



Navigating tumor angiogenesis: therapeutic perspectives and myeloid cell regulation mechanism

Fan Yang^{1,2,3} · Gloria Lee¹ · Yi Fan¹

Received: 31 December 2023 / Accepted: 4 March 2024
© The Author(s) 2024

Abstract

Sustained angiogenesis stands as a hallmark of cancer. The intricate vascular tumor microenvironment fuels cancer progression and metastasis, fosters therapy resistance, and facilitates immune evasion. Therapeutic strategies targeting tumor vasculature have emerged as transformative for cancer treatment, encompassing anti-angiogenesis, vessel normalization, and endothelial reprogramming. Growing evidence suggests the dynamic regulation of tumor angiogenesis by infiltrating myeloid cells, such as macrophages, myeloid-derived suppressor cells (MDSCs), and neutrophils. Understanding these regulatory mechanisms is pivotal in paving the way for successful vasculature-targeted cancer treatments. Therapeutic interventions aimed to disrupt myeloid cell-mediated tumor angiogenesis may reshape tumor microenvironment and overcome tumor resistance to radio/chemotherapy and immunotherapy.

Keywords Tumor angiogenesis · Anti-angiogenic therapy · Vessel normalization · Endothelial reprogramming · Macrophages · MDSCs · Neutrophils · Therapy resistance · Immunotherapy · Radiochemotherapy

Introduction

Angiogenesis, the formation of new blood vessels from pre-existing ones, is a hallmark of cancer. Tumor angiogenesis is a pivotal process that promotes cancer growth, progression, and metastasis and induces therapy resistance [1]. Over the past few decades, considerable efforts have been directed towards understanding the molecular and cellular mechanisms that underlie tumor angiogenesis. These lead to the development of promising anti-angiogenic therapeutic strategies that aim to inhibit overgrowth and sprouting of tumor endothelial cells (ECs). Beyond the traditional

anti-angiogenic concept that focuses on vessel delivery function, recent advances have revealed that the interactions between the tumor vasculature and the immune system are critical for regulation of tumor vascularity and immunity [2, 3]. The tumor microenvironment (TME) is a complex milieu composed of various non-neoplastic cell types, including ECs, stromal cells, and a diverse array of immune cells. The dynamic interplay between these cellular components in the vascular TME has significant implications for tumor development, immune evasion and the efficacy of cancer therapies, particularly immunotherapies. Therefore, development of efficient therapeutic strategies that reprogram the vascular TME will offer exciting opportunities for cytotoxic radio/chemotherapy and T cell-based immunotherapy.

In this review, we discuss the emerging strategies for tumor vasculature-targeting therapy. We provide a comprehensive overview of the complex regulation of tumor angiogenesis by myeloid cells, including macrophages, myeloid-derived suppressor cells (MDSCs), and neutrophils within the TME. We discuss the impact of these immune cells on tumor angiogenesis. We highlight that myeloid cells interact with ECs to regulate tumor angiogenesis and create a specialized niche that induces immune evasion and promotes tumor growth, providing crucial targets for vasculature-targeting therapy. These approaches may have the potential

✉ Fan Yang
fanyangforever@gmail.com

✉ Yi Fan
fanyi@upenn.edu

¹ Present Address: Department of Radiation Oncology, University of Pennsylvania, Philadelphia, PA 19104, USA

² Department of Obstetrics and Gynecology, Ren Ji Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai 200127, China

³ Shanghai Key Laboratory of Gynecologic Oncology, Ren Ji Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai 200127, China

to revolutionize cancer treatment, paving the way for more effective therapeutic strategies.

Tumor angiogenesis

Basic principle of aberrant tumor angiogenesis

Tumor angiogenesis is fundamental to cancer progression, metastasis, and therapy resistance. Tumor angiogenesis refers to the pathophysiological process where new blood vessels sprout from pre-existing ones to supply nutrients, oxygen, and cellular network for tumor growth [1, 4–7]. The intricate network of blood vessels also allows cancer cells to infiltrate the bloodstream and disseminate throughout the body, giving rise to metastasis. These collectively suggest anti-angiogenic therapy, a treatment that aims to inhibit EC overgrowth and sprouting, as a promising strategy for cancer treatment. Notably, the newly formed vessels are structurally and functionally abnormal—they are tortuous and leaky with a disorganized, haphazard architecture. This abnormal vasculature leads to a chaotic blood flow, which creates a heterogeneously hypoxic tumor microenvironment [8]. Such hostile conditions can foster cancer cells that are more aggressive and therapy resistant, further promoting tumor growth and metastasis. Furthermore, the abnormal vessels also form a barrier to the effective delivery of drugs to the tumor, thereby contributing to therapy resistance [9].

Vessel normalization has, therefore, joined anti-angiogenic treatment as promising strategies for solid tumor treatment.

Tumor angiogenesis is a complex process, subject to regulation by a balance between pro-angiogenic and anti-angiogenic factors within a solid tumor [10, 11]. When the equilibrium tilts toward pro-angiogenic factors, ECs are stimulated to proliferate and migrate towards the tumor, forming new blood vessels. The presence of excessive pro-angiogenic factors further stimulates vascular abnormalities [12, 13]. This imbalance drives both neovascularization and vascular aberrancy, serving as a critical therapeutic target for vessel normalization and cancer treatment. Finally, tumor ECs undergo genetic and metabolic alteration to acquire pro-tumor phenotypes including aberrant vessel structure and function, a rewired adhesome that reduces lymphocyte attachment, and local release of immunosuppressive molecules. Thus, anti-angiogenesis, vessel normalization, and endothelial programming serve as promising strategies for vasculature-targeting approaches for cancer treatment (Fig. 1).

Anti-angiogenic therapy

Anti-VEGF/VEGFR

Anti-angiogenic therapy represents a promising strategy for cancer treatment by inhibiting the formation of new blood vessels that nourish tumors, thus depriving them of

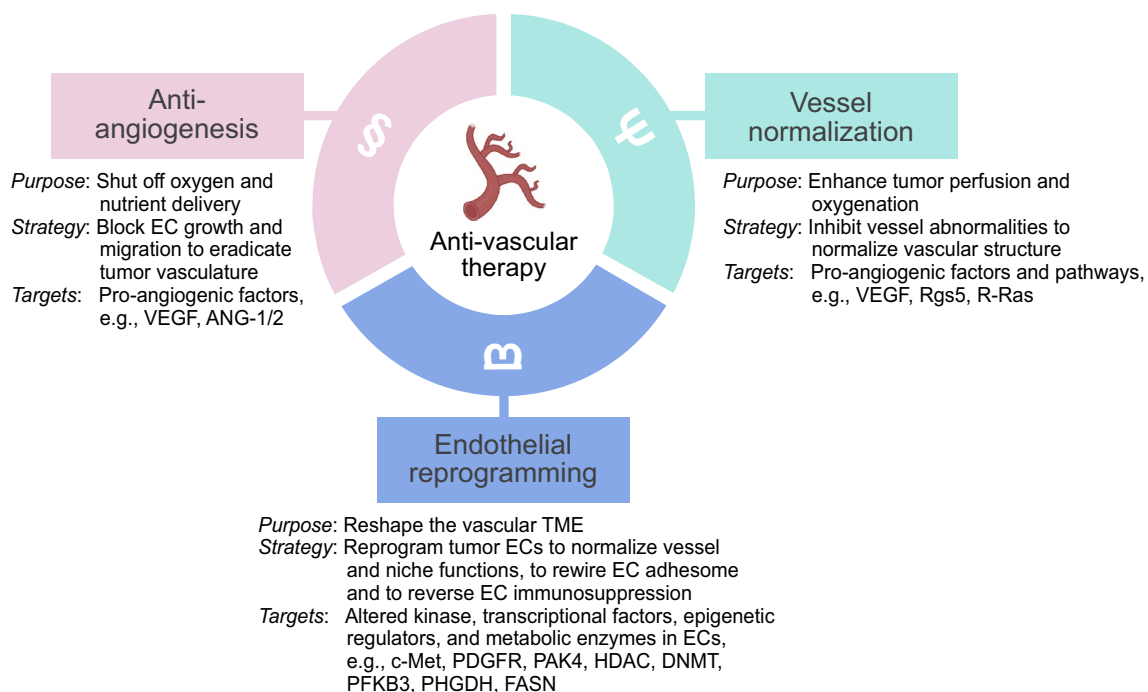


Fig. 1 Therapeutic strategies for vasculature-targeting anti-cancer treatment. Therapeutic strategies targeting tumor vasculature have emerged as transformative for cancer treatment, encompassing anti-angiogenesis, vessel normalization, and endothelial reprogramming

