

Osteoblast-induced EGFR/ERBB2 signaling in androgen-sensitive prostate carcinoma cells characterized by multiplex kinase activity profiling

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Abstract Bone metastases in prostate cancer are predominantly osteoblastic. To study regulatory mechanisms underlying the establishment of prostate cancer within an osteoblastic microenvironment, human androgen-sensitive prostate carcinoma cells (LNCaP) were treated with culture medium conditioned by human osteoblast-derived sarcoma

cells (OHS), and activated signalling pathways in the carcinoma cells were analyzed using microarrays with tyrosine kinase substrates. Network interaction analysis of substrates with significantly increased phosphorylation levels revealed that signalling pathways mediated by EGFR and ERBB2 were activated in LNCaP cells under OHS influence but also by androgen treatment. Activation of EGFR/ERBB2 signalling was also found in LNCaP cells in cocultures with OHS cells or osteoblastic cells that had been differentiated from human mesenchymal stem cells. Our experimental data suggests osteoblast-directed induction of signalling activity via EGFR and ERBB2 in prostate carcinoma cells and may provide a rationale for the use of EGFR or ERBB2 inhibition in systemic prevention or treatment of metastatic prostate cancer in the androgen-sensitive stage of the disease.

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Abbreviations

FBS Fetal bovine serum
EGF Epidermal growth factor

Introduction

Metastatic prostate cancer is a leading cause of cancer morbidity and mortality. The skeleton is the principal organ for metastasis formation in prostate cancer, and bone metastases are frequently painful and debilitating. Whilst androgens are critical regulators of prostate carcinoma growth and progression, most patients respond only temporarily to androgen ablation therapy. Skeletal metastases are predominantly osteoblastic, as apparent both

radiographically and histopathologically, and are associated with elevated serum level of bone-specific alkaline phosphatase, a marker of osteoblast proliferation [1, 2]. These observations suggest that the biological interaction between prostate carcinoma cells and osteoblasts contributes to the metastatic progression of prostate cancer.

In order to experimentally address how osteoblastic cells may influence prostate carcinoma cell biology upon formation of bone metastasis, the human, androgen-sensitive prostate carcinoma cell line LNCaP [3] was treated with culture medium conditioned by the human, osteoblast-derived sarcoma cell line OHS [4], and subsequent multiplex profiling of LNCaP kinase activity was performed using flow-through microarrays with peptide substrates (Tyrosine Kinase PamChip® Arrays; PamGene International BV, 's-Hertogenbosch, The Netherlands), a novel platform that allows rapid, real-time measurements of phosphopeptide signatures generated by biological samples. As supplementary approaches, we used LNCaP cells in cocultures with either OHS cells or with osteoblastic cells that had been differentiated from human mesenchymal stem cells.

Insight into regulatory mechanisms underlying the establishment of prostate carcinoma cells within an osteoblastic microenvironment may eventually lead to more effective therapies to prevent or treat metastatic disease. Hence, from a therapeutic perspective, we compared the intracellular LNCaP signalling pathways activated by influence of osteoblastic cells with pathways induced by androgen treatment, to identify signalling networks potentially accessible for therapeutic targeting.

Materials and methods

Cell cultures

The LNCaP and OHS cell lines were routinely held in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS) and 2.0 mM glutamine, defined as growth medium. Seventy-two hours before start of experimental incubations, monocultures of LNCaP and OHS were seeded in a total number of 1.0×10^6 cells in 75 cm² cell flasks in RPMI containing 2% charcoal-treated FBS and glutamine. After 48 h, this medium was changed to RPMI containing 0.5% charcoal-treated FBS and glutamine, defined as experimental medium, for another 24 h before experimental incubations were started (at time 0). All cell cultures were invariably held in 10 ml medium throughout different incubations.

At time 0 (start of the experimental incubations), LNCaP cells were refed with experimental medium supplemented with 100 nM of the synthetic androgen analog

R1881 (methyltrienolone; Biocompare, Inc., South San Francisco, CA), giving rise to the LNCaP entity denoted androgenic, or with medium conditioned by OHS cells. The conditioned medium had been collected from OHS monocultures that had been grown for 48 h (relative to time 0) in experimental medium. This medium was subsequently diluted 1:10 in either fresh experimental medium or in medium obtained from standard LNCaP monocultures after 48 h of incubation (relative to time 0) in experimental medium, before the application onto monocultured LNCaP cells, giving rise to the LNCaP entities denoted paracrine 1 and paracrine 2, respectively.

Cocultures of LNCaP/OHS were also seeded in a total number of 1.0×10^6 cells in 75 cm² cell flasks in 10 ml medium and were incubated identically to the monocultured cell lines prior to and during experimental incubations. Different ratios of LNCaP to OHS cells had been tested in a series of cocultures to find the optimal culturing conditions [5], and a 10:1 seeding ratio was chosen.

Differentiated mesenchymal stem cells from human bone marrow were prepared as previously described [6, 7]. Briefly, bone marrow (10–20 ml) was aspirated from the posterior iliac crest of healthy, adult volunteers. Mononuclear cells were isolated by density gradient centrifugation and resuspended in complete medium, consisting of MEM α supplemented with 20% FBS and 2.0 mM glutamine. After 24 h, non-adherent cells were discarded, and the adherent cells were thoroughly washed and further cultured in complete medium for 7 days. Subsequently, the cells were detached and replated in osteogenic differentiation medium, consisting of MEM α with 10% FBS, 1.0 mM glutamine, 10 nM dexamethasone, 10 μ M β -glycerol phosphate, and 200 μ M ascorbic acid. This medium was replaced every 3–4 days for about 20 days before LNCaP cells (1.0×10^6 cells in 75 cm² cell flasks) were seeded on top of the osteoblasts and the cocultures of LNCaP and differentiated stem cells were incubated identically to the other culture setups prior to and during experimental incubations.

