#### **ORIGINAL PAPER**



# Large-scale spatial patterns of riverine communities: niche versus geographical distance

Cássia Rocha Pompeu<sup>1</sup> · Francisco J. Peñas · José Barquín ·

Received: 8 August 2021 / Revised: 11 November 2022 / Accepted: 15 November 2022 / Published online: 27 November 2022 © The Author(s) 2022

#### Abstract

Freshwater ecosystems are among the most threatened ecosystems on Earth. Effective conservation strategies are essential to reverse this trend and should be based on sound knowledge of biodiversity patterns and the main drivers structuring them. In this study, we investigated the role of environmental and dispersal-connectivity controls on freshwater diatom and fish communities' variability. We used 441 biological samples obtained from Spanish biomonitoring datasets, which cover a highly variable environmental gradient across the national river network. We compared the taxonomic and trait-based spatial dependency of the two biotic groups using distance-decay relationships and variation partitioning with spatially constrained randomisations. Our findings showed that most of the diatoms and fish biological variation was attributed to pure spatial and spatially structured environmental variation. Compared to diatoms, fish community composition presented a stronger spatial dependency, likely because of their weaker dispersal ability. In addition, broad-scale environmental characteristics showed a higher predictive capacity for fish assemblages' variation. Trait-based similarities presented lower spatial dependency than taxonomic datasets, indicating that they are less susceptible to dispersal-connectivity effects. These findings contribute to understand the mechanisms underlying river community assembly at large spatial scales (i.e., at and beyond the river network) and point out the importance of dispersal-connectivity processes, which are usually neglected in traditional niche-based biomonitoring programmes but can influence their outcomes (e.g., masking the detection of anthropogenic impacts). Therefore, the integration of the dispersal-connectivity component, as well as information on organisms' dispersal abilities, are crucial when establishing effective conservation objectives and designing biomonitoring strategies.

 $\textbf{Keywords} \ \ Stream \ biota \cdot Diatoms \cdot Fish \cdot Functional \ traits \cdot Metacommunities \cdot Spatial \ ecology \cdot Bioassessment$ 

Communicated by Mike Kevin Joy.

Cássia Rocha Pompeu rochac@unican.es

IHCantabria - Instituto de Hidráulica Ambiental de la Universidad de Cantabria, Santander, Spain



#### Introduction

Streams and rivers sustain a broad range of habitats and biodiversity, supporting the delivery of valuable ecosystem services for human societies (Harrison et al. 2010; Vörösmarty et al. 2010; Grizzetti et al. 2016). Despite this, they face severe anthropogenic threats worldwide, such as effluent discharge, introduction of exotic species, and flow regime alteration with dams and reservoirs (Dudgeon et al. 2006; Reid et al. 2019). These increasing pressures do not change only the primary drivers defining riverine species' niche (e.g., temperature, water chemistry, benthic substrate, river morphology), but also lead to alterations in the connectivity of habitats, affecting their spatial dynamics at large spatial scales (McCluney et al. 2014).

To date, global trends in riverine ecosystem degradation call for urgent implementation of sustainable resource management, which should be underpinned by the most updated understanding of cause-effect relationships between anthropogenic impacts, biodiversity, and ecological processes (Dudgeon et al. 2006). In this regard, water management decisions are informed by cost-effective biological assessments (Bonada et al. 2006), which are required by the European Water Framework Directive (WFD; European Commission 2000) and environmental legislation elsewhere (e.g., National Water Initiative in Australia, Clean Water Act in the USA).

Recent advances in spatial ecology look at the influence of the riverine network structure on biological patterns and processes shaping metacommunities (Altermatt and Fronhofer 2018; Tonkin et al. 2018; Erős and Lowe 2019), and new techniques (e.g., virtual watersheds; Barquín et al. 2015) facilitate the design and improvement of monitoring approaches (Siqueira et al. 2014; Heino et al. 2015). Riverine biological assemblages differ in their dispersal abilities and ecological preferences, and, at large spatial extents (i.e., scale at which most monitoring programmes are designed), they can be shaped by different factors. For example, good dispersers (e.g., diatoms, flying macroinvertebrates) are usually driven by their niche (i.e., environmental characteristics; Hájek et al. 2011; Astorga et al. 2012), while weak dispersers (e.g., fish) are more constrained by geographical distances and connectivity among river reaches (Shurin et al. 2009). In this context, body size seems to be a good proxy of the dispersal capacity and, as such, an important biological characteristic to evaluate the contribution of environmental versus dispersal-connectivity factors in determining riverine assemblages (Astorga et al. 2012; De Bie et al. 2012). Moreover, functional traits have been recognised as an alternative to taxonomy, as they should be more dependent on the niche than on dispersal-connectivity constraints and allow a deeper understanding of stressor's impacts and mechanisms (Gayraud et al. 2003; Menezes et al. 2010; Vandewalle et al. 2010; Chen and Olden 2018).

Despite the rapid development of metacommunity concepts and the recognised importance of dispersal-connectivity processes structuring biological communities in rivers (Leibold et al. 2004), the vast majority of biological assessment programmes and biotic indices consider that abiotic environmental conditions (i.e., species sorting) are the main, if not solely, factors controlling community assembly (Heino 2013; but see Cid et al. 2020). Thus, anthropogenic impacts are detected based on the deviation from reference conditions (Downes et al. 2002; Siqueira et al. 2014). However, at large spatial extents (i.e., multiple drainage basins), the importance of dispersal-connectivity processes will likely increase (Verleyen et al. 2009), potentially masking anthropogenic environmental impacts (Heino 2013; Vilmi et al. 2016).



The distance-decay relationships (DDR; Nekola and White 1999) have been applied in a range of environments, organisms, and spatial extents (Brown and Swan 2010; Soininen et al. 2007) to investigate geographical distance effects on communities, and provide important insights on spatial dynamics, especially for fragmented and highly dynamic freshwater habitats (Cañedo-Argüelles et al. 2015; Cid et al. 2020). The decay of community similarities over geographical distances can be first attributed to niche differences, as environmental variables tend to be spatially autocorrelated (Legendre and Fortin 1989; Nekola and White 1999), and with spatial processes (e.g., organisms' dispersal, spatial configuration, isolation of habitats, mass effect; Hubbell 2001; Soininen et al. 2007) also contributing to it. Likewise, variation partitioning is a quantitative method frequently used in ecology to assess the likelihood of predictors (e.g., environment, geographical distances) in explaining community patterns (Peres-Neto et al. 2006). It allows partitioning the variance into uniquely and jointly explained fractions and inferring underlying community assembly processes (Diniz-Filho et al. 2012; Brown et al. 2017). However, since dependency between biological communities' distribution, environment, and space is extremely common, the method should be applied carefully, as it can fail to control for spatial autocorrelation biases, generating spurious correlations (Gilbert and Bennett 2010; Tuomisto et al. 2012).

To our knowledge, the importance of environmental and dispersal-connectivity factors in determining the taxonomic and functional structure of multiple river biological communities at large spatial scales (i.e., multiple neighbouring river networks) has been rarely evaluated simultaneously (but see Henriques-Silva et al. 2019; Keck et al. 2018). In this study, we selected diatoms and fish, organisms with contrasting dispersal abilities, to investigate the relative importance of environmental- (i.e., niche) and spatial-related (i.e., dispersal-connectivity) mechanisms on community composition based on taxonomy (C-TX) and community structure based on functional traits (C-FT) across continental Spain. We expect that (1) the importance of environmental and dispersal-connectivity processes will differ between diatoms and fish, with (2) dispersal-connectivity processes being a key determinant for fish C-TX while (3) environmental factors being more relevant for diatom C-TX (i.e., dispersal ability and spatial control inferred from body size; De Bie et al. 2012). In addition, we expect that (4) C-FT will have a lower spatial dependency in comparison to C-TX, as functional traits should be more responsive to niche characteristics than to dispersal-connectivity dynamics (Hoeinghaus et al. 2007). Finally, we will highlight the implications of our findings for riverine biomonitoring programmes.

