



Germination responses with different treatments, seed vigour and seedling growth of *Pterolobium stellatum* (Forssk.) provenances stored from 19 to 30 years

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Abstract Knowledge of the genetic variability of different species for conservation purposes and quality seedling production after longer storage time is important. The aim of the research paper was to examine the effect of longer seed storage periods (19–30 year) on seed viability, seed vigour and initial growth of seedlings from five provenances of *Pterolobium stellatum* (Forssk). Seeds collected from Dabena, Gambo, Agaro, Bishofitu, and Merti localities and stored for 19, 19, 21, 25, and 30 years, respectively used for the study. Different seed dormancy-breaking treatments such as; soaking in cold water for 48 h (CW-48); soaking in hot water for 10 min (HW-10); 20 min (HW-20) and mechanical scarification; i.e., nicking (NK) used for the germination of the seeds. A randomized complete block design was used for the experiment and the study undertaken in a greenhouse. For each treatment,

four replications and for each replication, 50 seeds were used. One-way analysis of variance was used for germination index, early seedling growth and seed vigour index data. The results showed that for all the provenance seeds stored for different periods, NK treatment resulted relatively in a higher mean germination percentage and germination indices as compared to the other pre-treatments. The seed vigour index result showed significant differences among the different provenances ($P < 0.001$). Similarly, there were significant differences among the early growth of seedlings germinated from different provenance seeds ($P < 0.001$). The seeds collected from Merti provenance and stored for 30 years and Dabena provenance, stored for 19 years, have relatively resulted in higher mean germination index, seed vigour and seedling growth relative to the other studied provenance seeds. We observed that provenance had a more significant influence on germination, seed vigour and seedling growth than the seed storage period for the *P. stellatum*. Finally it is recommended that despite there is inter population variation, the seeds of *P. stellatum* can be stored for a longer time and application of nicking as a dormancy breaking treatment could enhance seed germination of the species.

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Introduction

Pterolobium stellatum (Forssk.) Brenan, which belongs to the *Fabaceae* family, is a perennial tall scandent woody species. It is an evergreen dicotyledonous flowering plant and semierect multistemmed climbing woody thorny shrub that can reach 2–15 m high (Janson and Cardon 2005; Ward and Spellenberg 1988). The species is native to Africa, and its distribution extends from South Africa to Eastern Africa (Janson and Cardon 2005; Teketay 1998). Traditionally, the species is used as a medicinal plant to treat tuberculosis, pneumonia, and other related respiratory diseases (Kigen et al. 2016; Getahun 1976). Leaves and bark are used for ink production (Janson and Cardon 2005). It is also used to treat infertility, epilepsy, and neuroglia (Kigen et al. 2016; Ragunthan and Abay 2009) and to treat sexually transmitted diseases (Njoroge and Bussmann 2006). Some chemical analyses of the plant parts showed that the species possess cytotoxic and anti-diarrhoea properties (Andualem et al. 2014). The plant is also used for animal fodder, live fences, firewood, charcoal, ornamental tree, and soil conservation purposes to have a healthy environment (Alemayehu et al. 2015; Zegeye et al. 2005).

The storage of seeds for a long time is one of the strategies of ex-situ conservation. Keeping seed viability in a conducive environment is an excellent alternative for maintaining or minimizing the loss of physiological quality of seeds during storage until sowing as an ex-situ conservation strategy (Solberg et al. 2020a, b; Silva et al 2019; Vijay 2015; Pradhan and Badola 2008; Ellis et al. 1991). For the conservation of plant genetic resources through ex-situ conservation, knowledge of the characteristics of the seed is important (Pradhan and Badola 2012). Different factors, such as temperature, nature of the seeds, moisture, insects, etc., could affect the viability of the seeds in a storage room, and pre-storage conditions also affect seed longevity (Mdlalose et al. 2021; Solberg et al. 2020a, b; De Vitis et al. 2020; Rao et al. 2017; Pradhan and Badola 2012; Delouche et al. 1973). Seed moisture, vigour, germination percentage, rate of germination, etc., are some of the most important parameters for assessing the seed quality of a species after storage of the seed (Mdlalose et al. 2021). Seed dormancy problems could also affect the germination of the seeds of different species. Seed coat-imposed dormancy

is an ecological mechanism that allows the seed to germinate only when conditions are favourable for supporting seedling growth (Argel and Paton 1999). In the case of legume species, the impermeable seed coat is considered to be the main dormancy problem for the germination of seeds (Grubb and Coomes 1997; Albrecht 1993). Therefore, to break seed dormancy problems using pre-sowing treatments, rapid and uniform germination and high germination capacity can be obtained (Huang and Gutterman 2000; Schutz and Rave 1999; Teketay and Tigabu 1996; Teketay 1996).