In the experimental setups, LNCaP cells were harvested 48 h after time 0, either directly from the various monocultures or after immunomagnetic cell segregation of cocultures (see below), and lysed in M-PER Mammalian Extraction Reagent containing Halt Phosphatase Inhibitor Cocktail and EDTA-free Halt Protease Inhibitor Cocktail (Pierce Biotechnology, Inc., Rockford, IL). Reference lysates were made from monocultured LNCaP grown for 48 h (relative to time 0) in experimental medium only.

In the experimental setup involving epidermal growth factor (EGF; Sigma–Aldrich Norway AS, Oslo, Norway), EGF was added to LNCaP cells in a final concentration of 50 ng/ml.

Kinase activity profiling

The kinase substrate array technology allows functional comparison of biological samples without prior knowledge of which activity pathways are influenced by the experimental conditions. The high-throughput format of the Tyrosine Kinase PamChip[®] Array is based on the use of a porous, three-dimensional aluminum-oxide material as solid support for the substrates. The sample lysates are actively pumped through the interconnected capillary pores of the arrays to allow contact with the reactive surface for enzymatic reaction with the peptide substrates. The phosphorylation kinetics are therefore rapid and can be completed within few minutes, allowing the generation of spot images to be followed in real-time.

Each array contains 144 peptide substrates, and these target peptides consist of 13 or 14 amino acids with sites for phosphorylation, mainly tyrosine, representing 100 different proteins. Reaction mixtures consisted of Abl Reaction Buffer (50 mM Tris-HCl pH 7.5, 10 mM MgCl₂, 1 mM EGTA, 2 mM dithiothreitol, 0.01% Brij 35; New England BioLabs, Inc., Ipswich, MA), 1 mg/ml bovine serum albumin, 100 μM ATP, and 12.5 μg/ml of the monoclonal, FITC-conjugated anti-phosphotyrosine antibody (Exalpha Biologicals, Inc., Maynard, MA), added to each sample lysate, containing 2–5 μg total protein. The arrays were blocked with 20 μg/ml bovine serum albumin and subsequently washed twice with Abl Reaction Buffer before the reaction mixtures were applied for initiation of enzymatic reactions. Spot images were recorded after each completed pumping cycle by a charge-coupled device camera until the reactions were terminated.

Raw and processed data and investigation and array designs can be obtained from the EMBL-EBI ArrayExpress database (www.ebi.ac.uk) by accession number E-TABM-626.

Microarray data adaptation and statistical analysis

Six independent LNCaP reference samples (denoted LNCaP baseline), one of each LNCaP treated sample (paracrine 1, paracrine 2, androgenic), one sample each of LNCaP and OHS cells immunosegregated from coculture (see below), and two independent samples of monocultured OHS cells were analyzed, and each sample was measured in three independent reactions. The image information was converted using BioNavigator software (PamGene International BV, www.pamgene.com). For each spot on the array, signal intensity after background subtraction was calculated and used for further analysis. Data normalization of spot signal intensities and subsequent comparison analyses were conducted using GeneSpring software (Agilent Technologies, www.home.agilent.com). All values

were normalized to the calculated mean value of all substrate phosphorylation intensities in the LNCaP baseline sample. In the LNCaP treated samples, the criterion for selecting a substrate with significantly increased phosphorylation level compared to the corresponding value in the baseline sample was set at a statistical difference of $P < 0.05$ (Student's *t*-test). Using this criterion, the fold-increase (\log_2) of substrate phosphorylation levels was within a range of 0.85 to > 5.2 .

Following the statistical analysis, two different pathway visualization systems were used to create information about pathway connectivity. Peptide names were generated from corresponding gene name entries and protein sequences in SwissProt (<http://au.expasy.org/sprot>) to enable unambiguous annotation of the peptide sequences and phosphorylation sites. It should be noted that, for some proteins, multiple peptides have been derived and are present on the Tyrosine Kinase PamChip[®] Array. For the pathway connectivity analysis, one list of peptide identifications for each of the treated samples was generated. First, those lists were loaded into PathwayArchitect software (Stratagene Corp., www.stratagene.com; Strand Life Sciences Pvt. Ltd., www.avadis.strandgenomics.com), and the program was used to generate a new list that contained all peptides in the others, but where duplicates had been removed. The peptide identifications in the final list were visualized through a direct interaction network, defined to show all interactions between peptides that were of the following types: binding, expression, protein modification, and regulation. Some peptides were omitted from the network because the interactions were below maximum score/quality, which is defined as an interaction from manually curated sources in the database. The direct interaction network was exported, all interactions were manually checked, and uninteresting interactions were removed. These manual edits were incorporated into the interaction network. Subsequently, the peptide list identified for each of the treated samples was imported into PathwayStudio software (Ariadne Genomics, www.ariadnegenomics.com), and pathways were created directly from the ResNet database (Ariadne Genomics), which comprises both curated and canonical pathways. The peptide set was cross-checked with these pathways, and a pathway list was generated and ranked by the hypergeometric probability factor. All interactions were manually inspected and selected on the criterion of the highest number of proteins being involved in a linear pathway or sub-pathway, by superposition of the peptide set to a cartoon representation of the pathway reported.