essential oxygen and nutrients for growth. Numerous anti-angiogenic agents targeting pro-angiogenic factors, such as vascular endothelial growth factor (VEGF, i.e., VEGF-A), fibroblast growth factors (FGF), and epidermal growth factor (EGF), have been extensively explored [14–16]. Among the most widely used anti-angiogenic agents are the monoclonal antibodies and tyrosine kinase inhibitors (TKIs) that target VEGF and the VEGF downstream kinases, respectively. VEGF plays a crucial role in both physiological and pathological angiogenesis [4]. In tumors, overexpression of VEGF, mainly driven by hypoxia-inducible factor (HIF)-1 α , promotes abnormal blood vessel growth and acts as a vascular permeability factor [17]. VEGF usually binds to the tyrosine kinase receptor VEGFR2, in collaboration with neuropilin-1 and VEGFR3, and interacts with other modulating pathways such as Notch, angiopoietin/Tie2, and ephrin/Eph to facilitate vessel growth [18–22]. A number of anti-angiogenic agents have been approved by FDA for treating cancer, highlighting their role in current oncology therapeutics. For instance, bevacizumab (Avastin), a humanized monoclonal antibody that blocks VEGF, is a notable example of an anti-angiogenic agent demonstrating anti-tumor results in colon and kidney cancers [23]. Additionally, small-molecule pharmacological inhibitors of VEGF receptor tyrosine kinase, such as sunitinib and sorafenib, also offer a promising opportunity to cancer therapy [24, 25]. However, the overall efficacy of these anti-angiogenic therapies is often limited and does not produce long-term benefits in patients with most other cancer types, such as glioblastoma [26–30]. Both intrinsic and acquired mechanisms contribute to tumor resistance to anti-angiogenic therapy, driven by the existence of redundant angiogenic pathways and the adaptive mechanisms that lead tumor cells to survive an avascular and hypoxic TME, respectively [6, 28].

Inhibition of vascular maturation

Inhibition of vascular maturation, a key aspect of functional vascularity, represents another therapeutic strategy for cancer. The EC growth factor signaling pathways composed of angiopoietin (Ang)-1/2 and their receptor, Tie2, play a critical role in this process. Ang-1, mainly secreted by pericytes and smooth muscle cells, promotes vascular remodeling and stabilization. Ang-1 overexpression is often observed in tumor vasculature, which enhances EC proliferation and pericyte-mediated vascular maturation, and increased vascular functions intensify the malignancy of various cancers [31]. Conversely, Ang-2 can induce angiogenesis and destabilize vasculature by binding to Tie2 and integrin receptors [32]. Given the role of angiopoietins in vascular biology, antibodies targeting these angiopoietins and dual inhibitors

of Ang-2 and VEGF show promising results in various malignancies [20].

Targeting the development pathways of ECs

Tumor angiogenesis is tightly controlled by EC differentiation and growth. This process can be triggered by hypoxia [33], Notch [34], and Wnt signaling pathways [35]. Hypoxia, typically resulting from rapid tumor growth and disordered vasculature, initiates a survival response within tumors. To survive under these extreme conditions, tumors employ a host of mechanisms, primarily the activation of HIFs that induce transcription of hypoxia-adaptive metabolic enzyme and VEGF [36, 37]. Moreover, hypoxia can stimulate activation of mTOR, PI3K, and AKT through post-translational modifications of these proteins [38], which are central to EC metabolism, survival, and motility regulation in response to nutrient and oxygen depletion. Notch signaling emerges as a pivotal player in the orchestration of vessel sprouting, branching, and maturation. Aberrations in Notch signaling have been linked to tumor angiogenesis, positioning the Notch pathway as a potential target for anti-angiogenic cancer therapies [34]. A recent study on Dll4-induced Notch signaling in EC growth and development shows that antibodies against Dll4 and VEGF had strikingly different effects on tumor blood vessels [39], suggesting differing mechanisms underlying Notch- and VEGF-mediated tumor angiogenesis. Dll4-driven Notch signaling appeared vital during active blood vessel formation, but less so for maintaining normal vessels [39]. Beside its established role in developmental angiogenesis and vascular differentiation, Wnt pathway has been implicated in tumor angiogenesis. The interaction between Wnt and Frizzled receptors activates varied signaling outcomes in both canonical and non-canonical pathways, contributing to regulation of EC functions. For example, canonical Wnt-frizzled signaling produces a β -catenin/Lef/TCF complex which triggers brain angiogenesis during development [40]. In the context of GBM, activation of Wnt/ β -catenin signaling in ECs is associated with chemoresistance [41], highlighting a potential target in GBM treatment. On the other hand, in the non-canonical pathway, Ca²⁺/calmodulin-dependent protein kinase II (CAMKII) activation influences ventral cell fate [42], and other signaling cascades like JNK and Daam-1 drive EC proliferation and maintain cellular architecture [43–45].

Overall, these findings highlight multiple regulatory mechanisms, mediated through hypoxia, VEGF, Notch, and Wnt, for EC proliferation, migration, and differentiation during tumor angiogenesis. Understanding their dysregulation in cancer may help develop new targets for anti-angiogenic therapies.

Vessel normalization therapy

Anti-angiogenic therapy can adversely enhance tumor hypoxia and reduce drug delivery, resulting from destroyed tumor vasculature, leading to increased resistance of tumors to radio/chemotherapy and targeted molecular therapy. Considering structurally and functionally abnormal vascularity in cancer, i.e., tortuous, leaky vasculature due to dysfunctional EC sprouting and overgrowth, a different strategy, namely, vessel normalization, that aims to restore normal vessel function, has been exploited [11]. This could be achieved by re-balancing the pro- and anti-angiogenic factors presented in the TME, with reduced hypoxia, improved perfusion allowing for proper drug delivery, and enhanced immune cell infiltration [12, 13]. Previous preclinical studies show that vessel normalizing doses of anti-VEGF treatment improve T cell infiltration and enhance immunotherapy [46, 47], due to enhanced vessel delivery and reduced intratumoral hypoxia. Moreover, a recent clinical trial shows promising results for combining anti-VEGF bevacizumab with immune checkpoint blockade in liver cancer treatment [29].

Additional therapeutic strategies for vessel normalization include decreasing vascular leakiness, enhancing the structural integrity, increasing perfusion, and adding angiostatic factors, with multiple targets identified. For example, targeting regulator of G protein signaling 5 (Rgs5) protein leads to more typical vessel morphology and function in tumors, without reducing vessel density [48]. Inhibiting L1CAM, a neural adhesion protein in tumor ECs, results in pruning and fortification of vessels, thereby reducing tumor growth and metastases [49]. Inhibition of neuronal nitric oxide synthase (nNOS) in cancer cells restores proper NO gradients, leading to denser and more effective vessels for oxygen and drug delivery [50]. Restoring semaphorin-3A (SEMA3A) initially prunes immature vessel, and long-term application increases vessel maturation [51]. Activation of R-Ras or lysophosphatidic acid (LPA) in ECs promotes vascular normalization [52, 53]. Chloroquine, known for its antimalarial properties, also plays a role in vessel normalization through endosomal Notch1 trafficking and signaling in ECs [54]. Activation of transient receptor potential vanilloid-4 (TRPV4) in tumor ECs restores normal mechanosensitivity and increases drug delivery [55]. Further strategies include using thrombospondin-1 (TSP-1), an endogenous antiangiogenic factor, to normalize vessels, enhance drug delivery, and increase the effectiveness of treatments like cisplatin [56].

Another innovative approach in vessel normalization involves modulating various cells within the perivascular niche. For instance, eribulin, a chemotherapy agent, regulates endothelial-pericyte interactions to fortify vessels and improve treatment outcomes [57]. Desmoplasia,

characterized by fibrotic tissue growth, impairs vascular function by compressing vessels [13]. Therefore, normalizing the extracellular matrix (ECM) is crucial, as it can improve both vascular function and treatment outcomes. Strategies targeting cancer-associated fibroblasts and the extracellular matrix, such as inhibiting TGF- β [58] or sonic hedgehog pathways [59], and using Nab-paclitaxel [60], show promise in reducing vessel compression. Additionally, altering metabolic pathways in pro-tumor macrophages leads to the formation of well-organized and fortified vessels, thereby enhancing oxygen delivery [61]. Antitumor CD4⁺ T cells also play a role in normalizing vessels by modulating angiogenic gene expression in tumors [62]. Inhibiting VEGF expression from these T cells further suggests their role in promoting abnormal tumor vessel phenotypes [63]. These findings highlight the complex interactions among ECs, other cells, and ECM in the TME, which may induce vessel abnormalities. Understanding and targeting these interactions can normalize tumor vasculature and improve cancer therapy outcomes.

