



Novel pentacyclic derivatives and benzylidenes of the progesterone series cause anti-estrogenic and antiproliferative effects and induce apoptosis in breast cancer cells

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Abstract

The promising antitumor effects of progesterone derivatives have been identified in many studies. However, the specific mechanism of action of this class of compounds has not been fully described. Therefore, in this study, we investigated the antiproliferative and (anti)estrogenic activities of novel pentacyclic derivatives and benzylidenes of the progesterone series. The antiproliferative effects of the compounds were evaluated on hormone-dependent MCF7 breast cancer cells using the MTT test. Estrogen receptor α (ER α) activity was assessed by a luciferase-based reporter assay. Immunoblotting was used to evaluate the expression of signaling proteins. All benzylidenes demonstrated inhibitory effects with IC₅₀ values below 10 μ M, whereas pentacyclic derivatives were less active. These patterns may be associated with the lability of the geometry of benzylidene molecules, which contributes to an increase in the affinity of interaction with the receptor. The selected compounds showed significant anti-estrogenic potency. Benzylidene **1d** ((8 S,9 S,10R,13 S,14 S,17 S)-17-[(2E)-3-(4-fluorophenyl)prop-2-enoyl]-10,13-dimethyl-1,2,6,7,8,9,11,12,14,15-decahydrocyclopenta[a]phenanthren-3-one) was the most active in antiproliferative and anti-estrogenic assays. Apoptosis induced by compound **1d** was accompanied by decreases in CDK4, ER α , and Cyclin D1 expression. Compounds **1d** and **3d** were characterized by high inhibitory potency against resistant breast cancer cells. Apoptosis induced by the leader compounds was confirmed by PARP cleavage and flow cytometry analysis. Compound **3d** caused cell arrest in the G2/M phase. Further analysis of novel derivatives of the progesterone series is of great importance for medicinal chemistry, drug design, and oncology.

Keywords Antiestrogenic activity · Benzylidenes · Breast cancer · Estrogen receptor α · Pentacyclic steroids

Introduction

Steroids are widely distributed in nature, and they have various effects on plant and animal cells [1]. Studies conducted in the 20th century showed that steroids and their metabolism play an extremely important role in human health. Steroidogenesis in humans is a multi-stage process that results in the formation of necessary steroidal products and their delivery to sensitive tissues. Disruption of steroid hormone signaling and activities of steroidogenic enzymes lead to

the development of several serious pathologies. Steroids are used to treat many different diseases, including allergic rhinitis, asthma, chronic obstructive pulmonary disease [2, 3], hives and eczema [4], painful joints or muscles (such as arthritis, tennis elbow and frozen shoulder) [5], pain caused by an irritated or trapped nerve (such as sciatica) [6], inflammatory bowel disease (such as Crohn's disease) [7], systemic lupus erythematosus [8], and multiple sclerosis [9]. The onset of the COVID-19 pandemic in 2020 also highlighted the importance of steroids for human health and medical applications [10, 11]. Dexamethasone was declared a “major development” in the fight against coronavirus disease [12, 13]. Dexamethasone has been described as a recent advancement that significantly reduces the mortality rate among severe COVID-19 patients [12].

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Steroids and steroid-like compounds are of particular importance to the progression and treatment of cancers [14]. The transformation of a normal cell into a malignant one is a very complex process. In some cases, steroids play an important supporting role in the growth of malignant cells, including breast and prostate cancer. It has been shown that 60–70% of breast cancers are hormone-dependent, which means that their growth is supported by estrogens. Estrogens, which are steroid hormones and regulate many processes in normal tissues, are converted into “traitors” during the progression of female cancers. When estrogens reach the tumor tissue, rapid cell proliferation occurs. These features of breast cancer have become a focus for many researchers since the 1940s [15, 16]. The discovery of tamoxifen, an effective anti-estrogen, has significantly improved the treatment of breast cancers [17, 18]. Tamoxifen (2-[4-[(Z)-1,2-diphenylbut-1-enyl]phenoxy]-N,N-dimethylethanamine) is not a steroid molecule, but it has a potent anti-estrogenic effect on hormone-dependent cells [19]. Further development of the series of synthetic steroids produced several effective steroidal anti-estrogens, including fulvestrant [20, 21]. Tamoxifen, fulvestrant, and other anti-estrogens penetrate breast cancer cells and bind to estrogen receptor α (ER α), blocking its interaction with estrogens. “Turning off” estrogen receptor α leads to a decrease in the proliferation rate of breast cancer cells [22, 23].

It was later found that blocking estrogen receptor α is not always sufficient to inhibit the growth of hormone-dependent cancer. In particular, it is very important to reduce the synthesis of estrogens. Thus, inhibitors of aromatase, an enzyme converting androgens to estrogens, have been synthesized [24]. Steroidal and nonsteroidal aromatase inhibitors showing high efficacy in experimental and clinical studies were obtained [25, 26]. Aromatase inhibitor testolactone, which was used as an anticancer drug to treat advanced-stage breast cancer, was discontinued in 2008 [27, 28]. Novel steroidal aromatase inhibitors, including exemestane and formestane, which have been developed on a steroid framework, are currently widely used to treat breast cancer [29].