Different pre-sowing treatments, such as cold stratification, mechanical disruption, or acid and hot water treatments are widely used to break seed dormancy problems, and these techniques can improve the germination of seeds within a short period (Tadros et al. 2011). Research shows that the seeds of *P. stellatum* have a prolonged dormancy problem, which is associated with the hard seed coat cover of the species (Jemaneh 2019; Teketay 1998). Some authors have indicated that if the seed of *P. stellatum* is stored properly and placed in airtight containers, cool, dry areas and insect-free places, it can stay for a longer period without losing its viability (Teketay 1998; Bein et al. 1996). However, knowledge of the seed longevity of the species of the different provenances of *P. stellatum* and its effect on the germination, seed vigour and seedling growth are lacking. Such knowledge is necessary for conservation purposes for farmers and seed enterprises. The objectives of this study were (1) to assess the impacts of different seed dormancy-breaking treatments on the germination of the seeds of different provenances of *P. stellatum* that were stored for 19–30 years (2) to evaluate the viability of the seeds of different provenances of *P. stellatum* after different periods of storage (3) to examine the seed vigour of the different provenances seeds (4) to assess the quality of the seedlings from the different provenances that were stored for different periods. At the beginning of the research, it was hypothesized that (1) different seed dormancy-breaking treatments have different impacts on the germination of the seeds of *P. stellatum* that were stored for different periods and (2) the storage period and provenances do not have an influence on the viability of the seeds, on seed vigour and seedling quality of *P. stellatum*.

Materials and methods

Seed sources and the storage period

The seeds of *P. stellatum* were collected in Ethiopia from Merti, Bishoftu, Agaro, Dabena, and Gambo localities in 1990, 1996, 2000, 2002, and 2002, respectively. It means that the storage ages of the collected seeds from Dabena, Gambo, Agaro, Bishoftu, and Merti localities were 19, 19, 21, 25, and 30 years, respectively. The mean altitude (meters above sea level) of the area where the provenance seeds of Merti, Bishoftu, Agaro, Dabena and Gambo collected were about 1550 m, 1886 m, 1744 m, 1823 m and 2059 m, correspondingly. A map of the localities where the different seeds of *P. stellatum* were collected is presented in Fig. 1. The documented pieces of information showed that the collected seeds from Agaro, Dabena, Bishoftu, Gambo, and Merti localities were air-dried under shade and stored in a seed store that has a temperature of 5 °C. While storing the seeds have a moisture percentage of 5.6%, 5.4%, 6.3%, 5.3%, and 5%, respectively. According to the information gathered in the seed laboratory, the mean germination percentages of the seeds of the different provenances using nicking as a dormancy-breaking

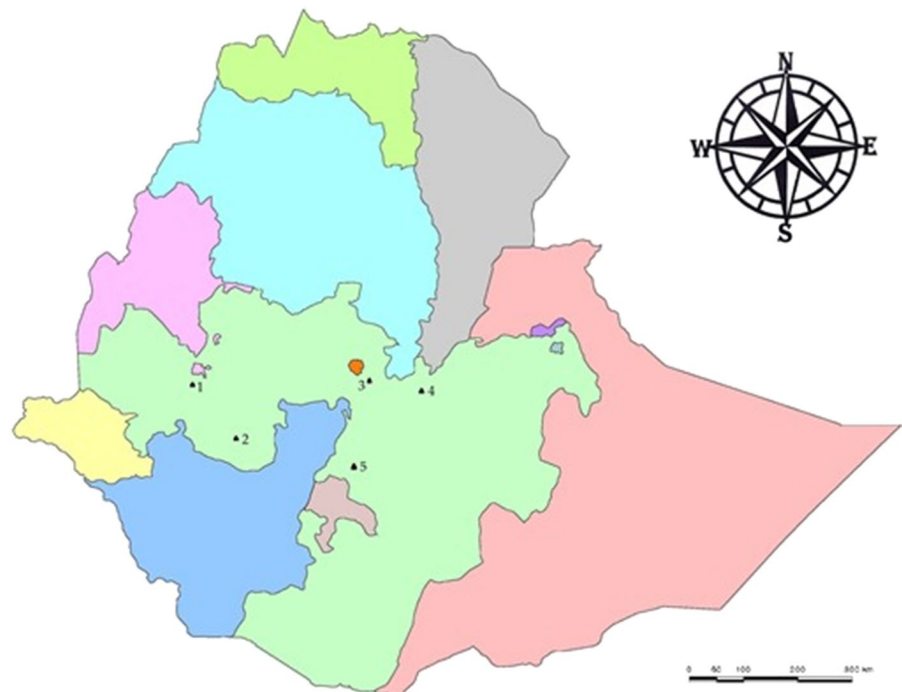
treatment at the time of the seed collection period for Agaro, Dabena, Bishoftu, Gambo, and Merti were 51%, 64%, 65%, 58%, and 65%, respectively.