Western blot analysis

Expression of phosphoproteins was measured by means of standard western immunoblots. Primary, polyclonal

antibodies, selected against the same phosphorylated epitopes as the corresponding target peptides on the microarrays, were anti-phospho(y1197)-EGFR (Abcam Plc., Cambridge, UK) and anti-phospho(y1248)-Neu (Santa Cruz Biotechnology, Inc., Santa Cruz, CA). Further, the monoclonal anti-EGFR antibody 425.3 [8], a polyclonal anti-ErbB2 antibody (R&D Systems Europe Ltd., Abingdon, UK), and a monoclonal anti- α -tubulin antibody (Calbiochem/Merck Biosciences Ltd., Nottingham, UK) were used.

Immunomagnetic cell separation

MOC-31 (IQ Corporation BV, Groningen, The Netherlands) is an IgG1 class antibody that binds to the EPCAM antigen, which is consistently expressed in most epithelial cells [9]. The high-affinity monoclonal antibody 9.2.27, which was originally developed against melanoma [10], recognizes an epitope on the high molecular weight melanoma-associated antigen and has also been shown to bind some subgroups of sarcoma, including osteosarcoma [11, 12]. The antibodies were conjugated to superparamagnetic particles coated with polyclonal sheep-antimouse IgG particles (Dynabeads; Dynal A.S., Oslo, Norway).

At the end of experimental incubations, cocultures of LNCaP cells with OHS cells and of LNCaP cells with differentiated mesenchymal stem cells were detached, and the resulting single cell suspensions were subjected to immunomagnetic target cell isolation, essentially as previously described [12–15]. The positive cell fractions were examined by light microscopy for the principal presence of cells with ≥ 5 immunobeads bound to their surface (bead rosettes). For the purpose of cell quantification and quality assurance, the immunosegregation procedure was repeated with 9.2.27-coated beads added to the negative cell fractions for isolation of OHS cells. The segregated cell fractions from LNCaP/OHS coculture samples (10^6 – 10^7 cells) contained 60–70% MOC-31-selected bead rosettes and $\sim 30\%$ 9.2.27-selected bead rosettes.

Results

Kinase activity evoked by osteoblastic and androgenic influence

Monocultured LNCaP cells were treated with OHS-conditioned medium to experimentally replicate the biological context of paracrine osteoblastic influence. Moreover, to simulate the complex processes involved in aberrant activation of the androgen signalling axis in prostate cancer [16, 17], the experimental setup included LNCaP cells treated with the synthetic androgen analog R1881 to

observe whether androgen receptor-mediated signalling pathways might differ from pathways activated by OHS-directed influence. Given the assumption that biologically relevant signalling events implicated in the metastatic phenotype require sustained activation, 48-h incubation times were used for the experimental LNCaP contexts.

From the tyrosine kinase microarray analysis, the substrate phosphorylation state generated by each LNCaP entity (paracrine, androgenic) was calculated with respect to baseline (untreated LNCaP cells). The identified substrates showed increases in phosphorylation level within a broad range (Table 1).

Each individual substrate phosphorylation signature was considered to represent a subset of the information flow through the globally activated signalling network of the particular LNCaP entity. Tools for analyzing interconnectivity of biological molecules have so far been used primarily to explore system-level gene expression data. However, such computational methods may also elucidate how kinase activity information is directed, and by using these algorithms, we assumed that phosphorylation events that appeared simultaneously might be interlinked and provide information about pathway connectivity [18, 19].

By applying these assumptions, the network interaction analysis omitted phosphorylated substrates that did not appear within any signalling pathway when defined by the interaction types delineated in “Materials and methods”. As illustrated in Fig. 1, the resulting network connectivity map indicated that signalling pathways involved in cell adhesion and motility as well as cell proliferation were activated in the paracrine LNCaP entity. Activation of similar but also completely unrelated proliferation pathways was observed in the androgenic entity. Interestingly, only the signalling pathway mediated by EGFR seemed to be activated by the influence of both osteoblastic cells and androgen treatment.

Since the technology for multiplex kinase activity profiling and computational tools for data analysis are at an early stage of development [20], no general consensus approach to data validation exists. We therefore performed conventional western immunoblotting for selected, individual phosphoproteins expressed by LNCaP cells [21, 22] that in addition are representative of signalling pathways and therapeutic targets under investigation in recently conducted trials of metastatic prostate cancer [23–26]. Markedly increased expression levels of EGFR phosphorylated on tyrosine 1197 and ERBB2 phosphorylated on tyrosine 1248 were found in both the paracrine and androgenic LNCaP entity (Fig. 2a). In contrast, EGFR was found to be non-phosphorylated following 48 h of incubation with either EGF or 10% FBS, whereas short-time EGF treatment caused transient increase in EGFR phosphorylation (Fig. 2b), which is in accordance with

Table 1 Peptide substrates with increased phosphorylation levels generated by the paracrine and androgenic LNCaP entities

