



Reproductive biology and population structure of the endangered shrub *Grevillea bedgoodiana* (Proteaceae)

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Abstract

Narrowly endemic species are particularly vulnerable to catastrophic events. Compared to widespread species, they may also be less capable of adapting to shifts in environmental pressures as a result of specialisation on a narrow range of local condition and limited ability to disperse. However, life-history traits, such as preferential outcrossing and high fecundity can maintain genetic diversity and evolutionary potential, and boost species resilience. The endangered *Grevillea bedgoodiana* (Enfield Grevillea) is an understorey shrub restricted to an area of ca. 150 km² in south-eastern Australia with a legacy of large-scale anthropogenic disturbance. Prior to this study little was known about its biology and population structure. Here, its breeding system was assessed through a controlled pollination experiment at one of its central populations, and eight populations were sampled for genetic analysis with microsatellite markers. The species was found to be preferentially outcrossing, with no evidence of pollination limitation. In most populations, allelic richness, observed heterozygosity and gene diversity were high (A_r : 3.8–6.3; H_o : 0.45–0.65, H_e : 0.60–0.75). However, the inbreeding coefficients were significant in at least four populations, ranging from F_i -0.061 to 0.259 despite high outcrossing rates. Estimated reproductive rates varied among sampled populations but were independent of gene diversity and inbreeding. Despite its small geographic range, the species' populations showed moderate differentiation (AMOVA: F_{ST} = 0.123), which was largely attributable to isolation by distance. We interpret these results as suggesting that *G. bedgoodiana* is reproductively healthy and has maintained high levels of genetic diversity despite recent disturbance.

Keywords Endangered species · Endemism · Genetic diversity · Microsatellites · Plant breeding system · Population structure

Introduction

Effective conservation of threatened species requires a thorough understanding of multiple interacting aspects of their biology (Aitken and Whitlock 2013; Allendorf et al. 2013; Scheele et al. 2018). The 'endangered' status is afforded to species that are at high risk of extinction, with one of the criteria for listing being acute restriction of the species'

geographic range. The rationale for this is that most threatening processes are spatially correlated, meaning that if all individuals of a species occur in a one small area, a single catastrophic event can impact its entire population (IUCN 2012). Moreover, compared to widespread congeners, geographically restricted species tend to have narrower niche breadth (Gaston et al. 1997; Slatyer et al. 2013) and be less genetically diverse (Gitzendanner and Soltis 2000; Cole 2003). When faced with novel challenges, such species may be unable to draw on standing genetic diversity to evolve and adapt (Aitken et al. 2008; Anderson et al. 2012). This is a precarious situation because narrow endemics tend to be poor dispersers (either due to lack of traits facilitating dispersal and/or because of being confined to a habitat 'island') (Gaston 1996), and so are unlikely to be able to track the conditions of their niche. However, the evaluation of extinction risk relating to the species' extent of occurrence is strongly context dependent. Different species with similar

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ranges may be vulnerable to different threats depending on their life history traits such as lifespan (Thrall et al. 2014), breeding (Willi et al. 2005) and mating systems (Yates et al. 2007; Nora et al. 2016; Ritchie et al. 2019), reproductive rates (Rymer et al. 2005) and seedbank dynamics (Kenny 2000; Pickup et al. 2003), as well as genetic diversity (Frankham 2015) and population size (Leimu et al. 2006; Caballero et al. 2017). Moreover, to add to the complexity of the task, all of the above should be considered in the context of the species' landscape, disturbance history and interspecific interactions, particularly, pollination ecology (Cranmer et al. 2012; Llorens et al. 2013).

In order to persist and evolve, outcrossing plant species need to maintain genetic diversity and produce enough viable offspring to replace the adults. At the level of local populations the plants' ability to do so depends on mate availability, but, in the case of animal-pollinated species, also on the local abundance and effectiveness of pollinators (Richardson et al. 2000; Devaux et al. 2014; Barrett and Harder 2017). Scarcity of effective pollinators may cause reduced levels of ovule fertilisation (pollination limitation) and reduced rates of outcrossing. This can lead to reduced seed set and/or increased inbreeding, potentially leading to low reproductive rates and low offspring fitness (inbreeding depression) (Copland and Whelan 1989; Wilcock and Neiland 2002; Coates et al. 2007; Bezemer et al. 2019). Moreover, for species without adaptations for long-distance seed dispersal, the maximum foraging range of the pollinators is expected to limit gene flow between geographically distinct populations (England et al. 2001). Low levels of gene flow can make small populations vulnerable to loss of genetic diversity through drift (Frankham 2015) and reduce the likelihood of the spread of adaptive traits (Lowe and Allendorf 2010; Aitken and Whitlock 2013; Ottewill et al. 2016). The knowledge of reproductive health and levels of inter-population gene flow is, therefore, crucial for evaluating resilience of a threatened species and deciding on appropriate conservation action. However, this knowledge cannot be gained solely by measuring seed set or by inference from population genetic parameters such as inbreeding coefficients and heterozygosity. This is particularly true for long-lived plants which may be reproducing sporadically but may employ reproductive strategies such as staggered germination of soil-stored seedbanks which may buffer local populations against loss of genetic diversity. Consequently, assessments of the reproductive health of plant populations should be based on estimates of actual reproductive rates followed by tests of whether they are correlated with measures of heterozygosity and inbreeding (Szulkin et al. 2010; Breed et al. 2015). This is relevant for two reasons. First, because the number of seeds produced by the adult plants in a population is a poor indicator of their reproductive potential as not all seeds will evade granivores, germinate and establish.

For example, in Proteaceae, most of the seeds produced are commonly lost and do not contribute to the next generation (Lamont and Groom 1998; Auld and Denham 1999). Second, while there is evidence that low fitness is associated with low population genetic diversity and high inbreeding, the reverse is not always the case (Reed and Frankham 2003; Frankham 2015).

Grevillea is a large and diverse genus of woody shrubs and trees in the Proteaceae (Makinson 2000). A wide array of reproductive strategies are known in grevilleas from full self-compatibility (Richardson et al. 2000; England et al. 2001; Gross et al. 2012) to obligate outcrossing (Hoebee and Young 2001; Caddy and Gross 2006; Holmes et al. 2008, 2009), and at least two species, *G. infecunda* (James et al. 2017) and *G. renwickiana* (James and McDougall 2014), are thought to be sterile and reproduce exclusively by root-suckering. The pollination strategies are also varied in this genus with native insect, bird (usually of the Meliphagidae), and mammal pollinators involved (Olde and Marriott 1994). The introduced honeybee, *Apis mellifera*, has become a common floral visitor to many grevilleas, leading to concerns over their potential to disrupt pollination and mating systems of native species (Vaughton 1996; Paton 2000; Smith and Gross 2002; Celebrezze and Paton 2004; Whelan et al. 2009). Pollination disruption is of particular concern for the primarily bird-pollinated forest understory grevilleas of south-eastern Australia, owing to habitat fragmentation and, in recent decades, the decline of forest birds (Ford et al. 2001; Ford 2011).

Since the early 2000s, several studies have assessed the genetic diversity and population structure of grevilleas (Hoebee and Young 2001; England et al. 2002; Llorens 2004; Nistelberger et al. 2015), including some species of the *Grevillea aquifolium* group (the 'holly-leaved grevilleas') from south-eastern Australia (Holmes et al. 2008, 2009; James and McDougall 2014; James et al. 2017). Several species in this group appear to be reproductively constrained as a result of pollinator and/or mate limitation (Holmes et al. 2008), variation in ploidy (Holmes et al. 2009) and sterility (Kimpton et al. 2002; James et al. 2017). This study focused on a narrowly endemic holly-leaved grevillea, *G. bedgoodiana* J.H. Willis ex McGill. This species is restricted to a small area of Palaeozoic sedimentary soils in central Victoria surrounded by soils derived from Quaternary volcanic activity (Joyce 2004). There are no historical collections of this species outside of its current range (Australasian Virtual Herbarium, accessed 08 June 2022) suggesting that the newer igneous soils are unsuitable for its growth. Currently, all Victorian grevilleas are protected under the state's Flora and Fauna Guarantee Act (1988), and the largest populations of *G. bedgoodiana* are managed for conservation within Enfield State Park.

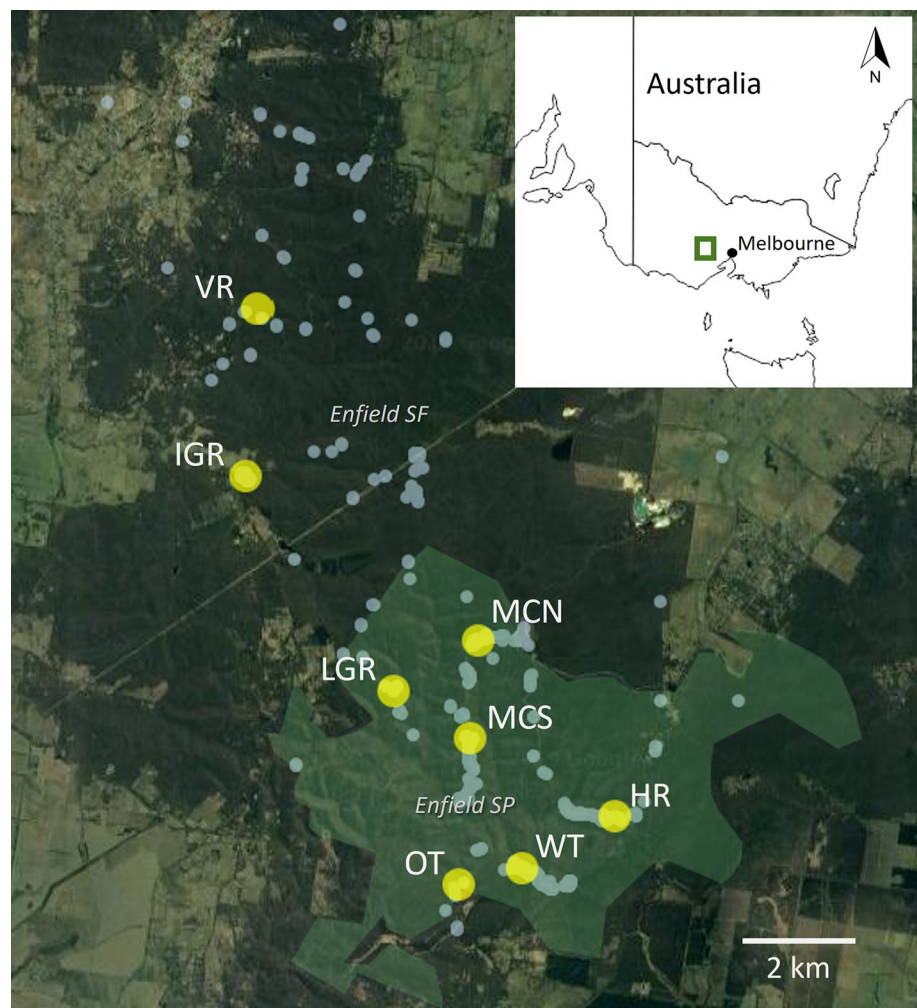
However, in the recent past (ca. 160 years), much of its habitat has been impacted by mining and logging while the surrounding landscape of fertile basaltic plains have been largely cleared for agriculture. This relatively recent disturbance likely caused population bottlenecks and disrupted pollination, potentially leaving a legacy of reduced genetic diversity which could affect the species' reproduction and long-term evolutionary potential. *Grevillea bedgoodiana* is listed as 'endangered' (Flora and Fauna Guarantee Act, 1988) but prior to this study it had not been a focus for research. Consequently, we sought to establish some of the fundamental aspects of its reproductive biology and population genetics, specifically, (1) to determine the species' breeding system through a controlled pollination experiment; (2) to assess the genetic diversity and structure of its populations through microsatellite-based analyses, and (3) to test for bottlenecks and reduction of reproductive rates in populations with lower-than-average genetic diversity.

