

Anthropogenic and natural causes influencing population genetic structure of *Juniperus procera* Hochst. ex Endl. in the Ethiopian highlands

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Abstract *Juniperus procera* is economically highly important but threatened tree species. It is the only species among 67 taxa in the genus *Juniperus* that naturally grows in Africa and south of the equator extending up to 18°S in Zimbabwe. Ethiopia is assumed to host the largest *J. procera* populations, which are also believed to have high genetic variation owing to their wide ecological amplitude. This study assessed genetic variation at AFLPs of *J. procera* populations in the Ethiopian highlands. In the study six populations, namely Chilimo, Goba, Menagesha-Suba, Wef-Washa, Yabelo and Ziquala were included. A total of 20–24 trees from each population were investigated based on 128 AFLP band positions. AMOVA revealed that most of the variation (94%) resided within populations of *J. procera* suggesting extensive gene flow among populations which is

attributable to the outcrossing mating system and effective gene transport mechanisms of the species. However, genetic differentiation among populations was still significant ($P < 0.05$), and the differentiation was significantly ($P < 0.05$) correlated with geographic distance. All population pairs were significantly ($P < 0.05$) differentiated except for Menagesha-Suba and Wef-Washa. These two populations also showed the highest gene diversity ($H_j = 0.301$ and $H_j = 0.297$, respectively). These results are in accordance with historical records that claim the establishment of the Menagesha-Suba juniper population as plantation of seedlings from Wef-Washa back in fifteenth century.

Keywords Anthropogenic impact · Gene flow · Genetic structure · *Juniperus procera* · Natural barriers

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Introduction

The history of the Ethiopian highland forests witnessed a huge impact of human activities and climate variability and changes (Gebu et al. 2009; Darbyshire et al. 2003; Teketay 1992). These activities and events have changed the forest ecosystems of the region; species in these ecosystems are believed to have undergone enormous changes in their genetic composition. According to recent findings, the Ethiopian highland forest cover has undergone four major phases

in the last 3,000 years: the pre-history *Podocarpus* dominant podo-juniper phase, man-induced decline in podo-juniper forest, forest recovery of the juniper dominant phase and the last human induced decline in forest cover (Darbyshire et al. 2003). A consequence of these phases is that nowadays the highlands of Ethiopia are characterized by highly fragmented small patches of remnant forests in remote and protected areas. Forest fragmentation is a significant threat to the maintenance of genetic resources (Broadbent et al. 2008; Tabarelli and Peres 2002), since it is often accompanied by an erosion of genetic variation and increased interpopulation genetic divergence due to increased random genetic drift, elevated inbreeding and reduced gene flow (Young et al. 1996).

High genetic variations for Ethiopian highland forest species such as *Hagenia abyssinica* (Bruce) J.F. Gmel., *Cordia africana* Lam., *Lobelia giberroa* Hemsl., and *Coffea arabica* L. have been reported by Ayele et al. (2009), Derero et al. (2010), Kebede et al. (2007) and Aga et al. (2005), respectively. The authors suggest both human and non-human impacts on genetic variation patterns and propose possible conservation strategies. For example, Ayele et al. (2009) and Aga et al. (2005) discuss human mediated gene transport as an important factor for the current genetic structure of *H. abyssinica* and *C. arabica*.

Juniperus procera Hochst. ex Endl. (Cupressaceae), commercially known as the African Pencil Cedar, is an evergreen tree that grows to a height of over 40 m and a diameter above 3 m (Pohjonen and Pukkala 1992; Negash 1995). *J. procera* is the only species among 67 taxa in the genus *Juniperus* that naturally grows in Africa. Its habitat stretches from the Arabian Peninsula in Asia to Zimbabwe in Africa (Adams 2004). *J. procera* is assumed to have evolved from *J. excelsa* M. Bieb. or from a common ancestor, and expanded southwards from the western mountains of the Arabian Peninsula to Ethiopia across the Red Sea and then further to the present southernmost populations in Zimbabwe (Adams 2004; Farjon 2005). The species grows in small scattered populations in most areas. It is one of the economically most important tree species in Ethiopia having favourable wood qualities such as fine texture, straight grains, workability and high resistance to termite and decay. It is used for a variety of purposes like constructions, furniture and outdoor works (Negash 1995; Bekele-Tesema 2007). Consequently, it is one of the tree

species that has been subjected to extensive logging (Negash 1995), leading its status to endangered in some provenances (Garzulia 2006; Borghesio et al. 2004) and threatened in its entire habitat (IUCN 2009).