Peptide substrate	Paracrine ^a	Androgenic
EGFR_y1197 ^b	1.84 ^c	1.94
EGFR_y1110	2.34	–
RB1_s807/s811	3.39	3.07
ERBB2_y877	1.67	–
ERBB2_y1248	1.56	–
CDK2_t14/y15	1.86	–
EPHB1_y778	2.11	–
IRS2_y919	5.56	–
JAK1_y1022/y1023	2.05	–
LAT_y200	3.49	–
LCK_y394	1.91	–
MET_y1230/y1234/y1235	1.57	–
MST1R_y1353	2.86	–
MST1R_y1356/y1360	3.38	–
PDPK1_y9	1.82	–
PDPK1_y373/y376	1.82	–
PTK2_y576/y577	1.39	–
PTK2B_y579/y580	1.21	–
RASA1_y460	1.63	–
RET_y1029	1.76	–
ZAP70_y492/y493	1.93	–
CREB1_y134/s133	–	1.47
ERBB4_y1284	–	3.31
GSK3B_y216	–	1.15
RAF1_s337/s338/y339/y340	–	1.14
CHRN1_y390	1.57	1.57
LTK_y772/y776/y777	–	1.20
DDR1_y792/y796/y797	1.20	–
MAPK10_t221/y223	1.32	–
MAPK12_t183/y185	–	–
PECAM1_y713	2.06	–
PRRX2_y214	1.48	–
ANXA1_y20/t23	2.41	–
CD79A_y182/y188	1.75	–
CTTN1_y477/y483	2.01	–
CTTN1_y499	2.13	–
ENO2_y43	1.85	–
EPHA2_y772	1.99	–
EPHA7_y608/y614	2.27	–
EPOR_y368	2.08	–
EPOR_y426	2.29	–
FER_y714	1.66	–
FES_y713	2.20	–
FGFR2_y769	1.77	–
FGFR3_y760	2.23	–
FRK_y387	2.02	–
LAT_y255	1.54	–

Table 1 continued

Peptide substrate	Paracrine ^a	Androgenic
NTRK2_y702/y706/y707	1.43	–
PDGFRB_y579/y581	3.87	–
PDGFRB_y716	1.78	–
PIK3R1_y607/s608	2.09	–
PXN_y31	1.88	–
PXN_y118	2.07	–
TEC_y519	1.68	–
PFKFB1_s33	–	1.36
PTPN11_y542	–	2.94
SYN1_s9	–	1.20

‘–’ denotes that the change in substrate phosphorylation level was not found to be significant ($P < 0.05$, Student's *t*-test)

^a Phosphopeptide values from the LNCaP paracrine 2 entity are given

^b For each substrate, position of phosphorylation sites within the protein is indicated

^c Fold changes (\log_2) relative to LNCaP baseline samples are listed

previously published data [21, 27, 28] and may indicate that functional signalling in prostate carcinoma cells under influence of osteoblastic cells is diverse and complex [21].

EGFR/ERBB2 phosphorylation in supplementary biological models

We used additional experimental setups as biological controls for the osteoblastic influence on prostate carcinoma cells; the first to model the direct interaction between LNCaP and OHS cells and the second to provide cells that might be more representative of physiological osteoblasts.

Culturing LNCaP cells with OHS cells caused substantial change in LNCaP morphology. The spindle-shaped feature of monocultured LNCaP cells (Fig. 3a) was rapidly lost upon direct contact with the OHS cells. In coculture, both cell types appeared rounded, although cytoplasmic processes were still apparent on LNCaP cells (Fig. 3b). Cellular morphology of the OHS cells (Fig. 3c), however, remained independent of the culturing conditions.

Non-hematopoietic stem cells in the bone marrow are capable of differentiating into a variety of tissue entities, including osteogenic cells of bone tissue [29]. Incubation of mononuclear cells isolated from adult, human bone marrow with mesenchymal stem cell-stimulating medium followed by osteogenic differentiation medium [6, 7] gave rise to cells with osteoblastic characteristics, for example mineral deposition (Fig. 3d, e) and alkaline phosphatase-secreting activity (not shown), to be used in coculture with LNCaP cells (Fig. 3f).

Fig. 1 Interconnected signalling pathways activated in LNCaP cells by influence of osteoblastic cells or androgen treatment. Two pathway visualization systems were applied to the data set (Table 1), which resulted in almost identical network connectivity maps. The substrate annotations are derived from gene name entries in SwissProt. The lines connecting nodes represent interactions of the following types: binding, expression, protein modification, and regulation. Yellow and blue nodes symbolize substrates phosphorylated by the paracrine and androgenic LNCaP entities, respectively. (Color figure online)