Materials and methods

Site and study species

Grevillea bedgoodiana is endemic to an area of approximately 150 km² comprised of Enfield State Forest (ESF) and Enfield State Park (ESP) (here, collectively referred to as Enfield Forest) in Victoria, Australia (Fig. 1). Since the 1850s areas of Enfield Forest were disturbed by gold mining (until the 1930s) and logging (until the 1960s) (Parks Victoria 1998). During the last 70 years, fires have occurred in Enfield Forest every 15–20 years. In 1995, a large wild-fire burnt through about 90% of the Forest (Parks Victoria 1998). Subsequently, fuel reduction burns were conducted in some parts of the forest, particularly near major roads and infrastructure (Irena Cassettari, Department of Environment, Land, Water and Planning, pers. comm.). The consequences of these multiple disturbances for the local flora and fauna assemblages are unknown. However, they likely contributed to the decline of the native vertebrate pollinators, (Ford et al.

Fig. 1 The recorded occurrences of *Grevillea bedgoodiana* (small grey dots) (Atlas of Living Australia, accessed Jan. 2020) and location of populations sampled in the current study (larger yellow dots). The six populations in the south are located within Enfield State Park (highlighted polygon). VR and IGR are in Enfield State Forest (map source: Google Earth)



2001; Recher et al. 2009) and may have led to bottlenecks and disruption of connectivity among plant populations (Young et al. 1996).

Grevillea bedgoodiana is an understory shrub of eucalypt-dominated open forest, growing on gravelly clay soils, predominantly on ridges and north-facing slopes in the southern and western parts of Enfield Forest. It tends to form discrete stands of several hundred individuals (Carter et al. 2006), with scattered individuals occurring occasionally particularly on roadsides (SW, pers. obs.). In the context of this study, the term ‘population’ is applied following a standard definition of a spatially discrete group of organisms where interbreeding is expected to occur primarily within the groups but not excluding the possibility of gene flow between groups. This is consistent with the National Recovery Plan for *Grevillea bedgoodiana* (Carter et al. 2006) which defined important populations of this species as those of over 1000 individuals (Carter et al. 2006). There are 11 such populations, containing an estimated total of 20,000 individuals (ca. 50% of all known plants), and all of them are within ESP (which has been established specifically for their protection) (Carter et al. 2006) (Fig. 1). The populations in ESF are considerably smaller and tend to be separated by greater distances (SW, pers. obs.). Consequently, this study focused primarily on the large populations in ESP (Table 1).

Grevillea bedgoodiana is a low to prostrate shrub, typically to 0.5 m tall with trailing lateral branches up to 1.9 m long (Fig. 2a), sometimes more upright. It flowers in spring from late October through November. Its confluences are secund (toothbrush-like) (Makinson 2000) and consist of 10–20 pairs of flowers (Fig. 2b) which open sequentially over 5–7 days (SW, pers. obs.). The individual zygomorphic flowers are protandrous. During the male phase of a flower, pollen is transferred to, and presented at the tip of a 12–16.5 mm-long style. Removal of pollen by visiting animals exposes the stigma, which becomes viscous and receptive to pollen ca. 2–3 days after the beginning of anthesis (SW, pers. obs.).

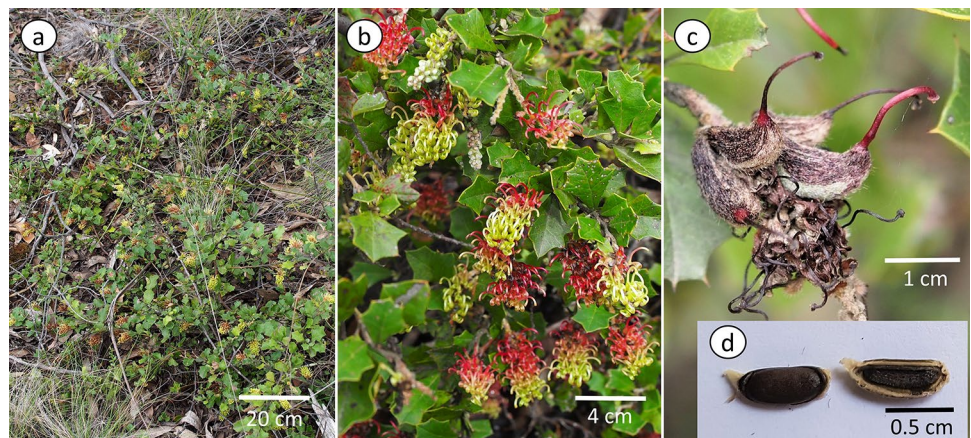
Animal visitors to *G. bedgoodiana* confluences were surveyed through direct observation and camera trapping in a concurrent study (Aristidou and Hoebee, unpublished data). The vertebrate visitors included one bird: the yellow-faced honeyeater (*Lichenostomus chrysops*) and several mammal species: antechinus (*Antechinus* sp.), eastern pygmy possum (*Cercartetus nanus*) and common brushtail possums (*Trichosurus vulpecula*). The invertebrate visitors of this species include native bees, wasps, and hoverflies (Emily Noble, Field Naturalists Club of Ballarat, pers. comm.). However, the most common day-time floral visitor at the sites during the study was the introduced European honeybee (*Apis*

Table 1 Information for populations of *Grevillea bedgoodiana* sampled in this study. ESP = Enfield State Park; ESF = Enfield State Forest, N_{est} = estimated adult population size (see main text)

Pop code	Nearby road/location	Latitude	Longitude	N_{est}	Herbarium voucher
OT ^a	Orchid Tk/ESP	– 37.780261	143.737549	1400	LTB07509
WT	Wattle Tk/ESP	– 37.7802116	143.7407774	4800	LTB07508
HR	Halls Rd/ESP	– 37.7691111	143.773387	2100	LTB07507
MCS	Misery Ck Rd South/ESP	– 37.7562726	143.7440631	3000	LTB07510
MCN	Misery Ck Rd North/ESP	– 37.739085	143.745302	1700	LTB07512
LGR	Long Gully Rd/ESP	– 37.7477043	143.7258619	1300	LTB07511
IGR ^a	Italian Gully Rd/ESF	– 37.712500	143.695783	400	LTB07673
VR	Victoria Rd/ESF	– 37.684450	143.698267	300	LTB07674

^aPopulations on sites with evidence of disturbance by gold mining

Fig. 2 *Grevillea bedgoodiana*, showing its **a** typical prostrate growth habit; **b** concolorous confluences with lime-green unopened flowers gradually turning red after the beginning of anthesis; **c** maturing follicles and **d** mature seeds with bordering elaiosomes



mellifera) (Aristidou and Hoebee, unpublished data). Selective exclusion of vertebrate visitors from flowers did not reduce fruit set relative to open pollination (Aristidou and Hoebee, unpublished data), suggesting vertebrates may play a minor role in contemporary pollination of *G. bedgoodiana*. The woody fruits (follicles) of *G. bedgoodiana* (Fig. 2c) contain up to two seeds, which are approximately 6 mm long when mature (Fig. 2d). The species is non-serotinous, and its seeds are thought to be dispersed primarily by gravity (Makinson 2000). However, the presence of an elaiosome (Fig. 2d) suggests that myrmecochory may contribute to secondary seed dispersal over short distances (Auld 1995; Edwards and Whelan 1995; Auld and Denham 1999). It is not known whether the seeds can germinate without fire/smoke (Carter et al. 2006) and how long they can persist in the soil. Field observations suggest that wild plants grown from seed are able to flower and set seed within five years (SW, pers. obs.). Older plants form a lignotuber and may be able to resprout after fire, but do not appear to reproduce clonally by root suckering (SW, pers. obs.).

Breeding system

A controlled pollination experiment was conducted at MCS (Fig. 1) between 25th of October and 4th of November 2018. Ten healthy adult plants with unopened flowers and growing at least 10 m apart were selected. Four pollination treatments were applied to each plant: autogamy (spontaneous selfing), geitonogamy (animal vector-mediated self-pollination), xenogamy (cross-pollination), and open pollination (control). The confluences in the autogamy treatment (two per plant) were bagged in bud using fine mesh organza bags to prevent visitation by pollinators and otherwise not manipulated. Those in the geitonogamy (one per plant) and xenogamy (two per plant) treatments were bagged in bud, had self-pollen removed shortly after anthesis and were hand-pollinated using either pollen from other flowers on the same plant (geitonogamy), or pollen from two different plants growing more than 10 m away (xenogamy). The open pollination confluences (three per plant) were tagged but otherwise not manipulated, then bagged several days after the last flowers had opened. The unbalanced design of this experiment was dictated by the amount of pollen that could be collected for geitonogamous pollination without affecting other treatments, while the use of three confluences in the open pollination treatment aimed to provide a more accurate estimate of natural levels of seed set. The developing confluences remained bagged until collection on 16th of December 2018. At that time, the follicles were well-developed but not yet open. They were counted, stored at room temperature for four days and then opened manually to assess seed set. The numbers of filled seeds were compared

between pollination treatments using paired *t*-tests in base R (R Core Team 2021).

Sampling, DNA extraction and genotyping

Eight representative populations were selected for the population genetic study: six in ESP and two in ESF (Table 1, Fig. 1). Fresh leaf tissue was collected from 30 adult plants at each of the eight sites (240 in total) – selecting plants growing at least 5 m apart and avoiding nearest neighbours to reduce the chance of sampling potential clones (Holmes et al. 2009) or closely related or individuals (Krauss et al. 2009). Sampled leaves were placed in individual paper envelopes and desiccated using silica gel beads (ChemSupply, Adelaide, Australia). The GPS coordinates of each plant sampled were recorded and representative voucher specimens for each population were collected and deposited at LTB with duplicates sent to MEL.

For each sample, 20 mg of dried leaf tissue was disrupted using a Tissue Lyser II (Qiagen, Hilden, Germany). DNA was then extracted using a DNeasy Plant Mini Kit (Qiagen, Hilden, Germany) following the manufacturer's protocol. The concentration of the DNA isolates was measured using a NanoDrop Lite (Thermo Fisher Scientific, Waltham, MA, USA). All samples were stored at $-20\text{ }^{\circ}\text{C}$ until required.

