### **ORIGINAL PAPER**



# Sunny, hot and humid nesting locations with diverse vegetation benefit *Osmia* bees nearby almond orchards in a mediterranean area

Carlo Polidori<sup>1</sup> · Sara Rodrigo-Gómez<sup>2</sup> · Federico Ronchetti<sup>3</sup> · Andrea Ferrari<sup>1</sup> · Jesús Selfa<sup>4</sup> · Diego Gil-Tapetado<sup>1,5</sup>

Received: 24 March 2023 / Accepted: 22 September 2023 / Published online: 16 October 2023 © The Author(s) 2023

### **Abstract**

Wild bees (Hymenoptera: Apoidea) play an important role as pollinators of many crops and managed populations of Osmia spp. (Megachilidae), through the installation of trap-nests, proved to be efficient in several fruit orchards. In order to optimize the trap-nest protocols, it is necessary to understand which environmental factors play a major role in the reproductive success of these bees. Here, we studied how climate, land use and vegetation affect nest occupation rate (OR, i.e. total number of colonized tunnels/total number of tunnels in the trap-nest), brood productivity (BP, i.e. total number of brood cells built in a completed nest tunnel) and parasitism rate (PR, i.e. total number of parasitized brood cells/BP) in Osmia bees nearby almond orchards in South-East Spain, a largely understudied Mediterranean area. We found that the summer solar radiation positively influenced all three parameters, while spring solar radiation positively affected OR and BP, and negatively PR. Higher abundance of pastures and forests compared with crops increased OR, though not BP, and reduced PR. Vegetation evenness and diversity of dominant plant species also positively affected OR and BP, while they were unimportant for PR. OR was not affected by climate, but BP increased with maximum temperature in the warmest month and decreased with temperature annual range. PR also increased with high temperature, as well as with precipitation. Arid conditions limited OR and BP and boosted parasitism. Overall, it seems that Osmia bees nearby almond field in this area would benefit from trap-nest installation in well solar-radiated, hot and humid sites with a diverse vegetation. Since we have also found a negative association between PR and BP in nests with at least one parasitized cell, environmental conditions which improve productivity will also likely reduce parasitism in these bees.

**Implications for insect conservation** Optimization of trap-nesting protocols for maintaining abundant *Osmia* populations is crucial to an effective use of these bees as managed pollinators. In our study we suggest that trap-nests locations should be chosen also taking into account a number of local climatic and habitat factors, given their importance in affecting key traits of reproductive success in these solitary bees.

**Keywords** Trap-nests · Managed solitary bees · bioclim · Corine · Spain

- ☐ Carlo Polidori carlo.polidori@unimi.it
- Dipartimento di Scienze e Politiche Ambientali, Università degli Studi di Milano, via Celoria 26, Milan 20133, Italy
- Delegación Provincial de Ciudad Real de la Consejería de Agricultura, Agua y Desarrollo Rural de la Junta de Comunidades de Castilla-La Mancha, Ciudad Real, Spain
- Dipartimento di Bioscienze, Università degli Studi di Milano, via Celoria 26, Milan 20133, Italy
- Departament de Zoologia, Universitat de València, c/Dr. Moliner, 50, Burjassot, València 46100, Spain
- Departamento de Bioaffiliationersidad, Ecología y Evolución, Universidad Complutense de Madrid, c/José Antonio Nováis 12, Madrid 28040, Spain

### Introduction

Pollination is an essential ecosystem service and is mostly provided by insects, especially bees (Hymenoptera: Apoidea), which have been reported to be key pollinators of more than 70% of the about 1500 world crops (Klein et al. 2007; Ollerton et al. 2011; Garibaldi et al. 2013). The economic contribution of pollinators to crop production reaches a large extent (Gallai et al. 2009; Potts et al. 2016). Unfortunately, such great importance parallels nowadays the evident signs of increasing pressure on European honeybees (*Apis mellifera* Linnaeus, 1758) and of the decline of native



wild bee species (van Engelsdorp and Meixner 2010; Potts et al. 2010), posing this ecosystem service at risk.

For wild bees, attempts to overcome such decline in agricultural areas is partially based on providing additional nesting support to that naturally occurring in the environment. For example, installing trap-nests, also called bee hotels, is effective to sustain cavity-nesting bees (Bosch and Kemp 2002; Junqueira et al. 2012; Maclvor 2017; Yamamoto et al. 2012), a guild including most of the (still few) non-Apis managed bee species (Garibaldi et al. 2017). By increasing nesting opportunities for such wild bees, together with sustainable honeybee management (i.e. that not limits wild bees' fitness (Dupont et al. 2004; Paini 2004; Mallinger et al. 2017; Lázaro et al. 2021)), farmers can help face the rising demand for pollinator-dependent crops and avoid experience pollination deficits due to decline of natural populations (Garibaldi et al. 2014; Isaacs et al. 2017). Such cavity-nesting bees possess a typical gregarious nesting behaviour and commonly accept man-made nesting structures, e.g. tunnels in wood, reeds, paper straws or cardboards, making these species ideal for their reproduction as well as subsequent successful mass rearing (Maclvor 2017). Plans to favour an increasing abundance of wild bees are also important, since for most crops the pollination contribution of wild bees is equally important or sometimes even superior to that of honeybees (Apis mellifera) (Garibaldi et al. 2013; Eeraerts et al. 2019).

However, to optimize trap-nesting programs, it is necessary to analyse which factors drive the reproductive success of the target bee species. In fact, the population dynamics of wild bees are affected by many biotic and abiotic factors, such as food resources abundance and diversity, climatic conditions, landscape features and pressure from natural enemies (Steffan-Dewenter and Schiele 2008; Franzén and Nilsson 2013; Woodard and Jha 2017; Roulston and Goodell 2011; Papanikolaou et al. 2017).

A wide range of studies show, for example, that parasitism can greatly affect reproductive success in wild bees and that this is often directly density-dependent, despite inversely density-dependent or density-independent relationships have also been found (Antonini et al. 2003; Steffan-Dewenter and Schiele 2008; Palladini and Maron 2014; Groulx and Forrest 2017). Bee parasites' occurrence, however, can also indicate good persistence of the host species population, that host species population has been present in a given area for a short time, or considerable abundance fluctuations. Furthermore, higher number and abundance of eleptoparasitic species may indicate the high quality of the environment (Sheffield et al. 2013). In addition, landscape features such as a high diversity and amount of semi-natural habitats often favour higher bee abundance (Steffan-Dewenter 2003; Steckel et al. 2014; Pascual 2022).

Climatic conditions also have a heavy impact on bee reproduction and development (Bosch and Blas 1994; Ulbrich and Seidelmann 2001). Since all these factors may have differential importance depending on the geographical areas, studies devoted to this topic should ideally cover as many different regions as possible.

Here, we investigated how climate, land use and vegetation affect reproductive success in Osmia (Hymenoptera: Megachilidae) solitary bees nearby almond orchards in South-East Spain, a largely understudied Mediterranean area. Specifically, we measured nest occupation rate, brood productivity and parasitism rate. The genus Osmia Panzer, 1806, has almost 360 species described to date and it is distributed in the Palaearctic and the Nearctic region, with only few species recorded from the Oriental and Neotropical region (Michener 2007; Ungricht et al. 2008; Ascher and Pickering 2020). Osmia bees are univoltine or, more rarely, semivoltine and include some of the earliest emerging bees in spring, with males starting to fly a bit earlier than females. Females forage for pollen and nectar to provide food for their larvae, which develop in brood cells within the nests (Felicioli and Pinzauti 2008; Westrich 1989). Once developed, larvae pupate and then egress as adults. In some species, the freshly emerged adults still remain in the cocoon until the spring of the following year (Bosch et al. 2008).

The few species of *Osmia* currently considered adequate for mass-rearing and management in agricultural areas all nest in pre-existing tunnels. Female bees of these species indeed promptly use man-made structures for nesting and are known to be effective in increasing fruit set in economically important species such as almond, peach, apricot, plum, cherry, apple and pear (Torchio 1976; Felicioli and Pinzauti 2008; Bosch and Kemp 2002; Bosch et al. 2000; Sheffield 2014; Monzón et al. 2004). Hence, Osmia bees are now managed in different agricultural areas (Bosch and Kemp 2002; Cane 2006; Artz et al. 2013). While most species of Osmia are generalist in pollen use (Westrich 1989), they show a good pollen constancy, making them ideal when managed in or nearby the target crop fields or orchards (Westrich 1989). We choose to carry out our study in areas greatly exploited for almond cultivation. Here, honeybee hives were proved to be insufficient in large acreages planted in eastern and southern Spain (Bosch et al., 1992), making inadequate pollination a key cause, together with frost damage, drought and poor soils, of low production of almond in this area (Vargas and Romero 1987). On these trees, Osmia bees are known to be very efficient pollinators (Bosch et al. 2021), at least partially because almonds bloom very early (February-March) and because these bees are often the only ones on flight along with honeybees (Westrich 1989). While Osmia bee management was initiated in Spain almost 40 years ago (Asensio 1984), studies on reproductive success



of these bees were confined to the North-Eastern part of the country (Bosch et al. 2021 and references therein). Thus, to our knowledge, this would be the first work to analyse the factors affecting the nesting and reproductive success of *Osmia* in South-Eastern Spain.

### **Materials and methods**

## Study area and trap-nesting protocol

Our study took place in a largely agricultural area including the municipalities of Bullas, within the Province of Murcia, and Elda and Monóvar, within the Province of Alicante (South-East Spain) (Fig. 1A-B). Within such sites, almond orchards are common and range from relatively small (826 trees in 5.9 hectares: Bullas) to very large (19,571 trees in 51.7 hectares: Elda and Monóvar) in size.