The Ethiopian highlands are described as the largest reservoir of the species (Bekele 1994; Yirdaw 2002). In Ethiopia, it is one of the dominant tree species in most remnant highland natural forests (Negash 1995). Historical evidences indicate that most of the northern Ethiopian highlands that are currently devoid of forests once were dominantly covered by *J. procera* (Logan 1946; White 1983; Friis 1992; Teketay 1992; Darbyshire et al. 2003). Today, remnants of the old stands have been reserved only around old churches and monasteries in these highlands (Bongers et al. 2006). These sanctuaries are presently the main source of information to reconstruct the historical forest status of the area (Wassie et al. 2005). In the Ethiopian highlands *J. procera* grows naturally in pure stands or mixed with *Podocarpus falcatus* (Thunb.) R. Br. ex Mirb. in lower altitudes (Negash 1995) and with *Hagenia abyssinica* at higher altitudes (Umer et al. 2007). *J. procera* is known to grow in Ethiopia as low as 1,700 m around Yabelo (Tigabu et al. 2007) and as high as over 3,300 m in the Bale Mountains (Umer et al. 2007).

Owing to its occurrence over a wide range of elevations, ecological heterogeneity is believed to have resulted in high genetic differentiation among populations of *J. procera* (Mamo et al. 2006; Demissie and Bjornstad 1997). On the other hand, obligatory outcrossing of this dioecious species (Negash 2002) and efficient gene flow via the wind-dispersed pollen and seed migration by frugivorous birds (Adams 2004) potentially decrease differentiation, but enhance high diversity within stands. Historical records indicate that Ethiopian highland forests in general and *J. procera* forests in particular have been subjected to over 3,000 years of anthropogenic influences (Gebru et al. 2009; Darbyshire et al. 2003; Teketay 1992). Anthropogenic gene transportation might have contributed remarkably to long distance seed dispersal and the establishment of *Juniperus* populations in the central highlands of Ethiopia in fifteenth century (Demissew 1988; Eshetu 2002).

Genetic variation of *J. procera* has rarely been studied. Mamo et al. (2006) and Tigabu et al. (2007) prove pronounced variation in seed morphology and

germination percentage among Ethiopian populations of the species. However, these results do not allow to assess the variation of heritable traits of the species within or among populations.

Adams et al. (1993) included samples from Ethiopia (*J. procera*) and Greece (*J. excelsa*) to study past migration of *Juniperus* in Arabia and East Africa through RAPD fingerprints and terpenoids. However, diversity and differentiation within samples from Ethiopia was not analyzed. Mlangeni (2005) investigated genetic variation of *J. procera* from three countries including Ethiopia using AFLP markers. However, his study included only three populations from Ethiopia and low sample sizes, which makes it difficult to draw conclusions on the genetic structure of the species in Ethiopia.

The objectives of the present study are to assess genetic variation within and between populations of *J. procera* in Ethiopia, covering its wide geographic distribution and to explore the contribution of anthropogenic and natural causes for the current genetic structure of *J. procera* populations in Ethiopia.

Materials and methods

Study area

Samples were collected from six populations: Chilimo, Menagesha-Suba, Wef-Washa, Ziquala, Goba and Yabelo. Presence of natural barriers and historical information were considered for the selection of these populations (Table 1; Fig. 1). Chilimo, Menagesha-Suba and Wef-Washa represent populations in the central highland plateaus Northwest of the Rift Valley. On the other hand, Goba and Yabelo represent the Southeastern massif of the Southeastern

and Southern highlands, respectively. Ziquala is an uplifted mountain within the Rift valley on the Northwestern side.

Sampling

From each population 20–24 trees were randomly sampled and needles (leaves) of each selected tree were collected for DNA analysis in separate plastic bags containing silica gel. In order to avoid the collection of related trees a minimum of 50 m distance was kept between trees within a population. Geographic coordinates of each population and distance from the preceding point were also measured using a GPS.

