

## Cardiopulmonary exercise testing in thalassemia

John-Paul Carpenter · Dudley J. Pennell

Received: 9 November 2009 / Accepted: 9 November 2009 / Published online: 24 November 2009  
© Springer Science+Business Media, B.V. 2009

In this issue of International Journal of Cardiac Imaging, researchers from the Onassis Cardiac Surgery Centre and Agia Sophia Hospital in Athens report their findings of cardiopulmonary exercise testing (CPET) in relation to cardiovascular magnetic resonance (CMR) in patients with thalassemia major (TM) [1]. In support of previously available evidence, they found raised cardiac iron and low left ventricular ejection fraction in patients with heart failure. In addition, CPET indices (VO<sub>2</sub>max, anaerobic threshold (AT), metabolic equivalents (METs) and exercise duration) were not only lower in TM patients with heart failure but were closely correlated with iron loading, measured by CMR T2\*.

Inherited hemoglobinopathies affect millions of patients worldwide, mostly due to the thalassemia syndromes and sickle cell disease [2]. TM patients have a particularly severe anemia due to the absence or severe reduction in beta globin chains, and regular blood transfusions are required from soon after birth to prevent fatal complications of the anemia. Although transfusion prevents these problems, the

trade-off comes at the expense of iron overload. Each unit of blood contains ~250 mg iron and additional body iron loading comes from overabsorption of iron from the gastro-intestinal tract due to hepcidin suppression. Iron accumulates in the tissues and organs of the body, predominantly to start with in the liver, and without chelation therapy, progressive cardiac iron overload leads to arrhythmias, cardiac failure and death before the age of 20. The introduction of iron-chelating agents in the 1970s has changed the natural history of the disease and survival has dramatically improved [3]. However, myocardial siderosis leading to impaired cardiac function remains the leading cause of mortality in these patients.

Understanding the pathophysiology of iron-induced cardiac dysfunction is key to the management of heart failure in these patients. When the iron transporting capacity of transferrin is exceeded, non-transferrin bound iron appears in the plasma. This labile iron is associated with the development of free radicals which cause cellular damage by affecting membrane lipids and proteins. In cardiac myocytes, the mitochondrial respiratory chain is affected and cellular dysfunction occurs, leading to impaired myocardial contractility and subsequently heart failure [4]. Unfortunately, the onset of decompensated heart failure from iron-overload cardiomyopathy is unpredictable and a long period of asymptomatic severe iron loading may be present before any clear depression of left ventricular ejection fraction is

---

Editorial comment on the article of Mavrogeni et al.  
(doi:10.1007/s10554-009-9491-9).

---

J.-P. Carpenter · D. J. Pennell (✉)  
Cardiovascular Magnetic Resonance Unit, Royal  
Brompton Hospital, Sydney Street, London SW3 6NP,  
UK  
e-mail: d.pennell@imperial.ac.uk

apparent. As a consequence, echocardiographic parameters can remain normal despite severe myocardial siderosis but once heart failure occurs, it carries a poor prognosis [5]. Early detection of cardiac iron therefore becomes important but, until recently, non-invasive assessment of cardiac iron loading has been difficult. The historical methods used, namely serum ferritin and biopsy measurements of liver iron, have proven to be poor predictors of myocardial iron content and myocardial biopsy is not only invasive but may be misleading due to patchy iron deposition [6, 7].

From our unit, we have implemented a CMR technique for the measurement of cardiac iron. This was first described in 2001 and involves measuring  $T2^*$  which is a magnetic resonance parameter [6].  $T2^*$  is significantly affected by the magnetic field perturbation caused by particulate iron in the myocardium and therefore  $T2^*$  falls as the amount of iron increases. These findings which have been confirmed in an animal model of iron overload [8]. The normal human  $T2^*$  value (measured in milliseconds) is greater than 20 ms. When  $T2^*$  falls to below 10 ms, there is a progressive and significant fall in left ventricular ejection fraction (LV EF). We have also shown that  $T2^*$  is a strong prognostic indicator for the development of heart failure [9]. With intensive chelation therapy, both  $T2^*$  and LV EF improve, showing that iron-related cardiomyopathy can be reversed [10]. This has been confirmed in trials of the iron-chelating agents deferoxamine and deferoxamine [11–13] and improvements in cardiac mortality have been demonstrated related to the use of cardiac iron monitoring [14]. There remain some unanswered questions. First, it is not clear why some patients with high cardiac iron develop heart failure and others do not; second, whether further testing in patients with high cardiac iron could identify those most at risk; and third, why some patients have high cardiac iron with low liver iron and vice versa. The latter may be related to differences in iron kinetics and iron-handling genes but the first two issues are more challenging.