A total of 76 trap-nests were installed in 75 stations; only one station included two trap-nests (Fig. 1C-D). The trap stations were located at distances ranging from 0.9 to 12,325 m (average= $391.4\pm171 \text{ m}$ ). Each of the 76 trapnests consisted in a group of 24 (all but one trap) or 27 (one

trap) stems of the common reed *Phragmites australis* (Cav.) Trin ex Steud (Poaceae). Hence, a total of 1827 stems were accommodated across the 75 trap-nest stations. Each reed stem was 15-20 cm long and had a hole diameter of 6 to 11 mm, and each trap-nest included an equilibrate combination of smaller and larger stem reeds to limit any possible bias in reproductive success due to differences in reed size (Bosch and Kemp 2000, 2002). Only one entrance per reed stem was offered for nesting, and we cut the stems in a way that they end at their nodes. The trap-nests were set at a height of 1.5-2 m above the ground, on available supporting structures, such as buildings' roofs or stone walls. Trap-nests were placed at the periphery of almond orchards. Trap-nests were covered with a plastic surface in order to protect them from rain and to help the reeds keeping well packed (Fig. 1C-D). The trap-nests were established in the field between 27 of January and 7 of February 2017 and were removed between 18 and 22 of April 2017, i.e. at the end of the flying season for most of the species of *Osmia* occurring in Europe during spring. Nests were subsequently carried to the laboratory for their opening and content analysis.

Once the trap-nests were collected at the end of the flight season, we first discard those reeds which resulted

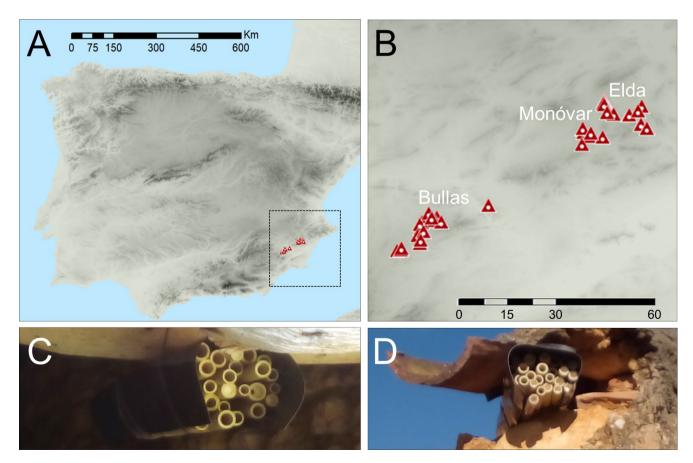


Fig. 1 Location of the study area in the Iberian Peninsula (A), and location of the trap-nesting stations installed during the study (B). Examples of trap-nests are shown in C-D.



unoccupied, i.e. those which had no signs of bee use as nests. The number of occupied nests by Osmia was used to calculate the nest occupation rate (OR, i.e. total number of colonized tunnels/total number of tunnels in the trap-nest). Since we wanted to analyse variables related with reproductive success, the opening and inspection of colonized reed tunnels were carried out only for those completed, i.e. those showing the typical nest plug at the entrance built by the occupying female at the end of provisioning and egg laying. Then, we recorded two further variables for the occupied nests. From each of them we recorded brood productivity (BP, i.e. total number of brood cells) and parasitism rate (PR, i.e. total number of parasitized brood cells/BP). Brood productivity was hence calculated *per* nest, not necessarily per female, since sometimes one female may build cells in more than one nest. However, this is not an individual-based study, and we were interested in studying if brood productivity in a nest depends on some environmental variables. In any case, the probability for a female to occupy more than one nest is not likely to be biased towards certain locations compared to others, and while we did not assume that the number of nests correspond to the number of females, in previous studies on Osmia and other trap-nesting bees (e.g. Palladini and Maron 2014; Wilson et al. 2020) this assumption was considered reasonable. Parasitism was recognized if individuals of non-bee or cuckoo bee species, both at immature (larvae, pupae) or adult stages, were found in the nest. Raw data of the variables used to calculate the response variables for the data analysis (OR, BP, PR) can be found in Table S1.

Osmia pupae from these nests were extracted from the brood cells in September-November 2017 and then given to the company WILDBIENE+PARTNER AG (Zurich, Switzerland) after cocoon identification, following a collaborative project aimed to implement mass-rearing of Osmia bees in agricultural areas in Southern Europe. However, to check species identity of nesters and natural enemies (see below), thirty reeds were kept in the lab at 4 °C to simulate winter and let the adults emerge the following spring.

# **Study species**

During the field work, only individuals (especially males) of *O. cornuta* were occasionally observed on flight. However, three species of *Osmia* emerged from a subset of 30 tunnels from the installed trap-nests that were kept in the lab during the winter following the field work (see above). These species were *Osmia cornuta* (Latreille, 1805) (19 tunnels), *Osmia tricornis* Latreille, 1811 (8 tunnels) and *Osmia latreillei* (Spinola, 1806) (3 tunnels). However, since all the other nests were opened and analysed in September to obtain pupae aimed to be used in subsequent managing

programs, it was not possible to ascertain which of the three species was associated with each of the colonized tunnel. All cocoons were, however, inspected to be sure they belong to Osmia, based on morphology, size and colour. Again, most of individuals could reasonably belong to O. cornuta or O. tricornis (82.8%, n of pupae = 530) since the cocoon was attached to both cell partitions (i.e. the walls build by the females to divide brood cells in the nests) and not only to the posterior one as in O. latreillei. Furthermore, the cocoon of O. latreillei lacks the silky outer layer typical of O. cornuta or O. tricornis (Bosch et al. 2001) and most of brood cells partitions were made from mud (as typical in O. cornuta and O. tricornis) and not chewed leaves (typical in O. latreillei) (Westrich 1989). Hence, while the data analysis was carried out without distinguishing the species (e.g.: Groulx et al. 2001), we can at least suggest that O. cornuta (which has a strong preference for Rosaceae as a pollen source and was proven to be an excellent almond pollinator (Westrich 1989; Bosch et al. 2021)) was likely the most abundant species in the studied area during our investigation.

# Geographical, climatic, land-use and vegetation variables

To determine which environmental variables influence the productivity and mortality of *Osmia* in our studied nests we have collected longitude, latitude and altitude (as well as 3 combinations of latitude and longitude, following Lomolino et al. (2017) (geographical variables), 24 climatic, 7 land use and 7 vegetation variables (Table 1). For each trap-nest, the value of each variable consists of either the intersection of the geographic coordinate of the nest with the different raster information or the spatial query in a buffer of a given size (only in the case of land use).

Climatic variables included the 19 ones related with temperature and precipitation available in WorldClim (https://www.worldclim.org/data/worldclim21.html), well as 4 mean seasonal values of solar radiation (also from WorldClim) and 1 variable related with aridity (an index in which lower values mean higher aridity and higher values mean lower aridity and indirectly higher humidity, see Trabucco and Zomer (2018). Land use variables included 6 ones derived, at both 250 m and 1 km of buffer from the trap-nest, from the CORINE dataset (retrieved from Copernicus: https://land.copernicus.eu/pan-european/ corine-land-cover). We choose these two buffer because Osmia cornuta and Osmia bicornis were reported, depending on calculations and methods, to have a maximum foraging distance of roughly 200-900 m (Gathmann and Tscharntke 2002; Hofmann et al. 2020). Furthermore, by using these two buffers we could verify whether spatial scale influences the effects of the habitat on the considered



**Table 1** Geographical, climatic, land-use and vegetation variables used in data analysis

Abbreviation	Variable	Source	Link		
pio1_PI	Annual Mean Temperature	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
oio2_PI	Mean Diurnal Range (Mean of monthly (max temp - min temp))	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io3_PI	Isothermality (bio2/bio7) (×100)	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io4_PI	Temperature Seasonality (standard deviation ×100)	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
nio5_PI	Max Temperature of Warmest Month	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io6_PI	Min Temperature of Coldest Month	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io7_PI	Temperature Annual Range (bio5-bio6)	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io8_PI	Mean Temperature of Wettest Quarter	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io9_PI	Mean Temperature of Driest Quarter	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io10_PI	Mean Temperature of Warmest Quarter	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io11_PI	Mean Temperature of Coldest Quarter	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io12_PI	Annual Precipitation	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io13_PI	Precipitation of Wettest Month	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io14_PI	Precipitation of Driest Month	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io15_PI	Precipitation Sea- sonality (Coefficient of Variation)	Fick and Hijmans	https://www.worldclim.org/data/worldclim21.html		
io16_PI	Precipitation of Wettest Quarter	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io17_PI	Precipitation of Driest Quarter	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io18_PI	Precipitation of Warmest Quarter	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
io19_PI	Precipitation of Coldest Quarter	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
RadSpri	Mean solar radiation in spring	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
RadSumm	Mean solar radiation in summer		https://www.worldclim.org/data/worldclim21.html		
adAutu	Mean solar radiation in Autumn		https://www.worldclim.org/data/worldclim21.html		
adWint	Mean solar radiation in winter	` /	https://www.worldclim.org/data/worldclim21.html		
levation	Altitude	Fick and Hijmans (2017)	https://www.worldclim.org/data/worldclim21.html		
CORINE	Land cover	Cover (2018)	https://land.copernicus.eu/pan-european/corine-land-cover		
CORINE_MOD	Land cover but with fewer categories grouping Corine's	Derived from Cover (2018)	. 1 1 2		
CORINE1kmMayor	Majority category of land cover at a scale of 1 km.				