Germination experiment and seed dormancy-breaking treatments

Randomized complete block design (RCBD) was used for the experiment and the study was undertaken in a greenhouse. Each treatment had four replications (tray boxes), and for each replication, 50 seeds were used. It means a total of 200 seeds were used for a single treatment. The different dormancy-breaking treatments applied for seed germination were (a) soaking in hot water for 10 min (HW-10), (b) soaking in hot water for 20 min (HW-20), (c) soaking in cold water for 48 h (CW-48), (d) mechanical scarification, i.e., nicking (NK), and e) control (CN), i.e., without any pre-sowing treatment. These dormancy breaking treatments are selected since the seed of the species has a hard seed coat cover, and the treatments are mostly recommended.

The germination experiment was performed using a tray box filled with sand (i.e., the sand has a thickness of approximately 10 cm) in a greenhouse. The sand that was used as a germination medium was

Fig. 1 Locations of the different provenances where the seeds of *Pterolobium stellatum* is collected. 1. Dabena 2. Agaro 3. Bishoftu 4. Merti 5. Gambo



cleaned using tap water through washing before the experiment. The seeds were sown in the sand-filled tray and then carefully covered by the sand. Watering of sown seeds was undertaken once a day in the morning till the end of the experiment. The germination study was attended between 17.03.2021 and 12.04.2021, i.e., for 26 days. After germination, the seedlings were kept in the tray to grow until the end of the experiment and used to evaluate the seed vigour (Fig. 2).

Data collection and analysis

Data on the germination of the emerged seedlings were collected daily. At the end of the experiment, i.e., 13/04/2021, each of the individual germinated and growing seedlings of the different provenances and different treatments were carefully uprooted. The root length and shoot length of uprooted individual seedlings were measured using a ruler, independently. Viability test of un-germinated seeds was not undertaken since the experiment was performed using sand, that serves as a media of germination, and for this reason it was difficult to find the buried and un-germinated seeds.

The collected germination and seedling shoot length and root length data were encoded in an MS

Excel sheet. The germination data was arc-sine transformed before the analysis. The normality of the distribution of the data was evaluated. One-way analysis of variance (ANOVA) was used for the handling of the data. When there were significant differences, Fisher's least significance difference (LSD) test was used for the mean separation at a significance level of $P=0.05$. SigmaPlot 13 (Systat Software, Inc., San Jose, CA, USA) program was used for the statistical analysis.

The mean final germination percentage (GP) using methodology of Scott et al. (1984), mean germination time (MGT) (Orchard 1977) and mean germination index (GI) (Bench et al. 1991) were analysed using the Eq. 1, 2, and 3, correspondingly. While the seed vigour index was calculated using the formula suggested by Abdul-Baki (1973) as presented in Eq. 4.

The final germination percentage (GP) was calculated using Formula (1):

$$GP = \frac{TG}{TS} \quad (1)$$

where TG = total number of seeds germinated.

TS = total number of sown seeds.

The mean germination time (MGT) of seeds was calculated using Formula (2):



Fig. 2 Germination tray boxes filled with sand that are used for the germination study and seedlings of *Pterolobium stellatum* in the greenhouse

$$\text{MGT} = \frac{\sum_{n=1}^n T1N1 + T2N2 + \dots + TkNk}{\sum_{n=1}^n N1 + N2 + \dots + Nk} \quad (2)$$

where N = number of newly germinated seeds.

T = time from the beginning of the experiment.

The germination index (GI) was calculated using Formula (3):

$$\text{GI} = \sum_{n=1}^n (T1 N1 + T2 N2 + \dots + Tk Nk) \quad (3)$$

where N = number of newly germinated seeds,

T = time from the beginning of the experiment.

The seed vigour Index (VI) was calculated using the formula (4)

$$\text{VI} = \frac{\text{SL}}{\text{GP}} \quad (4)$$

where SL = Seedling length (root length + shoot length).

GP = Germination percentage.

Results

Effect of pre-sowing treatments on seed germination

The effects of the different seed dormancy-breaking treatments on the mean germination percentage of

P. stellatum seeds stored for different periods is presented in Table 1. The results revealed that, despite their variation in their storage periods, for all the studied provenances seeds, the NK treatment resulted in a higher mean germination percentage relative to the other dormancy-breaking treatments applied to the seeds of *P. stellatum* (Table 1). Similarly, the mean germination index result, which combines the germination percentage and mean germination time of the seeds of the different provenances of *P. stellatum*, shows that seeds treated with NK also resulted in a higher mean germination index relative to the other seed dormancy-breaking treatments applied to the seeds of the species (Table 2). The seeds collected from Bishoftu provenance and stored for 25 years sown after NK treatment resulted in the highest mean germination percentage relative to the other provenances (Table 1). The seeds from Dabena provenance that is treated with CW-48, HW-20, HW-10 and CN have relatively higher mean germination percentages compared to the mean germination percentage of the seeds of other provenances (Table 1). However, the Gambo provenance which has the same period of storage as the Dabena provenance with all seed dormancy-breaking treatments has resulted in a relatively lower mean germination percentage as compared to the other provenance seeds (Table 1).