Laboratory analyses

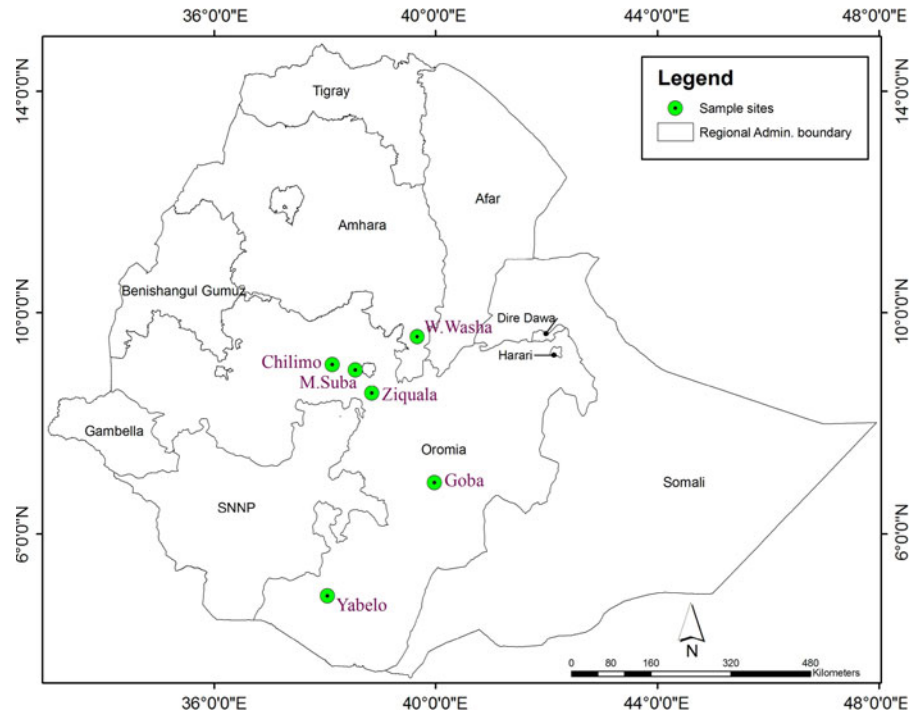
Total genomic DNA was extracted from a small amount of dried needles (about 20 mg) following the DNeasy 96 Plant Kit protocol of the manufacturer (Qiagen, Hilden, Germany). AFLP reactions were performed as described by Vos et al. (1995) with minor modifications. The genomic DNA was digested with the two restriction enzymes *EcoRI/MseI*. The corresponding double-stranded (ds) *EcoRI*-adapter and *MseI*-adapter were ligated to the sticky ends of the restriction fragments to generate template DNA for PCR amplification. Fragments were amplified with two primer pairs complementary to the adapters and restriction site sequences with one to three additional “selective” nucleotides at their 3′-ends. The *EcoRI* + 1 primer (E01) and *MseI* + 1 primer (M03) with selective nucleotides A and G, respectively, were used for a pre-selective amplification. The primer combination E35 and M69 (Keygene code), each +3 primers with selective nucleotides of ACA and GCG, respectively were used for selective amplification

Table 1 Sample sites information

| Population name | Altitude (m) | Latitude (0′) | Longitude (0′) | Annual rain fall (mm) | Geographic position to the Rift Valley |
|-----------------|--------------|---------------|----------------|------------------------|--|
| Chilimo | 2,461 | 09 04 | 38 08 | 1,610 ^a | Northwest (NW) |
| Goba | 2,956 | 06 56 | 39 59 | 925–1,150 ^b | Southeast (SE) |
| M. Suba | 2,633 | 08 58 | 38 33 | 1,250 ^a | Northwest (NW) |
| W. Washa | 2,837 | 09 34 | 39 40 | 1,047 ^a | Northwest (NW) |
| Yabelo | 1,880 | 04 53 | 38 03 | 744 ^a | Southeast (SE) |
| Ziquala | 2,767 | 08 33 | 38 51 | 1,215 ^c | Within Rift Valley (RV) |

Rainfall information source: ^a Mamo et al. 2006; ^b Umer et al. 2007; ^c Ayele et al. 2009