The Athens group have a longstanding interest not only in the assessment of iron loading but also of functional status in TM patients which has led to this latest research on CPET. Initial studies on the use of CMR in cardiac iron overload using older signal intensity ratio (SIR) and  $T2$  CMR techniques, showed that myocardial iron was inversely correlated with

both SIR and  $T2$  [15, 16]. Indeed  $T2$  has now become easier to do and may have a clinical role in the future [17]. An evaluation of myocardial and hepatic iron in patients who had undergone successful bone-marrow transplantation revealed that  $T2^*$  was more reliable than serum ferritin in monitoring iron loading and that the myocardial iron (measured using  $T2^*$ ) was the same as an age-matched normal population [18]. The Athens group went on to assess respiratory function in patients with thalassemia and iron overload. A restrictive ventilatory pattern on pulmonary function testing (i.e. low predicted total lung capacity (TLC)) was noted to be a common abnormality in TM patients. Although it was postulated that this might be due to iron deposition in the respiratory system, no correlation was found between reduced TLC and hepatic iron loading [19]. They have also described impaired oxygen kinetics and limited functional capacity of peripheral muscles in TM patients during cardiopulmonary exercise testing [20, 21]. This work has suggested that part of the exercise limitation in TM stems from a slower rate of high energy phosphate production and utilization with reduced oxidative capacity of myocytes.

Cardiopulmonary exercise testing (CPET) is a useful clinical technique for the assessment of exercise capacity and prognosis in patients with heart failure and other cardiovascular conditions [22]. It is non-invasive and provides information which not only combines the exercise-related responses of the lungs, heart, and skeletal muscle but also takes account of the effects of anaemia which can alter arterial or mixed venous blood oxygen levels. CPET involves an exercise protocol using a bicycle ergometer or a treadmill and can have an advantage over conventional techniques which rely mainly on the assessment of resting cardiac function. Many different protocols exist which are tailored to the patient's functional capacity. Workload is increased in stages, either by increasing the braking load on a bicycle or the slope of the treadmill ramp. A mouthpiece is used for ventilation and respiratory gas analysis with a non-rebreathe valve to prevent mixing of inspired and expired air. Respiratory volumes are calculated by integrating the air-flow/time curves. Gas analysers measure inspiratory and expiratory oxygen and carbon dioxide.

Oxygen uptake ( $VO_2$ ) is dependent on the product of the cardiac output and the difference between arterial oxygen and mixed venous oxygen content

( $VO_2 = CO \times (\text{arterial oxygen content} - \text{mixed venous oxygen content})$ ). It is usually indexed for weight and measured in mL  $O_2$ /kg/min. In healthy subjects,  $VO_2$  increases linearly with increasing exercise until it reaches a plateau at near-maximal exertion ( $VO_{2max}$ ). In patients with heart failure, the level of exercise may be limited by symptoms before  $VO_{2max}$  is reached and therefore, peak  $VO_2$  ( $PVO_2$ ) is used instead as an estimation of maximum oxygen uptake. With worsening exercise tolerance, peak  $VO_2$  falls. The ventilatory anaerobic threshold (AT) is a useful indicator of exercise capacity. During exercise, muscle metabolism is initially aerobic with a linear increase in ventilation and carbon dioxide production ( $VCO_2$ ) as  $VO_2$  increases. When the supply of oxygen cannot keep up with the demand for energy required by skeletal muscle, the adenosine triphosphate (ATP) production needs to be supplemented by anaerobic metabolism and hence the anaerobic threshold is reached. This leads to the production of lactic acid within the muscles which causes a rise in blood lactate and a fall in bicarbonate levels. As the switch to anaerobic metabolism results in increased  $CO_2$  production, the anaerobic threshold can be estimated by analysing the relative rate of production of  $CO_2$  compared to the oxygen uptake. There are a number of methods for doing this, all of which are confounded by considerable inter and intraobserver variability [23].