Although these strategies hold promise, the benefits of vessel normalization monotherapy have often been small and transient. For example, administration of low-dose bevacizumab to control excessive EC growth has been a central method used in vessel normalization. However, vessel normalization anti-VEGF therapies often lead to a transient window that is potentially open for additional therapies, after which tumors become resistant [64, 65]. Furthermore, the timing and dosing of vessel normalization therapy needs to be further optimized when combined with immunotherapies and other conventional cytotoxic therapies, as tumor immunogenicity and vascularity change over tumor development and treatment exposure [6, 13].

Endothelial reprogramming therapy

EC plasticity has been well characterized during embryogenesis [66, 67]. In pathological settings including cardiac, renal, and liver fibrosis, ossifying myositis, pulmonary hypertension, and cerebral cavernous malformation, ECs can take endothelial mesenchymal transition (Endo-MT) de novo to generate fibroblasts and stem-like cells [68–70]. Notably, cell plasticity plays a central role in the EC transcriptomic alteration and aberrant vascular phenotypes in cancer [71, 72]. As an alternative process to angiogenesis and vascular abnormality driven by pro-angiogenic factor-induced vessel sprouting and outgrowth, ECs retain key endothelial functions but undergo cell plasticity-mediated genetic reprogramming to induce aberrant vascularity in the tumor microenvironment. For example, ECs acquire partial Endo-MT, also known as endothelial transformation, to promote their ability to proliferate, migrate and secrete [71–73]. These transformed

ECs, unlike normal ECs, take transcriptomic alteration to drive distinct behaviors due to the influence of the TME, forming an abnormal architecture of tumor vasculature. This leads to poor perfusion and hypoxia within the TME, which fosters the selection of more aggressive, treatment-resistant cancer cells [74], and creates a physical barrier that shields tumor cells from immune cell attack and impedes the delivery of chemotherapeutic drugs, thereby inducing tumor resistance to chemo/radiotherapy and immunotherapy [7, 28]. The strategy for genetic reprogramming of tumor ECs, e.g., by targeting EC plasticity, aims to normalize these cells by reversing their abnormal traits of gene expression, making the vasculature resemble the normal one in structure and function, and, therefore, may eventually improve the efficacy of cytotoxic treatment and immunotherapy approaches [71]. In addition to transcriptomic alteration, tumor ECs also undergo metabolic changes in the TME [75]. Metabolic switches in tumor ECs are driven by genetic and epigenetic alteration of metabolism-associated genes in response to the cues in the TME, such as hypoxia. The adaptively rewired metabolism fosters EC survival and growth in the TME, contributing to aberrant tumor angiogenesis. Metabolic reprogramming of tumor ECs, therefore, serves as an additional strategy for vasculature-targeting cancer therapy [75].

Genetic reprogramming of ECs

The approach of genetic reprogramming of tumor ECs is initially termed as vascular de-transformation therapy, emphasizing its main target on EC plasticity [71]. Genetic reprogramming of tumor ECs would be expected to induce the formation of a stable, functionally normal, and structurally orderly vasculature, which reduces tumor hypoxia, improves drug delivery, and alleviates immunosuppression, thereby enhancing anti-tumor immune responses and the efficacy of other therapies [72]. Several strategies have been exploited for the genetic reprogramming of tumor ECs. HGF/c-Met is identified as a critical regulator of Endo-MT in cancer [73]. EC-specific c-Met knockout inhibits EC plasticity, reduces vascular aberrancy, and sensitizes tumor to chemotherapy [73]. Moreover, c-Met-mediated activation of Wnt signaling drives transformation of ECs into mesenchymal stem cell-like cells, leading to multidrug resistance in ECs and tumor chemoresistance [41]. Furthermore, platelet-derived growth factor (PDGF)-mediated EC plasticity controls VEGFR2 expression through Snail, which contributes to tumor resistance to anti-VEGF treatment [9]. Based on these results, combination of anti-PDGFR and anti-VEGFR therapy was explored in tumor, which shows promising synergistic anti-tumor effects [9]. A more recent whole kinome-wide screen identifies that p21-activating kinase 4 (PAK4) is a key driver of Endo-MT in

cancer [76]. Inactivation of PAK4 reprograms transcriptome in ECs and normalizes tumor vasculature. Notably, genetic and pharmacological ablation of PAK4 in ECs reshapes the immune landscape within the TME, improving T-cell infiltration and sensitizing tumor to CAR-T cell therapy [76]. Furthermore, several additional targets have been identified for endothelial reprogramming, including ERG, Myct1, and Lrg1. Forced expression of ERG, a transcription factor essential for endothelial homeostasis, restores the angiogenic balance in tumor ECs, thereby inhibiting tumor growth and vascular abnormalities [77]. Interestingly, Myct1, a downstream protein of ETV2 and Myc, which is primarily expressed in ECs, plays a crucial role in mesenchymal-like transcriptional activation. Myct1 deficiency in mouse tumor models decreases angiogenesis and increases antitumor immunity, thereby limiting tumor growth [78]. Lrg1 is exclusively expressed on tumor ECs rather than normal ECs or pericytes. Deletion or antibody-based neutralization of Lrg1 results in vessel normalization and promotes the TME toward an anti-tumor, immune-active state, enhancing the efficacy of various cancer therapies [79].

In addition to structural abnormalities, tumor vasculature is also characterized by altered EC adhesiveness. Tumor ECs undergo genetic alteration, often with downregulated adhesion proteins, such as intercellular adhesion molecule 1 (ICAM-1) and vascular cell adhesion molecule 1 (VCAM-1) that are necessary for immune cell attachment and extravasation [75, 80]. This leads to less T cell attachment to the endothelium, inhibiting T cell infiltration and contributing to tumor immune evasion. It is tempting to speculate that mesenchymal-like activation drives this dysfunctional adhesion in tumor ECs, induced by epithelial-mesenchymal transition (EMT)-associated transcriptional repressors including Snail, Slug, Twist-1/-2, and Zeb-1/-2. As such, inhibition of PAK4 reduces expression of Slug and Zeb-1, upregulating expression of VCAM-1 and Claudin-14 in tumor ECs, which eventually enhances T cell adhesion and improves CAR T cell immunotherapy [76]. Together, these findings underscore the potential of genetic reprogramming of tumor ECs as a promising approach for cancer treatment.

Epigenetic reprogramming of ECs

Epigenetic reprogramming in ECs represents another promising strategy for targeting tumor angiogenesis, considering tumor ECs undergo substantial epigenetic alterations to modulate their functionality in cancer. Acetylation of histone H3 has been well characterized in tumor ECs, which epigenetically regulates the expression of key genes essential for EC function and angiogenesis, including CLU, FBN1, TSPAN2, and ICAM1 [81]. The activity of histone deacetylases (HDACs), especially

HDAC1, is central to this process: they regulate MMP14 and VCAM-1 expression, driving EC growth and the formation of vascular structures [82, 83]. Inhibitors targeting HDACs, such as trichostatin A (TSA) and suberoylanilide hydroxamic acid (SAHA), hold promise in anti-angiogenesis therapy, as they modulate the transcription of several crucial pro-angiogenic signaling components, including receptors VEGFR1 and VEGFR2 [84], HIF-1 α , and VEGF [85]. HDAC inhibitors not only exhibit anti-angiogenic properties across various cancer types but also enhance leukocyte adherence and movement within tumor vessels, primarily through the upregulation of ICAM-1 [86], underscoring their potential to boost the effectiveness of immunotherapy. Moreover, histone methylation is also critical for tumor angiogenesis. EZH2, a key histone methyltransferase, reduces trimethylation of histone H3 at lysine 27 (H3K27me3), a repressive epigenetic mark, during Endo-MT induced by IL-1 β and TGF- β 2 [87]. Conversely, JMJD2B, a histone demethylase, epigenetically modulates Endo-MT by promoting repressive H3K9me3 occurring at the promoters of mesenchymal and TGF- β signaling genes, such as calponin (CNN1), AKT serine/threonine kinase 3 (AKT3), and sulfatase 1 (SULF1) [88].