The antitumor effects of progesterone derivatives have been identified in many studies. Cyproterone acetate is a progesterone derivative with antiandrogenic and progesterone-like activity that has been used in the treatment of advanced prostate cancer [30, 31]. It has not been approved by the Food and Drug Administration for use in the United States but has been approved in other countries. Various targets are considered for progesterone derivatives. Muafia Jabeen and colleagues conducted a pharmacological evaluation and docking studies of progesterone derivatives as anticancer agents [32]. They showed that progesterone derivatives exhibiting antiproliferative activity can bind to estrogen

receptor α . In a previous study [33], progesterone derivatives were studied as inhibitors of the 5 α reductase enzyme. A phase II study with the progesterone receptor antagonist lonaprisan (ZK 230,211) was performed on progesterone-positive breast cancers. That study examined the efficacy, safety, and tolerability of lonaprisan at two different doses (25 and 100 mg) before the start of phase III trials [34, 35]. The following clinical trials with lonaprisan were discontinued, and further development of the drug was terminated. The progesterone receptor antagonist onapristone has been used as first-line endocrine therapy in breast cancer patients; clinical trials of this drug have been restricted due to its liver toxicity [36]. Dosage forms of onapristone with extended release have been developed. New clinical trials are planned to study letrozole, palbociclib, and extended-release onapristone in patients with metastatic breast cancer (ClinicalTrials.gov ID: NCT04872608). In some cases, approved steroid drugs show adverse effects, low effectiveness, or are characterized by low bioavailability. For example, the clinical efficacy of the lone clinically approved estrogen receptor α degrader, fulvestrant, is limited by its poor oral bioavailability [20]. Although many steroid compounds with anticancer activity have been synthesized, the relevance of new developments in the steroid framework remains high. In this study, we obtained benzylidenes and pentacyclic derivatives of the progesterone series and evaluated their antiproliferative activities and potencies as anti-estrogens in hormone-dependent breast cancer cells.

Materials and methods

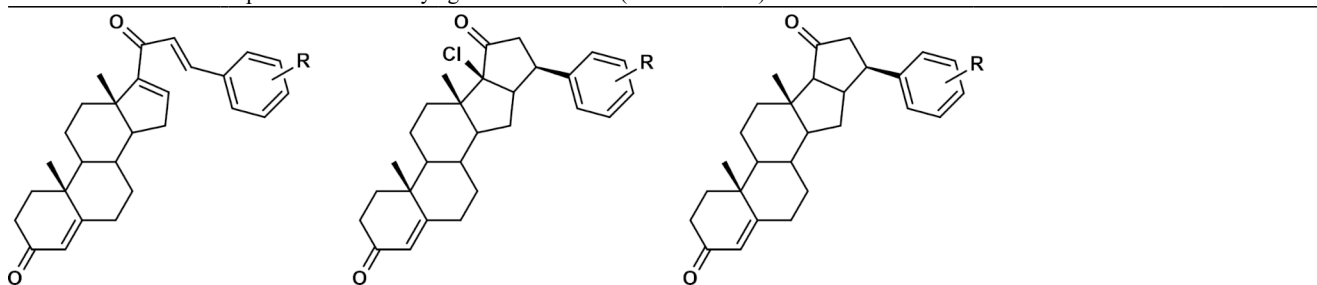
Chemistry

We studied the anticancer activities of three classes of steroids derived based on 16-dehydropregnenolone (Table 1). The desired compounds (benzylidenes **1a–h** [37] and D-annulated pentacyclic steroids **2a–h** [38] and **3b, c, e, and f** [39]) were synthesized by previously described methods. General information on these compounds is provided in Sect. 1 of the SI. Synthesis structural characterization including the ^1H NMR spectra of novel pentacyclic steroids **3a, d, g, and h** are described in Sect. 1.2 of the SI.

Biology

Evaluation of antiproliferative activity

The MCF7 human breast cancer cell line was purchased from the ATCC collection. MCF7 cells were cultured in standard DMEM medium (Gibco) supplemented with 10% fetal bovine serum (FBS, HyClone) at 37 °C, 5%

Table 1 Evaluation of antiproliferative activity against MCF7 cells (72 h treatment)