Table 1 (continued)

Abbreviation	Variable	Source	Link
CORINE1kmVary	Number of land cover categories at 1 km scale	Derived from Cover (2018)	
CORINE250mMayor	Majority category of land cover at a scale of 250 m.	Derived from Cover (2018)	
CORINE250mVary	Number of land cover categories at 250 m scale.	Derived from Cover (2018)	
CoefVariacionVegetation	Normalized dispersion of EVI (Enhanced Vegeta- tion Index)	Tuanmu and Jetz (2015)	https://www.earthenv.org/texture
HomogeneidadVegetation	Similarity of EVI between adjacent pixels	Tuanmu and Jetz (2015)	https://www.earthenv.org/texture
Simpson_index	Diversity of EVI applying the Simpson index	Tuanmu and Jetz (2015)	https://www.earthenv.org/texture
Shannon_index	Diversity of EVI applying the Shan- non index	Tuanmu and Jetz (2015)	https://www.earthenv.org/texture
EvennessVegetation	Evenness of EVI	Tuanmu and Jetz (2015)	https://www.earthenv.org/texture
q1	Diversity of effec- tive plant species	Derived from Tuanmu and Jetz (2015)	
q2	Diversity of domi- nant plant species	Derived from Tuanmu and Jetz (2015)	
Urbanization	Human density/ urbanization degree	SEDAC/NASA	https://sedac.ciesin.columbia.edu/data/set/ gpw-v4-population-density-rev11
AridityIP	Index of aridity	Trabucco and Zomer (2018)	https://cgiarcsi.community/2019/01/24/global-aridity-index-and-potential-evapotranspiration-climate-database-v2/
Latitude	Longitudinal coordinate of the nest	Original data	
Longitude	Latitudinal coordi- nate of the nest	Original data	
X*Y	Longitude × Latitude, following Lomolino et al. (2017)	Derived from original data	
X2Y	(2 × Longitude) × Latitude, follow- ing Lomolino et al. (2017)	Derived from original data	
XY2	Longitude $\times$ (2 $\times$ Latitude), following Lomolino et al. (2017)	Derived from original data	

bee traits. The land-use types used here to characterize the nesting sites were urban (0), crop (1), pasture (2), forest (3), other (4) and water (5). We then used as variables in the statistical analysis the type of land use which was more abundant (CORINEMayor) and the diversity of land use types (CORINEVary). In addition, we included 1 variable related with urbanization (human population density), retrieved from the SEDAC/NASA: https://sedac.ciesin.columbia.

edu/data/set/gpw-v4-population-density-rev11). Vegetation diversity was estimated by considering different measurements of the Enhanced Vegetation Index (EVI) (Tuanmu and Jetz 2015), retrieved from EarthEnv (https://www.earthenv.org/texture). In particular, we used the EVI's coefficient of variation, homogeneity, evenness and the Simpson and Shannon indices. To obtain more understandable measures of the Shannon and Simpson indices, we have



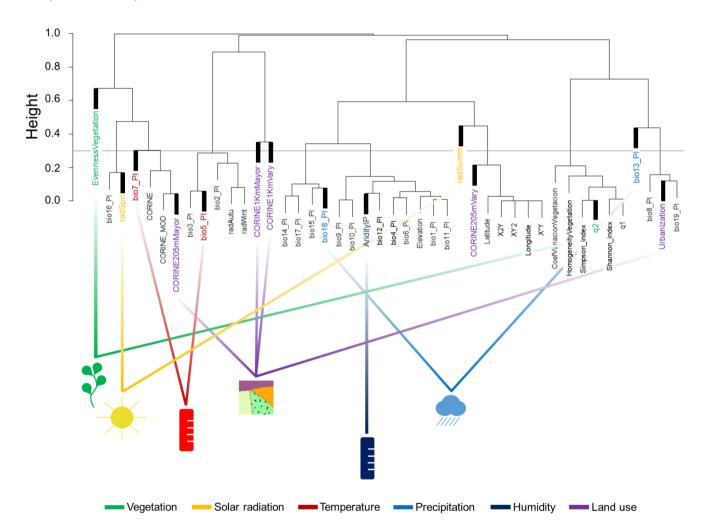
transformed these variables following Jost (2006), converting them into the true diversity variables of Hill's series (q1, effective species= $e^{Shannon index}$ ; q2, dominant species=1/Simpson index).

### Statistical analysis

Because the 44 geographical, climatic, land-use and vegetation variables may be variably inter-correlated, we first performed a hierarchical cluster analysis of similarity among all variables (through the Ward method, Harrell (2001), which depicted a dendrogram (Dormann et al. 2013) and then used to select those later entering in the subsequent statistical analysis of the data (Fig. 2). The distance threshold used to identify different clusters in the dendrogram was set at 0.3 (i.e., less than 70% correlation occurred). From each recognized cluster, we finally choose a single variable with, in a few cases, a second variable that we consider to

be equally biologically significant. The final set included 14 variables (Fig. 2).

We used two complementary approaches to detect which factors were relevant in explaining variation in the three dependent variables (OR, BP, PR). First, we used a Random Forest (RF) classifier algorithm to detect which selected variables are more important in explaining the variation in each of the three response variables. The RF classification and regression non-parametric approach, proposed by Breiman (2001), consists of an ensemble of decision trees. RF provides, through random permutation, an importance measure of the input variables which can be used to rank or select factors (e.g.: Gil-Tapetado et al. 2021). Here, we used the Conditional Inference Trees (CTree) algorithm (Hothorn et al. 2006) to fit to each of the trees to be grown for the forest. This procedure develops permutation tests, which address overfitting and variable selection biases by using a conditional distribution to measure the association between



**Fig. 2** Dendrogram obtained through the cluster analysis that was employed to select the relevant climatic, land-use and vegetation variables (highlighted by thicker branches and coloured names); the red horizontal line indicates the chosen distance-threshold to form the

clusters (0.3). Different colours identify different major categories of variables, and those belonging to the same category are connected by coloured lines to correspondent symbols



the output and the input variables, and considering the distributional properties (Williams 2011). For all RF analyses, we have considered 5000 trees as input. We performed 6 RF analyses considering the 14 selected independent variables and the four dependent variables. We arbitrarily decided to consider as "very important" the first four variables with decreasing values of importance.

Then, generalized linear mixed models (GLMMs) were used to assess the effect of the 14 selected independent variables on the four dependent variables. A Poisson error distribution was applied, and model selection was based on Akaike Information Criterion (AIC) (Akaike 1974). We first compared models carried out with and without latitude as random factor, by calculating and inspecting AICs and Bayesian information criterion (BIC, Schwarz 1978) values, and then performing  $\chi^2$  tests. While in their original forms a larger value of AIC and BIC for a given model indicates a better fit of the data, it is common to see use the "smalleris-better" form (i.e. when they are calculated directly from the  $-2 \times log$ -likelihood). Since AIC and BIC values were always lower in models with latitude (Table S1), we keep it as random factor. Categorical variables (i.e. CORINE-Mayor) which resulted significant in the GLMM were subjected to analysis of variance (ANOVA) to test for differences among categories. We also tested the relationship between parasitism rate and occupation rate and between parasitism rate and productivity by performing simple linear regressions. Since we recorded many nests with null mortality (see below), which strongly affected the goodness of these linear regressions, we decided to use both the complete dataset and a reduced dataset, by excluding all zeros from PR.

The statistical analysis was performed in R through the RStudio Software v 2022.02.2–485, R v 4.1.3 (R Core Team 2022). The RF analysis and the calculation of the variable importance was done in using the R packages *party* (Hothorn et al. 2006; Strobl et al. 2007, 2008) ('libcoin' (Hothorn 2021), 'modeltools' (Hothorn et al. 2020), 'strucchange' (Zeileis et al. 2022), 'coin' (Hothorn et al. 2021) and *rattle* (Williams 2011) ('tibble' (Müller et al. 2023), 'bitops' (Dutky and Maechler 2021). *rattle* was installed following the instructions of the developer of rattle package (https://rattle.togaware.com/). R package *lme4* (Bates et al. 2014) ('Matrix' (Bates et al. 2023) was used to perform both GLMMs and simple linear models. The R package *ggpubr* was used to create the graphics (Wickham 2016). In the text, mean values are reported ± Standard Error.

### **Results**

Out of the 1827 reeds from the 75 trap-nest stations, 530 were completed by the bees (leading to an overall occupation rate of 0.29) and were then used in all the subsequent analyses.

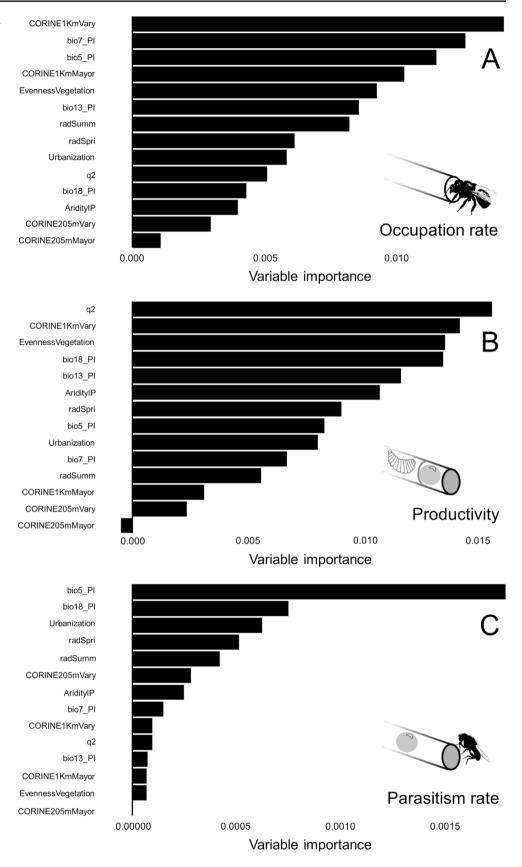
Occupation rate (OR) per trap-nest varied from 0.04 to 1 and was  $0.59 \pm 0.012$  on average. The RF analysis showed that the "majority" category of land cover at a scale of 1 km (CORINE1KmMayor), the number of land cover categories at 1 km scale (CORINE1KmVary), the maximum temperature of warmest month (bio5) and the temperature annual range (bio7) were relevant to explain variation in OR (Fig. 3A). The GLMM gave importance to two of these variables: higher values of CORINE1KmMayor and lower values of CORINE1KmVary (that is, habitats with certain abundant land cover types and little land cover type diversity at 1 km scale) lead to an increase in OR (Table 2; Fig. 4A-B). In particular, ANOVA showed that OR was highest in pastures, followed by forests, and very low in crops (SS = 2.28, MS = 1.14, F = 25.82, df = 2, P < 0.0001, Fig. S1A). The GLMM showed a positive effect also of humidity (i.e. greater values of aridity index), vegetation diversity and heterogeneity (q2 and EvennessVegetation) (Table 2; Fig. 4C), and of spring and summer solar radiation on OR (Table 2).