Despite, the NK treatment resulted in a relatively higher mean germination percentage (Table 1) and

Table 1 The mean germination percentage of the seeds of the different provenances of *Pterolobium stellatum* with different pre-sowing treatments. Means with similar letters are not significantly different ($P = 0.05$). Values in parentheses are standard errors

Treatments	Provenances				
	Dabena	Merti	Gambo	Bishoftu	Agaro
Cold water for 48 h	32 (4.5) ^{AB}	27.5 (1.7) ^A	26.5 (6.0) ^A	26.5 (1.3) ^B	24 (2.2) ^A
Hot water for 10 min	27.5 (0.5) ^B	26 (2) ^A	19.5 (3.9) ^A	22.5 (3.4) ^B	20 (2.2) ^A
Hot water for 20 min	37 (2.9) ^A	22 (2.9) ^A	20 (3.3) ^A	23 (3.1) ^B	25 (1.7) ^A
Nicking	40 (2.9) ^A	29 (1.7) ^A	28.5 (3.9) ^A	44 (2.7) ^A	31 (4.2) ^A
Control	29.5 (2.6) ^B	28 (2.7) ^A	19.5 (4.3) ^{φA}	25 (4.4) ^B	24 (2.4) ^A
<i>P</i> -value	0.050	0.27	0.443	<0.001	0.121

Table 2 The mean germination index result of the seeds of the different provenances with the different pre-sowing treatments. Values in parentheses are standard errors

Seed treatments	Agaro	Dabena	Gambo	Bishoftu	Merti
Coldwater for 48 h	326 (17)	433 (55)	370 (99)	315 (7)	365 (42)
Hot water for 10 min	306 (41)	401 (22)	239 (44)	311 (33)	369 (24)
Hot water for 20 min	379 (27)	537 (43)	257 (26)	307 (42)	341 (51)
Nicking	519 (73)	682 (41)	613 (105)	742 (49)	459 (30)
Control	375 (41)	399 (32)	224 (45)	303 (48)	350 (35)

germination index (Table 2) for all the provenance, the one-way ANOVA result revealed no significant differences among the different seed dormancy breaking treatments (HW-10, HW-20, CW-48, NK, and CN) within the Agaro provenance ($P=0.121$, Table 1), within Gambo provenance ($P=0.443$, Table 1), and Merti provenance ($P=0.27$, Table 1). On the other hand, the statistical analysis results showed a significant difference in the mean germination percentage between the seed dormancy-breaking treatments applied within the Dabena provenance ($P=0.05$, Table 1) and within Bishoftu provenance ($P=0.001$, Table 1). Figure 3 shows the one-way ANOVA result on the mean germination percentage of the different provenance seeds with different pre-sowing treatments. Accordingly, the one-way ANOVA analysis result revealed no significant differences in the mean germination percentage of the seeds among the Agaro, Dabena, Bishoftu, Gambo, and Merti provenances that were treated with CW-48 ($P=0.636$), HW-10 ($P=0.186$), CN seeds ($P=0.295$) (Fig. 3). However, the one-way ANOVA results revealed significant differences in the mean germination percentage of the seeds among the provenances of Agaro, Dabena, Bishoftu, Gambo and

Merti that were treated with HW-20 ($P=0.006$) and NK ($P=0.01$, Fig. 3).

Seed viability and seed vigour

The seed laboratory test result on the mean germination percentage of the different provenances of *P. stellatum* seeds in different years, with NK treatment, is presented in Fig. 4. At the time of seed collection, the mean germination percentage of the different provenance seeds of *P. stellatum* with NK treatment for Dabena, Agaro, Bishoftu, Merti and Gambo provenances was 64%, 51%, 65%, 65%, and 58%, respectively (Fig. 5). In the year 2021, the mean germination percentages of the seeds with NK treatment for the Dabena, Agaro, Bishoftu, Merti and Gambo provenances were 40%, 31%, 44%, 29%, and 29%, respectively. This result further indicated that the mean annual viability losses of the seeds of *P. stellatum* collected from Dabena, Agaro, Bishoftu, Merti, and Gambo provenances were 1.1%, 0.91%, 1.9%, 1.2% and 1.2%, respectively.