#### Materials and methods

#### Study area

The study area comprises continental Spain (Fig. 1; 40° 23′ N, 3° 33′ E), a country with great environmental variability within the Atlantic and the Mediterranean zones. The relief is characterised by an extensive high inland plateau (average of 650 m.a.s.l.) surrounded by relatively high mountain ranges: the Cantabrian Range in the north, the Iberian system in the east, and the Sierra Morena in the south. Outside this central plateau, the Pyrenees on the north-eastern border, and the Betico system in the southeast, are the highest mountain ranges, with peaks reaching 3,400 m.a.s.l. (Rivas-Martínez et al. 2004; Peñas and Barquín 2019).



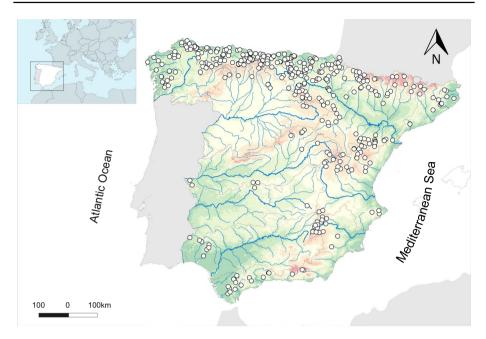


Fig. 1 Study area (505,000 km<sup>2</sup>) and sampling site selection, comprising a total of 441 samples

This complex orography plays a crucial role in defining the climate and hydrography. Five Köppen–Geiger climate zones are distributed along the area: mountain climate in high altitudes, with cold winters and abundant precipitations (annual averages reaching 3000 mm); the oceanic climate in the northwest region; hot steppe climate in the southeast, with minimum rainfall in Spain (annual averages lower than 150 mm); and several Mediterranean climate variations, with dry and hot summers (Rivas-Martínez et al. 2004; AEMET and IMP 2011). Thus, the twelve main catchments in the study area are conditioned by the combination of relief and climatic features, causing high variability in hydrological patterns (Peñas et al. 2014).

#### Characterisation of biological communities and functional traits

In this study, we used the national information exchange system on the status and quality of continental waters, called NABIA (Spanish Royal Decree 817/2015 on Water Policy; BOE 2015), compiled and provided by the Spanish Ministry for the Ecological Transition (MITECO). This database provides information on biological indicators from monitoring campaigns carried out since 2008 for assessing the ecological status and water quality of all water bodies in the country in compliance with the WFD (European Commission 2000).

From the NABIA database, we selected diatom and fish samples collected between May and October to maximise the spatial coverage while reducing "noise" caused by intraannual variability, as done in previous studies (e.g., Leathwick et al. 2005). The communities were surveyed annually between summer and early autumn depending on meteorological and hydrological conditions. Regarding diatoms, following the standard procedure UNE-EN13946:2014 (CEN 2014a), five to ten cobbles were selected randomly from the



benthos (ca. 100 cm² of exposed surface area) at a depth of ca. 10 cm to ensure that they were not exposed to air in the previous 4 weeks and that they were placed in the euphotic zone. Areas of heavy shade and close to the bank were avoided, as well as zones of slow current (approx. ≤20 cm s<sup>-1</sup>). Then, the upper part of the substratum was scrubbed with a dishwasher brush. Aliquots of the diatom samples collected were digested with hydrogen peroxide and permanent slides were mounted with Naphrax®. The material was decanted in a sample bottle and preserved using formaldehyde. From each sample, 400 diatom valves were identified in laboratory using a microscope (×1000 magnification) at the lowest feasible taxonomic level, according to the standard procedure UNE-EN 14407:2014 (CEN 2014b). Taxonomic identification was based on Delgado et al. (2013), Delgado and Pardo (2015), Krammer and Lange-Bertalot (1986–1991), (2004), Krammer (1997a, b), Lange-Bertalot (1993, 2001), Lange-Bertalot and Krammer (1989), Levkov (2009), Novais et al. (2009), Prygiel and Coste (2000), and Trobajo et al. (2013).

For fish communities, a single-pass sampling was performed using a portable electric fishing device (current generation 300–600 V, up to 1.5 A) in a representative area of the wadeable stream (ten times the average width of the stream and a minimum of 100 m<sup>2</sup>), following the standard procedure UNE-EN 14011:2003 (CEN 2003). After the pass removal, the captured fish were kept in oxygenated boxes, anaesthetised, counted, and identified at the species level. The fish were then released back to the stream alive after a recovery period.

Although macroinvertebrates are also widely used in the Spanish bioassessment programmes, we did not include them in our analysis due to their coarse taxonomic resolution (i.e., mainly at the family level) required for computing the Iberian Biological Monitoring Working Party (IBMWP) index (Alba-Tercedor et al. 2002).

Only samples surveyed in river reaches catalogued as 'good' or 'very good' ecological status were retained. The ecological status was determined using biological, physico-chemical, and hydromorphological characteristics and reference conditions for surface waters, as specified in the Spanish Hydrological Plan 2015–2021 within the European WFD (BOE 2015). Further, only sites unaffected by local anthropogenic pressures such as dams, embankments, or significant water abstractions upstream were kept for subsequent analyses. This site selection, covering only minimally disturbed streams, reduced the confounding effect of other factors (e.g., stressors), such as water pollution, hydromorphological pressures, or the presence of dams and reservoirs. After applying these criteria, we retained 177 sampling sites for diatoms and 264 for fish (Fig. 1).

Diatoms and fish were identified at the species level (see Tables S1 and S3 in Supplementary Information). The abundance of taxa was averaged in sites surveyed for multiple years to obtain a site-specific assemblage composition, following previous studies (Paavola et al. 2003; Filker et al. 2016). Due to the high number of diatom species identified, we eliminated rare taxa (those representing less than 2% of total sampled individuals when taking into account the total number of occurrences; Lavoie et al. 2009). Then, in the two biotic groups, C-TX was determined based on the presence/absence of each taxon.

To characterise C-FT, we assigned diatoms' species to ecological guilds, size (biovolume), and life forms according to Rimet and Bouchez (2012) (see Table S2 in Supplementary Information). For fish, we assigned the species to trait categories describing body size, feeding habits, tolerance to stressors, habitat use, and migration (see Table S4 in Supplementary Information), based on Iberian species information obtained from Doadrio (2002), Schmidt-Kloiber and Hering (2015), and Cano-Barbacil et al. (2020). It is important to note that trait categories can overlap and species may belong to multiple trait categories (e.g., *Amphora pediculus* is considered, at the same time, a low profile, pioneer, adnate,



and non-colonial species; *Phoxinus bigerri* is a non-migratory, omnivorous, rheophilic fish species inhabiting the water column). Then, for both organisms, the relative abundance corresponding to each trait category was calculated based on the abundance of the taxa contributing to it.

#### River network and environmental information

River reaches were characterised using hydrological and environmental information integrated within the virtual watersheds composing the Spanish river network (Barquín et al. 2015), constructed using flow direction inferred from a 10-m digital elevation model. Environmental variables describing topography, climate, land use and land cover, and geology (Table 1) were obtained from national and regional databases (Peñas and Barquín 2019).