Fig. 1 Geographic map of Ethiopia showing the sample sites



(nomenclature of AFLP primer enzyme combinations.). Primer E35 was labeled with the fluorescent dye 6-FAM. All PCR reactions were performed in the Peltier Thermal Cycler (PTC-200 version 4.0, MJ Research). PCR-amplification products were diluted in proportion of 1:20. Then, 2 μ l diluted product of each probe was mixed with 12 μ l Hi-Di™ formamide and the internal size standard GeneScan 500 ROX (2 μ l for 96 samples) from Applied Biosystems, and run on an automatic sequencer (ABI PRISM 3100 Genetic Analyser, Applied Biosystems). The size of the AFLP fragments was assessed with the software packages GeneScan 3.7 and Genotyper 3.7 (Applied Biosystems).

Data analysis

The reproducibility of amplification products was tested by performing the AFLP procedure and analyses twice for five samples. Only 100% reproducible fragments were considered. Accordingly, a total of 128 AFLP band positions within a readable range from 50 to 475 bps were selected and scored for band presence (1) and absence (0) at a particular position. Each band was manually monitored and edited. The band positions were transformed into a

binary matrix, '0' for absence and '1' for presence of a fragment. The matrices were saved in excel format for further analyses.

Within population genetic diversity

The percentage of polymorphic loci (PPL) at the 5% level and Nei's (1973) gene diversity (H_j) were computed using the software AFLP-Surv 1.0 (Vekemans et al. 2002) following a Bayesian approach with non-uniform prior distribution of allele frequencies (Zhitovskiy 1999). Computation was made after 10,000 permutations and 1,000 bootstraps. Shannon's index was estimated using the software POPGENE version 1.32 (Yeh et al. 2000).

Among population differentiation

Analysis of molecular variance (AMOVA) (Excoffier et al. 1992) and pairwise population differentiation were computed using Arlequin 3.11 (Excoffier et al. 2006) at 10,000 permutations. For AMOVA, populations were grouped into three groups based on their geographic location with reference of the Great Rift Valley (Goba and Yabelo-Southeast, Ziquala within and the rest Northwest/Central highlands). In order to

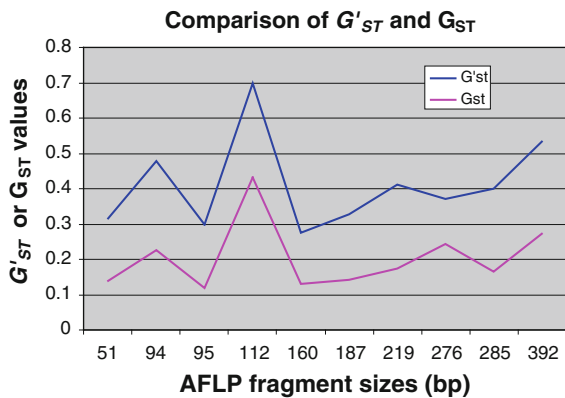


Fig. 2 Nei's differentiation (G_{ST}) and Hedrick's standardized differentiation (G'_{ST})

estimate the magnitude of variation among populations contributed by individual markers, standardized genetic differentiations (G'_{ST}) at each band positions were calculated according to Hedrick (2005) from Nei's (1973) coefficient of genetic differentiations (G_{ST}) of each band computed in POPGENE version

1.32 (Yeh et al. 2000) and pattern of changes in G_{ST} versus G'_{ST} values are shown in Fig. 2. Frequency distributions of ten AFLPs with the highest G'_{ST} values are shown in Fig. 3. Nei's (1978) genetic distance was computed using AFLP-Surv 1.0 from which an UPGMA (Unweighted pair group method using arithmetic averages) dendrogram was generated using the method of SAHN clustering in NTSYSpc 2.0 (Rohlf 1998). In order to estimate the influence of geographic distance on genetic distance between population pairs, a Mantel test (Mantel 1967) was performed using GenAlEx6.3 (Smouse et al. 2008; Peakall and Smouse 2006).