Entry	1a-h Code	2a-h R	3a-h IC ₅₀ values against MCF7 cells, μM		
			1 ^a	2 ^a	3
1	a	H	2.7 ± 0.3	10.0 ± 1.5	22.7 ± 2.6
2	b	4-Cl	3.1 ± 0.3	6.0 ± 0.8	8.8 ± 0.9
3	c	4-Br	3.6 ± 0.4	8.1 ± 0.9	10.9 ± 1.2
4	d	4-F	1.9 ± 0.2^b	7.1 ± 0.9	7.3 ± 0.8
5	e	2,4-Cl ₂	3.5 ± 0.3	8.2 ± 0.9	13.0 ± 1.8
6	f	2-Cl-6-F	2.4 ± 0.2	17.7 ± 2.1	18.8 ± 2.1
7	g	3-MeO	6.7 ± 0.5	7.2 ± 0.8	25.1 ± 2.9
8	h	3,4,5-(MeO) ₃	2.0 ± 0.2	10.6 ± 1.3	14.7 ± 1.8

^a Our recently published data

^b The most active compound

CO₂ and 80–85% humidity (NuAire CO₂ incubator). The growth of MCF7 cells was evaluated by the modified MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide) (AppliChem) test [40] as described earlier [41]. The resistant cell lines were cultivated according to the method previously described in our work [42]. The cells were seeded at a density of 40 × 10³ cells per well in 24-well plates (TPP) in 900 μL of medium. The obtained compounds were dissolved in DMSO (AppliChem) to 5 mM before experiments, and then, the resulting solutions were diluted in the medium to the required concentrations. The viability of the cells after 72-h treatments was assessed after subtraction of the blank value (the absorbance in the well w/o cells) from all wells. Dose–response curves were analyzed by regression analysis using sigmoidal curves (Log(concentration) vs. normalized absorbance). The half-maximal inhibitory concentrations (IC₅₀) were determined with GraphPad Prism.

Luciferase activity

Transfection was performed as described earlier [43] with some modifications. Briefly, MCF7 cells were seeded at a density of 170 × 10³ cells per well onto 24-well plates a day before transfection. The next day, the medium was changed from DMEM containing 10% fetal bovine serum (FBS) to phenol red-free DMEM supplemented with 2% dextran-coated charcoal-treated (DCC) serum (steroid-free conditions). Then, the cells were transfected with plasmid containing the luciferase gene controlled by estrogen

response elements (ERE). To normalize the transfection efficacy, the β-galactosidase plasmid was co-transfected to the cells along with ERE-Luc. The plasmids used in this study were kindly provided by Frank Gannon and colleagues [44]. Transfection was performed for 4 h with Lipofectamine 2000 (Thermo Fisher Scientific) according to the manufacturer's manual. Then, fresh 10% DCC serum-containing phenol red-free DMEM was added to the plates; the studied compounds, which were diluted in medium, were added to the wells as well as the vehicle control and tamoxifen at a concentration of 5 μM. To induce ERα activity, 10 nM 17β-estradiol (Sigma-Aldrich) was used, and it was added 40 min after treatments with the compounds. Cells were lysed within 18 h, and luciferase activity was measured according to the Promega protocol using a Tecan Infinite M200 Pro. The activity of β-galactosidase was analyzed by colorimetric assay using a MultiScan plate reader. The luciferase/β-galactosidase activities were normalized by the internal control values and represented as the mean ± SD value. The relative luciferase activity in 17β-estradiol-treated cells was taken as 100 units. ERα activities were calculated in relative units as the ratio of the luciferase/galactosidase activity.

Immunoblotting

MCF7 breast cancer cells were seeded on 100-mm dishes (Corning), and after 24 h of growth, the compounds were added to a fresh medium. To prepare the cell extracts, MCF7

cells were washed twice in phosphate buffer and incubated for 10 min on ice in modified lysis buffer containing 50 mM Tris-HCl at pH 7.5, 0.5% Igepal CA-630, 150 mM NaCl, 1 mM EDTA, 1 mM DTT, 1 mM PMSF, 0.1 mM sodium orthovanadate, aprotinin, leupeptin, and pepstatin (1 µg/mL each) as described earlier [45]. The protein content was determined using the Bradford method [46].

Cell lysates (40 µg of protein) were separated in 10% SDS-PAGE under reducing conditions, transferred to a nitrocellulose membrane (GE HealthCare), and processed according to a standard protocol. To prevent nonspecific absorption, the membranes were treated with 5% nonfat milk solution in TBS buffer (20 mM Tris and 500 mM NaCl at pH 7.5) with 0.1% Tween-20 and then incubated with primary antibodies overnight at 4 °C.

ER α , Cyclin D1, CDK2, CDK4, and PARP antibodies were obtained from Cell Signaling Technology; the antibodies against α -tubulin (Cell Signaling Technology) were added to standardize loading. Goat anti-rabbit IgGs (Jackson ImmunoResearch) conjugated to horseradish peroxidase were used as secondary antibodies. Signals were detected using ECL reagent (as described in Mruk and Cheng's protocol [47]) and an ImageQuant LAS4000 system (GE HealthCare).