**Table 1** Environmental variables describing topography, climate, land use and land cover, geology, and hydrology in the diatoms (n = 177) and fish (n = 264) sampling sites. Only uncorrelated variables in each category (Pearson's  $|r| \le 0.7$ ) are shown. Variables refer to the mean catchment values upstream of the sampling site

Description	Diatoms		Fish	
	Mean ± sd	Range	Mean ± sd	Range
Catchment size (km <sup>2</sup> )	$226 \pm 502.2$	0.9 to 4845	225.6±410.	1 1.6 to 4845
Distance to river outlet (km)	$394 \pm 272.1$	0 to 915	$176.3 \pm 252.2$	20 to 928.3
Elevation above sea level (m)	$653.1 \pm 351.7$	1 to 1734	$372.8 \pm 339$	1 to 1.435.3
Slope in segment (0 to 1)	$0.3 \pm 0.1$	0 to 0.7	$0.4 \pm 0.2$	0.1 to 1.0
Annual precipitation (mm)	$864.3 \pm 327.4$	329.3 to 1796.3	$1132.1 \pm 342.6$	6 455.9 to 1947.1
Annual temperature (°C)	$11.9 \pm 2.6$	6.2 to 18	$11.8 \pm 2.3$	5 to 18.6
Annual evapotranspiration (mm)	$477.2 \pm 66.1$	294.3 to 684.2	$511.4 \pm 75.5$	318.3 to 726.8
Fraction occupied by urbanisation	$0.0 \pm 0.0$	0 to 0.1	$0.0\pm0.0$	0 to 0.1
Fraction occupied by agriculture	$0.1 \pm 0.2$	0 to 0.6	$0.1 \pm 0.1$	0 to 0.6
Fraction occupied by pastures	$0.2 \pm 0.2$	0 to 0.8	$0.2 \pm 0.1$	0 to 0.7
Fraction occupied by broadleaf forests	$0.2 \pm 0.2$	0 to 0.9	$0.2\pm0.2$	0 to 0.8
Fraction occupied by coniferous forests	$0.1 \pm 0.2$	0 to 1	$0.1\pm0.2$	0 to 0.8
Fraction occupied by heathlands and shrubs	$0.2 \pm 0.2$	0 to 0.7	$0.3 \pm 0.2$	0 to 0.7
Fraction occupied by denuded areas	$0 \pm 0.1$	0 to 0.7	$0.1 \pm 0.1$	0 to 0.7
Fraction occupied by calcareous rocks	$0.5 \pm 0.4$	0 to 1.0	$0.4 \pm 0.4$	0 to 1.0
Fraction occupied by conglomerate rocks	$0.3 \pm 0.3$	0 to 1.0	$0.2 \pm 0.3$	0 to 1.0
Fraction occupied by sedimentary rocks	$0 \pm 0.1$	0 to 0.7	$0 \pm 0.0$	0 to 0.4
Fraction occupied by siliceous rocks	$0.1 \pm 0.3$	0 to 1.0	$0.2 \pm 0.3$	0 to 1.0
Fraction occupied by volcanic rocks	$0 \pm 0.1$	0 to 0.8	$0.1 \pm 0.3$	0 to 1.0
SI <sub>NAT</sub> 1	$-1.49 \pm 0.62$	-1.49 to 1.51	$0.10 \pm 0.45$	-1.55 to 1.59
$SI_{NAT}2$	$0.21 \pm 0.71$	-1.21 to 1.42	$-0.30 \pm 0.67$	-1.40 to $1.40$
$SI_{NAT}3$	$-0.04 \pm 0.49$	-1.79 to $1.10$	$-0.14 \pm 0.48$	-1.42 to $0.95$
SI <sub>NAT</sub> 4	$0.24 \pm 0.39$	-0.81 to 1.44	$-0.07 \pm 0.42$	-1.09 to 1.21



To characterise the flow regime, we used a set of non-correlated synthetic hydrological indices ( $SI_{NAT}$ ). They were calculated based on the normalised daily flow series recorded in natural flow gauges and predicted to all river reaches in the country using Random Forest models (Peñas and Barquín 2019). The four  $SI_{NAT}$  (Table 1) explained 84% of the total hydrological variance and set the critical natural hydrological patterns in continental Spain (Peñas and Barquín 2019).

#### Data analyses

We used DDR (Nekola and White 1999; Soininen et al. 2007) to assess the effects of geographical distances and environmental characteristics on diatoms and fish assemblages. We analysed the dispersal-connectivity and environmental control in both biotic groups and their C-TX and C-FT using the DDR linear regression coefficients (slope), which described the rate of species turnover, and the initial similarities (intercept), which described the species turnover at small spatial extents (Astorga et al. 2012).

The DDR was estimated regressing biological similarities as a function of geographical and environmental distances, with the Jaccard index for C-TX pairwise biological communities similarities and Bray-Curtis distances for C-FT. Since our study area was composed of multiple unconnected river networks, we computed Euclidean distances based on samples' latitude and longitude, ranging from 1 to 1,031 km. Similarly, to compute environmental distances, the environmental (topography, climate, geology, and LULC) and hydrological variables (Table 1) were first standardised (zero mean and unit variance), as they had different measurement units and ranges, and pairwise Euclidean distances between all sites were calculated. We used partial correlation analysis (Legendre and Legendre 1998) to factor out the effect of the geographical distances on environmental variables and test for independent correlations between environmental distances and the biological similarities. Given the different ranges described by geographical and environmental distances, we rescaled both distances to values between 0 and 1.

Then, to quantify and compare the relative contribution of environmental and geographical distance-related processes to the diatom and fish C-TX and C-FT variability, we used variation partitioning (Peres-Neto et al. 2006). The biological variation was decomposed into four fractions: the variation explained (1) uniquely by non-spatial environmental variation, i.e., species variation explained by the environmental dataset (Table 1: topography, climate, geology, LULC and hydrology) independently of any spatial structure, (2) uniquely by spatial patterns (i.e., geographical distance among sites) that are not shared by the environmental dataset, (3) by the spatial patterns in community data that are shared by the environmental dataset (i.e., shared or spatially structured environmental variation), and (4) the unexplained variation (Borcard et al. 1992). We used Moran's eigenvector maps (MEMs; Dray et al. 2006) as a proxy for the diatom and fish C-TX and C-FT spatial distribution patterns (i.e., geographical distances). The MEMs are linearly independent vectors capable of describing a wide range of spatial scales (Griffith and Peres-Neto 2006) and are linked to a spatial weighting matrix (SWM), which determines the spatial relationships between the sampled sites (Bauman et al. 2018a). We tested three graph-based schemes (Gabriel graph, relative neighbourhood graph, and minimum spanning tree) with two weighting matrices (binary and linearly decreasing as a function of pairwise-site distances) as potential SWM (Benone et al. 2020). All positively spatially correlated MEMs for all SWM candidates were computed, as a subset of MEMs is not able to remove the spatial autocorrelation from model residuals and may lead to inflated type I error of the



pure spatial fraction (Peres-Neto and Legendre 2010; Clappe et al. 2018). Then, an optimisation procedure (Bauman et al. 2018b) selected the SWM which MEMs yield to the maximisation of the adjusted R<sup>2</sup> (see detailed explanation in Table S7 in Supplementary Information). Regarding environmental predictors, we ran a forward selection procedure to retain the relevant environmental variables with a double-stopping criterion (Blanchet et al. 2008) to avoid overestimation of explained variance.

Finally, Moran Spectral Randomisation (MSR; Wagner and Dray 2015; Clappe et al. 2018) was applied to correctly account for spatial autocorrelation in the variation partitioning. This method performs spatially constrained randomisations in variation partitioning and maintains the data's spatial characteristics, avoiding type I error inflation and removing spurious correlations (Clappe et al. 2018).

All statistical analyses were performed in R software (R Core Team 2020) using the packages vegan (Oksanen et al. 2019) and adespatial (Dray et al. 2020).

#### Results

# Diatom and fish assemblages

A total of 95 diatoms species were retained in our analyses (Table S1 in Supplementary Information). The most abundant species were *Achnanthidium minutissimum* (corresponding to 19.2% of all sampled diatoms), *Achnanthidium pyrenaicum* (14.2%), *Cocconeis euglypta* (8.1%), *Amphora pediculus* (4.5%), and *Achnanthidium lineare* (4.2%). Other genera, such as *Gomphonema* (corresponding to 9.4% of all sampled diatoms) and *Nitzschia* (5.4%), were also present in samples. Regarding their functional traits, the high-profile guild, stalked, and small-sized diatoms dominated most of the sampling sites (Table S2 in Supplementary Information).

Fish assemblages had a total of 38 species (Table S3 in Supplementary Information). The most abundant taxa were *Salmo trutta* (corresponding to 29.8% of all sampled fish), *Phoxinus bigerri* (28.7%), *Parachondrostoma miegii* (6.7%), *Salmo salar* (5.5%), and *Anguilla anguilla* (4.2%). Regarding their functional traits, migratory, sensitive to stressors, and large-bodied species dominated most of the sampling sites (Table S4 in Supplementary Information).