Results

Within population genetic diversity

The lowest ($H_j = 0.23049$) and the highest ($H_j = 0.31874$) gene diversities, congruently Shannon

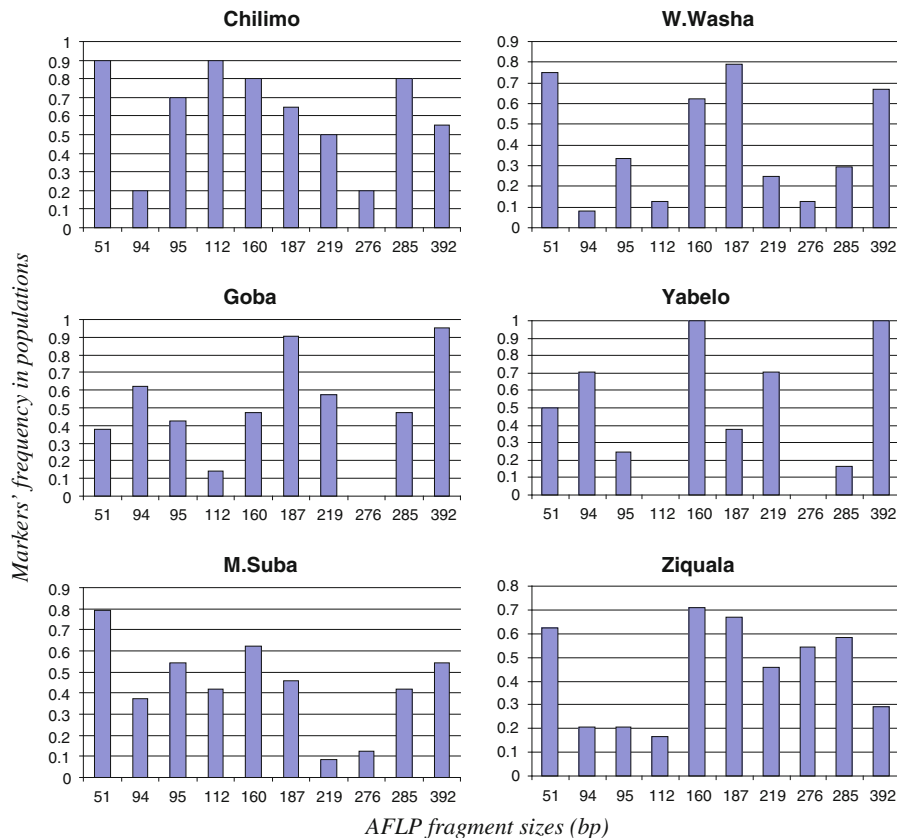


Fig. 3 Relative frequencies of AFLP fragments with the top highest G'_{ST} values in populations

Table 2 Within population genetic variation of six *J. procera* populations

| Population | <i>n</i> | Loc. | PL | PPL | H_j | <i>I</i> |
|------------|----------|------|-----|------|---------|----------|
| Chilimo | 20 | 128 | 119 | 93.0 | 0.26369 | 0.2490 |
| Goba | 21 | 128 | 63 | 49.2 | 0.23049 | 0.2139 |
| M.Suba | 24 | 128 | 119 | 93.0 | 0.31874 | 0.3064 |
| W.Washa | 24 | 128 | 116 | 90.6 | 0.31036 | 0.3039 |
| Yabelo | 24 | 128 | 113 | 88.3 | 0.26431 | 0.2474 |
| Ziquala | 24 | 128 | 117 | 91.4 | 0.28428 | 0.2761 |

n number of individuals, #*loc* number of loci, #*PL* polymorphic loci, *PPL* %PL, H_j , gene diversity, *I* Shannon index

indices were observed in Goba and Menagesha-Suba populations, respectively. The lowest PPL (49%) was computed for Goba. Generally, populations South East of the Rift valley (Goba and Yabelo) showed lower PPL and gene diversity than other populations (Table 2).

Among population variation

AMOVA

The AMOVA showed that only about 6% of the total variation was attributable to variation among popu-

lations (Table 3). A small, but significant amount of the genetic variation (3.16%) was distributed among geographic regions.