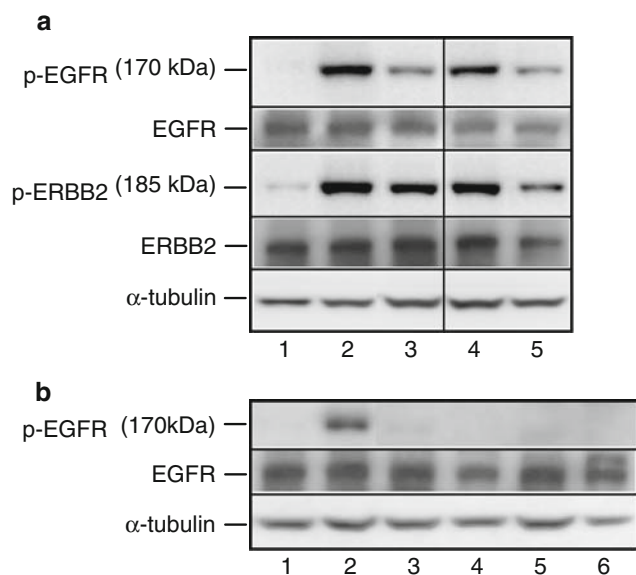
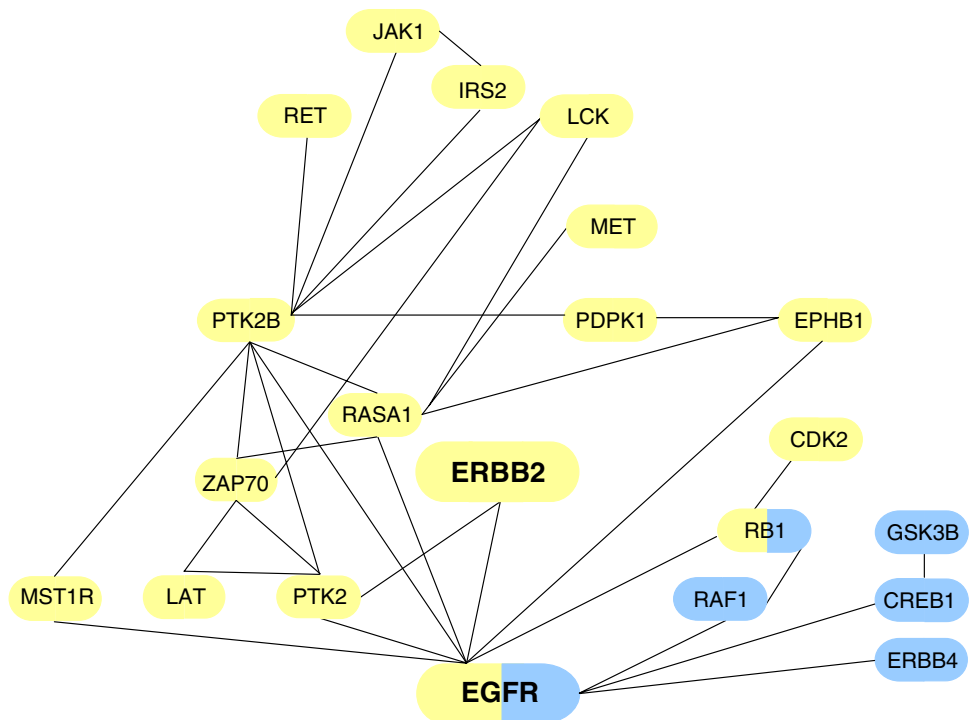


Fig. 2 Expression of phospho(y1197)-EGFR and phospho(y1248)-ERBB2 in LNCaP cells. **a** Cells were harvested after 48 h of experimental treatments. Lanes: (1) baseline, (2) paracrine entity, (3) androgenic entity, (4) cells immunosegregated from coculture with OHS cells, and (5) cells immunosegregated from coculture with osteoblastic cells differentiated from human mesenchymal stem cells. **b** Cells were treated with 50 ng/ml EGF or 10% FBS. Lanes: (1) baseline, (2) EGF for 15 min, (3) EGF for 24 h, (4) EGF for 48 h, (5) FBS for 48 h, and (6) 48 h untreated

We applied the immunomagnetic cell separation method for selective isolation of LNCaP cells from the cocultured osteoblastic cells (Fig. 3g) and subjected the isolated

carcinoma cells to western immunoblotting. Increased expression levels of EGFR phosphorylated on tyrosine 1197 and ERBB2 phosphorylated on tyrosine 1248 were found in LNCaP cells from coculture with both OHS cells and osteoblastic cells that had been differentiated from human mesenchymal stem cells (Fig. 2a).

Supplementary kinase activity profiling

To assess robustness of the biological models in the analytical application of multiplex kinase activity profiling, three experimental setups were exploited.

Kinase activity was compared in LNCaP cells treated with medium conditioned by OHS cells only and with the addition of medium obtained from standard LNCaP monocultures (LNCaP entities denoted paracrine 1 and paracrine 2, respectively). In this experimental setting, the profile of phosphorylated substrates generated by the LNCaP cells did not significantly change by the addition of medium conditioned by LNCaP monocultures (compare Table 1 with Table 2), strongly indicating that the signalling pathways of biological importance in the paracrine setting were activated by factors secreted by the OHS cells exclusively.

Kinase activity induced in LNCaP cells by direct contact with OHS cells was analyzed following carcinoma cell isolation from cocultures. The substrate phosphorylation state generated by such LNCaP cells overlapped the phosphopeptide signatures identified in the paracrine LNCaP entities (Table 2).