# Similarity decay over geographical and environmental distances

The biological similarities decreased with geographical and environmental distances in both fish and diatom communities (Fig. 2 and Table S5 in Supplementary Information). Diatoms presented lower initial similarities compared to fish, indicating higher beta diversity at small spatial scales. Fish biological similarities showed steeper decay rates when compared to diatoms (1.5 and 2.5 times steeper over geographical distances for C-TX and C-FT, respectively; Fig. 2a, d), indicating a higher explanatory capacity of the environmental variables and spatial patterns for fish. Fish C-TX similarities presented a large number of "zero" (24% of samples) or "one" (5% of samples) values, showing that these assemblages had no species in common or shared all species, respectively. In both organisms, the C-FT (Fig. 2, in grey) showed lower decay rates (-57% in diatoms and -38% in fish), higher initial similarities and higher scattering over geographical distances when compared to the



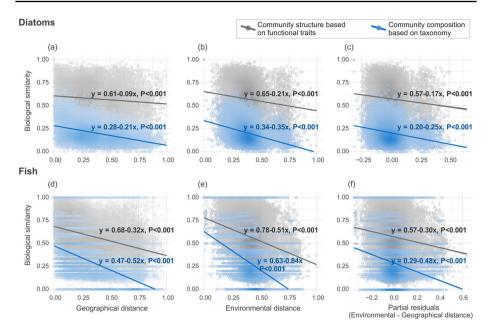


Fig. 2 Biological similarity decay for diatoms ( $\mathbf{a}$  to  $\mathbf{c}$ ) and fish ( $\mathbf{d}$  to  $\mathbf{f}$ ) over geographical distances, environmental distances, and environmental partial residuals after removing the spatial component. We used Jaccard index among pairs of samples for taxonomical community composition (in blue) and Bray–Curtis similarities for functional community structure (in grey). Equations were obtained with simple linear regression (see also Table S5 in Supplementary Information)

C-TX, indicating that geographical distances exerted a lower control on community similarity when using functional traits.

The geographical and environmental distances among samples showed positive correlations (see Fig. S1 in Supplementary Information), suggesting that environmental characteristics were spatially correlated. In both organisms, the DDR over environmental distances (Fig. 2b, e), compared to the DDR over geographical distances, showed higher initial similarities and decay rates (an increase of 100% for diatoms and 60% for fish, on average). In contrast, after splitting out the spatial component (Fig. 2c, f), the decay rates were 54% higher for diatoms and 7% lower for fish in relation to the DDR over geographical distances, indicating that spatial patterns exerted an important control on fish assemblage variation.

## Contribution of environment versus dispersal-connectivity factors

The variation partitioning (Fig. 3 and Table S6 in Supplementary Information) showed that environmental variables and MEMs explained larger fractions of the biological variation in fish assemblages (50.4%, on average) when compared to diatoms (31.1%), in agreement with the DDR analysis. The shared fraction between environment and spatial patterns (i.e., spatially structured environmental fraction) corresponded to the major proportion of explained variation in most cases (see also Fig. S2 in Supplementary Information).

Environmental variables and MEMs, together, accounted for 25.5% and 36.7% of the diatoms' taxonomical and functional variability, respectively (Fig. 3a, b). The



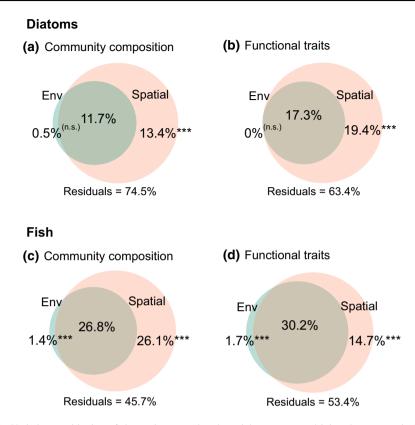


Fig. 3 Variation partitioning of the environmental and spatial components driving the taxonomical community composition and functional traits of diatoms ( $\mathbf{a}$ ,  $\mathbf{b}$ ) and fish ( $\mathbf{c}$ ,  $\mathbf{d}$ ). Fractions represent pure environmental (Env), spatialized environmental (shared fraction), and pure spatial (Spatial) fraction after MSR corrections. Significance of testable fractions was determined with ANOVA of RDA models; p-values are represented as \*\*\* $\leq$ 0.001; \*\* $\leq$ 0.01; \*\* $\leq$ 0.05; n.s. not significant (see also Table S6 in Supplementary Information)

contribution of the pure spatial component was similar in both cases, corresponding to 52.4% of the total explained variation in diatom C-TX and 52.8% in C-FT. The spatially structured environmental contribution was slightly higher when C-FT was considered (47.2% of the total explained variation, in comparison to 45.7% for C-TX). The fraction of community variability attributed to the non-spatialized environment was negligible in both cases.

Environmental variables and MEMs, together, accounted for 54.3% and 46.6% of the explained variation in fish C-TX and C-FT, respectively (Fig. 3c, d). In contrast to diatoms, the pure environmental fraction had small but significant contributions to fish C-TX and C-FT variability. When considering fish C-TX, the contributions of pure spatial and spatially structured environmental fractions were similar. In fish C-FT, the spatially structured environmental fraction explained twice the variation of the pure spatial fraction, suggesting that fish functional traits are less subject to dispersal-connectivity processes and are more responsive to environmental characteristics.



#### Discussion

This study indicates that environmental and spatial controls differ in their contribution to structuring riverine biological communities. Geographical distances exerted a higher control on fish (low dispersal ability) than on diatom (high dispersal ability) community similarities, while the non-spatially structured environment showed minimal contributions to both organisms' variability. Finally, community similarity based on functional traits showed a higher influence of environmental factors than geographical distances.

# Differences between biotic groups: the effects of distance and dispersal capacity on biological assemblages

Our findings showed a decrease in biological similarities over geographical and environmental distances, suggesting that diatom and fish assemblages are jointly driven by niche (e.g., environmental filtering; Leibold et al. 2004) and dispersal (Hubbell 2001) processes. The differences observed in initial similarities, decay rates, and the fraction of variation attributed to environment and space indicated that environmental and distance-related dynamics share the controlling the biological variation in the two communities, in agreement with our first hypothesis and previous studies (Brown and Swan 2010; Astorga et al. 2012; Keck et al. 2018; Leibold and Chase 2018).

Geographical distances in the DDRs and MEMs in the variation partitioning showed a higher control on fish assemblages in comparison to the non-spatialized environment, supporting our second hypothesis. The obligated aquatic dispersion through the river network and larger body size have been suggested to limit fish propagation and settlement along a larger range of habitat conditions (Cohen et al. 2003). These characteristics could contribute to the higher spatial control on fish C-TX found in this study and elsewhere (Shurin et al. 2009; Astorga et al. 2012). Besides, suitable environmental conditions in distant river reaches would not assure the colonisation by specific fish species, as connectivity (e.g., unconnected river basins, dams forming dispersal barriers) or the past biogeographical history have a key role in determining fish community patterns at large spatial scales (Leibold et al. 2010; Mazaris et al. 2010). These factors seem to be plausible explanations for the lower beta diversity and higher distance decay of fish communities.

Diatom C-TX showed a lower spatial control in comparison to fish, as demonstrated in the DDRs and variation partitioning analyses. These results agree with previous research (Finlay 2002; Vilmi et al. 2016) and could be linked to the ecological characteristics of diatoms (i.e., smaller size, shorter generation times, larger populations, and aerial dispersion), which allow them to overcome geographical barriers and colonise sites with favourable conditions. However, since environmental variables and MEMs together explained relatively low amounts of diatom community similarities, and the contribution of the pure environmental fraction was negligible, our findings could not support our third hypothesis. The low explanatory capacity of environmental variables for diatom C-TX, which has also been reported elsewhere (Liu et al. 2013; Keck et al. 2018), could be attributed to the large-scale definition of our environmental variables. Relevant local variables (e.g., water quality, physico-chemical characteristics, hydraulic and riparian conditions) that could be critical for determining diatom community composition (Soininen et al. 2009, 2016; Hájek et al. 2011) were not covered in our dataset. Furthermore, the complexity of diatom communities (i.e., a higher number of rare species in comparison to fish), biotic interactions,



and random and stochastic processes (e.g., ecological drift), which cannot be captured in our analyses, may also influence the community assembly dynamics (Hubbell 2001; Vellend et al. 2014; Vilmi et al. 2017).