Productivity (BP) varied from 1 to 21 cells *per* nest and was  $4.41 \pm 0.19$  on average. Following the RF analysis, the number of land cover categories at 1 km scale (CORINE1K-mVary), vegetation diversity and heterogeneity (q2 and EvennessVegetation) and the precipitation of Warmest Quarter (bio18) were relevant to explain variation in BP (Fig. 3B). The GLMM revealed that productivity was positively affected by humidity (AridityIP), the maximum temperature of warmest month (bio5), vegetation diversity and heterogeneity (q2 and EvennessVegetation) (Fig. 4D, E) and of spring and summer solar radiation (Table 2). On the other hand, lower values of BP were recorded in sites with increasing values of temperature annual range (bio7), CORINE1K-mVary (Fig. 4F) and urbanization level (Table 2).

From none to all of brood cells *per* nest were lost due to parasitism, and PR was  $2.22 \pm 0.45\%$  on average. Most of the nests (489 out of 530) showed no signs of parasitism. Insect parasites found in the studied nests included species of kleptoparasites in the genus *Cacoxenus* Loew, 1858 (Diptera: Drosophilidae) and kleptoparasitic bees in the genus *Stelis* Panzer, 1806 (Hymenoptera: Megachilidae). Overall, 72 individuals of *Cacoxenus* and 11 individuals of *Stelis* were detected in the nests. The maximum temperature of warmest month (bio5) was by far the most important variable shaping PR, more than doubling the second-highest importance value recorded for precipitation of Warmest



Fig. 3 Histograms of importances based on Random Forest for the 14 climatic, land-use and vegetation variables selected by the Cluster analysis. The variables are ordered top-to-bottom as most-to-least important in accounting for occupation rate (A), brood productivity (B) and parasitism rate (C)





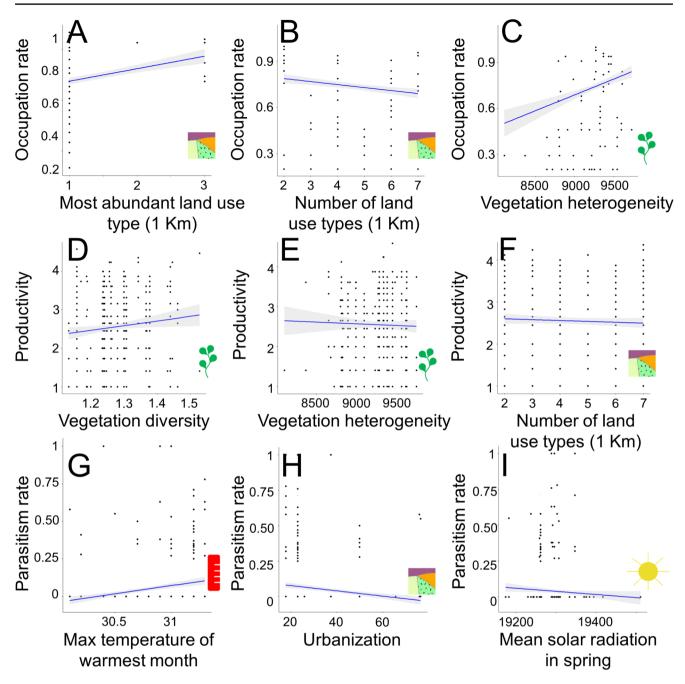
**Table 2** Results of Generalized linear mixed models (GLMM) for the four variables related with reproductive success in *Osmia* bees. Significant effects are in **bold** 

effects are in bold							
Dependent variable	AIC	Random effect (latitude)	Factor	Estimate		t	P
Occupation rate	-2133.9	Intercept = $0.014$ ; Residual = $0.002$	(Intercept)	-16.400	5.468	-3.000	0.003
			AridityIP	0.002	0.000	26.964	
			bio13_PI	0.096	0.068	1.406	0.160
			bio18_PI	0.026	0.026	1.015	0.310
			bio5_PI	-0.177	0.168	-1.057	0.290
			bio7_PI	-0.264	0.152	-1.730	0.084
			CORINE1kmMayor	0.427	0.147	2.910	0.004
			CORINE1kmVary	-0.164	0.012	-13.321	< 0.001
			CORINE250mMayor	0.006	0.135	0.048	0.962
			CORINE250mVary	-0.077	0.009	-8.603	< 0.001
			EvennessVegetation	0.001	0.000	91.951	< 0.001
			q2	2.172	0.495	4.387	< 0.001
			radSpri	0.000	0.000	117.720	< 0.001
			radSumm	0.000	0.000	70.814	< 0.001
			Urbanization	-0.007	0.005	-1.426	0.154
Productivity	1363.6	Intercept = 0.04; Residual = 0.69	(Intercept)	-86.380	10.460		< 0.001
		1,	AridityIP	0.010	0.000	62.308	
			bio13 PI	0.192	0.101	1.899	0.058
			bio18 PI	0.057	0.045	1.269	0.205
			bio5 PI	1.829	0.290	6.305	
			bio7 PI	-1.929	0.220		< 0.001
			CORINE1kmMayor	0.104	0.116	0.896	0.370
			CORINE1kmVary	-0.121	0.055	-2.215	0.027
			CORINE250mMayor	-0.035	0.320	-0.109	0.913
			CORINE250mVary	0.019	0.136	0.139	0.889
			EvennessVegetation	0.000	0.000		< 0.001
			q2	3.549	0.886		< 0.001
			radSpri	0.001	0.000	235.134	
			radSumm	0.001	0.000	309.114	
			Urbanization	-0.021	0.007	-3.064	0.001
Parasitism rate	-629.7	Intercept = 0.002; Residual = 0.016	(Intercept)	8.729	1.401		< 0.002
	-027.7	intercept = 0.002, Residual = 0.010	AridityIP	-0.001	0.000	-107.529	
			bio13 PI	0.036	0.000	2.785	0.001
			_	0.036	0.013	2.074	0.003
			bio18_PI bio5_PI	0.013	0.007	8.081	
				-0.089	0.032	-2.977	0.001
			bio7_PI CORINE1kmMayor	-0.056	0.030	-2.977	0.003
			-				
			CORINE1kmVary CORINE250mMayor	<b>0.029</b> 0.011	<b>0.013</b> 0.061	<b>2.292</b> 0.175	<b>0.022</b> 0.861
			-				
			CORINE250mVary	-0.003	0.027	-0.108	0.914
			Evenness Vegetation	0.000	0.000	0.042	0.967
			q2 <b>radSpri</b>	0.355	0.191	1.860	0.063
			<del>-</del>	-0.001	0.000	-621.158	
			radSumm	0.001	0.000	255.093	
			Urbanization	-0.003	0.001	-2.187	0.029

Quarter (bio18), following the RF analysis (Fig. 3C). This analysis also recognized as important variables the urbanization level and the spring solar radiation (Fig. 3C). The GLMM showed higher PR at sites with increasing precipitation in both wettest and warmest quarter (bio13, bio18), maximum temperature of warmest month (bio5) (Fig. 4G),

summer solar radiation and CORINE1KmVary (Table 2). Parasitism rate was instead lower at sites with increasing humidity, temperature annual range (bio7), CORINE1Km-Mayor, urbanization level (Fig. 4H) and spring solar radiation (Fig. 4I) (Table 2). PR was highest in crops and lowest in pastures and forests (SS=0.22, MS=0.11, F=5.4, df=2,





**Fig. 4** Linear regression plots showing the effects of selected climatic, land-use and vegetation variables on occupation rate (A-C), brood productivity (D-F) and parasitism rate (G-I). Grey areas around regression lines indicate 95% confidence envelopes. Correspondence of the x-axis variables and their codes (see Table 1). Most abundant land

use type (1 Km)=CORINE1KmMayor; Number of land use types (1 Km)=CORINE1KmVary; Vegetation heterogeneity=Evenness-Vegetation; Vegetation diversity=q2; Max temperature of warmest month=bio5 PI; Mean solar radiation in spring=radSpri

P=0.005, Fig. S1B). Additionally, nests with more brood cells were affected by lower parasitism rate, but only when nests with null parasitism rate were excluded (with parasitism only: linear regression,  $R^2$ =0.34, N=41, P=0.001; with all nests:  $R^2$ =0.0001, N=530, P=0.82). On the other hand, nests in blocks with higher OR did not suffer lower or greater PR (linear regression,  $R^2$ =0.007, N=530, P=0.61).

### **Discussion**

In this study, we explored which environmental conditions may boost reproductive success of *Osmia* bees in trap-nests located nearby almond orchards, which are economically relevant in South-East Spain (International Nut and Dried Fruit 2020). If farmers plan to manage almond pollination



with Osmia bees, selecting the best location for trap-nests in terms of bee occupation rate and brood productivity is a key point. Particularly since correctly integrating non-Apis pollinators into IPPM (integrated pest and pollination management) safeguards fruit production (Lundin et al. 2021). Indeed, the implementation of solitary bee management is hampered by both empirical studies and practical guidelines concerning, e.g.: the best nest location, density and spatial configuration (Eeraerts 2020; Garibaldi et al. 2020). Despite the intensification of agriculture and the loss of natural habitats are key drivers of the observed declines in bee populations in recent decades (van Engelsdorp and Meixner 2010; Potts et al. 2010), it is also true that mass-flowering crops (such as large almond orchards) seem to provide abundant resources for bees during blooming periods (Eeraerst et al. 2022).