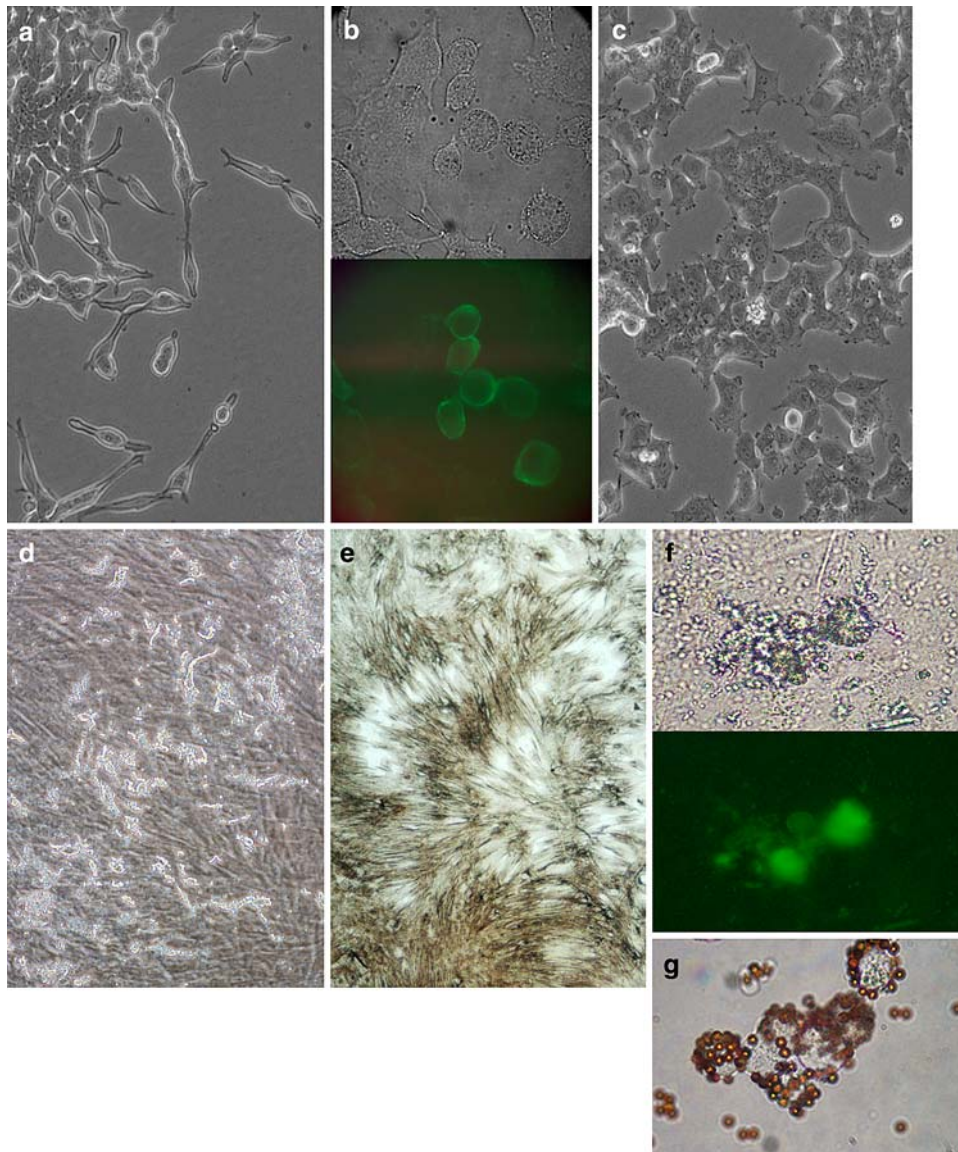


Fig. 3 Morphologic characteristics of LNCaP and osteoblastic cells. **a** Monocultured LNCaP cells, untreated (magnification $\times 300$). **b** LNCaP/OHS coculture, at start of experimental incubation (magnification $\times 400$). The fluorescent cells are stained with FITC-conjugated 9.2.27 antibody; accordingly, representing OHS cells. **c** Monocultured OHS cells (magnification $\times 300$). **d** Mononuclear cells isolated from adult human bone marrow were cultured in mesenchymal stem cell-stimulating medium followed by osteogenic differentiation medium for 20 days to obtain in vitro-differentiated osteoblasts (magnification $\times 300$). **e** The differentiated osteoblasts were fixed in ice-cold methanol for 30 min, incubated in dark with 5% silver nitrate

solution for 30 min, and washed thoroughly with deionized water and then with 5% Na_2CO_3 and 0.2% formaldehyde, according to the von Kossa staining protocol for calcium salts. Mineral deposits are visualized as dark brown stain (magnification $\times 300$). **f** Coculture of LNCaP cells and in vitro-differentiated osteoblasts, at start of experimental incubation (magnification $\times 400$). The fluorescent cells are stained with FITC-conjugated MOC-31 antibody; accordingly, representing LNCaP cells. **g** Suspended cells binding MOC-31-coated beads; accordingly, representing LNCaP cells, after immunomagnetic selection from LNCaP/OHS coculture (magnification $\times 400$) (Color figure online)

We have previously evaluated the immunomagnetic cell separation method for selective isolation of target cells and demonstrated that target cell populations are highly enriched [12–15]. But because kinase activity profiling might be sensitive to the possible presence of contaminants, we screened for phosphopeptide signatures generated by

immunoselected as well as monocultured OHS cells. Notably, the resulting substrate phosphorylation patterns were closely similar for the two conditions (Table 3) and also clearly distinguishable from those generated by the LNCaP entities, which argues against significant contamination of OHS cells in MOC-31-positive LNCaP cell isolates.

Table 2 Peptide substrates with increased phosphorylation levels generated by the LNCaP paracrine 1 entity and LNCaP cells immunosegregated from coculture with OHS cells (co-LNCaP)

Peptide substrate	Paracrine 1	Co-LNCaP
EGFR_y1197 ^a	1.50 ^b	2.16
EGFR_y1110	–	1.86
RB1_s807/s811	3.72	–
ERBB2_y877	1.06	–
ERBB2_y1248	–	–
CDK2_t14/y15	1.47	–
EPHB1_y778	1.78	–
IRS2_y919	4.88	–
JAK1_y1022/y1023	1.81	–
LAT_y200	2.70	2.94
LCK_y394	1.38	–
MET_y1230/y1234/y1235	1.08	–
MST1R_y1353	2.29	2.20
MST1R_y1356/y1360	2.60	2.01
PDPK1_y9	1.50	–
PDPK1_y373/y376	1.50	–
PTK2_y576/y577	1.33	1.03
PTK2B_y579/y580	0.85	0.85
RASA1_y460	1.27	0.96
RET_y1029	1.24	–
ZAP70_y492/y493	1.68	1.21
CREB1_y134/s133	–	–
ERBB4_y1284	–	–
GSK3B_y216	–	–
RAF1_s337/s338/y339/y340	–	–
CHRNB1_y390	1.00	0.95
LTK_y772/y776/y777	–	0.96
DDR1_y792/y796/y797	1.11	0.91
MAPK10_t221/y223	1.28	0.98
MAPK12_t183/y185	–	2.23
PECAM1_y713	1.51	0.87
PRRX2_y214	1.23	0.93
ANXA1_y20/t23	1.93	–
CD79A_y182/y188	1.39	–
CTTN1_y477/y483	1.45	–
CTTN1_y499	1.64	–
ENO2_y43	1.43	–
EPHA2_y772	1.60	–
EPHA7_y608/y614	1.64	–
EPOR_y368	1.67	–
EPOR_y426	1.77	–
FER_y714	1.36	–
FES_y713	1.82	–
FGFR2_y769	1.28	–
FGFR3_y760	1.63	–
FRK_y387	1.47	–
LAT_y255	1.13	–

