivery nanocarrier for anticarcinogenic drug: A B3LYP-D3 study. *J Nanoanalysis*. 2018;5:202–9.
- Sobh RA, Nasr HES, Mohamed WS. Formulation and in vitro characterization of anticancer drugs encapsulated chitosan/multi-walled carbon nanotube nanocomposites. *J Appl Pharm Sci*. 2019;9:32–40.
- Hou PX, Liu C, Cheng HM. Purification of carbon nanotubes. *Carbon*. 2008;46:2003–25.
- Ghoshal S, Kushwaha SKS, Srivastava M, Tiwari P. Drug loading and release from functionalized multiwalled carbon nanotubes loaded with 6-mercaptopurine using incipient wetness impregnation method. *AJADD*. 2014;2:213–23.
- Tarawneh MA, Ahmad SHJ, Rasid R, Yahya SY, Lau K, Kong I. Sonication effect on the mechanical properties of MWNT reinforced natural rubber. *J Compos Mater*. 2013;47:579–85.
- Zeng H, Gao C, Wang Y, Watts PCP, Kong H, Cui X, et al. In situ polymerization approach to multiwalled carbon nanotubes-reinforced nylon 1010 composites: Mechanical properties and crystallization behaviour. *Polymer*. 2006;47:113–22.
- Yang D, Rochette J, Sacher E. Functionalization of multiwalled carbon nanotubes by mild aqueous sonication. *J Phys Chem B*. 2005;109:7788–94.

29. Peng H, Alemany L, Margrave J, Khabashesku V. Sidewall carboxylic acid functionalization of single-walled carbon nanotubes. *J Am Chem Soc.* 2003;125:15174–82.
30. Joshi GV, Kevadiya BD, Patel HA, Bajaj HC, Jasra RV. Montmorillonite as a drug delivery system: intercalation and in vitro release of timolol maleate. *Int J Pharm.* 2009;374:53–7.
31. Moustafa AB, Sobh RA, Rabie AM, Nasr HE, Ayoub MMH. Synthesis and in vitro release of guest drugs-loaded copolymer nanospheres MMA/HEMA via differential microemulsion polymerization. *J Appl Polym Sci.* 2013;129:853–65.
32. Jia N, Lian Q, Tian Z, Duan X, Yin M, Jing L, et al. Decorating multi-walled carbon nanotubes with quantum dots for construction of multi-color fluorescent nanopores. *Nanotechnol.* 2010;21:045606.
33. Zhao G, Shao D, Chen C, Wang X. Synthesis of few-layered graphene by H₂O₂ plasma etching of graphite. *Appl Phys Lett.* 2011;98:183114–3.
34. Yannopoulos SN, Zouganelis GD, Nurmohamed S, Smith JR, Bouropoulos N, Calabrese G, et al. Physisorbed o-carborane onto lyso-phosphatidylcholine-functionalized, single-walled carbon nanotubes: a potential carrier system for the therapeutic delivery of boron. *Nanotechnol.* 2010;21:85–101.
35. Rohankumar RC, Somnath DB, Mangesh AB, Dheeraj SR, Ganesh HW, Sachin ST, et al. Characterization, antioxidant, antimicrobial and cytotoxic activities of green synthesized silver and iron nanoparticles using alcoholic *Blumea eriantha* DC plant extract. *Materials Today Communications.* 2020;24:101320.