Community similarities based on functional trait dominance were less spatially dependent than community similarities based on taxonomic data. C-FT showed lower decay rates and higher scattering in both fish and diatom DDRs, and fish C-FT pure spatial contribution had a decrease of 16.5% in the total variation explained in relation to C-TX. These findings support our last hypothesis and suggest that trait-based approaches are more stable across large spatial extents and multiple biogeographical units, grouping the complexity of species composition into a reduced number of traits. Therefore, functional trait community structure should be less subject to geographical distances since trait filters act in a similar way across large ecoregions (Statzner and Bêche 2010) and thus could provide more consistent ecological responses to environmental variation (i.e., niche) regardless of spatial distribution (Hoeinghaus et al. 2007). Our diatoms C-FT showed a higher contribution of pure spatial fraction when compared to C-TX; however, in relation to the total explained variation, their spatial control was similar. Previous studies (Passy 2007; Liu et al. 2013; Jamoneau et al. 2018) also reported important spatial control on diatom C-FT, e.g., strongly attached growth forms predominant in headwaters, while weakly attached forms predominant in higher-order streams. However, rather than indicating the effect of spatial structuring in the species distribution, high pure spatial fractions can also be inflated by missing relevant spatially structured environmental variables (Diniz-Filho et al. 2012).

## Implications for biomonitoring and conservation programmes

Our study supports the idea that geographical distance and environmental spatial autocorrelation (closer river reaches are more environmentally similar), which are rarely taken into account in riverine biomonitoring programmes (Siqueira et al. 2012; Cid et al. 2020), contributed significantly to explaining the variation in diatom and fish (i.e., organisms commonly used in biomonitoring) assemblages. Our analyses incorporate river networks from 12 distinct river basins in Spain, suggesting that riverine communities have not only an important spatial dependency on the river network structure itself (Altermatt 2013; Tonkin et al. 2018) but also across the multiple neighbouring river networks. Our results support the view that regional species pool and dispersal seem to be fundamental factors determining taxonomic differences in river biological assemblages, as has been shown in other studies (Leibold and Chase 2018; Viana and Chase 2019).

These findings have important consequences for river biomonitoring and conservation. For example, the use of organisms with higher dispersal limitation (e.g., fish) in biomonitoring might be relevant to capture connectivity issues caused by human activities (e.g., dam construction), but might not be as promising for detecting changes in niche characteristics (e.g., hydro-morphological changes), as not all potential species may be present and, therefore, accurate detection of anthropogenic impacts can be compromised (Siqueira et al. 2014). Another important implication of our results is that biomonitoring programmes may fail to detect restoration benefits if desired taxa are unable to reach new suitable locations; thus, the presence of protected or unaltered river reaches with source populations in the immediate vicinity of the restored sites (e.g., within a 5-km radius for potential recolonization of macroinvertebrates; Sundermann et al. 2011) is crucial (Siqueira et al. 2014; Swan and Brown 2017). This later issue reflects the need to incorporate multiple and well-connected river reaches in relatively good ecological status across the landscape matrix, so



that investment in recovery is actually successful. Finally, the spatial configuration of river typologies might also affect the establishment of reference conditions (e.g., 37 different river typologies and reference conditions identified over the Spanish river network; BOE 2015). Reference conditions should cover relatively small geographical areas in order to reduce the effects of dispersal-connectivity processes on taxonomic differences, ensuring that the targeted organisms are able to reach and persist at all sites and allowing proper detection of species sorting dynamics (Heino et al. 2017).

Thus, dispersal-connectivity processes can have important effects on the performance of bioindicators and biomonitoring efforts, as different organisms' dispersal abilities and mass effects (intense dispersion) can mask anthropogenic disturbances or environmental factors controlling species distribution (Smucker and Vis 2011; Siqueira et al. 2012; Cid et al. 2020). In order to reinforce river biomonitoring programmes and the design of effective conservation strategies, we encourage that potential spatial-related factors such as organism's dispersal and habitat connectivity, acting at and beyond the river network, should be considered in the selection of reference sites, targeted organisms, grain extent, and spatial scales (Bonada et al. 2006; Seymour et al. 2016; Cid et al. 2020).

The functional traits, compared to taxonomic approaches, showed a lower spatial dependency in our study. This result suggests that functional traits would be more stable across biogeographical regions or multiple river networks, as they can provide valuable insights into the processes and mechanisms structuring biological assemblages (Usseglio-Polatera et al. 2000; Vandewalle et al. 2010), while species composition tends to respond to historical and spatial factors (Soininen et al. 2016). Functional traits could also facilitate the development of river typologies and reference conditions often used in biomonitoring, as they are responsive to niche characteristics (McGill et al. 2006; Culp et al. 2011). Moreover, functional approaches would allow a better understanding of biodiversity, which is usually limited to taxonomic richness (Bêche and Statzner 2009), and the development of holistic assessments of anthropogenic impacts (e.g., taking into account biotic interactions in a multi-trophic perspective; Aubin et al. 2013).

In summary, our findings indicate that environmental and dispersal-connectivity processes contributed to determining the structure and composition of the riverine biota in continental Spain. Both environmental and spatial aspects should be considered in national multi-scale biomonitoring programmes to assess riverine ecosystem health accurately. Functional approaches, besides their known advantages over taxonomy-based methods, can also complement biomonitoring efforts with consistent responses across broad spatial scales (i.e., when incorporating multiple ecoregions and biogeographic areas).

**Supplementary Information** The online version contains supplementary material available at https://doi.org/10.1007/s10531-022-02514-6.

**Acknowledgements** We would like to thank the Spanish Ministry for the Ecological Transition for providing the biological monitoring data. This project has received funding from the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie Grant Agreement No 765553 and from the project "WATERLANDS", code PID2019-107085RB-I00, funded by MCIN/AEI/10.13039/501100011033/ and by ERDF "A way of making Europe". We also thank two anonymous reviewers for their valuable comments.

**Author contributions** CRP, FJP and JB conceived the ideas and designed the methods; FJP and JB obtained the data; CRP and JB analysed the data; CRP led the writing of the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

Funding Open Access funding provided thanks to the CRUE-CSIC agreement with Springer Nature. This project has received funding from the European Union's Horizon 2020 research and innovation programme



under the Marie Skłodowska-Curie Grant Agreement No 765553 and from the project "WATERLANDS", code PID2019-107085RB-I00, funded by MCIN/AEI/10.13039/501100011033/ and by ERDF "A way of making Europe".

**Data availability** The data that support the findings of this study are available from the Spanish Ministry for the Ecological Transition. Data will be available in a digital depository with the permission of the national authority.

Code availability Not applicable.

#### **Declarations**

Conflict of interest The authors declare no conflict of interest.

Ethical approval Not applicable.

Consent to participate Not applicable.