Table 2 continued

Peptide substrate	Paracrine 1	Co-LNCaP
NTRK2_y702/y706/y707	1.08	–
PDGFRB_y579/y581	3.52	–
PDGFRB_y716	1.59	–
PIK3R1_y607/s608	1.59	–
PXN_y31	1.44	–
PXN_y118	1.61	–
TEC_y519	1.23	–
PFKFB1_s33	–	–
PTPN11_y542	–	–
SYN1_s9	–	–

‘–’ denotes that the change in substrate phosphorylation level was not found to be significant ($P < 0.05$, Student’s *t*-test)

^a For each substrate, position of phosphorylation sites within the protein is indicated

^b Fold changes (\log_2) relative to LNCaP baseline samples are listed

Discussion

Given the considerable health care challenges generated by the prevalence of metastatic bone disease in prostate cancer, the apparent shortage of adequate experimental models to study this aspect of disease progression is striking. From a clinical point of view, prostate cancer metastasis to bone is a lengthy and complex disease process, which makes it difficult to establish adequate experimental models to recreate all steps involved. As highlighted in a comprehensive review [30], most of the experimental systems examining this phenomenon are based on rodent models. Importantly, the model systems used in this study exclusively utilize cell types of human origin, but caution should be exercised regarding any compelling conclusion based on specific in vitro experimental conditions, which can not reflect an intact microenvironmental in vivo setting. Nevertheless, given that regulatory mechanisms implicated in the metastatic phenotype are evoked when carcinoma cells settle within an osteoblastic microenvironment, the combination of our experimental models and analytical technology may provide relevant information about functional signalling networks that facilitate this biological process, and in addition may enable the identification of new targets for therapeutic intervention.

The OHS cell line was originally established from a patient with aggressive osteosarcoma [4], and it might be argued that it is not representative for physiological osteoblasts. However, formation of osteosclerotic (i.e., osteoblastic) lesions following intratibial OHS cell inoculation has previously been demonstrated by radiographic, scintigraphic, and morphologic assessments [31].

Table 3 Phosphopeptides generated by monocultured OHS cells (mono-OHS) and OHS cells immunosegregated from coculture with LNCaP cells (co-OHS)

Peptide substrates	Mono-OHS	Co-OHS
ANXA1_y20/t23 ^a	-1.56 ^b	-2.84
CD79A_y182/y188	-3.70	-4.54
CDK2_t14/y15	-0.94	-1.32
CHRN1_y390	-0.49	-0.64
CREB1_y134/s133	-0.81	-1.29
CTTN1_y477/y483	-0.86	-1.43
CTTN1_y499	-1.56	-2.00
DCX_y112/s116	-0.92	-0.86
DDR1_y792/y796/y797	-0.40	-0.79
EFS_y253	-0.86	-1.18
ENO2_y43	-3.72	-4.76
EPHA2_y772	-1.43	-2.32
EPHA7_y608/y614	-1.60	-2.18
EPHB1_y778	-1.60	-2.40
EPOR_y368	-0.94	-1.40
EPOR_y426	-0.94	-2.32
ERBB2_y877	-1.74	-2.18
ERBB2_y1248	-1.22	-2.06
FER_y714	-1.15	-1.64
FES_y713	-1.03	-2.25
FGFR2_y769	-1.89	-2.06
FGFR3_y760	-1.56	-0.89
FRK_y387	-1.22	-1.84
GSK3B_y216	-1.25	-0.97
JAK1_y1022/y1023	-2.18	-2.32
KRT6E_s59	-0.92	-1.47
LAT_y255	-1.29	-2.40
LCK_y394	-1.60	-2.25
LTK_y772/y776/y777	-0.86	-1.03
MAPK10_t221/y223	-0.94	-0.62
MET_y1230/y1234/y1235	-0.89	-1.12
NCF1_s315/s320	-0.67	-0.74
NTRK1_y496	-1.94	-2.84
NTRK2_y702/y706/y707	-2.94	-3.84
PDGFRB_y579/y581	-4.32	-3.84
PDGFRB_y716	-1.51	-1.00
PDGFRB_y771/y775/y778	-1.00	-1.25
PDPK1_y9	-1.40	-1.25
PDPK1_y373/y376	-1.36	-2.00
PECAM1_y713	-1.06	-1.47
PFKFB1_s33	-0.79	-0.92
PIK3R1_y607/s608	-1.25	-2.56
PLCG1_y771	-4.57	-4.64
PRRX2_y214	-1.36	-1.25
PTK2_y576/y577	-1.15	-0.81
PTK2B_y579/y580	-0.71	-1.51
PXN_y31	-1.25	-1.84