Cell cycle analysis and evaluation of apoptosis

Cell cycle distribution and apoptosis were analyzed as described previously [48]. MCF7 cells were seeded in 6-well plates in DMEM containing 10% FBS and treated with compounds **3d**, **2d**, and **1d** for 24 h. Cell sediments were lysed in a buffer containing 50 µg/mL propidium iodide (Sigma-Aldrich), 100 µg/mL RNase A (Qiagen), 0.1% sodium citrate, and 0.3% NP-40 (Helicon) for 30 min in the dark at 4 °C. Cell cycle and apoptosis data were acquired by measuring DNA content using a Cytotax flow cytometer 26 (Beckman Coulter) in the PerCP-A channel. At least 20,000 fluorescent 'events' were collected for each sample. Data analysis was performed using CytExpert Software (Beckman Coulter).

Statistical evaluation

MS Excel and GraphPad Prism were used to analyze the data. Every experiment was performed at least three individual times to ensure reproducibility, calculate average values and obtain the SD values. ANOVA tests were estimated between groups.

Results and discussion

The antiproliferative activities of the obtained compounds were tested on MCF7 hormone-dependent breast cancer cells. The growth of MCF7 cells is supported by estrogens; thus, anti-estrogens, including some steroids, show high activity against this cancer [49, 50]. To assess the activity of the compounds, the MTT test was performed after 72 h of cell growth with the compounds. The IC₅₀ values are shown in Table 1.

The IC₅₀ values for pentacyclic steroids **3a–h** ranged from 7 to 25 µM. The compound with the methoxyphenyl substituent showed weak activity against MCF7 cells. The introduction of halogens into the side chain enhanced the antiproliferative effects of the obtained compounds. The highest activity was shown by a compound containing a fluorophenyl substituent. The activities of chloro-substituted pentacyclic steroids **2a–h** were similar. The highest activities were detected for compounds containing 4-chloro- and 4-fluorophenyl substituents at 6 and 7.1 µM, respectively. Interestingly, the introduction of a phenyl group with two halogen substituents led to a decrease in antiproliferative activity (compound **2f**). The series of benzylidene derivatives **1a–h** was highly active, as shown in Table 1. All benzylidenes demonstrated inhibitory effects with IC₅₀ values below 10 µM. The least active was benzylidene-bearing methoxyphenyl substituent **1g** with an IC₅₀ value of 6.7 µM. As in the case of pentacyclic steroids, benzylidene **1d** with the fluorophenyl substituent showed the greatest antiproliferative potency. These patterns may be associated with the geometry of benzylidene molecules. In contrast to pentacyclic steroids, the aryl residue in benzylidenes is not rigidly bound to the steroid residue but has a labile structure, which increases the affinity of the interaction with the receptor due to the lability of the molecular geometry.

Chloro-substituted compounds **3d**, **2d**, and **1d** from the common set of pentacyclic and benzylidene steroids, which showed high activity in the antiproliferative assay, were selected for in-depth analysis. Estrogens in the cell bind to estrogen receptor α (ESR1) [51]. Activation of ER α leads to changes in the expression of many genes, including proliferation regulators. Anticancer compounds may have activating or inhibitory effects on estrogen receptor α . Their activating abilities can be attributed to adverse effects. Using a reporter assay, we tested whether compounds **3d**, **2d**, and **1d** can activate the estrogen receptor in hormone-dependent MCF7 breast cancer cells. A steroid-free medium was used for this assay to minimize the effects of steroids and steroid-like compounds on the transcriptional activity of ER α . As shown in Fig. 1, pentacyclic steroid **3d** did not increase the activity of estrogen receptor α , while the natural ligand of the estrogen receptor 17 β -estradiol (E2) caused a

Fig. 1 ER α agonist activity in MCF7 cells. Luciferase reporter assay was performed after 18-h treatments with the compounds at different concentrations. E2–17 β -estradiol; * p <0.05 versus the control sample