**Consent for publication** Not applicable.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

#### References

- AEMET, IMP (2011) Atlas climático ibérico/Iberian climate atlas. Agencia Estatal Meteorol. Minist. Medio Ambient. Medio Rural y Mar. Inst. Meteorol. Port. 79
- Alba-Tercedor J, Jáimez P, Álvarez M et al (2002) Índice IBMWP y estado ecológico de ríos mediterráneos ibéricos. Limnetica 21:175–185
- Altermatt F (2013) Diversity in riverine metacommunities: a network perspective. Aquat Ecol 47:365–377. https://doi.org/10.1007/s10452-013-9450-3
- Altermatt F, Fronhofer EA (2018) Dispersal in dendritic networks: ecological consequences on the spatial distribution of population densities. Freshw Biol 63:22–32. https://doi.org/10.1111/fwb.12951
- Astorga A, Oksanen J, Luoto M et al (2012) Distance decay of similarity in freshwater communities: do macro- and micro-organisms follow the same rules? Glob Ecol Biogeogr 21:365–375. https://doi.org/10.1111/j.1466-8238.2011.00681.x
- Aubin I, Venier L, Pearce J, Moretti M (2013) Can a trait-based multi-taxa approach improve our assessment of forest management impact on biodiversity? Biodivers Conserv 22:2957–2975. https://doi.org/10.1007/s10531-013-0565-6
- Barquín J, Benda LE, Villa F et al (2015) Coupling virtual watersheds with ecosystem services assessment: a 21st century platform to support river research and management. Wires Water 2:609–621. https://doi.org/10.1002/wat2.1106
- Bauman D, Drouet T, Dray S, Vleminckx J (2018a) Disentangling good from bad practices in the selection of spatial or phylogenetic eigenvectors. Ecography (Cop) 41:1638–1649. https://doi.org/10.1111/ecog.03380
- Bauman D, Drouet T, Fortin MJ, Dray S (2018b) Optimizing the choice of a spatial weighting matrix in eigenvector-based methods. Ecology 99:2159–2166. https://doi.org/10.1002/ecy.2469
- Bêche LA, Statzner B (2009) Richness gradients of stream invertebrates across the USA: taxonomy- and trait-based approaches. Biodivers Conserv 18:3909–3930. https://doi.org/10.1007/s10531-009-9688-1
- Benone NL, Soares BE, Lobato CMC et al (2020) How modified landscapes filter rare species and modulate the regional pool of ecological traits? Hydrobiologia. https://doi.org/10.1007/s10750-020-04405-9



- Blanchet FG, Legendre P, Borcard D (2008) Forward selection of explanatory variables. Ecology 89:2623–2632. https://doi.org/10.1890/07-0986.1
- BOE (Boletín Oficial del Estado) (2015) Real Decreto 817/2015, de 11 de septiembre, por el que se establecen los criterios de seguimiento y evaluación del estado de las aguas superficiales y las normas de calidad ambiental. MAPAMA 80582–80677
- Bonada N, Prat N, Resh VH, Statzner B (2006) Developments in aquatic insect biomonitoring: a comparative analysis of recent approaches. Annu Rev Entomol 51:495–523. https://doi.org/10.1146/annurev.ento.51.110104.151124
- Borcard D, Legendre P, Drapeau P (1992) Partialling out the spatial component of ecological variation. Ecology 73:1045–1055
- Brown BL, Swan CM (2010) Dendritic network structure constrains metacommunity properties in riverine ecosystems. J Anim Ecol 79:571–580. https://doi.org/10.1111/j.1365-2656.2010.01668.x
- Brown BL, Sokol ER, Skelton J, Tornwall B (2017) Making sense of metacommunities: dispelling the mythology of a metacommunity typology. Oecologia 183:643–652. https://doi.org/10.1007/s00442-016-3792-1
- Cañedo-Argüelles M, Boersma KS, Bogan MT et al (2015) Dispersal strength determines meta-community structure in a dendritic riverine network. J Biogeogr 42:778–790. https://doi.org/10.1111/jbi.12457
- Cano-Barbacil C, Radinger J, García-Berthou E (2020) Reliability analysis of fish traits reveals discrepancies among databases. Freshw Biol 65:863–877. https://doi.org/10.1111/fwb.13469
- CEN (2003) UNE-EN14011:2003. Water quality—sampling of fish with electricity
- CEN (2014a) UNE-EN13946:2014a. Water quality—guidance for the routine sampling and preparation of benthic diatoms from rivers and lakes
- CEN (2014b) UNE-EN 14407:2014b. Water quality—guidance for the identification and enumeration of benthic diatom samples from rivers and lakes
- Chen W, Olden JD (2018) Evaluating transferability of flow-ecology relationships across space, time and taxonomy. Freshw Biol 63:817–830. https://doi.org/10.1111/fwb.13041
- Cid N, Bonada N, Heino J et al (2020) A metacommunity approach to improve biological assessments in highly dynamic freshwater ecosystems. BioScience 70:427–438. https://doi.org/10.1093/biosci/biaa0 33
- Clappe S, Dray S, Peres-Neto PR (2018) Beyond neutrality: disentangling the effects of species sorting and spurious correlations in community analysis. Ecology 99:1737–1747. https://doi.org/10.1002/ecy. 2376
- Cohen JE, Jonsson T, Carpenter SR (2003) Ecological community description using the food web, species abundance, and body size. Proc Natl Acad Sci USA 100:1781–1786. https://doi.org/10.1073/pnas. 232715699
- Commission E (2000) Directive 2000/60/EC of the European Parliament and of the Council of 23 October 2000 establishing a framework for community action in the field of water policy. Off J Eur Parliam L327:1–82.
- Culp JM, Armanini DG, Dunbar MJ et al (2011) Incorporating traits in aquatic biomonitoring to enhance causal diagnosis and prediction. Integr Environ Assess Manag 7:187–197. https://doi.org/10.1002/ieam.128
- De Bie T, De Meester L, Brendonck L et al (2012) Body size and dispersal mode as key traits determining metacommunity structure of aquatic organisms. Ecol Lett 15:740–747. https://doi.org/10.1111/j. 1461-0248.2012.01794.x
- Delgado C, Pardo I (2015) Comparison of benthic diatoms from Mediterranean and Atlantic Spanish streams: community changes in relation to environmental factors. Aquat Bot 120:304–314. https://doi.org/10.1016/j.aquabot.2014.09.010
- Delgado C, Ector L, Novais MH et al (2013) Epilithic diatoms of springs and spring-fed streams in Majorca Island (Spain) with the description of a new diatom species *Cymbopleura margalefii* sp. nov. Fottea 13:87–104. https://doi.org/10.5507/fot.2013.009
- Diniz-Filho JAF, Siqueira T, Padial AA et al (2012) Spatial autocorrelation analysis allows disentangling the balance between neutral and niche processes in metacommunities. Oikos 121:201–210. https://doi.org/10.1111/j.1600-0706.2011.19563.x
- Doadrio I (ed) (2002) Atlas y libro rojo de los peces continentales de España, 2nd edn. Ministerio de Medio Ambiente, Madrid
- Downes BJ, Barmuta LA, Fairweather PG et al (2002) Applying monitoring designs to flowing waters. Monitoring ecological impacts. Cambridge University Press, Cambridge, pp 197–248
- Dray S, Legendre P, Peres-Neto PR (2006) Spatial modelling: a comprehensive framework for principal coordinate analysis of neighbour matrices (PCNM). Ecol Model 196:483–493. https://doi.org/10. 1016/j.ecolmodel.2006.02.015