The plants were genotyped using the three primer method implementing four fluorescently labelled universal primers (Schuelke 2000; Blacket et al. 2012) and fourteen sets of primers designed to amplify microsatellite loci in two closely related species (*Grevillea aquifolium* and *G. infecunda*; James et al. 2012). DNA was amplified in two multiplex reactions, each consisting of seven primer pairs. Sequence tails of the universal primers were added to the forward primers, such that each multiplex PCR amplified up to two loci labelled with the same fluorescent primer avoiding size overlap of the expected amplicons (James et al. 2012) (Table S1). Multiple Primer Analyzer (Thermo Fisher Scientific, Waltham, MA, USA) was used to predict potential primer dimerization. A third multiplex reaction was designed to most efficiently re-amplify samples which initially failed to amplify or produced ambiguous amplicon peaks for some loci (Table S1). A Type-it Microsatellite Kit (Qiagen, Germany) was used to amplify the loci in 15 μL reactions containing 10–20 ng of template DNA, 0.1 μM of each of the tailed forward primers, 0.2 μM of each of the reverse primers and 0.2 μM of the FAM, VIC, NED and PET fluorescent labelled primers. Polymerase chain reactions (PCR) were performed using BioRad MyCycler™ Thermal Cycler (BioRad, Hercules) with a denaturation step at 95 $^{\circ}\text{C}$ for 5 min, followed by 30 cycles of 95 $^{\circ}\text{C}$ for 30 s, 57 $^{\circ}\text{C}$ for 90 s, and 72 $^{\circ}\text{C}$ for 30 s, followed by a final extension step at 60 $^{\circ}\text{C}$ for 30 min. Resultant amplicons were processed by the Australian Genome Research Facility (AGRF,

Melbourne) using LIZ500 as an internal standard. Geneious Prime v.2019.0 (Biomatters Ltd, Auckland, New Zealand) was used in-house for manual genotype determination.

Population genetics analyses

Linkage disequilibrium between loci and the frequencies of null alleles were estimated using Genepop (Rousset 2008; R Core Team 2013). Basic population genetic parameters, including number of alleles per locus (N_a), number of private alleles (A_{priv}), observed heterozygosity (H_o), gene diversity (expected heterozygosity, H_e) were calculated using GenAlEx 6.5 (Peakall and Smouse 2006, 2012). Allelic richness (A_r , number of alleles per locus with rarefaction to the smallest sample size) were calculated using SPAGeDi (Hardy and Vekemans 2002). SPAGeDi was also used to estimate the individual inbreeding coefficients (F_i , with a test of significance based on 999 permutations) and population outcrossing rates ($1-s$) based on standardized identity disequilibrium using the kinship coefficient (Loiselle et al. 1995). The 95% confidence intervals for A_r , H_o , H_e and $1-s$ were approximated by multiplying the standard errors of the estimates by ± 1.96 . Bottleneck 1.2.02 (Piry et al. 1999) was used to test for excess of heterozygotes (evidence of recent population bottlenecks) employing the Wilcoxon signed rank test (owing to the low number of loci) with sequential Bonferroni correction (Holm 1979) and the infinite allele model (IAM) of marker evolution. Other models (stepwise mutation and two-phase) could not be used because our data included irregular amplicons (presumably resulting from mutations in the microsatellite flanking region England et al. 2002; Holmes et al. 2009; Putman and Carbone 2014)).

Population differentiation was assessed through F_{ST} -based analysis of molecular variance (AMOVA) (Excoffier et al. 1992) as implemented in GenAlEx. Resulting values of F_{ST} from this approach are equivalent to θ of Weir and Cockerham (1984) (Holsinger and Weir 2009). Isolation by distance was assessed by a Mantel test (GenAlEx) plotting pairwise F_{ST} against untransformed geographic (Euclidean) distance, repeating the analyses for standardised $F_{\text{ST}}^* = F_{\text{ST}} / (1 - F_{\text{ST}})$ (Rousset 1997; Séré et al. 2017). The genetic structure of the sampled populations was also assessed using Structure 2.3.4 (Pritchard et al. 2000, 2009) testing K values of 1–8 (1,000,000 MCMC steps, with 100,000 dememorization cycles, replicated ten times for each K value). Analyses were run under the admixture model of ancestry with sampling sites as priors and the correlated allele frequencies model. Structure Harvester (<http://taylor0.biology.ucla.edu/structureHarvester/>) was used to select the model that best explained the data using the Evanno (Delta K) method (Earl and VonHoldt 2012). Discriminant analysis of principal

components (DAPC, adegenet) was used as an alternative method to assess the clustering of the populations according to individual plants' genotypic similarity (Jombart, 2008; R Core Team, 2013).

Regression analysis: reproductive rates vs gene diversity and inbreeding

Reproductive rates were estimated for each of the eight populations by the number of juveniles (i.e., plants with flexible stems, fewer than about 40 leaves and not flowering) per 100 adults (N_j) in two 2×50 m belt transects. The transects were selected such that they passed through areas of typical plant density at each site (avoiding roadsides and population edges). The counts of adult plants in the transects were also used to estimate the total number of adult plants at the sampled populations (N_{est}). This was done by extrapolating the number of plants in the transects to the total area occupied by each population, where the area was estimated using QGIS v3.4.8 (QGIS Development Team 2017) as half that of the minimum convex polygons enclosing the plants sampled for the genetic analyses. Linear regression (R Core Team, 2013) was used to test for correlation between N_j and gene diversity (H_e) and inbreeding coefficients (F_i) of the adult populations. Because reproductive rates are potentially related to time since fire, detailed fire history data was obtained for Enfield Forest area from DataShare (<https://datashare.maps.vic.gov.au/search>; 'Fire history overlay of most recent fires'). The study sites were overlaid with the fire history data using QGIS v3.16.1, and linear regression used to test for correlation between population N_j values and time since last fire. The R package 'gvlma' (Peña and Slate 2006) was used to validate the regression models.

Results

Breeding system

Across the 80 experimental conflorescences on ten plants, 75 follicles were initiated. Of those, 57 had fully matured (although not all contained viable seeds), and 18 were aborted. The aborted follicles included the only follicle produced through autogamy, and three out of six of the follicles produced through geitonogamy. Consequently, no seeds were produced through autogamous pollination. Geitonogamy yielded a total of three seeds, mean = 0.30 ± 0.21 (SE) (median 0) seeds per conflorescence, which was significantly less than in the xenogamy treatment, mean = 1.55 ± 0.36 (SE) (median 1.5) (paired t-test, $n = 10$, mean of differences = -1.25 , $t = -2.521$, $df = 9$, $p = 0.033$) (Fig. 3). It was also less than in the open pollination treatment, mean = 1.33 ± 0.36

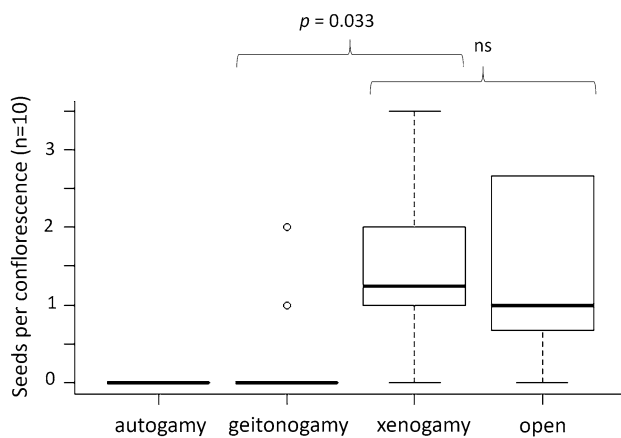


Fig. 3 Comparison of seed set from different pollination treatments indicating significance of the paired t-tests for the difference in seed set between pollination treatments ($n=10$ plants). The number of within-plant replicates (conflorescences per plant) varied between treatments: geitonogamous, $n=1$; autogamous and xenogamous, $n=2$; open pollinated, $n=3$

(median 1) seeds per conflorescence. However, due to high variance in the data, the difference was only significant at $\alpha=0.06$ (paired t-test, $n=10$, mean of differences -1.033 , $t=-2.151$, $df=9$, $p=0.060$). Xenogamy produced slightly more seeds per conflorescence than open pollination, but the difference was not significant (paired t-test, $n=10$, mean of differences 0.217 , $t=0.548$, $df=9$, $p=0.597$) (Fig. 3).

Microsatellite data

All of the 14 primer pairs used amplified *G. bedgoodiana* DNA. However, five loci (AQ11, AQ14, AQ34, IN32, IN41) were excluded from analyses due to missing or ambiguous PCR products for more than 75% of individuals in any population. The remaining nine loci were successfully amplified in at least a third of the samples (10 individuals) in all populations. Evidence of linkage was detected in one out of 36 loci pairs (IN18-IN34) when testing across all populations (Chi-sq > 49.6 , $df=16$, $p<0.001$, which remained significant after Bonferroni correction for 36 simultaneous tests, lowest adjusted $\alpha=0.00139$, all other p-values > 0.05). However, per population tests showed significant linkage between these two loci in only two of the eight populations (LGR and VR) and they were retained in the data set.

The frequencies of null alleles across all loci and populations were low: mean $= 0.10 \pm 0.02$ SE (median 0.06). However, locus-by-locus analysis showed high frequencies of null alleles in two out of nine loci, IN12: mean $= 0.21 \pm 0.06$ SE (median 0.18), and IN17: mean $= 0.25 \pm 0.07$ SE (median 0.17). Excluding these loci from the data had negligible effect on estimates of allelic richness and expected heterozygosity (H_e) but biased upward the estimates of observed

heterozygosity (H_o) in seven out of eight populations (mean difference: $+0.06$) and considerably lowered the mean population fixation indices in all populations, except OT (F_i , mean difference: -0.0875). It also lowered the estimates of global F_{ST} by 0.03 and increased by 10% the coefficient of determination (R^2) in the Mantel test of isolation by distance, but it had negligible effect on the topography of the inferred population structure (i.e., clustering of the populations). In the interest of retaining the information provided by these loci, both were retained in the data set, and two alternative estimates of F were reported (with and without IN12 and IN17).

Genetic diversity

Across the nine loci, a total of 107 alleles were identified (7–25 alleles per locus). The allelic diversity was distributed unevenly among the populations, with 1–6 private alleles per population and mean allelic richness (A_r) ranging from 3.8 to 6.3, mean 5.25 (after rarefaction to $n=10$) (Table 2). The population estimates of observed and expected heterozygosity (H_o and H_e) also varied considerably (from 0.45 to 0.63 and from 0.60 to 0.75 respectively). The inbreeding coefficient (F_i) ranged from -0.061 to 0.259 (0.149–0.260, when estimated excluding loci IN12 and IN17) and were significantly greater than 0 for all sampled populations except VR, when estimated with all 9 loci, and for four out of eight populations when estimated excluding IN12 and IN17 (Table 3). The estimates of outcrossing rates (1-s) were high for all populations, ranging from 0.90 to 1.0 (Table 2). Bottleneck analysis with sequential Bonferroni correction indicated significant excess of heterozygotes only at IGR (two-tailed Wilcoxon's test, $p=0.002$ compared to $\alpha=0.006$).