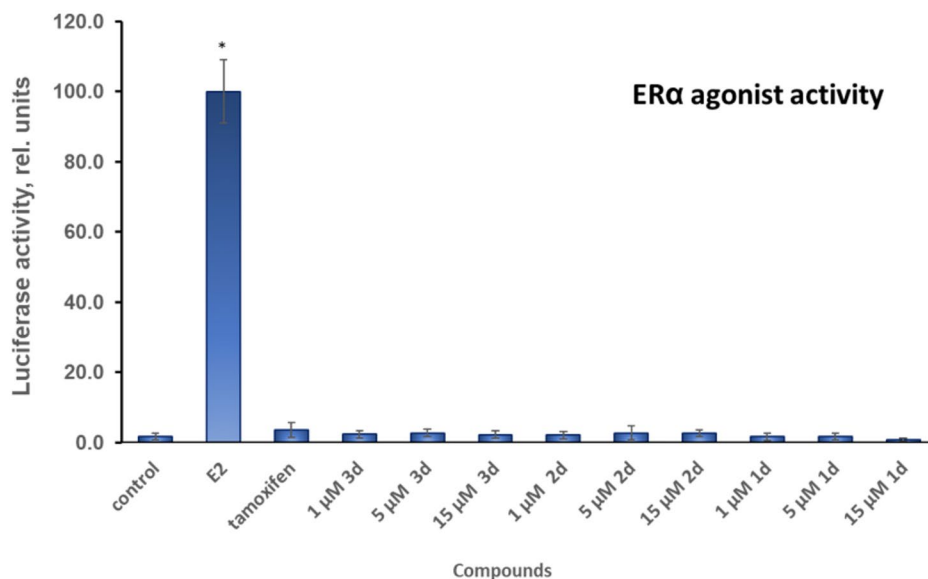
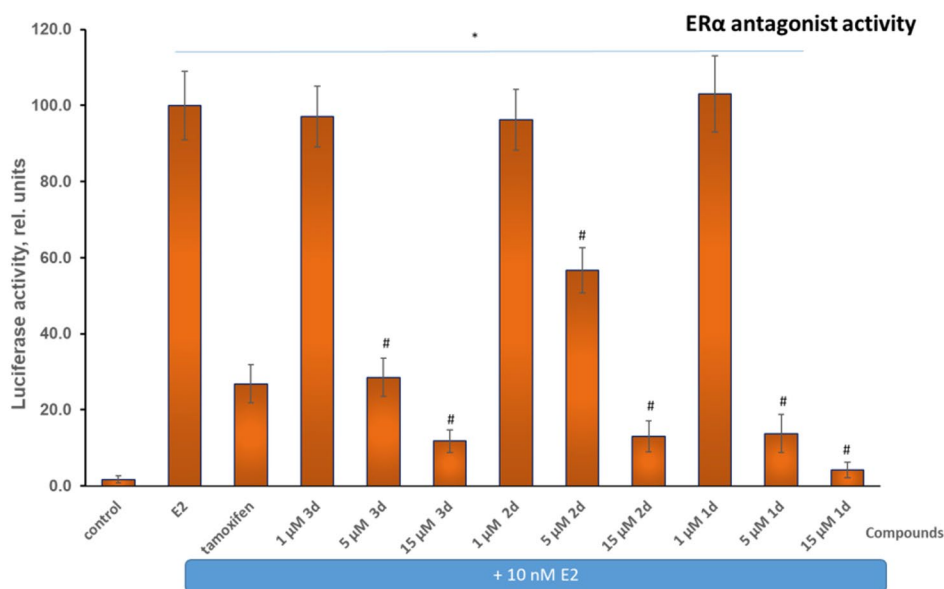


Fig. 2 ER α antagonist activity in MCF7 cells. Luciferase reporter assay were performed after 18-h treatments with the compounds at different concentrations (5 μ M tamoxifen was used as a reference drug; luciferase was induced by treatments with 10 nM E2). E2–17 β -estradiol; * p <0.05 versus the control sample; # p <0.05 versus MCF7 cells treated with 10 nM E2 alone



significant increase in ER α activity. Chloro-substituted pentacyclic steroid **2d** and benzylidene **1d** showed no agonistic activity. Additionally, the agonistic activity of the anti-estrogen tamoxifen, which in some conditions can significantly activate estrogen receptor α , was not observed [52].