To be confident that  $PVO_2$  is a good estimate of  $VO_{2max}$ , it is important to ensure that a test is maximal. This can be achieved by observing a rise in heart rate and blood pressure with exercise together with ensuring that the anaerobic threshold has occurred at 50–70% of  $PVO_2$ . However, a proportion of patients with heart failure will not reach the anaerobic threshold and  $PVO_2$  results need to be interpreted with this in mind. The  $VE/VCO_2$  slope, which represents the relationship between expired minute ventilation ( $VE$ ) and carbon dioxide production is an alternative, additional prognostic indicator. Based on the results of a number of trials, a risk stratification algorithm has been suggested for use in heart failure [24]. Patients with  $PVO_2 > 18$  ml/kg/min have a good long term outcome, whereas those with  $PVO_2$  of below 10 ml/kg/min have a high risk of cardiac events and death. With  $PVO_2$  between 10 and 18 ml/kg/min, there is an intermediate risk unless the  $VE/VCO_2$  slope is  $\geq 35$ , indicating a high risk.

In thalassemia patients with heart failure and poor ventricular function, exercise capacity may be limited by a number of factors. The cardiac output will be unable to increase sufficiently to provide skeletal muscles with adequate perfusion and therefore anaerobic metabolism will occur early. As cardiac iron impairs mitochondrial respiratory function, a higher level of cardiac siderosis is likely to have more of an impact on cellular metabolism. The degree of anemia, albeit mild, will also affect the oxygen supply to the muscles due to reduced arterial oxygen content.

However, CPET has its limitations. CPET can be a valuable tool in expert hands, but the technique is neither simple nor widely available. It requires expert supervision and a significant amount of training. While equipment costs are undoubtedly less than CMR, the reporting and interpretation of CPET is not easy. It is however possible that CPET might have the potential to determine which patients with a low  $T2^*$  are most likely to develop cardiac complications. At present, although patients with  $T2^*$  less than 10 ms are known to have a greatly increased relative risk, it is impossible to predict which of these patients will progress to left ventricular dysfunction and heart failure.

Further research is needed to establish whether CPET is an independent risk stratifier. Thus, we do not yet know if CPET adds anything to the  $T2^*$  or the ejection fraction. The fact that those patients with low cardiac  $T2^*$  have poor CPET indices provides further evidence to support cardiac  $T2^*$  as the current method of choice for monitoring cardiac iron loading in patients with transfusion-dependent thalassemia major.

## References

1. Mavrogeni S, Gotsis E, Verganelakis D, Berdoussis E, Dritsas A, Kolovou G, Toulas P, Ladis V (2009) Effect of iron overload on exercise capacity in thalassaemic patients with heart failure. *Int J Cardiovasc Imaging*. doi:[10.1007/s10554-009-9491-9](https://doi.org/10.1007/s10554-009-9491-9)
2. Modell B, Darlison M (2008) Global epidemiology of haemoglobin disorders and derived service indicators. *Bull World Health Organ* 86(6):480–487
3. Modell B, Khan M, Darlison M, Westwood MA, Ingram D, Pennell DJ (2008) Improved survival of thalassaemia major in the UK and relation to  $T2^*$  cardiovascular magnetic resonance. *J Cardiovasc Magn Reson* 10(1):42