We have found that both local (small-scale) climate and landscape features (i.e. vegetation diversity, urbanization level) affect nest occupation rate, brood productivity and parasitism rate in *Osmia* bees in the studied area. Previous studies converge in recognizing the influence of both climate and landscape on such bee parameters from trap nest-based studies.

Concerning landscape characteristics, a positive effect of increasing landscape complexity was observed, and was often associated with a greater vegetation diversity, i.e. a greater spectrum and abundance of floral resources. For example, previous studies showed that *Osmia* bees produce more females (the more expensive sex in terms of resource requirements) within landscapes with a higher proportion of semi-natural habitats (i.e. with greater supply of floral resources) (Galbraith et al. 2021; Zaragoza-Trello et al. 2021; Eeraerts et al. 2021). Accordingly, in our study, two parameters boosted occupation rate by the studied species of Osmia: q2 (diversity of dominant plant species) and Evenness Vegetation (evenness of the Enhanced Vegetation Index), both linked to vegetation diversity and heterogeneity. Hence, these bees would more likely prefer nesting locations with such vegetation characteristics to assure an adequate foraging success during the breeding period. In general, flower resources represent the primary energy and nutrients source for both adult and larval bees and hence are considered to be a major driver of wild bee populations (Roulston and Goodell 2011). Proximity to supplementary floral resources was seen to promote reproductive success of Osmia lignaria Say, 1837 in North-American almond orchards (Boyle et al. 2020).

This also agrees with the effect of habitat types (CORINE categories) that we have found on the occupation rate. Indeed, trap-nests were more successfully colonized in areas with increasing pastures and forests, rather than in areas dominated by crops. Forest superiority over crops is

somehow surprising. However, it may be related to the character of the local forests. For example, high abundance of spring flowering shrubs and trees, including the wind-pollinated tree species, like oaks, may boost the visits by females of some *Osmia* species for pollen collection. Not only abundance of certain habitats, but also a low heterogeneity of habitats (low values of CORINEVary) seems to boost both occupation rate and brood productivity. Also, accordingly with the importance of semi-natural habitats nearby nesting sites, we have found a negative effect of urbanization on productivity, though not on occupation rate. Possibly, urbanized areas provide a great availability of nesting substrates (e.g.: holes in walls) which may not compromise the overall abundance of nesting females.

In our study, vegetation diversity and heterogeneity increased brood productivity per nest as well; suggesting that such higher foraging success indeed happened under conditions of favourable vegetation. Landscapes with large quantities of foraging resources are likely to facilitate brood provisioning in bees (Roulston and Goodell 2011) and support larger source populations to colonize the trap nests. We suggest that vegetation diversity enhance Osmia reproductive success in conjunction with the great abundance of floral resources provided by the almond fields adjacent to the trap-nests stations. For example, Da Jauker et al. (2012) found that, in Osmia bicornis (Linnaeus, 1758), the number of produced offspring increased with availability of oilseed rape (as well as semi-natural habitats). Though, in the case studied by Jauker et al. (2012) oilseed rape likely benefits solitary bees in the form of abundant nectar (used more by adults for foraging flights) rather than pollen for larval provisioning, which was scarce in the brood cells. On the other hand, Eeraerst et al. (2021) found, for O. cornuta and O. bicornis, that most of the offspring (up to almost 90%) was produced during the period of mass-flowering of fruit trees (apple, pear and sweet cherry), decreasing afterwards. Based on their results, Eeraerst et al. (2021) suggested that, in fruit trees-rich landscapes, management needs to focus on providing additional floral resources to sustain viable populations of solitary bees after mass-flowering. In our case, the productivity can be maintained high at nesting locations with a diverse vegetation, as discussed above. Eckerter et al. (2022) also showed that semi-natural, flower-rich habitats such as hedgerows benefit the reproduction of Osmia bees in the vicinity of oilseed rape crops. In a multi-species (community-level) study, Dainese et al. (2018) also showed that trap-nesting bee populations grow markedly with the increasing availability of food resources in the landscape, including mass-flowering period of oilseed rape.

Concerning climatic conditions, we have found that temperature, and to a lesser extent precipitation and humidity, influenced occupation rate and brood productivity of *Osmia* 



bees. In megachilid bees, Fliszkiewicz et al. (2012) and Forrest and Chisholm (2017) suggested (the former) or showed (the latter) that the number of brood cells and the nest construction rate increase at warmer conditions, while Tobajas et al. (2021) found that the offspring production was higher in sunny aspects, i.e. that better heat up nests. On the other hand, in Osmia-pollinated orchards commercially high yields have been achieved also under bad weather conditions (Bosch and Kemp 1999). Humidity not only can affect access to water but is also critical because mud is required of females for nest construction, so it is not surprising that in our study more humid locations increased both occupation rate and productivity. Similar effects of humidity were observed in other studies on *Osmia* (e.g.: Boyle et al. 2020). However, it is not clear if the positive effect of higher temperatures on reproductive success necessarily implies that females select for hotter nesting sites, as these would jeopardize the larval survival due to overheating. For example, O. bicornis females were observed to avoid nest sites with temperatures above 28 °C and tend to choose nesting sites with 18–24 °C (Ostap-Chec et al. 2021). However, the study of Ostap-Chec et al. (2021) was conducted in Poland, where spring temperatures are not expected to be very high at the timing of O. bicornis occurrence, and the study was based on local managed population that was under selection for several generations.

We have found *Osmia* nests to be parasitized essentially by Cacoxenus flies, and to a minor extent by Stelis cuckoo bees. Both kleptoparasites consume the pollen-nectar provision within the brood cell, often killing the bee progeny through starvation, and were commonly found in Osmia nests (Krunić et al. 2005; Tlak Gajger et al. 2022; Zajdel et al. 2014; Shebl et al. 2018; Cane et al. 2007). Besides providing permanent forage and nesting sites, semi-natural elements seem to benefit pollinators also by mitigating negative effects of parasitism. It is thus not surprising that we have found parasitism rate to be lower when crop abundance increased. These results agree with a study on Osmia pumila Cresson, 1864, which found that brood cells experienced a 15-fold higher probability of parasitism in experimental cages with sparse bloom than in those with rich bloom (Goodell 2003). Brood parasites may also require a greater heterogeneity of habitats, since we have found higher parasitism rate at locations with higher CORI-NEVary. Furthermore, while parasitism seemed increased at higher temperatures such as their host bees, it was reduced at lower humidity levels and at higher solar radiation levels in spring, in contrast with their host bees. Previous studies reported variable results on the effects of landscape complexity and parasite infestation, with negative (Jauker et al. 2012), positive (Steckel et al. 2014) or no effects (Dainese et al. 2018; Eeraerts et al. 2022), suggesting that finer habitat elements besides complexity may be relevant, or that additional factors, including the occurrence of alternative hosts, may be relevant for parasite abundances.

Perhaps because of the partially opposite effects of climatic and land-use variables on brood productivity and parasitism rate, we have found a negative correlation between these two latter factors. This suggests an inverse density-dependent parasitism at the nest scale. Across nests of a given Osmia species, density-dependent parasitism was previously observed in several studies. For example, brood parasitism rate was negatively correlated with O. lignaria progeny density (Farzan 2018). However, such benefit is not suggested at population-level scale. That is, larger nesting aggregations (i.e. blocks with more occupied nests) did not provide in our study a lower impact from parasites. Nest abundance was either positively or negatively correlated with parasitism rate in Osmia bees. For example, nest cavity availability had a negative effect on Stelis cuckoo bee parasitism rate on O. lignaria, with lower parasitism rates occurring in nest blocks with more available cavities (Farzan 2018). On the other hand, parasitism of nest cells strongly increased with the number of actively nesting bees at a nesting block in a study involving multiple Osmia species (Groulx and Forrest 2018). Different reasons can be behind the variability in such results, which certainly are not contradictory or mutually exclusive. Indeed, besides nest density, nest architecture (Münster-Swendsen and Calabuig 2000), resource availability (Goodell 2003), foraging efficiency, age of the mother bee (Seidelmann 2006), choice of nesting material (MacIvor and Salehi 2014) and different parasitism strategies (Torchio 1979; Danforth and Visscher 1993) are known to affect parasitism rate in megachilid bees. As observed by data we retrieved from the literature (Medler 1967; Goodell 2003; Yokoi et al. 2012; Jauker et al. 2012; Prosi et al. 2016; Felicioli et al. 2017; Müller et al. 2020; Earaerts 2020), together with our own findings, a negative correlation between productivity and parasitism rate across species and populations of Osmia appeared (linear regression,  $R^2 = 0.31$ , N = 29, P = 0.001) (Fig. S2A), but a non-significant association appeared between occupation rate and parasitism rate (linear regression,  $R^2 = 0.17$ , N = 12, P = 0.18) (Fig. S2B).

In conclusion, it seems that *Osmia* bees nearby almond field in our Southern Mediterranean area would benefit from trap-nest installation in well solar-radiated, hot and humid sites with a diverse vegetation. Furthermore, since we have also found a negative association between brood productivity and parasitism rate, environmental conditions which improve brood productivity will also likely reduce parasitism in these bees. In addition to the above discussed factors influencing bee success from trap-nests, other factors are known to be important and should be taken into



account while managing wild bees. For example, the type of nesting material (e.g. bamboo reed, paper or cardboard tubes, wooden laminar nests) may influence trap-nesting bees (Eeraerts et al. 2022). As well, the diameter and the length of the nesting tunnel provided in the traps are relevant for bees' reproductive success (Seidelmann et al. 2016; Amininasab et al. 2021). However, in our study we have kept all such parameters essentially constant or in any case with no evident biases among sites, strongly suggesting that the observed variance in occupation rate, productivity and parasitism rate indeed depend primarily by the studied environmental factors.