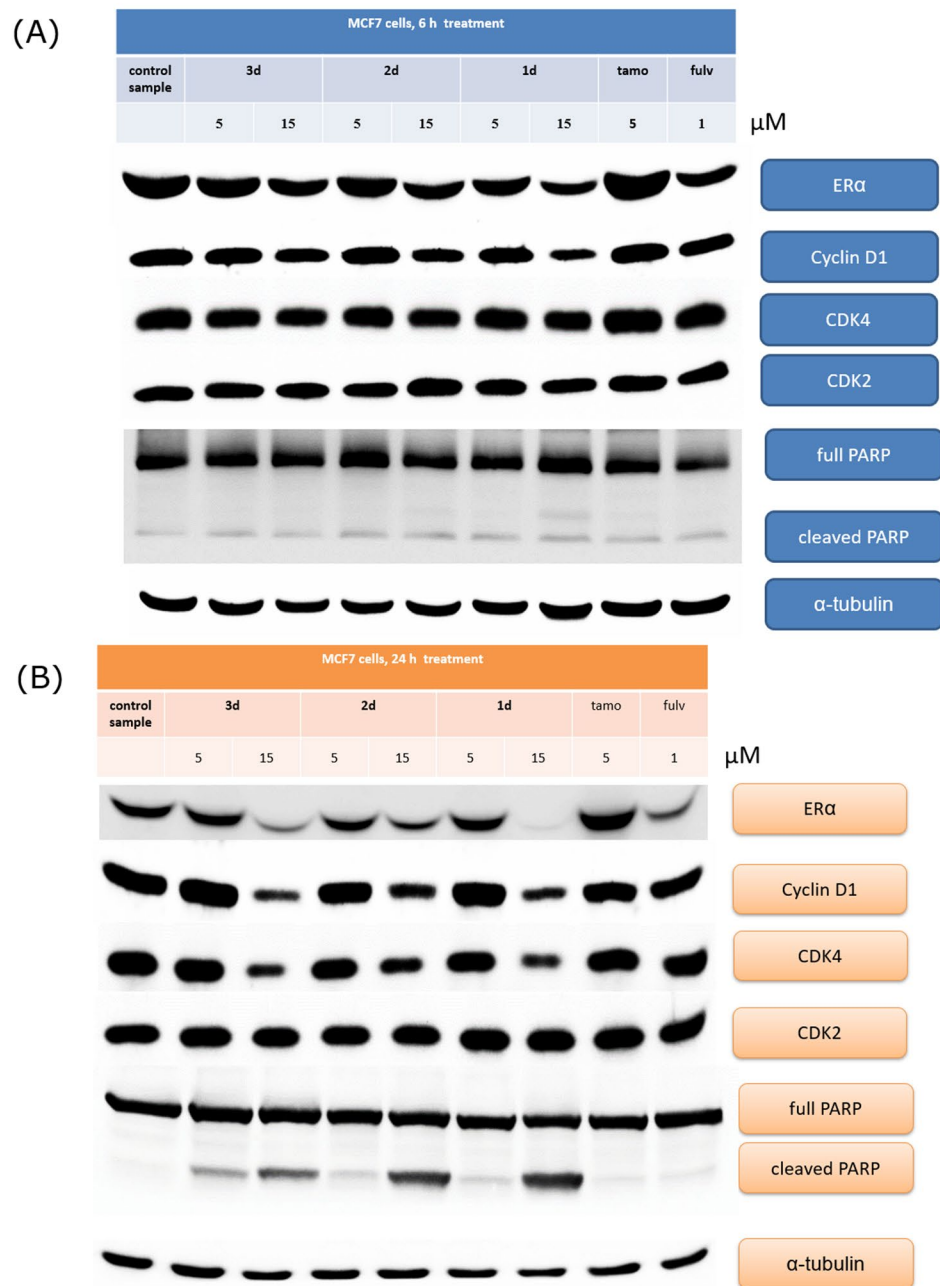
The inhibitory potencies of the compounds were evaluated after activation of the estrogen receptor with 17 β -estradiol (Fig. 2). 17 β -estradiol caused a significant increase in luciferase activity, which indicates a high activity of estrogen receptor α in MCF7 cells. Compounds **3d**, **2d**, and **1d** at 1 μ M did not affect the E2-induced activity of estrogen receptor α . Then, by increasing the concentration of compounds to 5 μ M, their anti-estrogenic properties were revealed. Compound **2d** at 5 μ M suppressed the activity of

estrogen receptor α by 43 relative units. Compound **3d** was more active, i.e., it blocked estrogen receptor α activity by 71 rel. units. Among the selected compounds, benzylidene **1d** showed the greatest activity at a concentration of 5 μ M, and estrogen receptor α retained only 14 units of activity. All three selected compounds at a concentration of 15 μ M significantly inhibited the activity of estrogen receptor α . The highest activity was observed for benzylidene **1d**.

Identification of signaling pathways was performed by immunoblotting. MCF7 cells were treated with the compounds for indicated periods, and then, protein expression was determined by immunoblotting (Fig. 3 A, B).

First, short-term 6-h incubation of MCF7 cells with the compounds was analyzed. In these experiments, no

Fig. 3 Identification of signaling pathways in MCF7 cells. MCF7 cells were treated with compounds or reference drugs and then subjected to immunoblotting; (A) 6-h and (B) 24-h treatment with compounds or reference drugs; tamo - tamoxifen, fulv - fulvestrant



significant changes in protein expression were shown (Fig. 3 A). With an increase in incubation duration to 24 h, compounds **3d** and **1d** caused a remarkable decrease in ERα expression at a concentration of 15 μM. Treatment with anti-estrogen fulvestrant also caused ERα degradation in MCF7 cells. Compound **1d** demonstrated the strongest ERα decrease. Compound **2d** triggered a slight decrease in ERα expression under the same conditions. Tamoxifen treatment resulted in a partial increase in ERα expression, which may be attributed to the stabilization of inactivated cytoplasmic ERα [53, 54]. Moreover, all three compounds led to

the cleavage of PARP (116 kDa) and the formation of an 89-kDa PARP fragment, which is a key marker of apoptotic cell death [55–57]. The effect grew with the increase of the compound concentration from 5 to 15 μM. Then, 89-kDa PARP was significantly increased in MCF7 cells treated with compound **1d** at a concentration of 15 μM. The anti-estrogens tamoxifen and fulvestrant did not cause apoptosis in MCF7 cells. Cyclin D1 was analyzed to be an important ERα-dependent gene and cell cycle regulator (Fig. 3B) [58, 59]. A decrease in ERα expression and PARP cleavage in MCF7 cells were accompanied by a slight decrease in cyclin

D1 expression at a concentration of 15 μM when fulvestrant treatment did not affect cyclin D1 expression. CDK2 and CDK4 are key regulators of the cell cycle. As shown in Fig. 3B, selected compounds down-regulated CDK4 expression, while CDK2 expression was unchanged.