The seed vigour index analysis result for the Agaro, Dabena, Bishoftu, Gambo, and Merti provenances is presented in Fig. 5. The analysis result showed that

Fig. 3 One-way analysis of variance results on the mean germination percentage of the different provenance seeds with different pre-sowing treatments. Means with similar letters are not significantly different ($P=0.05$)

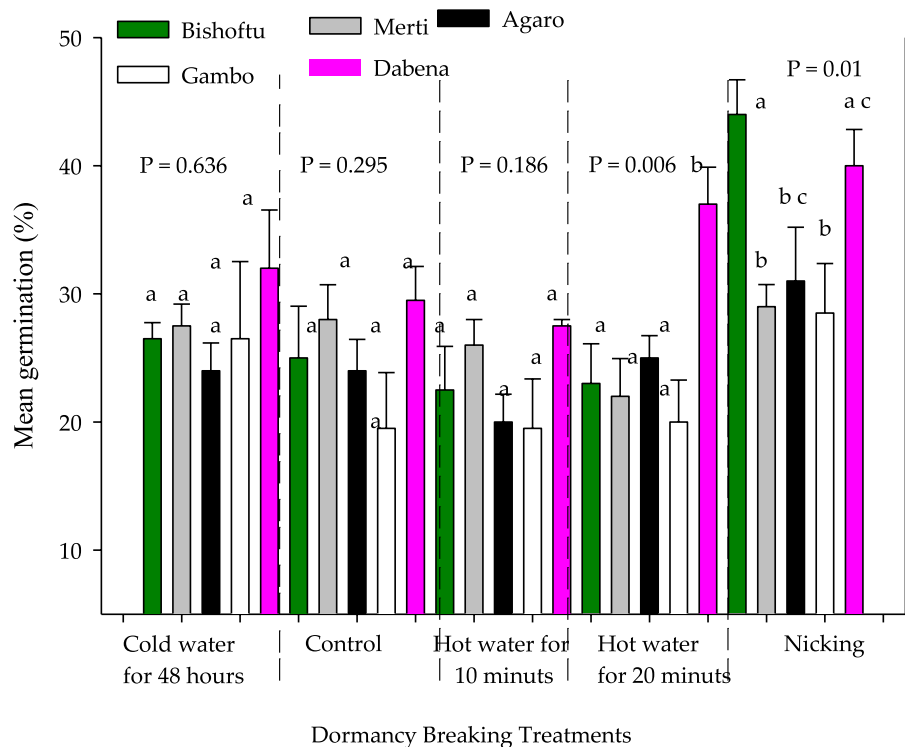


Fig. 4 Mean germination percentage of the different provenance seeds of *Pterolobium stellatum* using mechanical scarification; i.e., nicking, as a pre-sowing treatment in the seed laboratory (germination results between 1990 and 2020) and in the final germination with the present experiment in the year 2021

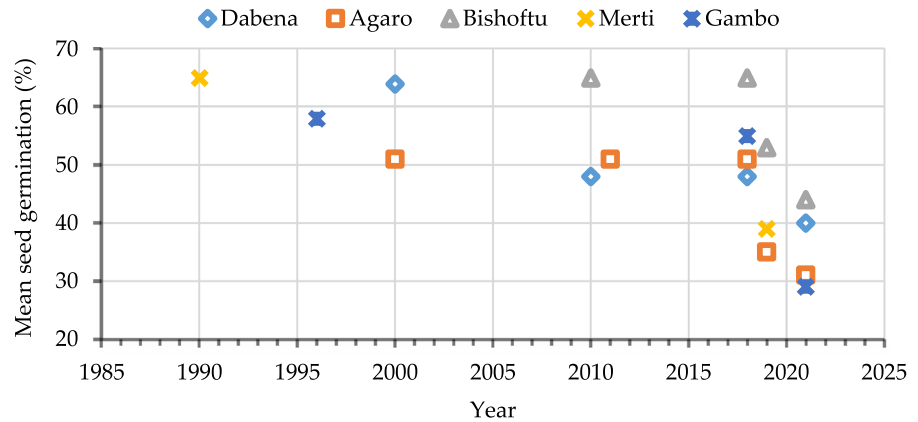
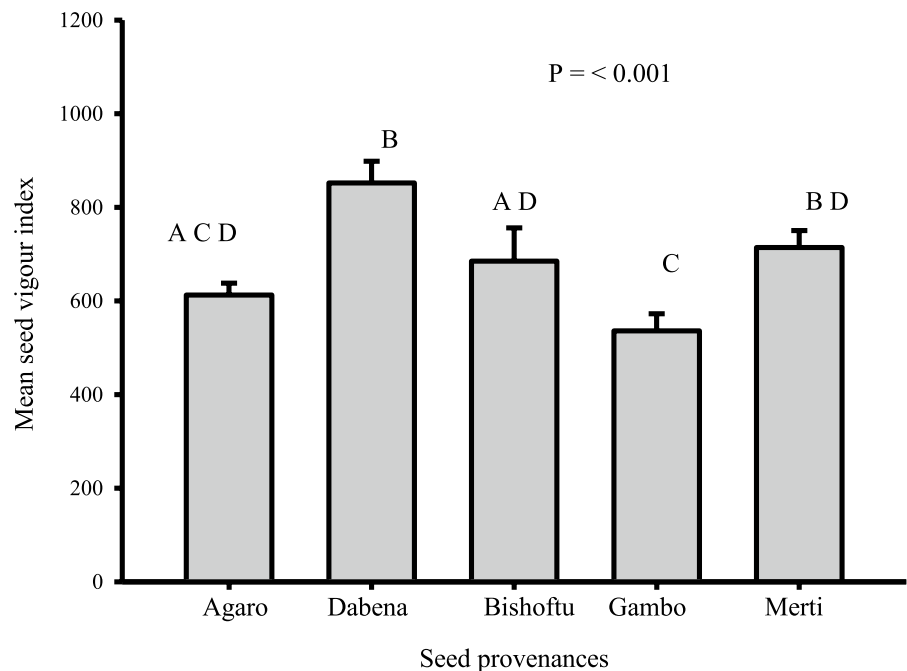