Population structure

AMOVA indicated that the populations were moderately differentiated with 12% of the genotypic variation due to among-population differences ($F_{ST}=0.123$, $p=0.001$). Pairwise F_{ST} values ranged from 0.012 to 0.286 (all p-values < 0.01) and were consistently highest for all pairs involving the small and peripheral VR population (Table 3). The Mantel test indicated moderate and statistically significant correlation between genetic differentiation and geographic distances between sampled populations ($R^2=0.49$, $p=0.01$) (Fig. 4); a similar correlation was found for standardised F_{ST}^* .

Structure Harvester's delta K plot indicated that the models with three and five clusters explained the data nearly equally well (Fig. 5a). In the K=3 model, the small peripheral VR population appeared genetically distinct while the remaining populations formed two clusters intermixing at

Table 2 Summary of genetic diversity parameters for *Grevillea bedgoodiana* by population based on nine microsatellite loci. N_{est} =estimated size of the adult population, N_a =number of alleles per locus, A_r =allelic richness (min. $n=10$), A_{priv} =private alleles; H_o =observed heterozygosity, H_e =gene diversity (expected heterozygosity) F_i =mean individual inbreeding coefficient, (F_i)=alternative

Pop	N_{est}	N_a	A_r	A_{priv}	H_o	H_e	F_i	(F_i)	1-s
OT	1400	5.3	4.5 _(1.1)	1	0.45 _(0.15)	0.60 _(0.16)	0.259	0.260	1.00 _(0.15)
WT	4800	7.1	5.9 _(1.4)	2	0.58 _(0.08)	0.73 _(0.09)	0.220	0.186	0.94 _(0.32)
HR	2100	6.9	5.6 _(1.4)	2	0.65 _(0.11)	0.69 _(0.10)	0.074	0.044	1.00 _(0.00)
MCS ^a	3000	7.9	6.3 _(1.0)	4	0.63 _(0.18)	0.75 _(0.07)	0.187	0.036	0.95 _(0.24)
MCN	1800	7.6	6.3 _(0.9)	2	0.60 _(0.19)	0.75 _(0.07)	0.225	0.035	1.00 _(0.19)
LGR	1300	6.7	5.5 _(0.9)	6	0.59 _(0.11)	0.70 _(0.10)	0.177	0.101	0.90 _(0.13)
IGR	400	4.9	4.1 _(0.6)	3	0.52 _(0.16)	0.66 _(0.05)	0.233	0.101	0.96 _(0.39)
VR	300	4.8	3.8 _(0.5)	2	0.64 _(0.15)	0.60 _(0.11)	-0.061	-0.149 ^b	0.99 _(0.17)
Mean	–	6.40	5.25	2.75	0.58	0.69	0.164	0.077	0.97
SE	–	0.41	0.33	0.52	0.02	0.02	0.035	0.040	0.01

Bold font indicates F_i values significantly greater than 0 (999 permutations, $p < 0.05$)

^aThis population was used in the pollination experiment

^bValue significantly lower than 0 (999 permutations, $p < 0.001$)

Table 3 Matrix of pairwise F_{ST} (AMOVA, 999 permutations, all p -values < 0.01) (below diagonal), and pairwise geographic (Euclidean) distances (km) between the sampled populations of *Grevillea bedgoodiana* (above diagonal)

	OT	WT	HR	MCS	MCN	LGR	IGR	VR
OT	–	1.2	3.2	2.7	4.5	3.8	8.5	11.3
WT	0.095	–	2.0	2.6	4.3	4.0	8.9	11.5
HR	0.114	0.012	–	3.1	4.1	4.7	9.3	11.5
MCS	0.117	0.036	0.047	–	1.8	1.7	6.4	8.9
MCN	0.115	0.045	0.041	0.021	–	1.8	5.3	7.4
LGR	0.196	0.109	0.127	0.072	0.073	–	4.8	3.1
IGR	0.202	0.112	0.122	0.112	0.088	0.134	–	3.1
VR	0.286	0.194	0.203	0.186	0.155	0.166	0.211	–

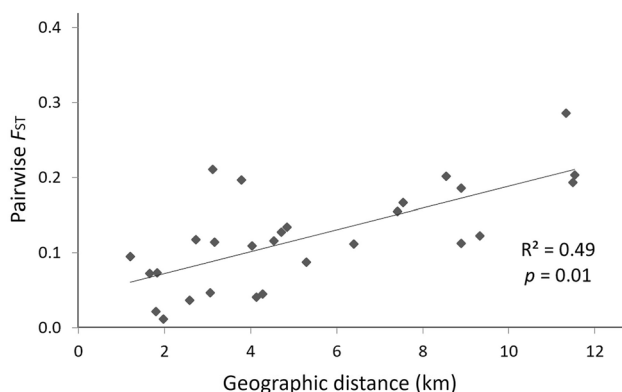


Fig. 4 Isolation by distance plot of pairwise F_{ST} vs geographic (Euclidean) distances among eight populations of *Grevillea bedgoodiana*

estimate of inbreeding coefficient following exclusion of markers IN12 and IN17 (see text), 1-s= outcrossing rate; values in parentheses indicate approximate $\pm 95\%$ confidence limits of the estimates; bold font indicates F_i values significantly greater than 0 (999 permutations, $p < 0.05$)

the MCS and MCN in the centre of ESP. The $K=5$ model further assigned distinct ancestry to the OT, LGR and IGR with the remaining WT, HR, MCS and MCN forming a broad fifth cluster variously intergrading with their geographic neighbours but virtually no gene flow between the populations that were ≥ 6 km apart (e.g., OT and VR) (Fig. 5b). The associated tree plots show relative genetic distances between the clusters (Fig. 5c).

The results of DAPC agreed with those of Structure and indicated genetic divergence of the two smaller populations from ESF (IGR and VR) from a main cluster consisting of the six larger populations from ESP (Fig. 6a). Removing VR and IGR from the analysis, allowed to resolve finer structuring of the remaining populations, whereby the two smaller and somewhat peripheral OT and LGR populations appeared to diverge from the four central populations (Fig. 6b).

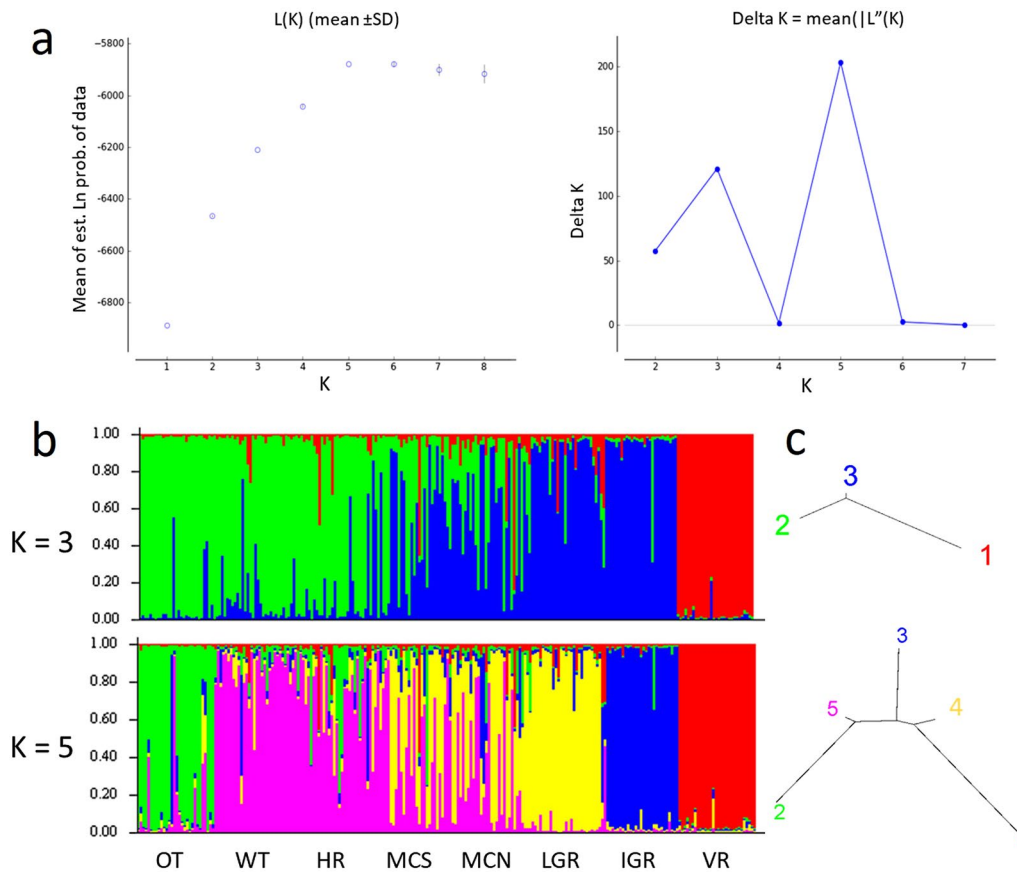


Fig. 5 **a** Structure Harvester’s L(K) plot and Delta K plot indicating K=3 and K=5 models explained the data nearly equally well (Evanno method); **b** Structure bar plots illustrating patterns of genetic admixture in the sampled *Grevillea bedgoodiana* populations under

the two models. Each vertical line represents one individual, different colours indicate proportion of an individual’s genotype deriving from one of the K ancestral groups; **c** Structure tree plots showing the relative genetic distances between the clusters