Beyond histone modifications, DNA methylation significantly influences the behavior of tumor ECs and, consequently, the immune profiles as well. For instance, deletion of DNA methyltransferase 1 (DNMT1) in ECs inhibits tumor growth and reshapes the immune environment, due to the increased expression of cytokines, chemokines, cell adhesion molecules in ECs, such as Cxcl9 and Cxcl10 that are crucial for infiltration of CD8⁺ T cells into the tumor [89]. DNMT1 silencing in ECs also enhances the expression of IL-33, Ccl21, and Ccl19 that are critical for neogenesis of high endothelial venule (HEV), a specialized postcapillary venule adapted for lymphocyte trafficking. Moreover, DNMT inhibitor treatment boosts leukocyte infiltration into tumors by upregulating ICAM1 expression in ECs [86]. Interestingly, proangiogenic factor FGF2 promotes ERK-mediated DNMT1 phosphorylation and nuclear translocation to repress Cxcl9 and Cxcl10 transcription [89], suggesting feedback loops that regulate angiogenic pathway activation and epigenetic regulation.

In summary, recent studies identifying an intricate network of epigenetic regulation in ECs during tumor angiogenesis provide profound insights into the mechanisms driving epigenetic regulation of EC functions, and opens new avenues for developing therapeutic strategies targeting these epigenetic alterations to inhibit tumor growth and enhance immunotherapy outcomes.

Metabolic reprogramming of ECs

Given metabolic adaptation is required for cell proliferation and migration, such as EC outgrowth and sprouting, targeting endothelial metabolism has emerged as a promising strategy for modulating tumor angiogenesis [75, 90–95]. This strategy may not only rewire tumor vasculature by targeting EC sprouting, but also recondition the metabolic TME as it changes the EC-derived metabolites that are locally released. A key regulatory pathway of endothelial metabolism is glycolysis, a process critical for EC survival and proliferation in the hypoxic TME, as it generates necessary energy and metabolites anaerobically. For instance, disruption of glycolysis via PFKFB3 inhibition stabilizes the vascular barrier by improving pericyte adhesion, reduces metastasis, and enhances the efficacy of cancer chemotherapy [90, 95]. Furthermore, decreasing aerobic glycolysis in tumor ECs reduces vascular abnormalities, increases T cell infiltration, and overcomes tumor resistance to immunotherapy [96]. Notably, PHGDH, which diverts glycolysis into a specific serine biosynthetic pathway, promotes aberrant tumor angiogenesis, through its role in regulating nucleotide synthesis and maintaining the redox balance essential for endothelial proliferation [94]. Endothelial metabolism also contributes to the immunosuppressive TME by providing immunomodulatory metabolites produced by the vascular niche. As such, inhibition of serine metabolism in tumors ECs reduces their production of lactate and 2-hydroxyglutarate, two immunosuppressants in the TME, leading to activation of anti-tumor immunity [94]. Beyond glucose metabolism, other metabolic pathways in ECs are also being explored as therapeutic targets. Loss of endothelial ADRB2, which encodes the β 2-adrenergic receptor, leads to angiogenesis inhibition through oxidative phosphorylation [97]. Similarly, disrupting fatty acid metabolism in ECs, as evidenced by that knockdown of fatty acid synthase (FASN) and the loss of CPT1A, a critical enzyme in fatty acid oxidation (FAO), limit vessel sprouting and proliferation through mTOR signaling and nucleotide synthesis, indicating the role of lipid metabolism in maintaining the physical structure of tumor vessels [91, 92]. Additionally, restricting glutamine metabolism through glutaminase 1 (GLS1) impairs vessel sprouting due to disrupted EC proliferation and migration [93], highlighting the importance of glutamine in sustaining macromolecule production necessary for angiogenesis. Collectively, the metabolic processes within ECs are fundamental not just for their energy and biosynthetic needs but also play a pivotal role in maintaining the structural and functional integrity of blood vessels in the TME. Understanding of these regulatory pathways offer key insights into how blood vessels adapt and grow in the TME, opening up new possibilities for targeted therapies aimed at modulating tumor angiogenesis.

In summary, genetic, epigenetic and metabolic reprogramming of tumor ECs represent promising advances in vasculature-targeting therapy, with the potential to improve the efficacy of conventional cytotoxic treatments and immunotherapies [75]. There are potential drugs that may be tested for endothelial reprogramming therapy (Table 1). In addition, a number of clinical trials are currently undergoing to evaluate the synergistic effects of combining conventional anti-angiogenic agents, such as Bevacizumab and axitinib, with immunotherapies, aiming to enhance treatment efficacy and patient outcomes (Table 2).

Regulation of tumor angiogenesis by myeloid cells

The vasculature is the avenue through which circulation-derived immune cells are recruited into the solid tumors. The infiltrating immune cells are exposed to the local vascular niche and interact with ECs mainly through paracrine mechanisms. The infiltrating immune cells locally regulate vascularity, potentially modulating

sprouting angiogenesis and vascular abnormalities. Here we discuss the regulatory mechanism underlying tumor angiogenesis by myeloid cells, which may serve as key therapeutic targets for vasculature-based cancer treatment.

Tumor-infiltrating myeloid cells, mainly including macrophages, MDSCs and neutrophils, regulate tumor angiogenesis by secretion of a variety of pro-angiogenic factors. For instance, TAMs and MDSCs are known to secrete pro-angiogenic factors that stimulate EC proliferation and sprouting, leading to tumor angiogenesis and progression [98–100]. Neutrophils serve as an additional source of released pro-angiogenic factors that regulate tumor growth and metastasis [101–103]. Myeloid cells can also indirectly enhance tumor angiogenesis by expressing matrix proteases and mesenchymal-associated factors that facilitate EC migration and vascular remodeling and maturation. In addition, myeloid cells, particularly perivascular macrophages, also contribute to dynamic vascular permeability in tumor [104]. Therefore, myeloid cells can regulate tumor angiogenesis through both direct secretion of pro-angiogenic factors and indirect modulation of the TME with multiple mechanisms potentially involved (Fig. 2).

Table 1 Potential drugs for endothelial reprogramming therapy

Name	Brand Name	Mechanism of action	Status
Ficlatuzumab/AV-299	N/A	Monoclonal antibody against HGF	Under clinical trials
YYB101	N/A	Monoclonal antibody against HGF	Under clinical trials
Cabozantinib	Cometriq, Cabometyx	Inhibitor of c-Met (and VEGFR2, AXL, and RET)	FDA-approved for medullary thyroid cancer, kidney cancer
Olaratumab	Lartruvo	Monoclonal antibody against PDGFR α	FDA-approved for soft-tissue sarcoma (STS)
Ripretinib	Qinlock	Inhibitor of PDGFR α (and KIT)	FDA-approved for advanced gastrointestinal stromal tumor (GIST)
Sunitinib	Sutent	Inhibitor of PDGFRs (VEGFRs and KIT)	FDA-approved for renal cell carcinoma (RCC), pancreatic cancer, and imatinib-resistant gastrointestinal stromal tumor (GIST)
KPT-9274	N/A	Inhibitor of PAK4 (and NAMPT)	Under clinical trials
Panobinostat/ LBH589	Farydak	HDAC inhibitor	FDA-approved for multiple myeloma
Vorinostat/SAHA	Zolinza	HDAC inhibitor	FDA-approved for cutaneous T-cell lymphoma (CTCL)
Belinostat/ PDX101	Beleodaq	HDAC inhibitor	FDA-approved for peripheral T-cell lymphoma
Romidepsin/ FK228	Istodax	HDAC inhibitor	FDA-approved for cutaneous T-cell lymphoma (CTCL) and other peripheral T-cell lymphomas (PTCLs)
Azacitidine	Vidaza	DNMT inhibitor	FDA-approved for myelodysplastic syndrome, myeloid leukemia, and juvenile myelomonocytic leukemia
Decitabine	Dacogen	DNMT inhibitor	FDA-approved for myelodysplastic syndromes and acute myeloid leukemia (AML)
TVB-2640	N/A	FASN inhibitor	Under clinical trials
Etomoxir	N/A	CPT1A inhibitor	Under clinical trials
IACS-6274	N/A	GLS1 inhibitor	Under clinical trials