Pairwise population differentiation

Significant ($P < 0.05$) genetic differentiation was observed for all pairwise comparisons in the F_{ST} analysis. Only populations Menagesha-Suba and Wef-Washa from the same geographic region showed no significant differentiation ($P = 0.32031$) (Table 4). Most of the population pairs showed highly significant differentiation ($P < 0.01$). The highest genetic differentiation ($F_{ST} = 0.12758$) was observed between Yabelo from the Southeast and Chilimo from the Northwest.

Population clustering

The Mantel test revealed a significant correlation ($P < 0.05$) between geographic distances and pairwise genetic distances (Fig. 4). The UPGMA dendrogram based on Nei's (1978) genetic distance also reflected the correlation between geographic and genetic distances (Fig. 5). However, the relatively

Table 3 Partitioning of the total diversity based on an AMOVA

| Source of variation | <i>df</i> | Sum of squares | Variance components | Percent of variation | <i>P</i> value |
|----------------------------------|-----------|----------------|---------------------|----------------------|----------------|
| Among populations | 5 | 139.495 | 0.72843 | 6.06 | 0.000 |
| Within populations | 131 | 1,478.271 | 11.28451 | 93.94 | 0.000 |
| Among regions | 2 | 75.504 | 0.38228 | 3.16 | 0.024 |
| Among populations within regions | 3 | 63.991 | 0.44598 | 3.68 | 0.000 |
| Within population | 131 | 1,478.271 | 11.28451 | 93.16 | 0.000 |

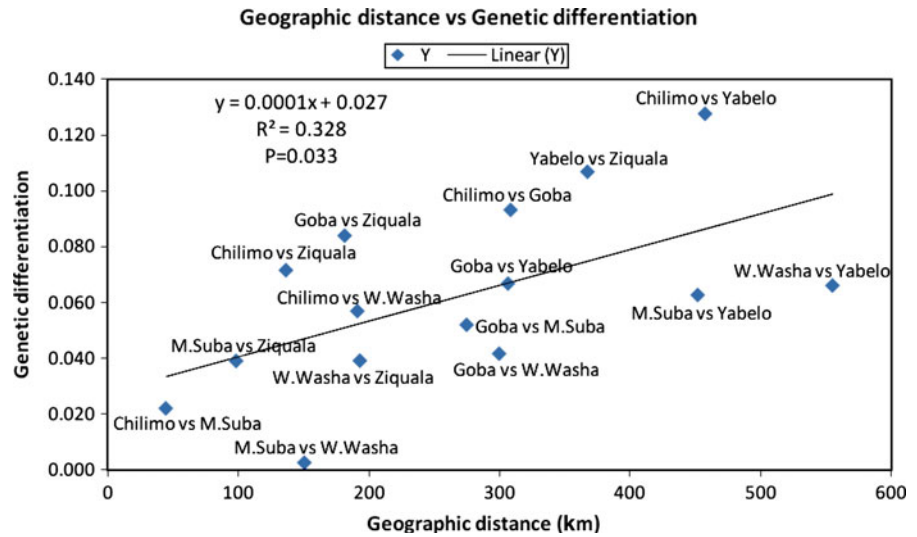
df degree of freedom

Table 4 Pairwise genetic differentiation (F_{ST}) between *J. procera* populations

| Population | Chilimo | Goba | M.Suba | W.Washa | Yabelo | Ziquala |
|------------|---------|---------|---------|---------|---------|---------|
| Chilimo | | *** | * | *** | *** | *** |
| Goba | 0.09311 | | *** | ** | *** | *** |
| M. Suba | 0.02191 | 0.05189 | | NS | *** | ** |
| W. Washa | 0.05683 | 0.04152 | 0.00244 | | *** | ** |
| Yabelo | 0.12758 | 0.06667 | 0.06258 | 0.06598 | | *** |
| Ziquala | 0.07148 | 0.08387 | 0.03893 | 0.03902 | 0.10678 | |

* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; ^{NS} $P > 0.05$

Fig. 4 Mantel test between pairwise genetic differentiation (F_{ST}) and geographic distances



distant northwestern populations (Fig. 1) Menagesha-Suba and Wef-Washa grouped together.

Markers contributing to high genetic differentiation

Out of the 128 markers ten had G'_{ST} above 0.27. South eastern populations (Goba and Yabelo) tended to be unique in harboring 94 and 392 bps size markers at a higher frequency, a 51 bp marker at a relatively lower frequency and lacking the 276 bp marker. Yabelo further lacked the marker with the highest G'_{ST} of 112 bp (Fig. 3).