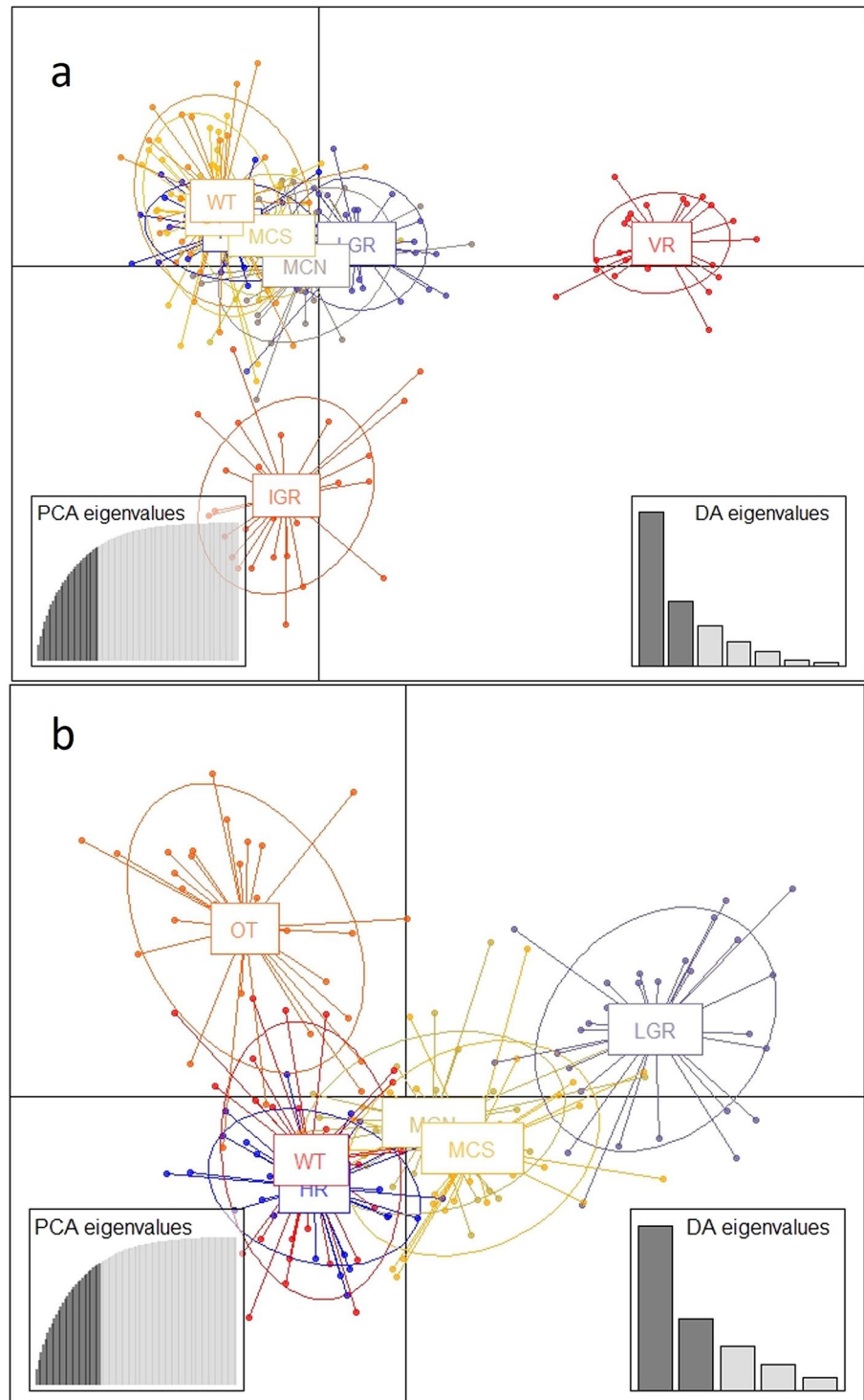
Reproductive rates vs gene diversity and inbreeding

The estimates of population areas ranged from 2500 and 22,000 m² and population sizes (N_{est}) from 300 to 4800 adult plants (Table 4). Reproductive rates (N_j) ranged from 5 to 93 juveniles per 100 adults (Table 4). Because N_j for IGR was a clear outlier, the data was log-transformed to achieve linearity. Regression analysis revealed no significant correlation between $\log_{10}(N_j)$ and gene diversity (H_e) (adjusted $R^2 = -0.50$, $df=6$, 95% CI: $-0.89 - 0.31$) or between $\log_{10}(N_j)$ and population mean inbreeding coefficients (F_i) estimated based on both full data set (9 loci) (adjusted $R^2 = -0.04$, $df=6$, 95% CI: $-0.89 - 0.31$) and without IN12 and IN17 (adjusted $R^2 = -0.06$, $df=6$, 95% CI: $-0.73 - 0.67$). The fire history data for the area revealed that two the study sites have been control-burnt since the 1995 wild-fire (OT and WT, 10–9 and 7–6 years ago, respectively). A third site (WT) was partially burnt 14 years ago (Table 4). The differences in time since last fire did not appear to influence N_j values (adjusted $R^2 = -0.001$, $df=6$, $p=0.36$). The models satisfied the assumptions of linear regression.

Discussion

This study aimed to determine the breeding system of the endangered *Grevillea bedgoodiana*, and to assess the genetic diversity and population structure across most of its range. It also aimed to quantify the reproductive output of the eight sampled populations, and to test whether it correlated with the estimates of gene diversity and inbreeding coefficients. Based on controlled pollinations of ten plants in a large central population, the species was found to be incapable of reproduction via autogamy. The small number of seeds produced through geitonogamy indicated the species is strongly and preferentially outcrossing but capable of self-fertilisation (at least at the MCS population). This result was supported by the genetic analyses which suggested high outcrossing rates, despite low-to-moderate levels of inbreeding in at least four of the populations studied. Open pollination and hand cross pollination resulted in equivalent levels of seed set indicating that seed production was not limited by pollen

Fig. 6 Discriminant analysis of the principal components (DAPC) scatter plots: **a** including all populations sampled; **b** excluding the two outlying populations from ESF for better resolution of the structure of the ESP populations. In both analyses retained were the first 30 principal components (dimensions of variation in the genotypic data) and all discriminant functions (synthetic variables optimizing the clustering of groups)



availability. Thus, the low number of seeds produced relative to total numbers of flowers per conflorescence (typical in Proteaceae; Hermanutz et al. 1998) is indicative of the conflorescence carrying capacity and/or resource limitation. Allelic richness, heterozygosity and gene diversity of the populations were mostly high. There was a moderate degree of differentiation among the sampled

populations, which could be largely attributed to isolation by distance. Analyses exploring population structure indicated that the large central populations in Enfield State Park were well connected but there was considerably less gene flow among the populations at the periphery of the species range and private alleles were detected in all populations. Numbers of seedlings per hundred adults were not

Table 4 Estimated population area, estimated population size (N_{est}), estimated number of juveniles per 100 adults (N_j) and time since last fire at the sampled populations of *Grevillea bedgoodiana*

Pop	Area ($\times 1000 \text{ m}^2$)	N_{est}	N_j	Time since last fire (years)
OT	11	1400	8	10, 9 ^a
WT	22	4800	16	23, 14 ^a
HR	17	2100	5	7, 6 ^a
MCS	18	3000	6	23
MCN	16	1800	5	23
LGR	6.0	1300	23	23
IGR	5.5	400	93	23
VR	2.5	300	25	23

^a These populations were control-burnt in two stages (using a road as a fire break) or only a portion of the population was burnt since the 1995 wildfire. In each case, the lower of the two values was used in the regression analysis

correlated with gene diversity or inbreeding coefficients. This result suggests there was no reduction in reproductive rates in populations with lower-than-average gene diversity or elevated estimates of inbreeding as could be expected if they were declining due to inbreeding depression (Szulkin et al. 2010).

Realised breeding systems are known to vary among populations in a number of angiosperms (Whitehead et al. 2018) including *Grevillea* (Hermanutz et al. 1998; Richardson et al. 2000). Likewise, the flowering intensity, abundance of pollinators and reproductive output are likely to vary both spatially and temporally (e.g., *G. repens*, a species closely related to *G. bedgoodiana*, Holmes et al. 2008; see also Copland and Whelan 1989). Consequently, the results of the controlled pollination experiment presented here need to be interpreted with caution as they are based on a small sample of plants from a single, central location and the experiment was conducted during a single flowering season. Moreover, the viability and fitness of the seeds produced through geitonogamy was not assessed. However, preferential outcrossing with a possibility of low rates of selfing (mixed mating) was supported by the high estimates of multilocus outcrossing rates in all populations. The proportion of initiated follicles that matured was high in *G. bedgoodiana* compared to, for example, *G. barklyana* (Vaughton 1996), in which consistently fewer than half of all initiated fruit developed to maturity. Furthermore, the presence of juveniles at all study sites indicates the species produces viable seeds capable of germinating at least sporadically under favourable conditions and without requiring fire/smoke cues (given that the most recent major wildfire in Enfield Forest occurred 23 years prior to this study and most of the populations have not been burnt since then). This is in contrast to for example

G. barklyana (Vaughton 1998) which has very low recruitment rates in the absence of fire, and several other species in which germination is strongly cued by fire and smoke (Kenny 2000).

The floral characters of *G. bedgoodiana* suggest the species should be primarily bird-pollinated (Olde and Marriott 1994; Cronk and Ojeda 2008; Willmer 2011) whereby the flowers have no obvious scent, do not reflect UV light (Aristidou and Hoebee, unpublished data) and produce copious amounts of somewhat dilute nectar ($^{\circ}\text{Bx}$: $25.8 \pm 0.9 \text{ SE}$, $n=6$ conflorescences; SW, unpublished data). Although, classical pollination syndromes may not always reliably predict actual primary pollinators (Waser et al. 1996; Rosas-Guerrero et al. 2014). A single bird species (the Yellow-faced honeyeater; *Lichenostomus chrysops*) was recorded visiting flowers during two weeks of remote camera trapping, there were also visits by small nocturnal mammals which may also be effective pollinators (Aristidou and Hoebee, unpublished data). However, the frequency of visitation by vertebrates was very low (only 16 records in total) and the most common floral visitor was the European honeybee (*Apis mellifera*, Aristidou and Hoebee, unpublished data), which was large enough to occasionally touch the pollen presenters/stigma while foraging for nectar (SW, pers. obs.). Thus, the species' potential vertebrate pollinators appear to have been effectively replaced by the honeybees. This is consistent with the well-documented decline of the woodland- and forest-dependent birds in south-eastern Australia (Ford et al. 2001; MacNally et al. 2009; Ford 2011) and with the finding that selective exclusion of vertebrates from *G. bedgoodiana* conflorescences had no effect on seed set (Aristidou and Hoebee, unpublished data). However, recent studies have raised concerns regarding the consequences of pollination by honeybees for reproduction and gene flow of native plants adapted to vertebrate pollination. Due to their high mobility and limited grooming of pollen, avian pollinators appear to play an important role in maintenance of genetic diversity and long-distance pollen dispersal of species they pollinate (Hoebee 2002; Breed et al. 2015; Krauss et al. 2009b; Bezemer et al. 2019; Kestel et al. 2021). For example, evidence from the self-compatible *Grevillea macleayana* (another species likely adapted to bird-mediated pollination) suggests that honeybees are more likely than birds to distribute pollen among flowers within individual plants, potentially reducing outcrossing rates (England et al. 2001). While in self-compatible population of *G. barklyana*, selective exclusion of vertebrate pollinators (allowing honeybee visitation) resulted in 50% reduction in seed set (Vaughton 1996). Our results of high outcrossing rates and low geitonogamous seed set in *G. bedgoodiana*, suggest that its mating system at the population level should be comparatively resilient to disruption by honeybees.