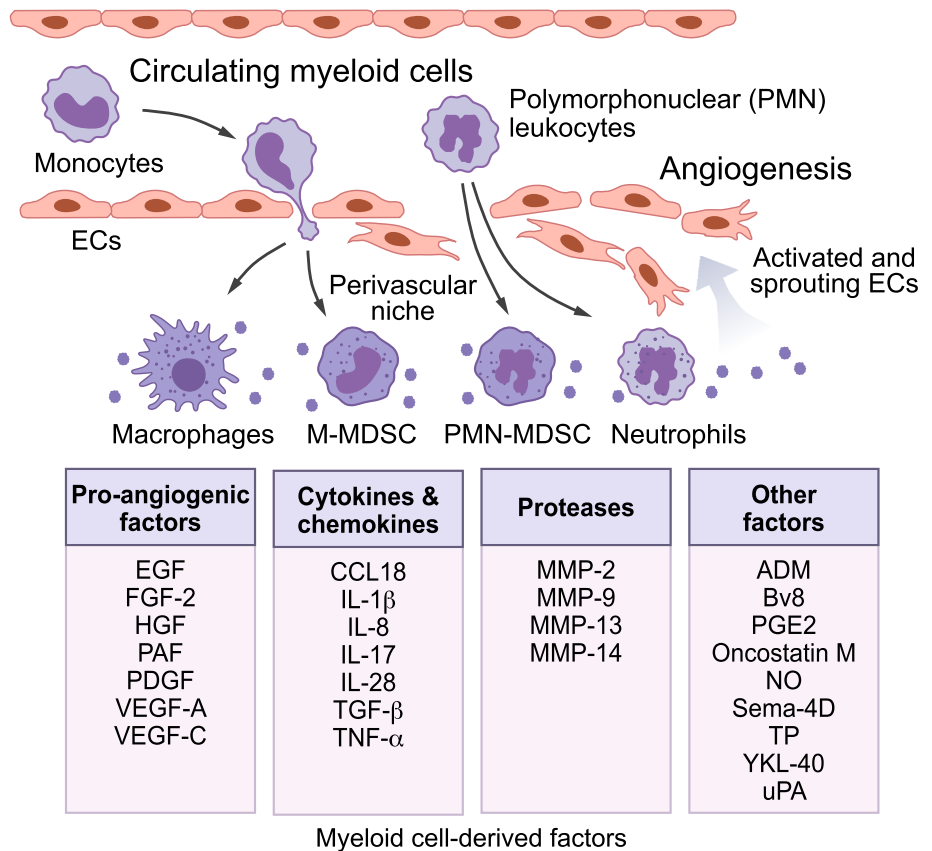
Table 2 Clinical trials evaluating combination of anti-angiogenic and immunotherapies

Clinical Trial Identifier	Phase	Status	Angiogenic Therapy	Immunotherapy	Other Drugs	Cancer
NCT05488522	I	Recruiting	Bevacizumab	Atezolizumab	Stereotactic body radiotherapy (SBRT)	Advanced hepatocellular carcinoma (HCC)
NCT02873195	II	Active	Bevacizumab	Atezolizumab	Capecitabine	Refractory metastatic colorectal cancer
NCT04356729	II	Recruiting	Bevacizumab	Atezolizumab	–	Unresectable or metastatic stage II or IV cutaneous melanoma
NCT03762018	III	Active	Bevacizumab	Atezolizumab	Standard chemotherapy	Malignant pleural mesothelioma
NCT03074513	II	Active	Bevacizumab	Atezolizumab	–	Rare solid tumors
NCT02210117	I	Active	Bevacizumab	Nivolumab, Ipilimumab	–	Resectable metastatic kidney cancer
NCT06083844	II	Recruiting	Bevacizumab	Pembrolizumab	Low-dose cyclophosphamide	High grade ovarian cancer with minimal residual disease after frontline treatment
NCT03175432	II	Active	Bevacizumab	Atezolizumab	Cobimetinib	Untreated melanoma with brain metastasis
NCT04721132	II	Recruiting	Bevacizumab	Atezolizumab	–	Resectable liver cancer
NCT02921269	II	Completed	Bevacizumab	Atezolizumab	–	Recurrent, persistent, or metastatic cervical cancer
NCT03141684	II	Recruiting	Bevacizumab	Atezolizumab	–	Advanced unresectable alveolar soft part sarcoma
NCT01950390	II	Active	Bevacizumab	Ipilimumab	–	Stage III-IV melanoma
NCT04981509	II	Recruiting	Bevacizumab, Erlotinib	Atezolizumab	–	Advanced stage kidney cancer
NCT05211323	II	Recruiting	Bevacizumab	Atezolizumab	Gemcitabine, cisplatin	Advanced unresectable liver cancer
NCT02997228	III	Recruiting	Bevacizumab	Atezolizumab	Combination chemotherapy	Mismatch repair deficient, metastatic, colorectal cancer
NCT02853318	II	Completed	Bevacizumab	Pembrolizumab	Low-dose cyclophosphamide	Recurrent ovarian, fallopian tube, or primary peritoneal cancer
NCT05468359	I/II	Recruiting	Bevacizumab	Atezolizumab	Cyclophosphamide, sorafenib	Pediatric solid tumors
NCT03396926	II	Active	Bevacizumab	Pembrolizumab	Capecitabine	Locally advanced, metastatic, or nonresectable microsatellite stable colorectal cancer
NCT03172754	I/II	Recruiting	Axitinib	Nivolumab	–	Advanced renal cell carcinoma
NCT04996823	II	Recruiting	Axitinib	Ipilimumab	–	Advanced melanoma
NCT04338269	III	Active	Cabozantinib	Atezolizumab	–	Inoperable, locally advanced, or metastatic renal cell carcinoma
NCT05805501	II	Recruiting	Axitinib	Triagolumab, Tobemstomig, Pembrolizumab	–	Previously untreated, unresectable locally advanced or metastatic clear-cell renal cell carcinoma

Table 2 (continued)

Clinical Trial Identifier	Phase	Status	Angiogenic Therapy	Immunotherapy	Other Drugs	Cancer
NCT04493203	II	Recruiting	Axitinib	Nivolumab	–	Unresectable stage III or IV melanoma
NCT02133742	Ib	Completed	Axitinib	Pembrolizumab	–	Advanced renal cell cancer
NCT04919629	II	Recruiting	Bevacizumab	Pembrolizumab	APL-2	Recurrent ovarian, fallopian tube or primary peritoneal cancer
NCT05231122	II	Recruiting	Bevacizumab	Pembrolizumab, anti-CD40 CDX-1140	–	Recurrent ovarian cancer
NCT02636725	II	Completed	Axitinib	Pembrolizumab	–	Advanced alveolar soft part sarcoma and soft tissue sarcomas
NCT04370509	II	Recruiting	Axitinib	Pembrolizumab	–	Locally advanced metastatic clear cell kidney cancer
NCT03092856	II	Active	Axitinib	Anti-OX40	–	Metastatic kidney cancer

Fig. 2 Regulation of tumor angiogenesis by myeloid cells. Tumor-infiltrating myeloid cells, including macrophages, MDSCs, and neutrophils, interact with ECs and modulate tumor angiogenesis through secreted factors



Macrophages

Macrophage-produced pro-angiogenic factors

Macrophages are a major cellular component of solid tumors

[105]. TAMs promote tumor angiogenesis by secreting a plethora of pro-angiogenic growth factors, cytokines, and chemokines that induce EC proliferation and migration, including EGF[106], FGF-2/bFGF [107], platelet-activating factor (PAF) [108], PDGF [109], VEGF [110–112], TNF-α

[113], IL-1 [114, 115], IL-8/CXCL8 [116, 117], and CCL18 [118]. TAMs undergo alternative polarization in the TME to stimulate tumor angiogenesis [107, 119], which is characterized by elevated expression of these pro-angiogenic factors [120]. Macrophages are a major source of pro-angiogenic factors, particularly VEGF, that are present in tumors. Macrophages employ diverse mechanisms to express VEGF, mainly induced by hypoxia through HIFs-mediated transcriptional activation and further stimulated by multiple cytokines like IL-1 β [121–123] and CCL18 [118]. Moreover, TAMs significantly contribute to production of proteases, particularly matrix metalloproteinase (MMP)-9, presented in the TME [124], which directly facilitates EC overgrowth and sprouting by remodeling ECM and indirectly activates ECs by providing the active form of VEGF as a result of cleaving VEGF and releasing it from the binding to ECM [125–129]. Bone marrow-derived, MMP-9-expressing macrophages also participate in tumor neovascularization together with vascular endothelial progenitor cells [130], providing an additional mechanism for tumor angiogenesis. TAMs also release various additional factors that have pro-angiogenic activity, such as adrenomedullin (ADM), PGE₂, Sema-4D, thymidine phosphorylase (TP), urokinase-type plasminogen activator (uPA), and YKL-40. For instance, ADM induces EC proliferation and tumor angiogenesis and growth [131], PGE₂ enhances EC motility and survival, contributing to tumor angiogenesis [132], Sema-4D binds to its receptor Plexin-B1 on ECs to induce tumor angiogenesis [133], TP stimulates EC migration [134], uPA promotes ECM degradation and vascular invasion [135], and YKL-40 activates MAPK signaling in ECs, leading to increased expression of VEGFR-2 that facilitates vessel sprouting [136]. Targeting tumor macrophage-released pro-angiogenic factors represent a promising strategy for therapeutic modulation of tumor angiogenesis.