4. Herschko C, Link G, Cabantchik I (1998) Pathophysiology of iron overload. *Ann NY Acad Sci* 850:191–201
5. Felker GM, Thompson RE, Hare JM, Hruban RH, Clemetson DE, Howard DL, Baughman KL, Kasper EK (2000) Underlying causes and long-term survival in patients with initially unexplained cardiomyopathy. *N Engl J Med* 342:1077–1084
6. Anderson LJ, Holden S, Davis B, Prescott E, Charrier CC, Bunce NH, Firmin DN, Porter J, Walker JM, Pennell DJ (2001) Cardiovascular T2-star (T2\*) magnetic resonance for the early diagnosis of myocardial iron overload. *Eur Heart J* 22:2171–2179
7. Fitchett DJ, Coltart DJ, Littler WA, Leyland MJ, Trueman T, Gozzard DI, Peters TJ (1980) Cardiac involvement in secondary haemochromatosis: a catheter biopsy study and analysis of myocardium. *Cardiovasc Res* 14(12):719–724
8. Wood JC, Otto-Duessel M, Aguilar M, Nick H, Nelson MD, Coates TD, Pollack H, Moats R (2005) Cardiac iron determines cardiac T2\*, T2 and T1 in the gerbil model of iron cardiomyopathy. *Circulation* 112(4):272–280
9. Kirk P, Roughton M, Porter JB, Walker JM, Tanner MA, Patel J, Wu D, Taylor J, Westwood MA, Anderson LJ, Pennell DJ (2009) Cardiac T2\* magnetic resonance for prediction of cardiac complications in thalassemia major. *Circulation* Oct 2 [Epub ahead of print]
10. Anderson LJ, Westwood MA, Holden S, Davis B, Prescott E, Wonke B, Porter JB, Walker JM, Pennell DJ (2004) Myocardial iron clearance during reversal of siderotic cardiomyopathy with intravenous desferrioxamine: a prospective study using T2\* cardiovascular magnetic resonance. *Br J Haematol* 127:348–355
11. Tanner MA, Galanello R, Dessi C, Smith GC, Westwood MA, Agus A, Roughton M, Assomull R, Nair V, Walker JM, Pennell DJ (2007) A randomized, placebo-controlled, double blind trial of the effect of combined therapy with deferoxamine and deferiprone on myocardial iron in thalassemia major using cardiovascular magnetic resonance. *Circulation* 115:1876–1884
12. Pennell DJ, Berdoukas V, Karagiorga M, Ladis V, Piga A, Aessopos A, Gotsis ED, Tanner MA, Smith GC, Westwood MA, Wonke B, Galanello R (2006) Randomized controlled trial of deferiprone or deferoxamine in beta-thalassemia major patients with asymptomatic myocardial siderosis. *Blood* 107:3738–3744
13. Tanner MA, Galanello R, Dessi C, Smith GC, Westwood MA, Agus A, Pibiri M, Nair SV, Walker JM, Pennell DJ (2008) Combined chelation therapy in thalassemia major for the treatment of severe myocardial siderosis with left ventricular dysfunction. *J Cardiovasc Magn Reson* 10:12
14. Modell B, Khan M, Darlison M, Westwood MA, Ingram D, Pennell DJ (2008) Improved survival of thalassaemia major in the UK and relation to T2\* cardiovascular magnetic resonance. *J Cardiovasc Magn Reson* 10:42
15. Mavrogeni SI, Gotsis ED, Markussis V, Tsekos N, Politis C, Vretou E, Kremastinos D (1998) T2 relaxation time study of iron overload in b-thalassemia. *MAGMA* 6(1):7–12
16. Mavrogeni SI, Markussis V, Kaklamanis L, Tsiapras D, Paraskevaidis I, Karavolias G, Karagiorga M, Douskou M, Cokkinos DV, Kremastinos DT (2005) A comparison of magnetic resonance imaging and cardiac biopsy in the evaluation of heart iron in patients with beta-thalassemia major. *Eur J Haematol* 75:241–247
17. He T, Kirk P, Firmin DN, Lam WM, Chu WC, Au WY, Chan GC, Tan RS, Ng I, Biceroglu S, Aydinok Y, Fogel MA, Cohen AR, Pennell DJ (2008) Multi-center transferability of a breath-hold T2 technique for myocardial iron assessment. *J Cardiovasc Magn Reson* 10:11
18. Mavrogeni S, Gotsis ED, Berdousi E, Ladis V, Verganelakis K, Toulas P, Cokkinos DV (2007) Myocardial and hepatic T2\* magnetic resonance evaluation in ex-thalassaemic patients after bone-marrow transplantation. *Int J Cardiovasc Imaging* 23(6):739–745
19. Dimopoulou I, Kremastinos DT, Maris TG, Mavrogeni S, Tzelepis GE (1999) Respiratory function in patients with thalassaemia and iron overload. *Eur Respir J* 13(3):602–605
20. Vasileiadis I, Roditis P, Dimopoulos S, Ladis V, Pangalis G, Aessopos A, Nanas S (2009) Impaired oxygen kinetics in beta-thalassaemia major patients. *Acta Physiol (Oxf)* 196(3):357–363
21. Nanas S, Vasileiadis I, Dimopoulos S, Sakellariou D, Kapsimalakou S, Papazachou O, Tasoulis A, Ladis V, Pangalis G, Aessopos A (2009) New insights into the exercise tolerance of beta-thalassaemia major patients. *Scand J Med Sci Sports* 19(1):96–102
22. Albouaini K, Eged M, Alahmar A, Wright DJ (2007) Cardiopulmonary exercise testing and its application. *Heart* 93:1285–1292
23. Yeh MP, Gardner RM, Adams TD, Yanowitz FG, Crapo RO (1983) “Anaerobic threshold”: problems of determination and validation. *J Appl Physiol* 55(4):1178–1186
24. Corra U, Mezzani A, Bosimini E, Giannuzzi P (2004) Cardiopulmonary exercise testing and prognosis in chronic heart failure: a prognosticating algorithm for the individual patient. *Chest* 126(3):942–950