The genetic diversity of *G. bedgoodiana* was not uniformly distributed over its geographic range. The three largest populations (WT, MCS and MCN) had higher allelic richness than the two smallest populations (IGR and VR; Table 2). The observed heterozygosity and gene diversity also tended to be higher in the larger populations, however, the differences were not significant. OT appeared to break the trends in the data. Despite its relatively large size, its genetic diversity parameters, particularly allelic richness, were closer to those of the two small population from ESF (IGR and VR; Table 2), suggesting it may have gone through a recent bottleneck (Allendorf et al. 2013). This is consistent with OT and IGR bearing evidence of severe disturbance associated with historic gold mining (abandoned mine shafts and mounds of disturbed soil). Our Bottleneck analysis detected significant excess of heterozygotes at IGR, but not at OT. However, we note that because the short generation time and largely overlapping generations of *G. bedgoodiana* (the plants reach sexual maturity within five years), the bottleneck may have occurred too long ago (up to ca. 160 years prior to this study) to be reliably detected by this method (Allendorf et al. 2013). Nonetheless, the presence of *G. bedgoodiana* at these highly impacted sites, suggests that the species is either able to recover after severe and prolonged (several decades) disturbance, or can colonise disturbed areas relatively quickly. Additionally, the lack of correlation between the reproductive rates and gene diversity suggests that even the least genetically diverse populations were not affected by inbreeding depression. In fact, contrary to expectation, the population with the highest ratio of juveniles to adult plants (indicative of high fitness of the adult plants) was the visibly disturbed, small, and putatively bottlenecked IGR population. While it must be noted that the power of our test was sufficient only to detect large effect size with high probability (80%, https://www.statskingdom.com/sample_size_regression.html), this result suggests that *G. bedgoodiana* is capable of vigorous reproduction, possibly stimulated by soil disturbance breaking seed dormancy (Edwards and Whelan 1995; Pickup et al. 2003; Holmes et al. 2008). Taken together, our results suggest that the sampled populations of *G. bedgoodiana* are reproductively healthy, resilient to disturbance and have life history traits likely to promote genetic diversity, such as preferential outcrossing and overlapping generations (due to the staggered germination of soil-stored seeds). However, the species may still be at risk of reproductive failure should large wild-fires occur repeatedly in short succession (England et al. 2003). Frequent small fires such as the prescribed fuel reduction burns undertaken by the land managers in parts of the species range may also impact negatively on the species.

The genetic diversity and population structure of other species of *Grevillea* has been studied by several authors (Hoebee and Young 2001; England et al. 2002, 2003;

Hoebee et al. 2008; Holmes et al. 2009; Nistelberger et al. 2015; James et al. 2017; Kanjere and Hoebee, unpublished data). The methodological differences among these studies (molecular markers used, population genetic parameters reported and variation in sample size) make comparisons with our results difficult. Nonetheless, some comparisons with closely related congeners can be made. *Grevillea bedgoodiana* appeared to be less genetically diverse than either of the two grevilleas for which the microsatellite markers used in this study were originally developed: *G. infecunda*, a clonal narrow endemic, and *G. aquifolium*, a species with considerably wider distribution ($H_e = 0.78$ and 0.79 respectively, compared to 0.68 for *G. bedgoodiana*) (James 2012). However, *G. bedgoodiana* was considerably more diverse than two other narrowly endemic holly-leaved grevilleas: *G. obtecta* (mean $H_e = 0.57$ (Kanjere and Hoebee, unpublished data)) and *G. repens* (mean $H_e = 0.58$ (Holmes et al. 2009)). Despite this, the population inbreeding coefficients (F_i) were on average higher in *G. bedgoodiana* than in the latter two species (mean $F_i = 0.030$ and 0.024 , respectively, compared to 0.164 for *G. bedgoodiana*). This is likely a consequence of the differences in the three species' breeding systems: *G. obtecta* and *G. repens* are self-incompatible (Kanjere and Hoebee, unpublished data; Holmes et al. 2008), while *G. bedgoodiana*, although preferentially outcrossing, appears to be capable of low levels of selfing.

Given the restricted distribution of *G. bedgoodiana* and considering that birds and potentially other vertebrates appear to contribute to its pollination, one could reasonably expect the species to be panmictic, or nearly so. However, our results indicate that the species' populations are moderately structured with at least three distinct clusters of ancestry detected by the Structure and DAPC methods. It must be noted that Structure may overestimate the number of genetic clusters when inbreeding or isolation by distance are significant (Pritchard et al. 2000, 2009). In such cases the authors of the program note that the may not be a single 'correct' value of K and the Structure results need to be interpreted in the context of the species biology (Pritchard et al. 2000, 2009). Nonetheless, DAPC analysis of the main cluster of central five populations suggested further structuring even among the populations separated by only a few kilometres. Compared to two other holly-leaved grevilleas for which microsatellite-based population genetic studies were undertaken, the global F_{ST} for *G. bedgoodiana* was intermediate: considerably lower than that reported for *G. repens* – a species with strongly disjunct distribution (Holmes et al. 2009) – but higher than for *G. obtecta*, which has a continuous distribution across a similar geographic range and showed little genetic differentiation (Kanjere and Hoebee, unpublished data). Our results also suggest that considerable amount of genetic diversity of *G. bedgoodiana* is attributable to variation among its small peripheral populations, as

indicated by AMOVA and presence of private alleles in all populations (Table 2). This suggests a degree of reproductive isolation. However, we acknowledge that our data does not allow us to infer inter-population migration rates (Holsinger and Weir 2009; Lowe and Allendorf 2010). Given the importance of nectarivorous birds for long-distance pollen dispersal in other species (Hoebee 2002; Llorens et al. 2012; Phillips et al. 2014) and their current decline (Ford et al. 2001), future studies should test whether avian and/or mammal pollinators are essential and currently sufficiently abundant to maintain gene flow and demographic connectivity across the range of *G. bedgoodiana* (e.g., Sork and Smouse 2006; Krauss et al. 2009; Lowe and Allendorf 2010; Bezemer et al. 2016).

Implications for conservation

Currently, conservation management of *Grevillea bedgoodiana* is focused on protecting its largest populations in Enfield State Park (ESP). The management plan for ESP (Parks Victoria 1998) outlines the key conservation actions which include implementation of a fire management program to mitigate the risk of large-scale wildfires and control of invasive weeds and pest animal species (Parks Victoria 1998). It also postulates maintenance of genetic diversity of the native plant communities. Our results indicate that the most genetically diverse populations of *G. bedgoodiana* are already protected within ESP. However, the smaller populations in ESF were found to be considerably distinct. Conservation management should include provision of greater protection to the ESF populations to ensure maintenance of their unique genetic diversity. Moreover, while our results suggest *G. bedgoodiana* can flower and reproduce within six years post-fire, populations may require more time to replenish their soil-stored seedbanks and to build up their genetic diversity. In relation to this, our result of significant isolation by distance leads us to recommend that managers allow at least five years between burning adjacent patches of the species habitat to allow unburnt older plants an opportunity to breed with the younger plants grown from seeds in the burnt areas. This would increase the degree of generational overlap and mitigate the risk of loss of genetic diversity through drift. This is particularly relevant for the small, geographically isolated populations which are less likely to receive pollen migrants from the large core populations. Finally, the apparent decline of the vertebrate pollinators, is a cause for concern. While the European honeybees appear to be effective pollinators of this species, pollination solely by honeybees may be insufficient for maintenance of high outcrossing rates and gene flow among populations. Consequently, an integrated conservation action should aim to

protect and increase local abundance of native vertebrate pollinators, particularly nectarivorous birds.

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Author contributions SW and SH: study design. SW and GH: collections. SW: performed the laboratory and analyses under guidance from GH and SH; all authors participated in the development and writing of the manuscript.

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Data availability Microsatellite data generated during this study are available in Figshare repository, <https://doi.org/10.26181/20967757>.

Code availability R code used for DAPC and regression analyses including formatted data files are available in Figshare repository, <https://doi.org/10.26181/20967757>.

Declarations

Competing interests The authors declare no competing interests.

Conflict of interest The authors declare that they have no competing interest.

Ethical approval This research was conducted under the conditions of research permits 10007717 and 10008914 issued by the Department of Environment Land Water and Planning, Victorian Government. Animal ethics permit was not required.

Consent to participate SW, GH and SH have each consented to participation.

Consent for publication SW, GH and SH have each given full consent for publication.