**Supplementary Information** The online version contains supplementary material available at <a href="https://doi.org/10.1007/s10841-023-00523-6">https://doi.org/10.1007/s10841-023-00523-6</a>.

Acknowledgements Thanks are due to Sara Muntada for helping in trap-nest sampling and opening, and to Jordi Bosch (CREAF, Spain) for the kind support, cocoon identification and helpful suggestions during the study. The field study was possible through the permit AUF20160118 from Consejería de Agua, Agricultura y Medio Ambiente – Región de Murcia (Spain) and the permit 638/16(FAU16\_049) from Direcció General de Medi Natural i d'Avaluació Ambiental – Generalitat Valenciana.

**Author contributions** CP and JS conceived the project. CP designed the study, analysed part of the data and wrote the first draft of the manuscript. SRG conducted the field and the lab work. DG-T and FR performed part of the data analysis. AF retrieved literature data used in some analyses. All authors contributed in preparing the final version of the manuscript.

Funding Open access funding provided by Università degli Studi di Milano within the CRUI-CARE Agreement. The study was partially funded by a Research and Development Agreement between the Universitat de València and WILDBIENE+PARTNER AG (2016). Open access funding provided by Università degli Studi di Milano within the CRUI-CARE Agreement.

**Data Availability** All data reported in this paper are available in the manuscript and in the supplementary material.

### **Declarations**

**Competing interests** The authors declare no competing interests.

Conflict of interest The authors declare that they have no conflict of interest.

**Ethical approval** All authors have approved the manuscript and agree to submit it to the Journal of Insect Conservation.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless

indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <a href="http://creativecommons.org/licenses/by/4.0/">http://creativecommons.org/licenses/by/4.0/</a>.

### References

- Akaike H (1974) A new look at the statistical model identification. IEEE Trans Automat Contr 19(6):716–723
- Amininasab SM, Hosseini-Moosavi SM, Xu CC (2021) Influence of breeding time, nest size, and egg size on the breeding success of the common Moorhen *Gallinula chloropus*. Acta Oecologia 113:103779. https://doi.org/10.1016/j.actao.2021.103779
- Antonini Y, Martins RP, Rosa CA (2003) Inverse density-dependent and density-independent parasitism in a solitary ground-nesting bee in Southeast Brazil. Trop Zool 16:83–92. https://doi.org/10.1080/03946975.2003.10531185
- Artz DR, Allan MJ, Wardell GI, Pitts-Singer TL (2013) Nesting site density and distribution affect Osmia lignaria (H ymenoptera: Megachilidae) reproductive success and almond yield in a commercial orchard. Insect Conserv Divers 6:715–724. https://doi. org/10.1111/icad.12026
- Ascher JS, Pickering J (2020) Discover Life bee species guide and world checklist (Hymenoptera: Apoidea: Anthophila). http://www.discoverlife.org/mp/20q?guide=Apoidea species
- Asensio E (1984) *Osmia (Osmia) cornuta* latr pollinisateur potentiel des arbres fruitiers en espagne (Hymenoptera, Megachilidae), in: Proc V Symp Int sur la Pollinisation, 27–30 September 1983, Versailles, pp. 461–465
- Bates D, Maechler M, Jagan M, Davis TA, Oehlschlägel J, Riedy J, R Core Team (2023) Matrix: Sparse and Dense Matrix Classes and Methods. R package. Available at: https://cran.r-project.org/web/ packages/Matrix/index.html
- Bates D, Mächler M, Bolker B, Walker S (2014) Fitting linear mixed-effects models using lme4. J Stat Softw 67:1–48. https://doi.org/10.18637/jss.v067.i01
- Bosch J (1992) Parasitism in wild and managed populations of the almond pollinator *Osmia cornuta* Latr.(Hymenoptera: Megachilidae). J Apic Res 31:77–82. https://doi.org/10.1080/00218839. 1992.11101266
- Bosch J, Blas M (1994) Effect of over-wintering and incubation temperatures on adult emergence in *Osmia cornuta* Latr (Hymenoptera, Megachilidae). Apidologie 25:265–277. https://doi.org/10.1051/apido:19940301
- Bosch J, Kemp WP (1999) Exceptional cherry production in an orchard pollinated with blue orchard bees. Bee World 80:163–173. https:// doi.org/10.1080/0005772X.1999.11099452
- Bosch J, Kemp WP (2002) Developing and establishing bee species as crop pollinators: the example of *Osmia* spp.(Hymenoptera: Megachilidae) and fruit trees. Bull Entomol Res 92:3–16. https://doi.org/10.1079/BER2001139
- Bosch J, Kemp WP, Peterson SS (2000) Management of *Osmia lignaria* (Hymenoptera: Megachilidae) populations for almond pollination: methods to advance bee emergence. Environ Entomol 29:874–883. https://doi.org/10.1603/0046-225X-29.5.874
- Bosch J, Maeta Y, Rust R (2001) A phylogenetic analysis of nesting behavior in the genus *Osmia* (Hymenoptera: Megachilidae). Ann Entomol Soc Am 94(4):617–627
- Bosch J, Osorio-Canadas S, Sgolastra F, Vicens N (2021) Use of a managed solitary bee to pollinate almonds: Population sustainability and increased fruit set. Insects 12:56. https://doi.org/10.3390/insects12010056



- Bosch J, Sgolastra F, Kemp WP (2008) Ecophysiology of *Osmia* Mason Bees used as Crop pollinators. In: James R, Pitts-Singer TL (eds) Bee Pollination in Agricultural Ecosystems. Oxford University Press, Oxford, UK, pp 83–104. https://doi.org/10.1093/acprof:oso/9780195316957.003.0006
- Boyle NK, Artz DR, Lundin O, Ward K, Picklum D, Wardell GI et al (2020) Wildflower plantings promote blue orchard bee, *Osmia lignaria* (Hymenoptera: Megachilidae), reproduction in California almond orchards. Ecol Evol 10:3189–3199. https://doi. org/10.1002/ece3.5952
- Breiman L (2001) Random forests. Mach Learn 45:5–32. https://doi. org/10.1023/A:1010933404324
- Cane JH (2006) The Logan BeeMail shelter: a practical, portable unit for managing cavity-nesting agricultural pollinators. Am Bee J 146:611–613
- Cane JH, Griswold T, Parker FD (2007) Substrates and materials used for nesting by north american Osmia bees (Hymenoptera: Apiformes: Megachilidae). Ann Entomol Soc Am 100:350–358. https://doi.org/10.1603/0013-8746(2007)100[350:SAMUFN]2.0 .CO:2
- Cover CL (2018) European Union, Copernicus Land Monitoring Service. Eur Environ Agency
- Dainese M, Riedinger V, Holzschuh A, Kleijn D, Scheper J, Steffan-Dewenter I (2018) Managing trap-nesting bees as crop pollinators: spatiotemporal effects of floral resources and antagonists. J Appl Ecol 55:195–204. https://doi.org/10.1111/1365-2664.12930
- Danforth BN, Visscher PK (1993) Dynamics of a host-cleptoparasite relationship: *Holcopasites ruthae* as a parasite of *Calliopsis pugi-onis* (Hymenoptera: Anthophoridae, Andrenidae). Ann Entomol Soc Am 86:833–840. https://doi.org/10.1093/aesa/86.6.833
- Dormann CF, Elith J, Bacher S, Buchmann C, Carl G, Carré G et al (2013) Collinearity: a review of methods to deal with it and a simulation study evaluating their performance. Ecography 36:27–46. https://doi.org/10.1111/j.1600-0587.2012.07348.x
- Dupont YL, Hansen DM, Valido A, Olesen JM (2004) Impact of introduced honey bees on native pollination interactions of the endemic *Echium wildpretii* (Boraginaceae) on Tenerife, Canary Islands. Biol Conserv 118:301–311. https://doi.org/10.1016/j. biocon.2003.09.010
- Dutky S, Maechler M (2021) Bitops: Bitwise Operations. R package. Available at: https://cran.r-project.org/web/packages/bitops/index.html
- Eckerter PW, Albrecht M, Herzog F, Entling MH (2022) Floral resource distribution and fitness consequences for two solitary bee species in agricultural landscapes. Basic Appl Ecol 65:1–15. https://doi.org/10.1016/j.baae.2022.09.005
- Eeraerts M (2020) Cardboard nesting cavities may promote the development of *Osmia cornuta* and reduce infestation of kleptoparasitic mites. J Appl Entomol 144:751–754. https://doi.org/10.1111/jen.12793
- Eeraerts M, Clymans R, Van Kerckvoorde V, Belien T (2022) Nesting material, phenology and landscape complexity influence nesting success and parasite infestation of a trap nesting bee. Agric Ecosyst Environ 332:107951. https://doi.org/10.1016/j.agee.2022.107951
- Eeraerts M, Piot N, Pisman M, Claus G, Meeus I, Smagghe G (2021) Landscapes with high amounts of mass-flowering fruit crops reduce the reproduction of two solitary bees. Basic Appl Ecol 56:122–131. https://doi.org/10.1016/j.baae.2021.07.005
- Eeraerts M, Vanderhaegen R, Smagghe G, Meeus I (2020) Pollination efficiency and foraging behaviour of honey bees and non-*Apis* bees to sweet cherry. Agric for Entomol 22:75–82. https://doi.org/10.1111/afe.12363
- Farzan S (2018) Progeny density and nest availability affect parasitism risk and reproduction in a solitary bee (*Osmia lignaria*)