Table 3 continued

Peptide substrates	Mono-OHS	Co-OHS
PXN_y118	-1.03	-1.29
RAF1_s337/s338/y339/y340	-0.89	-1.18
RASA1_y460	-1.36	-1.09
RET_y1029	-1.22	-2.18
SYN1_s9	-1.06	-1.40
TEC_y519	-1.51	-1.64
TYRO3_y686	-1.40	-1.29
ZAP70_y492/y493	-1.56	-1.74

^a For each substrate, position of phosphorylation sites within the protein is indicated

^b Fold changes (\log_2) relative to LNCaP baseline sample are listed

Kinase activity microarrays represent an emerging technology that may become a powerful tool for signal transduction profiling of biological samples. A key advantage of this technology lies in its ability to provide signalling pathway maps that indicate the state of information flow through intracellular networks. Essentially, the peptide substrate array technology determines kinase activity profiles, which is different from but complementary to mass spectrometry, which identifies phosphorylated proteins that represent the end products of kinase activity. An additional advantage of kinase activity microarrays is their robustness with small sample quantities, typically 2–5 μg total protein being sufficient for Tyrosine Kinase PamChip[®] Array analysis, in contrast to several milligrams of total protein usually required for mass spectrometry analysis.

Of note, the phosphopeptide signatures generated by the LNCaP paracrine 1 and paracrine 2 entities and by OHS cells cultured alone or with LNCaP cells, respectively, were essentially identical. This indicates low variation between biologically similar samples and a high degree of reproducibility using this peptide substrate array technology.

Computational methods for mapping of phosphorylation networks are being developed, and reconstruction of EGFR-mediated pathways have been used to test applicability of such models [32]. Correct network definition may be restricted by the influence of contextual factors, such as subcellular compartmentalization or temporal expression of the proteins involved [33]. Therefore, to predict pathway connectivity with sufficient accuracy, the use of more than one computational technique on the dataset of interest has been recommended [20]. In this study, two different pathway visualization systems identified signalling networks that were essentially identical.

The signalling pathway mediated by EGFR was found to be activated in androgen-sensitive LNCaP cells under the influence of either osteoblastic cells or androgen treatment.

Analysis by conventional western immunoblotting also revealed that ERBB2-dependent signalling was activated in both the paracrine and androgenic LNCaP entities, although the latter did not seem to generate this particular substrate phosphorylation profile on the microarray analysis. The kinase activity and protein expression data together strongly indicates a central involvement of both signalling pathways in the interaction between prostate carcinoma cells and osteoblasts.

Additionally, given that EGFR and ERBB2 are phosphorylated in androgen-sensitive LNCaP cells upon influence of osteoblasts, a functional androgen signalling axis [16, 17] may appear to be permissive for activity of these particular pathways in prostate cancer. Not the less, EGFR phosphorylation resulting from short-term incubation with bone stromal conditioned medium (containing 6% FBS) and suppression of *in vivo* bone metastasis formation following targeted inhibition of EGFR-dependent signalling have been demonstrated in androgen-independent prostate carcinoma PC3 cells [34]. Separately, EGFR and ERBB2 may facilitate androgen receptor-driven activity in prostate cancer at the level of target gene transcription in the absence or at low concentrations of androgens [35, 36]. Yet, androgen receptor pathway genes, identified by system-level analysis of gene expression in primary tumor specimens from therapy-naïve prostate cancer patients, were reported to be down-regulated, with a few exceptions, in lymph-node metastases from the patients [37]. This finding further supports the assumption that the regulatory control by the androgen receptor on carcinoma cell biology is lost in the process of prostate cancer metastasis, even in the presence of activating receptor ligands.

Of importance, our experimental data suggests that targeted inhibition of the signalling pathways directed by EGFR or ERBB2 may simultaneously ablate androgen-driven proliferation of prostate carcinoma cells and the survival responses within an osteoblastic microenvironment. It equally provides a biological rationale for the use of EGFR or ERBB2 inhibition in systemic prevention or treatment of metastatic prostate cancer in the androgen-sensitive stage of the disease. Intriguingly, the therapeutic concept of EGFR or ERBB2 inhibition in hormone-refractory prostate cancer has recently been evaluated; however, in initial studies addressing the use of single-agent therapies in patients with androgen-resistant disease, neither receptor-blocking antibodies nor small-molecular tyrosine kinase inhibitors showed clinically significant activity [23–26].

Of note, the tumor suppressor PTEN phosphatase is frequently found to be functionally inactivated in prostate cancer, leading to increased activity of AKT kinase signalling independent of the up-stream EGFR/ERBB2 and to insensitivity to EGFR/ERBB2 inhibitors [16, 38–40]. The

LNCaP cells are deficient in PTEN [41, 42], questioning the relevance of extrapolating from our data that EGFR/ERBB2 signalling is involved in the maintenance of osteoblast-induced survival responses in prostate cancer. On the other hand, development of androgen independence and more advanced disease is associated with increased PTEN loss [43–45], favoring therapeutic EGFR/ERBB2 inhibition in the initial, androgen-sensitive stage of the disease. Thus, if exploitable in patients with therapy-naïve prostate cancer, inhibitory EGFR/ERBB2 targeting might be incorporated into treatment schedules with a potential reduction in the alternative requirement of long-term androgen depletion, a reduction in related side effects, and, intriguingly, the potential for an improvement in patient survival.

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Conflict of interest statements A. H. Ree, Å. Bratland, P. J. Boender, and R. Ruijtenbeek are owners of a patent for diagnostic application of the Tyrosine Kinase PamChip® Array technology in prostate cancer (WIPO no. WO/2008/125633). P. J. Boender and R. Ruijtenbeek are employees of PamGene International BV.

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