The cell cycle of the MCF7 cells treated with the leader compounds was analyzed using flow cytometry after staining the cells with a DNA-binding dye propidium iodide (PI). MCF7 cells were incubated with compounds **3d**, **2d**, and **1d** for 24 h, and afterwards, their distribution in different phases of the cell cycle was assessed; MCF7 cells incubated with DMSO were assayed as a control sample (Fig. 4). Apoptotic cells were defined as a sub-G1 peak in the histograms [60, 61].

As shown in Fig. 4, the fraction of cells in the sub-G1 phase of the cell cycle increases in proportion to the increase in the concentrations of compounds **3d**, **2d**, and **1d**. Thus, the antiproliferative activity of the tested compounds is associated precisely with the death of tumor cells of the MCF7 cell line as detected by the cell fraction in the sub-G1 phase. These data correlate well with the apoptosis identified above (Fig. 3B). Interestingly, compound **3d** (15 μM) caused cell arrest in the G2/M phase. Several effective chemotherapy drugs and drug candidates cause cell arrest in this phase. These effects have been detected for the following agents: doxorubicin [62], curcumin [63], ribociclib in combination with cisplatin [64], cinobufagin [65], timosaponin AIII [66], apigenin [67], and others.

One of the key problems in oncology is resistance. A course of chemotherapy is usually prescribed after surgery. Many patients are initially sensitive to standard chemotherapy, but with time, cancer cells may develop resistance to the antiproliferative effect of the drugs. In such cases, it is impossible to continue treatment with the chosen drug because the tumor does not respond to this therapy. At this stage, the chemotherapist has to find other drugs that can inhibit tumor growth. Importantly, some drugs can contribute to the development of multidrug resistance. We were interested in whether the resistant cells would be sensitive to the obtained compounds. Cells resistant to several drugs were chosen for the experiments. Cell lines were obtained by prolonged cultivation with the appropriate drug. MCF7/HT, MCF7/DCT, MCF7/CP, and MCF7/RAP cells were characterized by resistance to hydroxytamoxifen, docetaxel, cisplatin, and rapamycin, respectively. Data on the antiproliferative activity of the compounds are shown in Table 2. Compound **1d** was less active against resistant lines than against the MCF7 line. Nevertheless, the IC_{50} value did not exceed 10 μM , indicating a sufficiently high activity of this steroid. Similar data were obtained for compound **2d**. The IC_{50} values for the resistant lines ranged from 11.1 to 17.8 μM . The most interesting data were obtained for compound

3d. This compound was active against MCF7/HT and the parent breast cancer cells (MCF7). The MCF7/DCT and MCF7/RAP breast cancer cells showed higher sensitivity to compound **3d**. The activity of compound **3d** toward MCF7/CP cells was slightly lower. Thus, the selected compounds are characterized by high activity against resistant cells, exhibiting IC_{50} values below 10 μM (**1d**, **3d**) and selectivity (**3d**).

Conclusion

A series of benzylidenes and pentacyclic derivatives of the progesterone series was obtained, and the antiproliferative effects of the compounds on hormone-dependent breast cancer cells were evaluated. The synthesized benzylidenes demonstrated inhibitory effects with IC_{50} values below 10 μM , and the activities of the benzylidenes were higher than the activities of their pentacyclic analogues. The activities of benzylidene derivatives compared to pentacyclic steroids are likely associated with the lability of the geometry of their molecules, leading to an increase in the affinity of interaction with the receptor.

Estrogen receptor α was studied as a potential target for the selected compounds. No agonistic activity of the compounds on estrogen receptor α was detected in MCF7 cells. The studied compounds showed significant anti-estrogenic properties, with benzylidene **1d** being the most active. In addition to potent anti-estrogenic effects, compounds **3d**, **2d**, and **1d** induced apoptosis in MCF7 cells as confirmed by flow cytometry analysis and PARP cleavage. Compound **3d** caused cell arrest in the G2/M phase. Steroids **1d** and **3d** exhibited inhibitory potency toward breast cancer cells with acquired resistance to various anticancer drugs, which makes them important for targeted drug design. Further study of the obtained compounds is of great interest to medicinal chemistry and cancer research.