- (Hymenoptera: Megachilidae). Environ Entomol 47:70–76. https://doi.org/10.1093/ee/nvx189
- Felicioli A, Ambroselli S, Cilia G, Sagona S (2017) Parasitization of a wild and reared population of the solitary bee *Osmia cornuta* latr. By the parasitoid *Anthrax anthrax* Schrank (Diptera, Bombyliidae): comparison between two types of artificial nest. J Apic Res 56:598–605. https://doi.org/10.1080/00218839.2017.1343019
- Felicioli A, Pinzauti M (2008) Pollination by *Osmia* Bees (Hymenoptera: Megachilidae), pp. 2971 2978 In: Capinera JL (ed) Encyclopedia of Entomology 2nd edition, Springer, 4346. https://doi.org/10.1007/978-1-4020-6359-6 3029
- Fick SE, Hijmans RJ (2017) WorldClim 2: new 1km spatial resolution climate surfaces for global land areas. Int J Climatol 37:4302– 4315. https://doi.org/10.1002/joc.5086
- Fliszkiewicz M, Giejdasz K, Wasielewski O, Krishnan N (2012) Influence of winter temperature and simulated climate change on body mass and fat body depletion during diapause in adults of the solitary bee, *Osmia rufa* (Hymenoptera: Megachilidae). Environ Entomol 41:1621–1630. https://doi.org/10.1603/EN12004
- Forrest JR, Chisholm SP (2017) Direct benefits and indirect costs of warm temperatures for high-elevation populations of a solitary bee. Ecology 98:359–369. https://doi.org/10.1002/ecy.1655
- Franzén M, Nilsson SG (2013) High population variability and source sink dynamics in a solitary bee species. Ecology 94:1400–1408. https://doi.org/10.1890/11-2260.1
- Galbraith SM, Cane JH, Rivers JW (2021) Wildfire severity influences offspring sex ratio in a native solitary bee. Oecologia 195:65–75. https://doi.org/10.1007/s00442-020-04809-3
- Gallai N, Salles JM, Settele J, Vaissière BE (2009) Economic valuation of the vulnerability of world agriculture confronted with pollinator decline. Ecol Econ 68:810–821. https://doi.org/10.1016/j.ecolecon.2008.06.014
- Garibaldi LA, Carvalheiro LG, Leonhardt SD, Aizen MA, Blaauw BR, Isaacs R et al (2014) From research to action: enhancing crop yield through wild pollinators. Front Ecol Environ 12:439–447. https://doi.org/10.1890/130330
- Garibaldi LA, Requier F, Rollin O, Andersson GK (2017) Towards an integrated species and habitat management of crop pollination. Curr Opin Insect Sci 21:105–114. https://doi.org/10.1016/j. cois.2017.05.016
- Garibaldi LA, Sáez A, Aizen MA, Fijen T, Bartomeus I (2020) Crop pollination management needs flower-visitor monitoring and target values. J Appl Ecol 57(4):664–670.
- Garibaldi LA, Steffan-Dewenter I, Winfree R, Aizen MA, Bommarco R, Cunningham SA et al (2013) Wild pollinators enhance fruit set of crops regardless of honey bee abundance. Science 339:1608–1611. https://doi.org/10.1126/science.1230200
- Gathmann A, Tscharntke T (2002) Foraging ranges of solitary bees. J Anim Ecol 71:757–764. https://doi.org/10.1046/j.1365-2656.2002.00641.x
- Gil-Tapetado D, Castedo-Dorado F, Lombardero MJ, Martel J, Álvarez-Álvarez P (2021) Spatial propagation and patterns of abundance of *Dryocosmus kuriphilus* throughout an invaded region. J Appl Entomol 145:10–25. https://doi.org/10.1111/jen.12836
- Goodell K (2003) Food availability affects *Osmia pumila* (Hymenoptera: Megachilidae) foraging, reproduction, and brood parasitism. Oecologia 134:518–527. https://doi.org/10.1007/s00442-002-1159-2
- Groulx AF, Forrest JR (2018) Nesting aggregation as a predictor of brood parasitism in mason bees (*Osmia* spp). Ecol Entomol 43:182–191. https://doi.org/10.1111/een.12484
- Harrell FE (2001) Regression modeling strategies. Springer New York, New York. https://doi.org/10.1007/978-1-4757-3462-1
- Hofmann MM, Fleischmann A, Renner SS (2020) Foraging distances in six species of solitary bees with body lengths of 6 to 15 mm, inferred from individual tagging, suggest 150 m-rule-of-thumb



- for flower strip distances. J Hymenoptera Res 77:105–117. https://doi.org/10.3897/jhr.77.51182
- Hothorn T (2021) libcoin: Linear Test Statistics for Permutation Inference. R package, available at: https://cran.r-project.org/web/packages/libcoin/index.html
- Hothorn T, Hornik K, Zeileis A (2006) Unbiased recursive partitioning: a conditional inference framework. J Comput Gr Stat 15:651–674. https://doi.org/10.1198/106186006X133933
- Hothorn T, Leisch F, Zeileis A (2020) modeltools: Tools and Classes for Statistical Models. R package, available at: https://cran.r-project.org/web/packages/modeltools/index.html
- Hothorn T, Winell H, Hornik K, van de Wiel MA, Zeileis A (2021) coin: Conditional Inference Procedures in a Permutation Test Framework. R package, available at: https://cran.r-project.org/web/packages/coin/index.html
- International Nut and Dried Fruit (2020) Statistical Year-book 2020. Available online: https://inc.nutfruit.org/wp-content/uploads/2021/09/1594640174\_INC\_Statistical\_Yearbook 2019-2020.pdf
- Isaacs R, Williams N, Ellis J, Pitts-Singer TL, Bommarco R, Vaughan M (2017) Integrated crop pollination: combining strategies to ensure stable and sustainable yields of pollination-dependent crops. Basic Appl Ecol 22:44–60. https://doi.org/10.1016/j.baae.2017.07.003
- Jauker F, Peter F, Wolters V, Diekötter T (2012) Early reproductive benefits of mass-flowering crops to the solitary bee *Osmia rufa* outbalance post-flowering disadvantages. Basic Appl Ecol 13:268–276. https://doi.org/10.1016/j.baae.2012.03.010
- Jost L (2006) Entropy and diversity. Oikos 113:363–375. https://doi. org/10.1111/j.2006.0030-1299.14714.x
- Junqueira CN, Hogendoorn K, Augusto SC (2012) The use of trapnests to manage carpenter bees (Hymenoptera: Apidae: Xylocopini), pollinators of passion fruit (Passifloraceae: Passiflora edulis f. flavicarpa). Ann Entomol Soc Am 105:884–889. https://doi.org/10.1603/AN12061
- Klein AM, Vaissière BE, Cane JH, Steffan-Dewenter I, Cunningham SA, Kremen C, Tscharntke T (2007) Importance of pollinators in changing landscapes for world crops. Proc R Soc B: Biol Sci 274:303–313. https://doi.org/10.1098/rspb.2006.3721
- Krunić M, Stanisavljevic L, Pinzauti M, Felicioli A (2005) The accompanying fauna of Osmia cornuta and Osmia rufa and effective measures of protection. Bullet Insec 58(2):141–152.
- Lazaro A, Müller A, Ebmer AW, Dathe HH, Scheuchl E, Schwarz M et al (2021) Impacts of beekeeping on wild bee diversity and pollination networks in the Aegean Archipelago. Ecography 44:1353– 1365. https://doi.org/10.1111/ecog.05553
- Lomolino MV, Riddle BR, Whittaker RJ (2017) Biogeography. Sinauer Associates, Sunderland Mass
- Lundin O, Rundlöf M, Jonsson M, Bommarco R, Williams NM (2021) Integrated pest and pollinator management—expanding the concept. Front Ecol Environ 19:283–291. https://doi.org/10.1002/fee.2325
- MacIvor JS (2017) Cavity-nest boxes for solitary bees: a century of design and research. Apidologie 48:311–327. https://doi.org/10.1007/s13592-016-0477-z
- Macivor JS, Salehi B (2014) Bee species-specific nesting material attracts a generalist parasitoid: implications for co-occurring bees in nest box enhancements. Environ Entomol 43:1027–1033. https://doi.org/10.1603/EN13241
- Mallinger RE, Gaines-Day HR, Gratton C (2017) Do managed bees have negative effects on wild bees? A systematic review of the literature. PLoS ONE 12. https://doi.org/10.1371/journal.pone.0189268
- Medler JT (1967) Biology of *Osmia* in trap nests in Wisconsin (Hymenoptera: Megachilidae). Ann Entomol Soc Am 60:338–344. https://doi.org/10.1093/aesa/60.2.338