Perivascular macrophages

Macrophages expressing Tie2 receptor (also known as Tek) often reside near vasculature and exhibit high angiogenic potential, playing a significant role in physiological and pathological angiogenesis [137]. A subpopulation of Tie2⁺ macrophages show a pro-angiogenic activity during embryogenesis [138]. These Tie2⁺ tissue macrophages release VEGF-C and soluble VEGFR1 to bridge between EC tip cells and modulate vessel branching in development [139, 140]. Similar to these findings, bone marrow-derived TAMs cluster around tumor vasculature and co-express Tie2 and CD206, showing more robust pro-angiogenic activity than Tie2⁻ monocytes or macrophages [141, 142]. Tie2⁺ monocytes express a number of tumor-promoting genes including *Mmp9*, *Vegfa*, *Cxcl12*, *Tlr4*, *Nrp1*, and *Pdgfb* at a high level [143], and their pro-angiogenic

potential could be further stimulated by EC-derived factors in the perivascular niche [144]. The presence of Tie2⁺ macrophages in tumor regions has been linked to increased tumor microvascular density, enhanced tumor grade and distant metastasis, and reduced survival rates in human patients [145–147]. Perivascular macrophages accumulate in the tumor microenvironment following chemotherapy, radiotherapy, and anti-angiogenic therapy, contributing to vascular reconstruction, and potentially leading to tumor relapse [112, 148–151]. These macrophages originate from the precursors of a subset of Tie2⁺ circulating monocytes and are attracted to tumors by chemotaxis, induced by EC-derived angiopoietin-2 (ANG-2), a ligand of Tie2 [141, 144, 152]. Interaction with ECs stimulates Tie2 expression in TAMs and enhances the production of pro-angiogenic factors by these macrophages [144]. Ang-2 also drives Tie2⁺ macrophages to express IL-10 and CCL17, which inhibit T-cell proliferation and disrupt vascular homeostasis [153].

Recent studies have shed light on the importance of perivascular macrophages in the TME. In addition to promoting angiogenesis, these macrophages, residing near blood vessels, promote the formation of the vascular niche that contributes to tumor progression. Activation of these macrophages by extracellular matrix proteins, such as TNC (tenascin-C), through toll-like receptor 4 (TLR4) signaling, leads to the secretion of nitric oxide (NO) and TNF- α [154]. These factors, in turn, induce the expression of niche components in ECs, facilitating the establishment of a supportive TME for tumor growth and metastasis. These macrophages usually acquire anti-inflammatory phenotypes, contributing to spatially restrict immunosuppression in the vascular niche. As such, tumor EC-derived IL-6 induces alternative polarization and immunosuppressive phenotypes in perivascular macrophages [155]. In addition, Lyve-1⁺ macrophages have a critical role in creating a pro-angiogenic TME through maintaining and expanding a perivascular mesenchymal cell population, ultimately establishing a specialized niche that supports tumor progression [156]. Macrophage-derived TNF- α and endothelial TNF receptor are identified as crucial components of this regulatory mechanism. Perivascular macrophages, activated via TNC and TLR4 to induce the formation of pro-tumor vascular niche that drives tumor metastasis [154]. The spatial interaction between macrophages and ECs provides strong evidence for the intricate crosstalk that stimulates angiogenesis and tumor progression, metastasis, and therapy resistance [109].

Macrophage-mediated vascular maturation

Macrophages regulate vascular maturation under physiological and pathological conditions. As a resident macrophage population in brain, microglia maintain the

integrity of blood–brain barrier that mainly consist of tightly associated ECs [157, 158]. Loss of NG2 proteoglycan in myeloid-specific and pericyte-specific cells leads to significant reductions in early-stage intracranial tumor growth [159, 160]. Myeloid-specific NG2 loss-induced vascular deficits, characterized by poor pericyte coverage on ECs and immature vessel, result in smaller vessel diameter, lower patency, increased leakiness, inefficient blood flow in tumor vasculature, and elevated intratumoral hypoxia [159]. TAMs promote pericyte coverage and stabilize tumor vasculature through the secretion of PDGF-B, contributing to vascular maturation [161]. Adenosine deaminase 2 (CECR1) is highly expressed by TAMs, contributing to tumor angiogenesis [161]. Increased CECR1 expression correlates with higher microvascular density in GBM tissues. Inhibition of CECR1 reduces new vessel formation, while CECR1 stimulation promotes vascular maturation through paracrine activation of pericytes via PDGFB-PDGFR β signaling [161].

Macrophage-mediated vascular permeability

VEGF was originally identified as vascular permeability factor (VPF) as a result of its potent ability to enhance vessel permeability, resulting in vascular leakage [162]. TAM-derived VEGF-A may, therefore, induce local vascular permeability in tumors. Consistent with this hypothesis, real-time intravital imaging reveals that dynamic vascular permeability occurs concurrently with cancer cell invasion and Tie2⁺ macrophage infiltration in the perivascular niche [163]. Genetic deletion of VEGF in TAMs reverses vascular permeability and cancer cell intravasation [163], suggesting a role of TAMs for regulation of vascular permeability. TAMs regulate vascular permeability through VEGF-induced downregulation of vascular junction proteins ZO-1 and VE-cadherin and through VLA4-mediated disruption of vascular adhesion proteins VCAM1 in ECs [163, 164]. In addition, M2-like polarized macrophage-derived exosomes containing miR-23a, miR-155 and miR-221 induces angiogenesis and vessel leakiness [165, 166], serving as an alternative mechanism for regulating tumor vascular permeability.

MDSCs

MDSCs are pathologically activated granulocytes (granulocytic or polymorphonuclear MDSCs, PMN-MDSCs) and monocytes (monocytic MDSCs, M-MDSCs) with potent immunosuppressive activity [167, 168]. MDSCs regulate immune responses in physiological and pathological conditions, including pregnancy, cancer, chronic infection, sepsis and autoimmunity [169]. In addition to their well-established role for direct suppression of lymphocyte activity,

MDSCs secrete various pro-angiogenic molecules to induce tumor angiogenesis [100, 170, 171]. Tumor-associated Gr1⁺CD11b⁺ mouse MDSCs produce MMP-9 and release VEGF-A to promote angiogenesis [172]. Consistent with these findings, tumor-infiltrating MDSCs express MMP-2,-13,-14 at a high level [173], and overexpression of MMP inhibitor TIMP-2 reduces MDSC infiltration and vascular density in tumor [174], suggesting a critical role of protease for MDSC-mediated tumor angiogenesis. Moreover, G-CSF stimulates Stat3-dependent MDSC expression of Bv8 [175], a potent driver of myeloid cell-dependent tumor angiogenesis [176]. MDSCs also express FGF-2 [171], PDGF [177], IL-1 β [178], IL-28/IFN- λ [179, 180], TGF β , EGF, and HGF [181] that can stimulate EC proliferation and migration, contributing to tumor angiogenesis [182]. In addition, MDSCs could directly differentiate into ECs [172] and induce tumor cell formation of vascular mimicry (VM) [183], serving as alternative processes to sprouting angiogenesis.