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References

- Aitken SN, Whitlock MC (2013) Assisted gene flow to facilitate local adaptation to climate change. *Annu Rev Ecol Evol Syst* 44:367–388. <https://doi.org/10.1146/annurev-ecolsys-110512-135747>
- Aitken SN, Yeaman S, Holliday JA et al (2008) Adaptation, migration or extirpation: climate change outcomes for tree populations. *Evol Appl* 1:95–111. <https://doi.org/10.1111/j.1752-4571.2007.00013.x>
- Allendorf FW, Luikart G, Aitken SN (2013) Conservation and the Genetics of Populations. Wiley-Blackwell, Chichester
- Anderson JT, Panetta AM, Mitchell-Olds T (2012) Evolutionary and ecological responses to anthropogenic climate change. *Plant Physiol* 160:1728–1740. <https://doi.org/10.1104/pp.112.206219>
- Auld TD (1995) Burning grevilleas, ants, rats and wallabies. *CALM Sci Suppl* 4:159–164
- Auld TD, Denham AJ (1999) The role of ants and mammals in dispersal and post-dispersal seed predation of the shrubs *Grevillea* (Proteaceae). *Plant Ecol* 144:201–213. <https://doi.org/10.1023/A:1009817132378>
- Barrett SCH, Harder LD (2017) The ecology of mating and its evolutionary consequences in seed plants. *Annu Rev Ecol Evol Syst* 48:135–157. <https://doi.org/10.1146/annurev-ecolsys-110316-023021>
- Bezemer N, Krauss SL, Phillips RD et al (2016) Paternity analysis reveals wide pollen dispersal and high multiple paternity in a small, isolated population of the bird-pollinated *Eucalyptus caesia* (Myrtaceae). *Heredity* (edinb) 117:460–471. <https://doi.org/10.1038/hdy.2016.61>
- Bezemer N, Hopper SD, Krauss SL et al (2019) Primary pollinator exclusion has divergent consequences for pollen dispersal and mating in different populations of a bird-pollinated tree. *Mol Ecol* 28:4883–4898. <https://doi.org/10.1111/mec.15264>
- Blacket MJ, Robin C, Good RT et al (2012) Universal primers for fluorescent labelling of PCR fragments - an efficient and cost-effective approach to genotyping by fluorescence. *Mol Ecol Resour* 12:456–463. <https://doi.org/10.1111/j.1755-0998.2011.03104.x>
- Breed MF, Ottewill KM, Gardner MG et al (2015) Mating system and early viability resistance to habitat fragmentation in a bird-pollinated eucalypt. *Heredity* 115:100–107. <https://doi.org/10.1038/hdy.2012.72>
- Caballero A, Bravo I, Wang J (2017) Inbreeding load and purging: Implications for the short-term survival and the conservation management of small populations. *Heredity* 118:177–185. <https://doi.org/10.1038/hdy.2016.80>
- Caddy HAR, Gross CL (2006) Population structure and fecundity in the putative sterile shrub, *Grevillea rhizomatosa* Olde & Marriott (Proteaceae). *Proc Linn Soc New South Wales* 127:11–18
- Carter O, Murphy AH, Downe J (2006) National Recovery Plan for the Enfield Grevillea, *Grevillea bedgoodiana*. Department of Sustainability and Environment, Melbourne
- Celebrezze T, Paton DC (2004) Do introduced honeybees (*Apis mellifera*, Hymenoptera) provide full pollination service to bird-adapted Australian plants with small flowers? an experimental study of *Brachyloma ericoides* (Epacridaceae). *Austral Ecol* 29:129–136. <https://doi.org/10.1111/j.1442-9993.2003.01328.x>
- Coates DJ, Sampson JF, Yates CJ (2007) Plant mating systems and assessing population persistence in fragmented landscapes. *Aust J Bot* 55:239–249. <https://doi.org/10.1071/BT06142>
- Cole CT (2003) Genetic variation in rare and common plants. *Annu Rev Ecol* 34:213–237. <https://doi.org/10.1146/annurev.ecolsys.34.030102.151717>
- Copland BJ, Whelan RJ (1989) Seasonal variation in flowering intensity and pollination limitation of fruit set in four co-occurring *Banksia* species. *J Ecol* 77:509. <https://doi.org/10.2307/2260766>
- Cranmer L, McCollin D, Ollerton J (2012) Landscape structure influences pollinator movements and directly affects plant reproductive success. *Oikos* 121:562–568. <https://doi.org/10.1111/j.1600-0706.2011.19704.x>
- Cronk Q, Ojeda I (2008) Bird-pollinated flowers in an evolutionary and molecular context. *J Exp Bot* 59:715–727. <https://doi.org/10.1093/jxb/ern009>
- Devaux C, Lepers C, Porcher E (2014) Constraints imposed by pollinator behaviour on the ecology and evolution of plant mating systems. *J Evol Biol* 27:1413–1430. <https://doi.org/10.1111/jeb.12380>
- Earl DA, VonHoldt BM (2012) Structure Harvester: a website and program for visualizing Structure output and implementing the Evanno method (Web version v0.6.94). *Conserv Genet Resour* 4:359–361. <https://doi.org/10.1007/s12686-011-9548-7>
- Edwards W, Whelan R (1995) The size, distribution and germination requirements of the soil-stored seed-bank of *Grevillea barklyana* (Proteaceae). *Aust J Ecol* 20:548–555. <https://doi.org/10.1111/j.1442-9993.1995.tb00574.x>
- England PR, Beynon F, Ayre DJ, Whelan RJ (2001) A molecular genetic assessment of mating-system variation in a naturally bird-pollinated shrub: contributions from birds and introduced honeybees. *Conserv Biol* 15:1645–1655. <https://doi.org/10.1046/j.1523-1739.2001.00236.x>
- England PR, Usher AV, Whelan RJ, Ayre DJ (2002) Microsatellite diversity and genetic structure of fragmented populations of the rare, fire-dependent shrub *Grevillea macleayana*. *Mol Ecol* 11:967–977. <https://doi.org/10.1046/j.1365-294X.2002.01500.x>
- England PR, Whelan RJ, Ayre DJ (2003) Effects of seed bank disturbance on the fine-scale genetic structure of populations of the rare shrub *Grevillea macleayana*. *Heredity* (edinb) 91:475–480. <https://doi.org/10.1038/sj.hdy.6800308>
- Excoffier L, Smouse PE, Quattro JM (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics* 131:479–491
- Ford HA (2011) The causes of decline of birds of eucalypt woodlands: advances in our knowledge over the last 10 years. *Emu - Austral Ornithol* 111:1–9. <https://doi.org/10.1071/MU09115>
- Ford HA, Barrett GW, Saunders DA, Recher HF (2001) Why have birds in the woodlands of southern Australia declined? *Biol Conserv* 97:71–88. [https://doi.org/10.1016/S0006-3207\(00\)00101-4](https://doi.org/10.1016/S0006-3207(00)00101-4)
- Forester BR, Lasky JR, Wagner HH et al (2018) Comparing methods for detecting multilocus adaptation with multivariate genotype-environment associations. *Mol Ecol* 27:2215–2233. <https://doi.org/10.1111/mec.14584>
- Frankham R (2015) Genetic rescue of small, inbred populations: meta-analysis reveals large and consistent benefits of gene flow. *Mol Ecol* 24:2610–2618. <https://doi.org/10.1111/mec.13139>
- Gaston KJ (1996) Species-range-size distributions: patterns, mechanisms and implications. *Trends Ecol Evol* 11:197–201. [https://doi.org/10.1016/0169-5347\(96\)10027-6](https://doi.org/10.1016/0169-5347(96)10027-6)
- Gaston KJ, Blackburn TM, Lawton JH (1997) Interspecific abundance-range size relationships: an appraisal of mechanisms. *J Anim Ecol* 66:579. <https://doi.org/10.2307/5951>

- Gitzendanner MA, Soltis PS (2000) Patterns of genetic variation in rare and widespread plant congeners. *Am J Bot* 87:783–792. <https://doi.org/10.2307/2656886>
- Gross CL, Nelson PA, Haddadchi A, Fatemi M (2012) Somatic mutations contribute to genotypic diversity in sterile and fertile populations of the threatened shrub, *Grevillea rhizomatosa* (Proteaceae). *Ann Bot* 109:331–342. <https://doi.org/10.1093/aob/mcr283>
- Hardy OJ, Vekemans X (2002) SPAGeDi: a versatile computer program to analyse spatial genetic structure at the individual or population levels. *Mol Ecol Notes* 2:618–620. <https://doi.org/10.1046/j.1471-8278.2002.00305.x>
- Hermanutz LAC, Innes DA, Denham AB, Whelan RB (1998) Very low fruit:flower ratios in *Grevillea* (Proteaceae) are independent of breeding system. *Aust J Bot* 46:465–478. <https://doi.org/10.1071/BT97046>
- Hoebee SE, Young AG (2001) Low neighbourhood size and high interpopulation differentiation in the endangered shrub *Grevillea iaspicula* McGill. (Proteaceae). *Heredity* (edinb) 86:489–496. <https://doi.org/10.1046/j.1365-2540.2001.00857.x>
- Hoebee SE, Thrall PH, Young AG (2008) Integrating population demography, genetics and self-incompatibility in a viability assessment of the Wee Jasper *Grevillea* (*Grevillea iaspicula* McGill., Proteaceae). *Conserv Genet* 9:515–529. <https://doi.org/10.1007/s10592-007-9366-3>
- Hoebee SE (2002) Conservation genetics of the endangered shrub *Grevillea iaspicula* McGill. (Proteaceae). Doctoral Dissertation, The Australian National University
- Holm S (1979) A simple sequentially rejective multiple test procedure. *Scand J Stat* 6:65–70. <https://www.jstor.org/stable/4615733>
- Holmes GD, James EA, Hoffmann AA (2008) Limitations to reproductive output and genetic rescue in populations of the rare shrub *Grevillea repens* (Proteaceae). *Ann Bot* 102:1031–1041
- Holmes GD, James EA, Hoffmann AA (2009) Divergent levels of genetic variation and ploidy among populations of the rare shrub, *Grevillea repens* (Proteaceae). *Conserv Genet* 10:827–837. <https://doi.org/10.1007/s10592-008-9643-9>
- Holsinger KE, Weir BS (2009) Genetics in geographically structured populations: defining, estimating and interpreting F_{ST} . *Nat Rev Genet* 10:639–650. <https://doi.org/10.1038/nrg2611>
- IUCN (2012) IUCN Red List Categories and Criteria: Version 3.1. Second edition. IUCN, Gland, Switzerland and Cambridge, UK. <https://portals.iucn.org/library/sites/library/files/documents/RL-2001-001-2nd.pdf>
- James EA, McDougall KL (2014) Spatial genetic structure reflects extensive clonality, low genotypic diversity and habitat fragmentation in *Grevillea renwickiana* (Proteaceae), a rare, sterile shrub from south-eastern Australia. *Ann Bot* 114:413–423. <https://doi.org/10.1093/aob/mcu049>
- James EA, Brown GK, Citroen R, Blacket MJ (2012) Microsatellite marker development for two species of holly-leaved *Grevillea* and cross-species amplification in the *Aspleniifolia*/hookeriana subgroup (Proteaceae). *Conserv Genet Resour*. <https://doi.org/10.1007/s12686-011-9493-5>
- James EA, Jordan R, Brown GK, Griffin PC (2017) Assessing the genetic legacy of a rare, clonal Australian shrub *Grevillea infecunda* (Proteaceae). *Folia Geobot* 52:387–400. <https://doi.org/10.1007/s12224-016-9258-8>
- Jombart T (2008) adegenet: a R package for the multivariate analysis of genetic markers. *Bioinformatics* 24:1403–1405. <https://doi.org/10.1093/bioinformatics/btn129>
- Jombart T, Devillard S, Balloux F (2010) Discriminant analysis of principal components: a new method for the analysis of genetically structured populations. *BMC Genet* 11:94. <https://doi.org/10.1186/1471-2156-11-94>
- Joyce EB (2004) The young volcanic regions of southeastern Australia: early studies, physical volcanology and eruption risk. *Proc R Soc Victoria* 116:1–13
- Kenny BJ (2000) Influence of multiple fire-related germination cues on three Sydney *Grevillea* (Proteaceae) species. *Austral Ecol* 25:664–669. <https://doi.org/10.1111/j.1442-9993.2000.tb00072.x>
- Kestel JH, Phillips RD, Anthony J et al (2021) Unexpectedly low paternal diversity is associated with infrequent pollinator visitation for a bird-pollinated plant. *Oecologia*. <https://doi.org/10.1007/s00442-021-04906-x>
- Kimpton SK, James EA, Drinnan AN (2002) Reproductive biology and genetic marker diversity in *Grevillea infecunda* (Proteaceae), a rare plant with no known seed production. *Aust Syst Bot* 15:485–492. <https://doi.org/10.1071/SB01029>
- Krauss SL, He T, Barrett LG et al (2009) Contrasting impacts of pollen and seed dispersal on spatial genetic structure in the bird-pollinated *Banksia hookeriana*. *Heredity* (edinb) 102:274–285. <https://doi.org/10.1038/hdy.2008.118>
- Lamont BB, Groom PK (1998) Seed and seedling biology of the woody-fruited Proteaceae. *Aust J Bot* 46:387–406. <https://doi.org/10.1071/BT96135>
- Leimu R, Mutikainen P, Koricheva J, Fischer M (2006) How general are positive relationships between plant population size, fitness and genetic variation? *J Ecol* 94:942–952. <https://doi.org/10.1111/j.1365-2745.2006.01150.x>
- Llorens TM, Byrne M, Yates CJ et al (2012) Evaluating the influence of different aspects of habitat fragmentation on mating patterns and pollen dispersal in the bird-pollinated *Banksia sphaerocarpa* var. *caesia*. *Mol Ecol* 21:314–328. <https://doi.org/10.1111/j.1365-294X.2011.05396.x>
- Llorens TM, Yates CJ, Byrne M et al (2013) Complex interactions between remnant shape and the mating system strongly influence reproductive output and progeny performance in fragmented populations of a bird-pollinated shrub. *Biol Conserv* 164:129–139. <https://doi.org/10.1016/j.biocon.2013.05.011>
- Llorens TM (2004) Conservation genetics and ecology of two rare *Grevillea* species. Doctoral Dissertation, University of Wollongong
- Loiselle BA, Sork VL, Nason J, Graham C (1995) Spatial genetic structure of a tropical understory shrub, *Psychotria officinalis* (Rubiaceae). *Am J Bot* 82:1420–1425. <https://doi.org/10.2307/2445869>
- Lowe WH, Allendorf FW (2010) What can genetics tell us about population connectivity? *Mol Ecol* 19:3038–3051. <https://doi.org/10.1111/j.1365-294X.2010.04688.x>
- Mac Nally R, Bennett AF, Thomson JR et al (2009) Collapse of an avifauna: Climate change appears to exacerbate habitat loss and degradation. *Divers Distrib* 15:720–730. <https://doi.org/10.1111/j.1472-4642.2009.00578.x>
- Makinson RO (2000) *Grevillea*. Flora of Australia Vol.17A. CSIRO Publishing, Melbourne
- MBG, Ayre DJ, Whelan RJ, (2000) Pollinator behaviour, mate choice and the realised mating systems of *Grevillea mucronulata* and *Grevillea sphacelata*. *Aust J Bot* 48:357–366. <https://doi.org/10.1071/BT98078>
- Nistelberger HM, Byrne M, Coates D, Roberts JD (2015) Genetic drift drives evolution in the bird-pollinated, terrestrial island endemic *Grevillea georgeana* (Proteaceae). *Bot J Linn Soc* 178:155–168. <https://doi.org/10.1111/boj.12270>