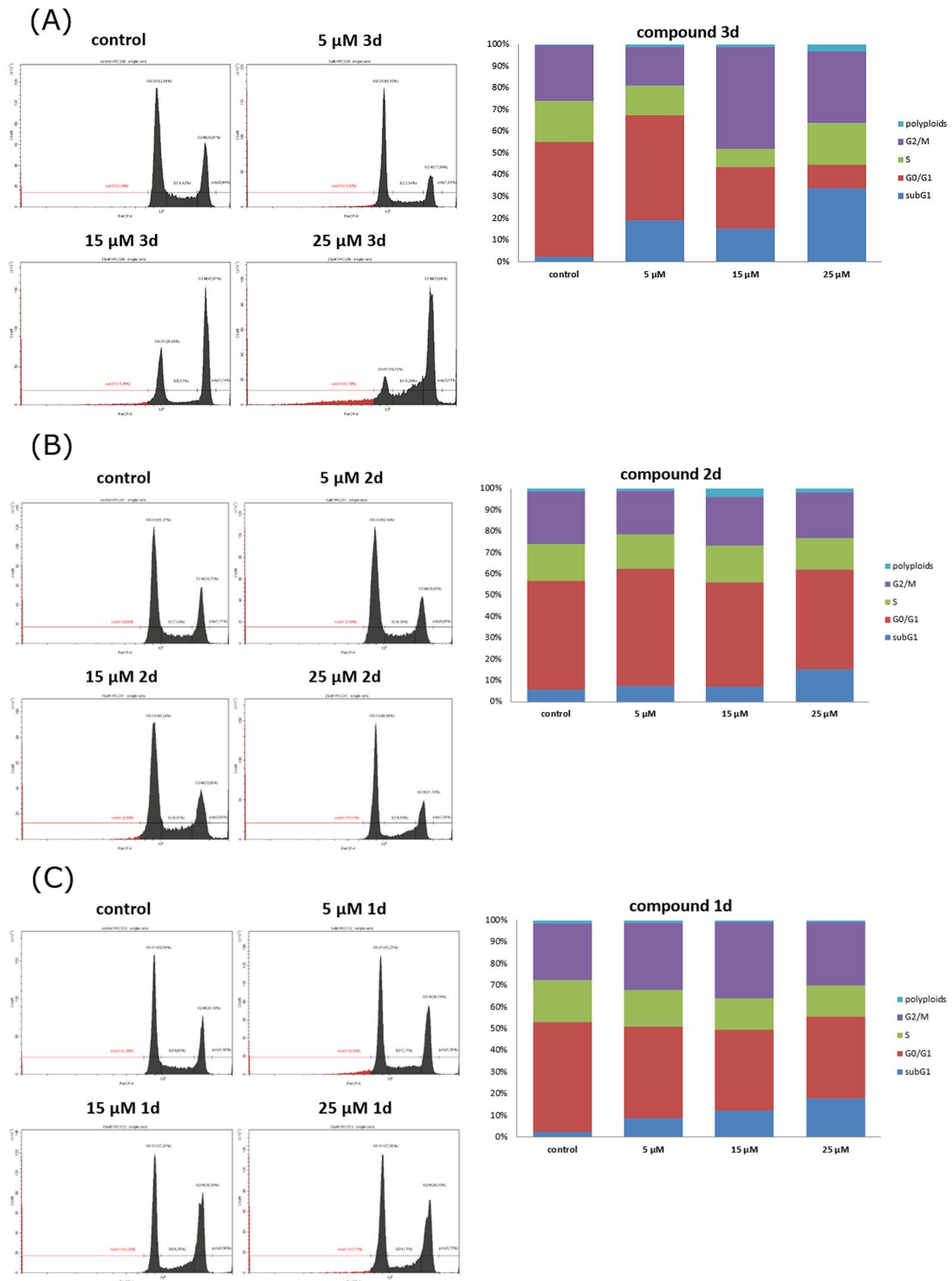


Fig. 4 Graphs and histograms showing the proportion of MCF7 cells in each phase of the cell cycle after incubation with the leader compounds for 24 h; (A) – 3d, (B) – 2d, (C) – 1d

Table 2 Antiproliferative activity of the leader compounds against resistant breast cancer cells

Compound	IC ₅₀ values, μ M				
	MCF7	MCF7/ HT	MCF7/ DCT	MCF7/ CP	MCF7/ RAP
1d	1.9±0.2	4.2±0.5	3.9±0.5	5.4±0.6	3.1±0.4
2d	7.1±0.9	17.7±1.2	11.1±1.0	17.8±1.4	12.0±1.0
3d	7.3±0.8	7.0±0.8*	3.1±0.3*	13.0±1.5	5.2±0.6*

* The activity against resistant cell lines was higher or the same as determined for the MCF7 line

HT – hydroxytamoxifen, DCT- docetaxel, CP- cisplatin, RAP - rapamycin

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10637-023-01332-z>.

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Data availability Synthesis and structural characterization of pentacyclic steroids are described in Appendix A. Any other information may be requested directly from the authors (Alex.Scherbakov@gmail.com).

Declarations

Conflict of interest The authors have no relevant financial or non-financial interests to disclose.

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