- Dray S, Bauman D, Blanchet G et al (2020) adespatial: Multivariate multiscale spatial analysis. R package version 0.3-20. https://CRAN.R-project.org/package=adespatial
- Dudgeon D, Arthington AH, Gessner MO et al (2006) Freshwater biodiversity: importance, threats, status and conservation challenges. Biol Rev 81:163. https://doi.org/10.1017/S1464793105006950
- Erős T, Lowe WH (2019) The landscape ecology of rivers: from patch-based to spatial network analyses. Curr Landsc Ecol Rep 4:103–112. https://doi.org/10.1007/s40823-019-00044-6
- Filker S, Sommaruga R, Vila I, Stoeck T (2016) Microbial eukaryote plankton communities of high-mountain lakes from three continents exhibit strong biogeographic patterns. Mol Ecol 25:2286–2301. https://doi.org/10.1111/mec.13633
- Finlay BJ (2002) Global dispersal of free-living microbial eukaryote species. Science (80-) 296:1061–1063. https://doi.org/10.1126/science.1070710
- Gayraud S, Statzner B, Bady P et al (2003) Invertebrate traits for the biomonitoring of large European rivers: an initial assessment of alternative metrics. Freshw Biol 48:2045–2064. https://doi.org/10.1046/j. 1365-2427.2003.01139.x
- Gilbert B, Bennett JR (2010) Partitioning variation in ecological communities: do the numbers add up? J Appl Ecol 47:1071–1082. https://doi.org/10.1111/j.1365-2664.2010.01861.x
- Griffith DA, Peres-Neto PR (2006) Spatial modeling in ecology: the flexibility of eigenfunction spatial analyses. Ecology 87:2603–2613. https://doi.org/10.1890/0012-9658(2006)87[2603:SMIETF]2.0.CO;2
- Grizzetti B, Lanzanova D, Liquete C et al (2016) Assessing water ecosystem services for water resource management. Environ Sci Policy 61:194–203. https://doi.org/10.1016/j.envsci.2016.04.008
- Hájek M, Roleček J, Cottenie K et al (2011) Environmental and spatial controls of biotic assemblages in a discrete semi-terrestrial habitat: comparison of organisms with different dispersal abilities sampled in the same plots. J Biogeogr 38:1683–1693. https://doi.org/10.1111/j.1365-2699.2011.02503.x
- Harrison PA, Vandewalle M, Sykes MT et al (2010) Identifying and prioritising services in European terrestrial and freshwater ecosystems. Biodivers Conserv 19:2791–2821. https://doi.org/10.1007/ s10531-010-9789-x
- Heino J (2013) The importance of metacommunity ecology for environmental assessment research in the freshwater realm. Biol Rev 88:166–178. https://doi.org/10.1111/j.1469-185X.2012.00244.x
- Heino J, Melo AS, Siqueira T et al (2015) Metacommunity organisation, spatial extent and dispersal in aquatic systems: patterns, processes and prospects. Freshw Biol 60:845–869. https://doi.org/10.1111/fwb.12533
- Heino J, Alahuhta J, Ala-Hulkko T et al (2017) Integrating dispersal proxies in ecological and environmental research in the freshwater realm. Environ Rev 25:334–349. https://doi.org/10.1139/er-2016-0110
- Henriques-Silva R, Logez M, Reynaud N et al (2019) A comprehensive examination of the network position hypothesis across multiple river metacommunities. Ecography (Cop) 42:284–294. https://doi.org/10.1111/ecog.03908
- Hoeinghaus DJ, Winemiller KO, Birnbaum JS (2007) Local and regional determinants of stream fish assemblage structure: inferences based on taxonomic vs. functional groups. J Biogeogr 34:324–338. https://doi.org/10.1111/j.1365-2699.2006.01587.x
- Hubbell SP (2001) the unified neutral theory of biodiversity and biogeography. Princeton University Press, Princeton
- Jamoneau A, Passy SI, Soininen J et al (2018) Beta diversity of diatom species and ecological guilds: response to environmental and spatial mechanisms along the stream watercourse. Freshw Biol 63:62–73. https://doi.org/10.1111/fwb.12980
- Keck F, Franc A, Kahlert M (2018) Disentangling the processes driving the biogeography of freshwater diatoms: a multiscale approach. J Biogeogr 45:1582–1592. https://doi.org/10.1111/jbi.13239
- Krammer K (1997a) Die cymbelloiden Diatomeen. Teil 1. Allgemeines und Encyonema Part. Bibl Diatomol 36:1–382
- Krammer K (1997b) Die cymbelloiden Diatomeen. Teil 2. Encyonema part., Encyonopsis and Cymbellopsis. Bibl Diatomol 37:1–469
- Krammer K, Lange-Bertalot H (1986) Süßwasserflora von Mitteleuropa, Bacillariophyceae, vol 1–5. Gustav Fischer Verlag, Stuttgart
- Krammer K, Lange-Bertalot H (2004) Süßwasserflora von Mitteleuropa, Bd. 02/4: Bacillariophyceae: Teil 4: Achnanthaceae, kritische Ergänzungen zu Achnanthes sl, Navicula s. str. Spektrum Akademischer Verlag, Heidelberg
- Lange-Bertalot H (1993) 85 Neue Taxa und über 100 weitere neu definierte Taxa erganzend zur Süßwasserflora von Mitteleuropa. Bibl Diatomol 27:1–454
- Lange-Bertalot H (2001) Navicula sensu stricto, 10 genera separated from Navicula sensu lato. In: Lange-Bertalot H (ed) Diatoms of Europe 2. A.R.G. Gantner Verlag K.G., Ruggell, p 526



- Lange-Bertalot H, Krammer K (1989) Achnanthes, eine Monographie der Gattung. Schweizerbart Science Publisher, Stuttgart
- Lavoie I, Dillon PJ, Campeau S (2009) The effect of excluding diatom taxa and reducing taxonomic resolution on multivariate analyses and stream bioassessment. Ecol Indic 9:213–225. https://doi.org/10.1016/j.ecolind.2008.04.003
- Leathwick JR, Rowe D, Richardson J et al (2005) Using multivariate adaptive regression splines to predict the distributions of New Zealand's freshwater diadromous fish. Freshw Biol 50:2034–2052. https://doi.org/10.1111/j.1365-2427.2005.01448.x
- Legendre P, Fortin MJ (1989) Spatial pattern and ecological analysis. Vegetatio 80:107–138. https://doi.org/ 10.1007/BF00048036
- Legendre P, Legendre L (1998) Numerical ecology, 2nd edn. Elsevier, Amsterdam
- Leibold MA, Chase JM (2018) Metacommunity ecology. Princeton University Press, Oxford
- Leibold MA, Holyoak M, Mouquet N et al (2004) The metacommunity concept: a framework for multi-scale community ecology. Ecol Lett 7:601–613. https://doi.org/10.1111/j.1461-0248.2004.00608.x
- Leibold MA, Economo EP, Peres-Neto P (2010) Metacommunity phylogenetics: separating the roles of environmental filters and historical biogeography. Ecol Lett 13:1290–1299. https://doi.org/10.1111/j.1461-0248.2010.01523.x
- Levkov Z (2009) Amphora sensu lato. In: Lange-Bertalot H (ed) Diatoms of Europe 5. Diatoms of the European inland waters and comparable habitats. A.R.G. Gantner Verlag K.G., Ruggell
- Liu J, Soininen J, Han BP, Declerck SAJ (2013) Effects of connectivity, dispersal directionality and functional traits on the metacommunity structure of river benthic diatoms. J Biogeogr 40:2238–2248. https://doi. org/10.1111/jbi.12160
- Mazaris AD, Moustaka-Gouni M, Michaloudi E, Bobori DC (2010) Biogeographical patterns of freshwater micro- and macro-organisms: a comparison between phytoplankton, zooplankton and fish in the eastern Mediterranean. J Biogeogr 37:1341–1351. https://doi.org/10.1111/j.1365-2699.2010.02294.x
- McCluney KE, Poff NL, Palmer MA et al (2014) Riverine macrosystems ecology: sensitivity, resistance, and resilience of whole river basins with human alterations. Front Ecol Environ 12:48–58. https://doi.org/10.1890/120367
- McGill BJ, Enquist BJ, Weiher E, Westoby M (2006) Rebuilding community ecology from functional traits. Trends Ecol Evol 21:178–185. https://doi.org/10.1016/j.tree.2006.02.002
- Menezes S, Baird DJ, Soares AMVM (2010) Beyond taxonomy: a review of macroinvertebrate trait-based community descriptors as tools for freshwater biomonitoring. J Appl Ecol 47:711–719. https://doi.org/10.1111/j.1365-2664.2010.01819.x
- Nekola JC, White PS (1999) The distance decay of similarity in biogeography and ecology. J Biogeogr 26:867–878. https://doi.org/10.1046/j.1365-2699.1999.00305.x
- Novais MH, Blanco S, Hlúbiková D et al (2009) Morphological examination and biogeography of the *Gomphonema rosenstockianum* and *G. tergestinum* species complex (Bacillariophyceae). Fottea 9:257–274. https://doi.org/10.5507/fot.2009.026
- Oksanen J, Blanchet FG, Friendly M et al (2019) vegan: Community ecology package. R package version 2.6-2. https://CRAN.R-project.org/package=vegan
- Paavola R, Muotka T, Virtanen R et al (2003) Are biological classifications of headwater streams concordant across multiple taxonomic groups? Freshw Biol 48:1912–1923. https://doi.org/10.1046/j.1365-2427. 2003.01131.x
- Passy SI (2007) Diatom ecological guilds display distinct and predictable behavior along nutrient and disturbance gradients in running waters. Aquat Bot 86:171–178. https://doi.org/10.1016/j.aquabot.2006.09.018
- Peñas FJ, Barquín J (2019) Assessment of large-scale patterns of hydrological alteration caused by dams. J Hydrol 572:706–718. https://doi.org/10.1016/j.jhydrol.2019.03.056
- Peñas FJ, Barquín J, Snelder TH et al (2014) The influence of methodological procedures on hydrological classification performance. Hydrol Earth Syst Sci 18:3393–3409. https://doi.org/10.5194/hess-18-3393-2014
- Peres-Neto PR, Legendre P (2010) Estimating and controlling for spatial structure in the study of ecological communities. Glob Ecol Biogeogr 19:174–184. https://doi.org/10.1111/j.1466-8238.2009.00506.x
- Peres-Neto PR, Legendre P, Dray S, Borcard D (2006) Variation partitioning of species data matrices: estimation and comparison of fractions. Ecology 87:2614–2625. https://doi.org/10.1890/0012-9658(2006) 87[2614:VPOSDM]2.0.CO;2
- Prygiel J, Coste M (2000) Guide méthodologique pour la mise en oeuvre de l'Indice biologique diatomées
- R Core Team (2020) R: a language and environment for statistical computing. R Core Team, Vienna
- Reid AJ, Carlson AK, Creed IF et al (2019) Emerging threats and persistent conservation challenges for freshwater biodiversity. Biol Rev 94:849–873. https://doi.org/10.1111/brv.12480
- Rimet F, Bouchez A (2012) Life-forms, cell-sizes and ecological guilds of diatoms in European rivers. Knowl Manag Aquat Ecosyst. https://doi.org/10.1051/kmae/2012018