- Nora S, Aparicio A, Albaladejo RG (2016) High correlated paternity leads to negative effects on progeny performance in two mediterranean shrub species. *PLoS ONE* 11:1–15. <https://doi.org/10.1371/journal.pone.0166023>
- Olde P, Marriott N (1994) *The Grevillea book*, vol 2. Kangaroo Press, Sydney
- Ottewell KM, Bickerton DC, Byrne M, Lowe AJ (2016) Bridging the gap: a genetic assessment framework for population-level threatened plant conservation prioritization and decision-making. *Divers Distrib* 22:174–188. <https://doi.org/10.1111/ddi.12387>
- Paton DC (2000) Disruption of bird-plant pollination systems in southern Australia. *Conserv Biol* 14:1232–1234. <https://doi.org/10.1046/j.1523-1739.2000.00015.x>
- Peakall R, Smouse PE (2006) GENALEX 6: genetic analysis in Excel population genetic software for teaching and research. *Mol Ecol Notes* 6:288–295. <https://doi.org/10.1111/j.1471-8286.2005.01155.x>
- Peakall R, Smouse PE (2012) GenAlix 6.5: genetic analysis in Excel population genetic software for teaching and research—an update. *Bioinformatics* 28:2537–2539. <https://doi.org/10.1093/bioinformatics/bts460>
- Peña EA, Slate EH (2006) Global validation of linear model assumptions. *J Am Stat Assoc* 101:341–354. <https://doi.org/10.1198/016214505000000637>
- Phillips RD, Steinmeyer F, Menz MHM et al (2014) Changes in the composition and behaviour of a pollinator guild with plant population size and the consequences for plant fecundity. *Funct Ecol* 28:846–856. <https://doi.org/10.1111/1365-2435.12237>
- Pickup M, McDougall KL, Whelan RJ (2003) Fire and flood: Soil-stored seed bank and germination ecology in the endangered carrington falls *Grevillea* (*Grevillea rivularis*, Proteaceae). *Austral Ecol* 28:128–136. <https://doi.org/10.1046/j.1442-9993.2003.01255.x>
- Piry S, Luikart G, Cornuet J-M (1999) BOTTLENECK: a computer program for detecting recent reductions in the effective population size using allele frequency data. *J Hered* 90:502–503
- Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using multilocus genotype data. *Genetics* 155:945–959. <https://doi.org/10.1093/genetics/155.2.945>
- Pritchard JK, Wen X, Falush D (2009) Documentation for structure software: Version 2.3. https://www.ccg.unam.mx/~vinausa/tlem09/docs/structure_doc.pdf
- Putman AI, Carbone I (2014) Challenges in analysis and interpretation of microsatellite data for population genetic studies. *Ecol Evol* 4:4399–4428. <https://doi.org/10.1002/ece3.1305>
- R Core Team (2021) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna
- Recher HF, Lunney D, Matthews A (2009) Small mammal populations in a eucalypt forest affected by fire and drought. I. Long-term patterns in an era of climate change. *Wildl Res* 36:143–158. <https://doi.org/10.1071/WR08086Richardson>
- Reed DH, Frankham R (2003) Correlation between fitness and genetic diversity. *Conserv Biol* 17:230–237. <https://doi.org/10.1046/j.1523-1739.2003.01236.x>
- Ritchie AL, Dyer RJ, Nevill PG et al (2019) Wide outcrossing provides functional connectivity for new and old *Banksia* populations within a fragmented landscape. *Oecologia* 190:255–268. <https://doi.org/10.1007/s00442-019-04387-z>
- Richardson MBG, Ayre DJ, Whelan RJ (2000) Pollinator behaviour, mate choice and the realised mating systems of *Grevillea mucronulata* and *Grevillea sphaecelata*. *Aust J Bot* 48:357–366. <https://doi.org/10.1071/BT98078>
- Rosas-Guerrero V, Aguilar R, Martín-Rodríguez S et al (2014) A quantitative review of pollination syndromes: do floral traits predict effective pollinators? *Ecol Lett* 17:388–400. <https://doi.org/10.1111/ele.12224>
- Rousset F (1997) Genetic differentiation and estimation of gene flow from F-Statistics under isolation by distance. *Genetics* 145:1219–1228. <https://doi.org/10.1177/000992287301200933>
- Rousset F (2008) genepop'007: a complete re-implementation of the GENEPOP software for Windows and Linux. *Mol Ecol Resour* 8:103–106. <https://doi.org/10.1111/j.1471-8286.2007.01931.x>
- Rymer PD, Welan RJ, Ayre DJ et al (2005) Reproductive success and pollinator effectiveness differ in common and rare *Persoonia* species (Proteaceae). *Biol Conserv* 123:521–532. <https://doi.org/10.1016/J.BIOCON.2005.01.002>
- Scheele BC, Legge S, Armstrong DP et al (2018) How to improve threatened species management: an Australian perspective. *J Environ Manage* 223:668–675. <https://doi.org/10.1016/j.jenvman.2018.06.084>
- Schuelke M (2000) An economic method for the fluorescent labeling of PCR fragments. *Nat Biotechnol* 18:233–234. <https://doi.org/10.1038/72708>
- Séré M, Thévenon S, Belem AMG, De Meeûs T (2017) Comparison of different genetic distances to test isolation by distance between populations. *Heredity (edinb)* 119:55–63. <https://doi.org/10.1038/hdy.2017.26>
- Slatyer RA, Hirst M, Sexton JP (2013) Niche breadth predicts geographical range size: a general ecological pattern. *Ecol Lett* 16:1104–1114. <https://doi.org/10.1111/ele.12140>
- Smith JA, Gross CL (2002) The pollination ecology of *Grevillea beadleana* McGillivray, an endangered shrub from northern New South Wales, Australia. *Ann Bot* 89:97–108. <https://doi.org/10.1093/aob/mcf015>
- Sork VL, Smouse PE (2006) Genetic analysis of landscape connectivity in tree populations. *Landsc Ecol* 21:821–836. <https://doi.org/10.1007/s10980-005-5415-9>
- Szulkin M, Bierne N, David P (2010) Heterozygosity-fitness correlations: a time for reappraisal. *Evolution* 64:1202–1217. <https://doi.org/10.1111/j.1558-5646.2010.00966.x>
- Thrall PH, Encinas-Viso F, Hoebee SE, Young AG (2014) Life history mediates mate limitation and population viability in self-incompatible plant species. *Ecol Evol* 4:673–687. <https://doi.org/10.1002/ece3.963>
- Vaughton G (1996) Pollination disruption by European honeybees in the Australian bird-pollinated shrub *Grevillea barklyana* (Proteaceae). *Plant Syst Evol* 200:89–100. <https://doi.org/10.1007/BF00984750>
- Vaughton G (1998) Soil seed bank dynamics in the rare obligate seeding shrub, *Grevillea barklyana* (Proteaceae). *Austral Ecol* 23:375–384. <https://doi.org/10.1111/j.1442-9993.1998.tb00742.x>
- Victoria P (1998) Enfield State Park Management Plan. Parks Victoria, Kew
- Waser NM, Chittka L, Price MV et al (1996) Generalization in pollination systems, and why it matters. *Ecology* 77:1043–1060. <https://doi.org/10.2307/2265575>
- Weir BS, Cockerham CC (1984) Estimating F-statistics for the analysis of population structure. *Evolution (NY)* 38:1358–1370. <https://doi.org/10.2307/2408641>
- Whelan RJ, Ayre DJ, Beynon FM (2009) The birds and the bees: pollinator behaviour and variation in the mating system of the rare shrub *Grevillea macleayana*. *Ann Bot* 103:1395–1401. <https://doi.org/10.1093/aob/mcp091>

- Whitehead MR, Lanfear R, Mitchell RJ, Karron JD (2018) Plant mating systems often vary widely among populations. *Front Ecol Evol* 6:38. <https://doi.org/10.3389/fevo.2018.00038>
- Wilcock C, Neiland R (2002) Pollination failure in plants: why it happens and when it matters. *Trends Plant Sci* 7:270–277. [https://doi.org/10.1016/S1360-1385\(02\)02258-6](https://doi.org/10.1016/S1360-1385(02)02258-6)
- Willi Y, Van Buskirk J, Fischer M (2005) A threefold genetic allee effect: population size affects cross-compatibility, inbreeding depression and drift load in the self-incompatible *Ranunculus reptans*. *Genetics* 169:2255–2265. <https://doi.org/10.1534/genetics.104.034553>
- Willmer P (2011) *Pollination and floral ecology*. Princeton University Press, Princeton
- Yates CJ, Coates DJ, Elliott C, Byrne M (2007) Composition of the pollinator community, pollination and the mating system for a shrub in fragments of species rich kwongan in south-west Western Australia. *Biodivers Conserv* 16:1379–1395. <https://doi.org/10.1007/s10531-006-6736-y>
- Young A, Boyle T, Brown T (1996) The population genetic consequences of habitat fragmentation for plants. *Trends Ecol & Evol* 11:413–418

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