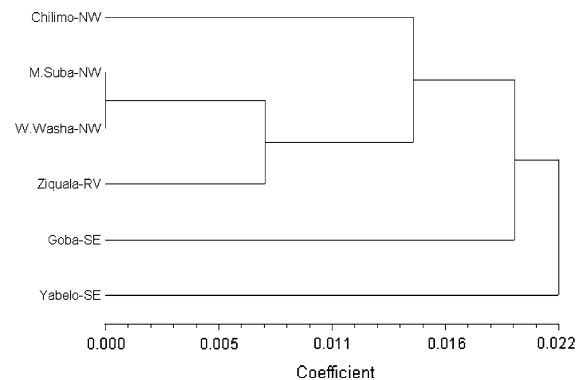


Fig. 5 UPGMA dendrogram based on unbiased Nei's genetic distance

Discussion

Within population variation and overall differentiation

The high within population variation and low population differentiation is most likely attributable to the reproductive biology and gene transportation mechanism of the species. *J. procera* is an obligatory outcrossing species (Negash 1995; Adams 2004; Farjon 2005) with efficient means of gene dispersal by pollen and seed; this promotes heterozygosity and the maintenance of high within population genetic diversity (White et al. 2007), but results in low differentiation among populations. The results obtained in this study are in accordance with many earlier studies that have reported high within population genetic variation as typical genetic structure for dioecious species (Billington 1991; Hamrick and Godt 1996; Weidema et al. 2000; Gerard et al. 2004). The production of large amounts of pollen by *J. procera* (Coetzee 1967) and the formation of pure stands in Ethiopia (Negash 1995) promote the efficiency of wind pollination.

Unlike most other conifers that produce stony cones, juniper seeds (except *J. drupacea* Labill.) are frequently dispersed by frugivorous birds (Adams et al. 1993; Adams 2004) which further contributes to gene exchange among distant populations. The relatively higher gene diversity of Ziquala, which is located on an uplifted isolated mountain in the rift valley, is

possibly linked to the high movement of birds that have promoted reproductive contact of this stand with other populations (Adams 2004). Ziquala is known as haven of many bird species (BirdLife International 2009, available on <http://www.birdlife.org>).

In accordance with the present results, Mlangeni (2005) observed high within population genetic variation investigating samples of *J. procera* that originated from different countries including Ethiopia based on AMOVA of 264 AFLP markers which ranged from 83.32% for Kenyan populations to 98.65% for Malawian populations and where the Ethiopian population scored 85.34%.

Contribution of geographic distance and natural barriers for population genetic variability

Although the majority of the genetic variation resided within populations, the AMOVA revealed that there was significant ($P < 0.05$) variation among populations as well (Table 3). The highest differentiation ($F_{ST} = 0.1278$, Table 4) was realized between Yabelo and Chilimo; which is in agreement with the long geographic distance between these two populations. Yabelo is the most divergent population as shown in the UPGMA dendrogram (Fig. 5). This population also appeared unique lacking an AFLP fragment of 112 bp that is most frequent at Chilimo and also present in all the other populations (Fig. 3). The strong differentiation of the Yabelo population from other populations is in line with the findings of Tigabu et al. (2007) and Mamo et al. (2006) who observed a high germination rate and unique morphology of *Juniperus* seeds from this population.

The significant ($P < 0.05$) variation among groups based on their location with reference to the Great Rift Valley suggests limited gene flow across this natural barrier. This view is supported by the strong differentiation of southeastern population (Goba and Yabelo) from the other populations on the northwestern side of the valley.

Yabelo has the second lowest gene diversity possibly attributable to its geographical isolation from other *J. procera* populations. The Yabelo population is considered as 'lowland' juniper in Ethiopia (Tigabu et al. 2007) where the species is less frequent. Similarly, Chilimo harbored a comparatively low genetic diversity as compared to the other populations in the central highlands reflecting its

marginal geographic position for species of dry evergreen forests such as *J. procera* (Negash 1995; Couralet et al. 2005).