Fig. 5 Mean seed vigour index of the different provenances of *Pterolobium stellatum* seeds. Means with the same letters are not significantly different ($P=0.05$)



the mean seed vigour index for the Agaro, Dabena, Bishoftu, Gambo, and Merti provenances were 613, 852, 685, 536, and 714, respectively. The statistical one-way ANOVA result revealed significant differences among the different seed provenances stored for different periods (Fig. 5). The significant differences in the seed vigour indices were between the provenances of Dabena and Gambo; Dabena and Agaro; Dabena and Bishoftu; Gambo and Merti and Bishoftu and Gambo seeds ($P<0.05$, Fig. 5). However, the statistical analysis results showed no significant differences in the seed vigour index between the Dabena and Merti; Merti and Agaro; Merti and Bishoftu;

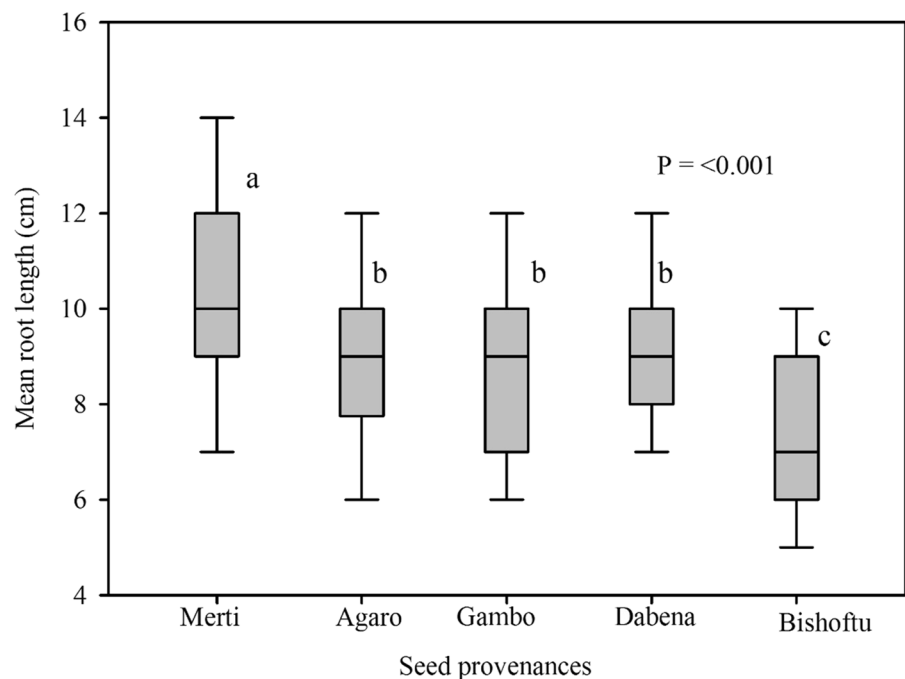
Bishoftu and Agaro; and Agaro and Gambo provenances ($P>0.05$, Fig. 5).

Early growth of seedlings from the different provenances

The analysis result showed that the mean root lengths of the seedlings germinated and grown from Gambo, Dabena, Agaro, Bishoftu and Merti provenances where their seeds stored for 19, 19, 21, 25 and 30 years were 8.6 cm, 9.3 cm, 8.8 cm, 7.5 cm and 10.2 cm, correspondingly. The one-way ANOVA analysis result showed significant differences in the

mean root lengths of the germinated seedlings from the seeds of the different provenances ($P = <0.001$, Fig. 6). Accordingly, the significant differences in the mean seedling root lengths were between the provenance seeds of Merti and Agaro; Merti and Gambo and Merti and Bishoftu (Fig. 6). However, the statistical one-way ANOVA result showed no significant differences in the mean root lengths of the seedlings that were germinated and grown from Agaro, Gambo and Dabena provenances (Fig. 6, $P > 0.05$). The result on the mean shoot lengths of the seedlings germinated and grown from the provenance seeds of Gambo, Dabena, Agaro, Bishoftu and Merti provenances were 16.2 cm, 17.4 cm 17.5 cm, 17.8 cm and 18.4 cm, respectively. The one-way ANOVA analysis results on the mean shoot lengths of the seedlings grown from the different provenance seeds are presented in Fig. 7. The analysis result showed significant differences in the mean shoot lengths of seedlings grown from the different provenance seeds ($P < 0.001$), and the significant differences were between Merti and Gambo; and Merti and Dabena provenances (Fig. 7). However, the statistical analysis result revealed no significant differences in the mean shoot lengths of the seedlings grown from the provenances seeds between Merti and Agaro; Merti and Bishoftu and Dabena and Bishoftu (Fig. 7, $P > 0.05$).