- Michener CD (2007) The bees of the World. John Hopkins University Press, Baltimore
- Müller A, Prosi R, Taylor S, Richter H, Herrmann M, Weibel U (2020)
  Unique nesting biology of *Osmia (Melanosmia) uncinata*, a palaearctic osmiine bee specialized on thick-barked conifers (Hymenoptera, Megachilidae). Alp Entomol 4:157–171. https://doi.org/10.3897/alpento.4.53489
- Müller K, Wickham H, Francois R, Bryan J, RStudio (2023) tibble: Simple Data Frames. R package, available at: https://cran.r-project.org/web/packages/tibble/index.html
- Münster-Swendsen M, Calabuig I (2000) Interaction between the solitary bee *Chelostoma florisomne* and its nest parasite *Sapyga clavicornis* empty cells reduce the impact of parasites. Ecol Entomol 25:63–70. https://doi.org/10.1046/j.1365-2311.2000.00225.x
- Monzón VH, Bosch J, Retana J (2004) Foraging behavior and pollinating effectiveness of *Osmia cornuta* (Hymenoptera: Megachilidae) and *Apis mellifera* (Hymenoptera: Apidae) on comice pear. Apidologie 35:575–585. https://doi.org/10.1051/apido:2004055
- Ollerton J, Winfree R, Tarrant S (2011) How many flowering plants are pollinated by animals? Oikos 120:321–326. https://doi.org/10.1111/j.1600-0706.2010.18644.x
- Ostap-Chec M, Kierat J, Kuszewska K, Woyciechowski M (2021) Red mason bee (*Osmia bicornis*) thermal preferences for nest sites and their effects on offspring survival. Apidologie 52:707–719. https://doi.org/10.1007/s13592-021-00858-6
- Paini DR (2004) Impact of the introduced honey bee (*Apis mellifera*) (Hymenoptera: Apidae) on native bees: a review. Austral Ecol 29:399–407. https://doi.org/10.1111/j.1442-9993.2004.01376.x
- Palladini JD, Maron JL (2014) Reproduction and survival of a solitary bee along native and exotic floral resource gradients. Oecologia 176:789–798. https://doi.org/10.1007/s00442-014-3028-1
- Papanikolaou AD, Kühn I, Frenzel M, Schweiger O (2017) Semi-natural habitats mitigate the effects of temperature rise on wild bees. J Appl Ecol 54:527–536. https://doi.org/10.1111/1365-2664.12763
- Pascual S (2022) Landscape composition and configuration affect bees in the olive agroecosystem. J Appl Entomol 146:1–18. https://doi.org/10.1111/jen.12934
- Potts SG, Biesmeijer JC, Kremen C, Neumann P, Schweiger O, Kunin WE (2010) Global pollinator declines: trends, impacts and drivers. Trends Ecol Evol 25:345–353. https://doi.org/10.1016/j.tree.2010.01.007
- Potts SG, Imperatriz-Fonseca V, Ngo HT, Aizen MA, Biesmeijer JC, Breeze TD et al (2016) Safeguarding pollinators and their values to human well-being. Nature 540:220–229. https://doi.org/10.1038/nature20588
- Prosi R, Wiesbauer H, Müller A (2016) Distribution, biology and habitat of the rare european osmiine bee species *Osmia (Melanosmia) pilicornis* (Hymenoptera, Megachilidae, Osmiini). J Hym Res 52:1. https://doi.org/10.3897/jhr.52.10441
- R Core Team (2022) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Venna, Austria. http://www.R-project.org/
- Roulston TAH, Goodell K (2011) The role of resources and risks in regulating wild bee populations. Ann Rev Entomol 56:293–312. https://doi.org/10.1146/annurev-ento-120709-144802
- Schwarz G (1978) Estimating the dimension of a model. Ann Stat 1:461–464. https://www.jstor.org/stable/2958889
- SEDAC/NASA = Center for International Earth Science Information Network (CIESIN) Columbia University (2018) Documentation for the Gridded Population of the World, Version 4 (GPWv4), Revision 11 Data Sets. Palisades NY: NASA Socioecon Data Appl Cent (SEDAC). Accessed February 2020 https://doi.org/10.7927/H45Q4T5F
- Seidelmann K (2006) Open-cell parasitism shapes maternal investment patterns in the Red Mason bee Osmia rufa. Behav Ecol 17:839–848. https://doi.org/10.1093/beheco/arl017



- Shebl MA, Hassan HA, Kamel SM, Osman MA, Engel MS (2018) Biology of the mason bee *Osmia latreillei* (Hymenoptera: Megachilidae) under artificial nesting conditions in Egypt. J Asia-Pacific Entomol 21:754–759. https://doi.org/10.1016/j.aspen.2018.05.008
- Sheffield CS (2014) Pollination, seed set and fruit quality in apple: studies with *Osmia lignaria* (Hymenoptera: Megachilidae) in the Annapolis Valley, Nova Scotia, Canada. J Pollinat Ecol 12:120–128. https://doi.org/10.26786/1920-7603(2014)11
- Sheffield CS, Pindar A, Packer L, Kevan PG (2013) The potential of cleptoparasitic bees as indicator taxa for assessing bee communities. Apidologie 44:501–510. https://doi.org/10.1007/s13592-013-0200-2
- Steckel J, Westphal C, Peters MK, Bellach M, Rothenwoehrer C, Erasmi S et al (2014) Landscape composition and configuration differently affect trap-nesting bees, wasps and their antagonists. Biol Conserv 172:56–64. https://doi.org/10.1016/j. biocon.2014.02.015
- Steffan-Dewenter I (2003) Importance of habitat area and landscape context for species richness of bees and wasps in fragmented orchard meadows. Conserv Biol 17:1036–1044. https://doi.org/10.1046/j.1523-1739.2003.01575.x
- Steffan-Dewenter I, Schiele S (2008) Do resources or natural enemies drive bee population dynamics in fragmented habitats. Ecol 89:1375–1387. https://doi.org/10.1890/06-1323.1
- Strobl C, Boulesteix AL, Kneib T, Augustin T, Zeileis A (2008) Conditional variable importance for random forests. BMC Bioinform 9:1–11. https://doi.org/10.1186/1471-2105-9-307
- Strobl C, Boulesteix AL, Zeileis A, Hothorn T (2007) Bias in random forest variable importance measures: illustrations, sources and a solution. BMC Bioinform 8:1–21. https://doi.org/10.1186/1471-2105-8-25
- Tlak Gajger I, Laklija I, Jurković M, Košćević A, Dar SA, Ševar M (2022) The impact of different Biotopes and Management Practices on the Burden of Parasites in Artificial Nests of *Osmia* spp. (Megachilidae) Bees Diversity 14:226. https://doi.org/10.3390/d14030226
- Tobajas E, Rosas-Ramos N, Asís JD, de Paz V, Tormos J, Baños-Picón L (2021) Effects of hillside aspect, landscape features, and kleptoparasitism on the reproductive success of the solitary bee *Osmia caerulescens*. Ecol Entomol 46:541–551. https://doi.org/10.1111/een.13000
- Torchio PF (1976) Use of *Osmia lignaria* Say (Hymenoptera: Apoidea, Megachilidae) as a pollinator in an apple and prune orchard. J Kans Entomol Soc 1:475–482. https://www.jstor.org/stable/25082856
- Torchio PF (1979) An eight-year field study involving control of *Sapyga pumila* Cresson (Hymenoptera: Sapygidae), a wasp parasite of the alfalfa leafcutter bee, *Megachile pacifica* Panzer. J Kans Entomol Soc 1:412–419. https://www.jstor.org/stable/25083923
- Trabucco A, Zomer RJ (2018) Global aridity index and potential evapotranspiration (ET0) climate database v2. https://doi.org/10.6084/m9.figshare.7504448.v3. CGIAR Consort Spat Inf
- Tuanmu MN, Jetz W (2015) A global, remote sensing-based characterization of terrestrial habitat heterogeneity for biodiversity and ecosystem modelling. Glob Ecol Biogeogr 24:1329–1339. https://doi.org/10.1111/geb.12365

- Ulbrich K, Seidelmann K (2001) Modeling population dynamics of solitary bees in relation to habitat quality. Web Ecol 2:57–64. https://doi.org/10.5194/we-2-57-2001
- Ungricht S, Mueller A, Dorn S (2008) A taxonomic catalogue of the Palaearctic bees of the tribe Osmiini (Hymenoptera: Apoidea: Megachilidae). Zootaxa 1865:1–253. https://doi.org/10.11646/ zootaxa.1865.1.1
- VanEngelsdorp D, Meixner MD (2010) A historical review of managed honey bee populations in Europe and the United States and the factors that may affect them. J Invertebr Pathol 103:S80–S95. https://doi.org/10.1016/j.jip.2009.06.011
- Vargas MJ, Romero MA (1987) Mejora del almendro en cataluna. Fruticult Prof 11:93–98
- Westrich P (1989) Die Wildbienen Baden-Württembergs. Ulmer. https://doi.org/10.1002/biuz.19900200117
- Wickham H (2016) ggplot2: Elegant Graphics for Data Analysis.

  Springer-Verlag New York. ISBN 978-3-319-24277-4, https://ggplot2.tidyverse.org
- Williams G (2011) Data mining with rattle and R: the art of excavating data for knowledge discovery. Springer New York, New York. https://doi.org/10.1007/978-1-4419-9890-3
- Wilson RS, Leonhardt SD, Burwell CJ, Fuller C, Smith TJ, Kaluza BF, Wallace HM (2020) Landscape simplification modifies trapnesting bee and wasp communities in the subtropics. Insects 11(12):853. https://doi.org/10.3390/insects11120853
- Woodard SH, Jha S (2017) Wild bee nutritional ecology: predicting pollinator population dynamics, movement, and services from floral resources. Curr Opin Insect Sci 21:83–90. https://doi.org/10.1016/j.cois.2017.05.011
- Yamamoto M, da Silva CI, Augusto SC, Barbosa AAA, Oliveira PE (2012) The role of bee diversity in pollination and fruit set of yellow passion fruit (*Passiflora edulis* forma flavicarpa, Passifloraceae) crop in Central Brazil. Apidologie 43:515–526. https://doi.org/10.1007/s13592-012-0120-6
- Yokoi T, Hirooka T, Terada T, Sugimoto S, Tamaru M, Satoh S, Kandori I (2012) Strongly biased sex ratio in cuckoo wasp *Chrysura hirsuta* (Hymenoptera: Chrysididae), a parasitoid of the mason bee *Osmia orientalis*. Entomol Sci 15:250–252. https://doi.org/10.1111/j.1479-8298.2011.00500.x
- Zajdel BA, Kucharska K, Kucharski D, Fliszkiewicz MO, Gąbka JA (2014) Accompanying fauna of red mason bees in annual and perennial nesting sites. Medycyna Weterynaryjna 70:746. https:// doi.org/10.2478/jas-2021-0009
- Zaragoza-Trello C, Vilà M, Bartomeus I (2021) Interaction between warming and landscape foraging resource availability on solitary bee reproduction. J Anim Ecol 90:2536–2546. https://doi.org/10.1111/1365-2656.13559
- Zeileis A, Leisch F, Hornik K, Kleiber C, Hansen B, Merkle EC, Umlauf N (2022) strucchange: Testing, Monitoring, and Dating Structural Changes. R package, available at: https://cran.r-project.org/web/packages/strucchange/index.html

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