Neutrophils

Neutrophils are the most abundant innate immune cells in bone marrow and peripheral blood [184]. Neutrophils have emerged as an important component of the TME, but their functional role in cancer is still controversial [185]. In accordance with their critical functions in developmental angiogenesis [186, 187], neutrophils modulate tumor angiogenesis by providing pro-angiogenic factors in a time- and tumor context-dependent manner, contributing to tumor growth and metastasis [101, 102]. Tumor-associated neutrophils secrete a plethora of pro-angiogenic molecules including VEGF [188, 189], FGF-2 [190], Bv8 [191, 192], IL-17 [193], and MMP-9 [188, 194]. Neutrophil-derived oncostatin M also up-regulates the secretion of VEGF [195], and reprograms and degrades the ECM which then primes the environment for angiogenesis [195]. MMP-9 released by neutrophils promotes the activation of VEGF and subsequent angiogenesis and tumor progression [129, 194, 196]. Neutrophils also carry an intracellular pool of VEGF and mediate its rapid secretion [197]. Interestingly, IFN- β inhibits the infiltration of proangiogenic neutrophils that express VEGF, MMP-9, and CXCR4 and reduces tumor growth, suggesting a potential therapeutic approach for targeting neutrophil-mediated tumor angiogenesis [188]. These findings together suggest that neutrophils support the pro-angiogenic switch during cancer development [194]. As such, neutrophils display different states based on the expression of markers such as HIF-1 α , arginase 1, and MMP-9, in which HIF1 α ⁺ neutrophils significantly correlate with greater angiogenesis and worse overall survival [198]. In addition to their role in directly driving pro-angiogenic functions, neutrophils can also indirectly promote

angiogenesis by activating pro-angiogenic functions of other immune cells [103]. For example, neutrophils reprogram T cells to acquire regulatory-like phenotypes and support their expression of IL-10, IL-17, and VEGF to promote angiogenesis [199].

Neutrophils also contribute to tumor vascularization through several non-angiogenic mechanisms, such as neutrophil extracellular trap (NET) formation [174, 175], vessel co-option, and VM mechanisms [200, 201]. NETs, the release of web-like DNA structures, constitute an important mechanism by which neutrophils prevent pathogen dissemination or deal with microorganisms of larger size [202]. Cancer cells can induce NET formation by neutrophils, leading to tumor angiogenesis [203, 204]. NET-associated myeloperoxidase produces H₂O₂ released to ECM and activates NF- κ B-mediated signaling in ECs, resulting in enhanced EC proliferation and migration [205]. Angiopoietins (ANG-1/-2) also induce NETs formation and promote neutrophil adhesion to endothelium and stimulated EC proliferation [206]. Finally, VM structures provide vascular channels for neutrophil infiltration and activation, leading to their expression of arginase, CCL2, CXCR4, and MMP-9 to promote angiogenesis and evade anti-angiogenic therapy [201], collectively suggest a critical role of neutrophils for tumor angiogenesis.

Conclusion remarks and future perspectives

Anti-angiogenesis, vessel normalization, and endothelial reprogramming stand out as promising strategies for targeting the vasculature in cancer treatment. They hold significant potential when combined with various anti-cancer approaches including, but not limited to, radiotherapy, chemotherapy, molecular targeted therapy, and immunotherapy. The application of these strategies in clinical settings might require optimization based on factors like tumor type, size, stage, location, and pathology to achieve the maximal efficacy in combination therapy. Particularly, genetic, epigenetic and metabolic reprogramming of tumor ECs may offer unique opportunities to empower T cell-based immunotherapy, considering that endothelial reprogramming could (1) inhibit excessive angiogenesis and suppress vascular aberrancy, leading to increase vessel delivery function to improve lymphocyte infiltration as well as to relieve intratumoral hypoxia and to activate these lymphocytes, (2) regulate adhesion molecule expression on ECs to promote lymphocyte attachment to endothelium and their recruitment to the tumors, and (3) reverse pro-tumor immunity that is induced by locally EC-derived immunosuppressive molecules, facilitating lymphocyte activation.

Among these innovative strategies, the induction of HEV neogenesis has emerged as a promising strategy to augment anti-tumor immunity and vessel functionality. HEVs play a vital role in lymphocyte trafficking and activation, serving as a critical target for therapeutic modulation of immunocyte infiltration [207]. Recent single-cell RNA-seq analyses suggest their significant involvement in upregulated expression of EC activation markers and co-stimulatory molecules that regulate dendritic cell function and T cell activation [208]. Activation of lymphotoxin β receptor (LT β R) signaling induces the formation of HEVs and T cell activation, and thereby sensitizes tumors to anti-angiogenic and anti-PD-L1 therapy [209], collectively suggesting that better understanding the immunostimulatory functions of HEVs may open new avenues for immunotherapeutic interventions in the future.

Despite these advancements, there are still several major limitations and challenges that restrict vascular-targeting therapy in cancer, due to treatment toxicity, vascular heterogeneity in cancer, and the lack of reliable biomarkers hinder the effectiveness and applicability of current treatments. Notably, tumors can develop resistance to traditional anti-angiogenic treatments by compensatory activation of additional pro-angiogenic pathways to sustain tumor vascularization and by activating HIF-1 α to support tumor growth and progression in low-oxygen conditions. One particular challenge is to develop a vascular-targeting strategy to selectively promote the infiltration of cytotoxic T cells and/or NK cells, but not immunosuppressive myeloid cells. Future spatiotemporal analysis of tumor specimens at single-cell transcriptome, epigenome, and metabolome levels will help address these challenges, as they need a deep understanding of the complex interplay between tumor vasculature and immune system in the tumor microenvironment.

Further research into the interaction of infiltrating myeloid cells with ECs regulates tumor angiogenesis, providing insights into the resolution of vascular formation, maturation, and aberrancy in cancer. Understanding of the underlying regulatory mechanism for tumor angiogenesis may lead to identification of new therapeutic targets for anti-vascular therapy, contributing to development of more efficient approaches for anti-angiogenesis, vessel normalization, and endothelial reprogramming therapy. It also remains largely unclear how ECs spatiotemporally regulate the immunity of these macrophages, MDSCs, and neutrophils in the vascular niche. New knowledge filling this gap may help understand tumor immunosuppression and lead to development of new myeloid cell-based immunotherapy for cancer treatment.

Author contributions FY and YF wrote the manuscript and produced the figures. FY performed literature research. GL contributed to the concepts and produced the tables. YF conceived the concepts described in this review. All authors commented on the manuscript.

Funding This work was partly financially supported by American Association for Cancer Research Judah Folkman Award (to Y.F.) and American Heart Association Innovation Project Award (to Y.F.).

Data availability Data sharing is not applicable to this article as no datasets were generated or analyzed for this review article.