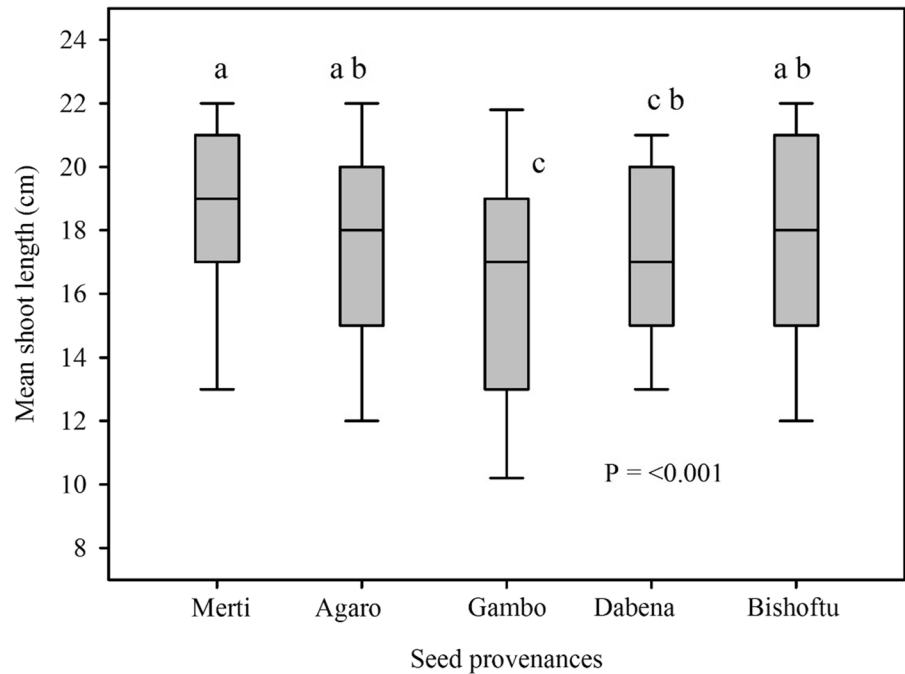
Fig. 6 One-way analysis of variance results on the mean root length of the seedlings grown from the seeds of the studied provenances. Means with the same letters are not significantly different ($P = 0.05$)



Discussion

The present study finding showed that for all the provenance of *P. stellatum* seeds that were stored for different periods, NK as a dormancy-breaking treatment, resulted, in relatively a higher mean germination percentage and mean germination index relative to the other seed dormancy-breaking treatments applied for the study. This result corresponds with the findings of Teketay et al. (1998) and Jemaneh (2019), who also in their findings indicated that nicking is the best treatment to break the seed dormancy problem of freshly collected *P. stellatum* seeds. Rodrigues-Junior et al. (2020) and Burrows et al. (2019) indicated that most of the *Fabaceae* family has a problem of physical dormancy, which is related to blockage of water uptake during the dormant state. Similar to many other *Fabaceae* species, the seeds of *P. stellatum* have a dormancy problem, which can be solved using NK. The species (*P. stellatum*) belongs to the *Fabaceae* family and different findings also recommended that scarification (mechanical treatment) as the recommended pre-sowing preparation for the dormancy breaking of this family species (Parera and Ruiz 2003; Seng and Cheong 2020; Schnadelbach et al. 2016; Iroko et al. 2021; Perez-Garcia 2009; Fulbright and Flenniken 1987). Some species have a problem with

Fig. 7 One-way analysis of variance results on the mean shoot length of the seedlings grown from the seeds of the studied provenances. Means with the same letters are not significantly different ($P=0.05$)



seed germination since part of the seed testa slows the embryo water intake, which slows seed germination (Burrows et al. 2019). Generally, the present results showed that the seeds of *P. stellatum* had a hard seed coat cover that inhibits water entrance to the cotyledon, which affects seed germination, and this problem can be solved with NK treatment.