- Rivas-Martínez S, Penas A, Díaz TE (2004) Bioclimatic and biogeographic maps of Europe. Bioclimates. Cartographic Service, University of León, León
- Schmidt-Kloiber A, Hering D (2015) Www.freshwaterecology.info—an online tool that unifies, standardises and codifies more than 20,000 European freshwater organisms and their ecological preferences. Ecol Indic 53:271–282. https://doi.org/10.1016/j.ecolind.2015.02.007
- Seymour M, Deiner K, Altermatt F (2016) Scale and scope matter when explaining varying patterns of community diversity in riverine metacommunities. Basic Appl Ecol 17:134–144. https://doi.org/10.1016/j.baae.2015.10.007
- Shurin JB, Cottenie K, Hillebrand H (2009) Spatial autocorrelation and dispersal limitation in freshwater organisms. Oecologia 159:151–159. https://doi.org/10.1007/s00442-008-1174-z
- Siqueira T, Bini LM, Roque FO, Cottenie K (2012) A metacommunity framework for enhancing the effectiveness of biological monitoring strategies. PLoS ONE 7:e43626. https://doi.org/10.1371/journal.pone. 0043626
- Siqueira T, Durães LD, de Oliveira RF (2014) Predictive modelling of insect metacommunities in biomonitoring of aquatic networks. In: Ferreira CP, Godoy WAC (eds) Ecological modelling applied to entomology. Springer, Cham, pp 109–126
- Smucker NJ, Vis ML (2011) Spatial factors contribute to benthic diatom structure in streams across spatial scales: considerations for biomonitoring. Ecol Indic 11:1191–1203. https://doi.org/10.1016/j.ecolind. 2010.12.022
- Soininen J, McDonald R, Hillebrand H (2007) The distance decay of similarity in ecological communities. Ecography (Cop) 30:3–12. https://doi.org/10.1111/j.2006.0906-7590.04817.x
- Soininen J, Paavola R, Kwandrans J, Muotka T (2009) Diatoms: unicellular surrogates for macroalgal community structure in streams? Biodivers Conserv 18:79–89. https://doi.org/10.1007/s10531-008-9447-8
- Soininen J, Jamoneau A, Rosebery J, Passy SI (2016) Global patterns and drivers of species and trait composition in diatoms. Glob Ecol Biogeogr 25:940–950. https://doi.org/10.1111/geb.12452
- Statzner B, Bêche LA (2010) Can biological invertebrate traits resolve effects of multiple stressors on running water ecosystems? Freshw Biol 55:80–119. https://doi.org/10.1111/j.1365-2427.2009.02369.x
- Sundermann A, Stoll S, Haase P (2011) River restoration success depends on the species pool of the immediate surroundings. Ecol Appl 21:1962–1971. https://doi.org/10.1890/10-0607.1
- Swan CM, Brown BL (2017) Metacommunity theory meets restoration: isolation may mediate how ecological communities respond to stream restoration. Ecol Appl 27:2209–2219. https://doi.org/10.1002/eap.1602
- Tonkin JD, Altermatt F, Finn DS et al (2018) The role of dispersal in river network metacommunities: patterns, processes, and pathways. Freshw Biol 63:141–163. https://doi.org/10.1111/fwb.13037
- Trobajo R, Rovira L, Ector L et al (2013) Morphology and identity of some ecologically important small Nitzschia species. Diatom Res 28:37–59. https://doi.org/10.1080/0269249X.2012.734531
- Tuomisto H, Ruokolainen L, Ruokolainen K (2012) Modelling niche and neutral dynamics: on the ecological interpretation of variation partitioning results. Ecography (Cop) 35:961–971. https://doi.org/10.1111/j. 1600-0587.2012.07339.x
- Usseglio-Polatera P, Bournaud M, Richoux P, Tachet H (2000) Biological and ecological traits of benthic freshwater macroinvertebrates: relationships and definition of groups with similar traits. Freshw Biol 43:175–205. https://doi.org/10.1046/j.1365-2427.2000.00535.x
- Vandewalle M, de Bello F, Berg MP et al (2010) Functional traits as indicators of biodiversity response to land use changes across ecosystems and organisms. Biodivers Conserv 19:2921–2947. https://doi.org/10.1007/s10531-010-9798-9
- Vellend M, Srivastava DS, Anderson KM et al (2014) Assessing the relative importance of neutral stochasticity in ecological communities. Oikos 123:1420–1430. https://doi.org/10.1111/oik.01493
- Verleyen E, Vyverman W, Sterken M et al (2009) The importance of dispersal related and local factors in shaping the taxonomic structure of diatom metacommunities. Oikos 118:1239–1249. https://doi.org/10.1111/j.1600-0706.2009.17575.x
- Viana DS, Chase JM (2019) Spatial scale modulates the inference of metacommunity assembly processes. Ecology 100:1–9. https://doi.org/10.1002/ecy.2576
- Vilmi A, Karjalainen SM, Hellsten S, Heino J (2016) Bioassessment in a metacommunity context: are diatom communities structured solely by species sorting? Ecol Indic 62:86–94. https://doi.org/10.1016/j.ecolind. 2015.11.043
- Vilmi A, Tolonen KT, Karjalainen SM, Heino J (2017) Metacommunity structuring in a highly-connected aquatic system: effects of dispersal, abiotic environment and grazing pressure on microalgal guilds. Hydrobiologia 790:125–140. https://doi.org/10.1007/s10750-016-3024-z
- Vörösmarty CJ, McIntyre PB, Gessner MO et al (2010) Global threats to human water security and river biodiversity. Nature 467:555–561. https://doi.org/10.1038/nature09440



Wagner HH, Dray S (2015) Generating spatially constrained null models for irregularly spaced data using Moran spectral randomization methods. Methods Ecol Evol 6:1169–1178. https://doi.org/10.1111/2041-210X.12407

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