Anthropogenic impact on genetic variability of *J. procera*

The Ethiopian highland forest cover has passed different phases of alterations due to human activities (Darbyshire et al. 2003). According to recent findings, gymnosperms (*P. falcatus* and *J. procera*) dominated forests in the Ethiopian highlands since 7,000 years ago where these species have been enormously subjected to human influences (Gebru et al. 2009). In the present study, the genetic similarity between Menagesha-Suba and Wef-Washa populations (Table 4) clearly illustrates anthropogenic effects on the genetic structure of *J. procera* in the Ethiopian highlands. Despite their isolation by larger geographic distance, they show a very low genetic differentiation ($F_{ST} = 0.00244$). According to written and oral records, Menagesha-Suba was established as plantation of wildlings from Wef-Washa in the fifteenth century by an Abyssinian king Zere-Yacob (Gilbert 1970; Demissew 1988). Through carbon isotopes analyses, Eshetu and Högberg (2000) confirmed that *J. procera* at Menagesha-Suba was originally established as plantation over 550 years ago. However, their study gave no indication from where exactly the original planting material originated. Despite all efforts to trace the origin of this plantation, the aforementioned historical records could neither be supported nor rejected so far. Hence, our results appear to be the first to confirm century-years old records claiming Wef-Washa as seed source of *Juniperus* at Menagesha-Suba.

The high genetic diversity in the presumably planted population Menagesha-Suba (Table 2) is likely to be due to a high number and diversity of the founder plants during establishment (Tang et al. 2003; Parisod et al. 2005). The Menagesha-Suba population has been protected since the 1600s by Imperial decrees (Demissew 1988) until to date as State forest which enabled the occurrence of refuge trees as old as about 600 years (Eshetu and Högberg 2000; Eshetu 2002), young regeneration and hence the presence of all the age classes in this population. Less disturbed populations harbor higher genetic diversity than populations subjected to frequent

disturbances (Lefevre et al. 2004; White et al. 2007). Anthropogenic gene transportation might also have contributed to the relatively high genetic diversity at Ziquala, a population under an old and big Monastery, which was reportedly established in the late twelfth century (Tamene 1998). It has very likely been visited by monks of other ancient monasteries that are sanctuaries of tree species in the Northern highlands (Bongers et al. 2006).

Even though the Goba population is in close distance to other populations of the Bale Mountains (Malcolm and Evangelista 2005; Umer et al. 2007) which might have favored higher gene diversity than in smaller and fragmented populations (Hamrick et al. 1992; Finkeldey and Hattermer 2007; White et al. 2007), it had the lowest gene diversity and a very low proportion of polymorphic loci (Table 2). On the other hand, the highest gene diversity was observed in this region for other species such as for *Hagenia abyssinica* (Ayele et al. 2008), *Lobelia gimberroa* (Kebede et al. 2007) and *Coffea arabica* (Aga et al. 2005). The Goba population has been highly disturbed by human activities as compared to the other population analyzed in this study. The proximity of this population to the Goba town might contribute for its sever disturbance. Thus, the Goba population possibly has experienced population bottlenecks (Nei et al. 1975) due to excessive disturbances.

Genetic structure of *J. procera* and its implication for conservation

Efficient mechanisms of gene dispersal via pollen and seed contributed to the maintenance of high genetic diversity within populations in the past, but significant genetic differentiation, a significant correlation between spatial and geographic distances, and evidence for natural barriers limiting gene dispersal was also observed. Accordingly, potentially unique genetic information might be harbored at least in some of the populations. This calls for conservation activities in multiple populations with special focus on populations characterized by high genetic differentiation. The result suggests to consider the geographic location and environmental barriers to set conservation priorities where genetic data are lacking.

The low diversity observed in the Goba population suggests that anthropogenic activities leading to heavy population disturbances can affect the genetic

composition of the species considerably. Anthropogenic activities therefore appear to be potential threats for the loss of genetic information particularly in spatially isolated small populations where genetic drift is possible. Gene flow from larger populations possibly enhances diversity in disturbed neighboring populations. The occurrence of large juniper forests close to the Goba population suggests the possibility to restore its genetic diversity by natural processes (gene flow of pollen and seed migration) if conservation measures allowing natural regeneration are implemented.

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