Knowledge of inter-population variation can improve the understanding of the mechanisms regulating seed germination (Bhatt 2021). The present result shows that despite the Gambo provenance and Dabena provenance seeds were stored for equal periods (19 years), their mean germination percentage and mean germination index of the Gambo provenance seeds with all the dormancy breaking treatments were relatively lower than those of the Dabena provenance seeds. This differences could be associated with the inter-population variation of the two provenances. The seed vigour index result also proved that the seeds of Dabena provenance were more vigorous than the Gambo provenance despite both having equal storage time. These results further could show that provenance affects the seed germination and seed vigour of *P. stellatum* which shows a variation on the provenances of the species. The differences in the mean germination of the *P. stellatum* seeds collected from the Dabena provenance relative

to Gambo provenance could be associated with their differences in the imbibition patterns of the seeds of the provenances. Differences in the texture of the seed, temperature, seed coat thickness, seed coat colour affects the imbibition pattern of a seed. Burrows et al. (2009) indicated that variability in the germination rate of three Tasmanian provenances of *Acacia melanoxylon* was associated with a difference in their imbibition patterns. Similarly, Burrows et al. (2019) found a variation in their imbibition in seeds of 48 *Acacia* species, which might be the cause of the differences in the germination of the different provenances. Fredrick et al. (2017) also revealed significant differences in different provenances of *Faidherbia albida* germination percentage that was associated with the variation of the seeds in imbibition.

Similarly, despite the storage period of the seeds of the Gambo provenance (19 years) was lower than that of the Merti (30 years), Bishoftu (25 years), and Agaro (21 years) provenances, its mean germination percentage and mean germination index with all the different seed treatments was lower than that of the other studied provenances. This may show that provenance may have a more significant effect on the germination of the *P. stellatum* species instead of the storage period. The results also showed that the mean annual seed viability loss for the seeds collected from Dabena, Agaro,

Bishoftu, Merti and Gambo provenances was on average 1.1%, 0.9%, 1.9%, 1.2%, and 1.2%, respectively. This could indicate that the mean storage time of the seeds has an impact on the mean germination capacity of the seeds of different provenances of *P. stellatum*. Research shows that seed storage of *Fabaceae* species for a longer time alters the germination capacity of different species (Jayasuriya et al. 2013; Colombe et al. 2019). The study showed that the viability of the seeds of the different provenances declined in its germination about 1.3% per year. The species seed can be stored for a longer time. Merritt et al. (2014) examined 172 Australian wild species and found many of the legume (*Fabaceae*) species to be stored for a longer time. Bhatt et al. (2021) found a variation in the germination percentage of three *Astragalus sieberi* D.C. (*Fabaceae*) populations.

Seed vigour refers to seed properties with the potential for rapid, uniform emergence and development of healthy and vigorous quality seedlings (Mangena 2021). Abdul-Baki and Anderson (1973) also indicated that the seed lot that shows the higher seed vigour index is considered to be more vigorous. The present study results revealed that the seeds collected from Dabena provenance have a higher vigour index compared to all other studied provenances, which shows that the seed lot is more vigorous. This might be associated with the lower storage period of the seeds of the provenance. De Vitis et al. (2020) indicated that increasing seed age can reduce germination vigour as the seed metabolic system begins to break down, resulting in seeds being slow or even unable to germinate, poor seedling development, and lower establishment for aged seeds that germinate. However, the Gambo provenance which has equal storage age with the Dabena provenance has resulted in the lowest seed vigour index even lower than those provenances where their seeds stored longer than the seeds of other studied provenances. Some provenances could result in a higher seed vigour index despite their long time of seed storage. This result could be associated with inter population variation of the provenance seeds.

The seed storage period is an important factor that affects seed viability and seedling growth of different plant species. For both the root and shoot growths, seedlings grown from Merti provenances which were stored for 30 years were relatively greater than the other studied provenance where their seeds were stored for less period. Despite the Gambo and Dabena

provenance seeds were stored for an equal period (19 years) the results showed that seedlings grown from Dabena provenances achieved higher root length and shoot length relative to the Gambo provenance seedlings. The one-way ANOVA result also showed significant differences in the mean root lengths and shoot lengths of the seedlings grown from different provenance seeds that were stored for different periods. All these results could show that in the case of *P. stellatum* provenance has more effect on the seedling growth than storage time which might be associated with genetic factors that influence the seed quality. Different research also showed that the effect of the seed storage period on seedling growth may vary depending on the provenance of the seeds, as different environmental and genetic factors may influence the seed quality and dormancy (Molla and Desta 2022; Wawrzyniak et al. 2020).

Conclusion

Overall, the present study shows that (1) the seeds of *P. stellatum* have a physical dormancy problem that can be solved using the nicking as a presowing treatment and (2) It is observed that provenance had a significant influence on germination, seed vigour and seedling growth than seed storage period for the *P. stellatum* which shows the genetic variability of the species (3) the mean annual viability loss of the seeds of *P. stellatum* collected from different provenances was 1.3%. Finally, based on the present study it is recommended that the *P. stellatum* species has inter-population variation and for the purpose of ex-situ conservation their viability period as the storage time increases varies.

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Declarations

Conflict of interest The authors declare no conflict of interest.

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