Declarations

Competing interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Carmeliet P, Jain RK (2011) Molecular mechanisms and clinical applications of angiogenesis. *Nature* 473(7347):298–307
- Fridman WH et al (2012) The immune contexture in human tumours: impact on clinical outcome. *Nat Rev Cancer* 12(4):298–306
- Tian L et al (2017) Mutual regulation of tumour vessel normalization and immunostimulatory reprogramming. *Nature* 544(7649):250–254
- Folkman J (1971) Tumor angiogenesis: therapeutic implications. *N Engl J Med* 285(21):1182–1186
- Weis SM, Cheresh DA (2011) Tumor angiogenesis: molecular pathways and therapeutic targets. *Nat Med* 17(11):1359–1370
- Cao Y, Langer R, Ferrara N (2023) Targeting angiogenesis in oncology, ophthalmology and beyond. *Nat Rev Drug Discov* 22(6):476–495
- Ebos JM, Kerbel RS (2011) Antiangiogenic therapy: impact on invasion, disease progression, and metastasis. *Nat Rev Clin Oncol* 8(4):210–221
- Jain RK (2005) Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science* 307(5706):58–62
- Liu T et al (2018) PDGF-mediated mesenchymal transformation renders endothelial resistance to anti-VEGF treatment in glioblastoma. *Nat Commun* 9(1):3439
- Bergers G, Benjamin LE (2003) Tumorigenesis and the angiogenic switch. *Nat Rev Cancer* 3(6):401–410
- Jain RK (2001) Normalizing tumor vasculature with anti-angiogenic therapy: a new paradigm for combination therapy. *Nat Med* 7(9):987–989
- Carmeliet P, Jain RK (2011) Principles and mechanisms of vessel normalization for cancer and other angiogenic diseases. *Nat Rev Drug Discov* 10(6):417–427
- Martin JD, Seano G, Jain RK (2019) Normalizing function of tumor vessels: progress, opportunities, and challenges. *Annu Rev Physiol* 81:505–534
- Ellis LM, Hicklin DJ (2008) VEGF-targeted therapy: mechanisms of anti-tumour activity. *Nat Rev Cancer* 8(8):579–591
- Turner N, Grose R (2010) Fibroblast growth factor signalling: from development to cancer. *Nat Rev Cancer* 10(2):116–129
- Ciardiello F, Tortora G (2008) EGFR antagonists in cancer treatment. *N Engl J Med* 358(11):1160–1174
- Semenza GL (2003) Targeting HIF-1 for cancer therapy. *Nat Rev Cancer* 3(10):721–732
- Lee P et al (2002) Neuropilin-1 is required for vascular development and is a mediator of VEGF-dependent angiogenesis in zebrafish. *Proc Natl Acad Sci USA* 99(16):10470–10475
- Benedito R et al (2012) Notch-dependent VEGFR3 upregulation allows angiogenesis without VEGF-VEGFR2 signalling. *Nature* 484(7392):110–114
- Daly C et al (2013) Angiopoietin-2 functions as a Tie2 agonist in tumor models, where it limits the effects of VEGF inhibition. *Cancer Res* 73(1):108–118
- Lobov IB, Brooks PC, Lang RA (2002) Angiopoietin-2 displays VEGF-dependent modulation of capillary structure and endothelial cell survival in vivo. *Proc Natl Acad Sci USA* 99(17):11205–11210
- Wang Y et al (2010) Ephrin-B2 controls VEGF-induced angiogenesis and lymphangiogenesis. *Nature* 465(7297):483–486
- Ferrara N et al (2004) Discovery and development of bevacizumab, an anti-VEGF antibody for treating cancer. *Nat Rev Drug Discov* 3(5):391–400
- Faivre S et al (2006) Safety, pharmacokinetic, and antitumor activity of SU11248, a novel oral multitarget tyrosine kinase inhibitor, in patients with cancer. *J Clin Oncol* 24(1):25–35
- Keating GM (2017) Sorafenib: a review in hepatocellular carcinoma. *Target Oncol* 12(2):243–253
- Wick W et al (2017) Lomustine and bevacizumab in progressive glioblastoma. *N Engl J Med* 377(20):1954–1963
- Gilbert MR et al (2014) A randomized trial of bevacizumab for newly diagnosed glioblastoma. *N Engl J Med* 370(8):699–708
- Bergers G, Hanahan D (2008) Modes of resistance to anti-angiogenic therapy. *Nat Rev Cancer* 8(8):592–603
- Finn RS et al (2020) Atezolizumab plus bevacizumab in unresectable hepatocellular carcinoma. *N Engl J Med* 382(20):1894–1905
- Batchelor TT et al (2013) Phase III randomized trial comparing the efficacy of cediranib as monotherapy, and in combination with lomustine, versus lomustine alone in patients with recurrent glioblastoma. *J Clin Oncol* 31(26):3212–3218
- Thomas M, Augustin HG (2009) The role of the Angiopoietins in vascular morphogenesis. *Angiogenesis* 12(2):125–137
- Maisonpierre PC et al (1997) Angiopoietin-2, a natural antagonist for Tie2 that disrupts in vivo angiogenesis. *Science* 277(5322):55–60
- Harris AL (2002) Hypoxia—a key regulatory factor in tumour growth. *Nat Rev Cancer* 2(1):38–47
- Li JL, Harris AL (2005) Notch signaling from tumor cells: a new mechanism of angiogenesis. *Cancer Cell* 8(1):1–3
- Zerlin M, Julius MA, Kitajewski J (2008) Wnt/Frizzled signaling in angiogenesis. *Angiogenesis* 11(1):63–69
- Keith B, Johnson RS, Simon MC (2011) HIF1 α and HIF2 α : sibling rivalry in hypoxic tumour growth and progression. *Nat Rev Cancer* 12(1):9–22
- Ferrara N, Gerber HP, LeCouter J (2003) The biology of VEGF and its receptors. *Nat Med* 9(6):669–676
- Wouters BG, Koritzinsky M (2008) Hypoxia signalling through mTOR and the unfolded protein response in cancer. *Nat Rev Cancer* 8(11):851–864

39. Ridgway J et al (2006) Inhibition of Dll4 signalling inhibits tumour growth by deregulating angiogenesis. *Nature* 444(7122):1083–1087
40. Stenman JM et al (2008) Canonical Wnt signaling regulates organ-specific assembly and differentiation of CNS vasculature. *Science* 322(5905):1247–1250
41. Huang M et al (2020) Wnt-mediated endothelial transformation into mesenchymal stem cell-like cells induces chemoresistance in glioblastoma. *Sci Transl Med*. <https://doi.org/10.1126/scitranslmed.aay7522>
42. Kühl M et al (2000) Ca(2+)/calmodulin-dependent protein kinase II is stimulated by Wnt and Frizzled homologs and promotes ventral cell fates in *Xenopus*. *J Biol Chem* 275(17):12701–12711
43. Habas R, Kato Y, He X (2001) Wnt/Frizzled activation of Rho regulates vertebrate gastrulation and requires a novel Formin homology protein Daam1. *Cell* 107(7):843–854
44. Yamanaka H et al (2002) JNK functions in the non-canonical Wnt pathway to regulate convergent extension movements in vertebrates. *EMBO Rep* 3(1):69–75
45. Cirone P et al (2008) A role for planar cell polarity signaling in angiogenesis. *Angiogenesis* 11(4):347–360
46. Huang Y et al (2012) Vascular normalizing doses of antiangiogenic treatment reprogram the immunosuppressive tumor microenvironment and enhance immunotherapy. *Proc Natl Acad Sci USA* 109(43):17561–17566
47. Dong X et al (2023) Anti-VEGF therapy improves EGFR-vIII-CAR-T cell delivery and efficacy in syngeneic glioblastoma models in mice. *J Immunother Cancer* 11(3):e005583
48. Hamzah J et al (2008) Vascular normalization in Rgs5-deficient tumours promotes immune destruction. *Nature* 453(7193):410–414
49. Magrini E et al (2014) Endothelial deficiency of L1 reduces tumor angiogenesis and promotes vessel normalization. *J Clin Invest* 124(10):4335–4350
50. Goel S et al (2011) Normalization of the vasculature for treatment of cancer and other diseases. *Physiol Rev* 91(3):1071–1121
51. Maione F et al (2009) Semaphorin 3A is an endogenous angiogenesis inhibitor that blocks tumor growth and normalizes tumor vasculature in transgenic mouse models. *J Clin Invest* 119(11):3356–3372
52. Sawada J et al (2012) Small GTPase R-Ras regulates integrity and functionality of tumor blood vessels. *Cancer Cell* 22(2):235–249
53. Takara K et al (2017) Lysophosphatidic acid receptor 4 activation augments drug delivery in tumors by tightening endothelial cell-cell contact. *Cell Rep* 20(9):2072–2086
54. Maes H et al (2014) Tumor vessel normalization by chloroquine independent of autophagy. *Cancer Cell* 26(2):190–206
55. Adapala RK et al (2016) Activation of mechanosensitive ion channel TRPV4 normalizes tumor vasculature and improves cancer therapy. *Oncogene* 35(3):314–322
56. Campbell NE et al (2010) The thrombospondin-1 mimetic ABT-510 increases the uptake and effectiveness of cisplatin and paclitaxel in a mouse model of epithelial ovarian cancer. *Neoplasia* 12(3):275–283
57. Funahashi Y et al (2014) Eribulin mesylate reduces tumor microenvironment abnormality by vascular remodeling in preclinical human breast cancer models. *Cancer Sci* 105(10):1334–1342
58. Chauhan VP et al (2013) Angiotensin inhibition enhances drug delivery and potentiates chemotherapy by decompressing tumour blood vessels. *Nat Commun* 4:2516
59. Stylianopoulos T et al (2012) Causes, consequences, and remedies for growth-induced solid stress in murine and human tumors. *Proc Natl Acad Sci USA* 109(38):15101–15108
60. Alvarez R et al (2013) Stromal disrupting effects of nab-paclitaxel in pancreatic cancer. *Br J Cancer* 109(4):926–933
61. Wenes M et al (2016) Macrophage metabolism controls tumor blood vessel morphogenesis and metastasis. *Cell Metab* 24(5):701–715
62. De Palma M, Jain RK (2017) CD4(+) T cell activation and vascular normalization: two sides of the same coin? *Immunity* 46(5):773–775
63. Voron T et al (2015) VEGF-A modulates expression of inhibitory checkpoints on CD8+ T cells in tumors. *J Exp Med* 212(2):139–148
64. Willett CG et al (2004) Direct evidence that the VEGF-specific antibody bevacizumab has antivasular effects in human rectal cancer. *Nat Med* 10(2):145–147
65. Kerbel RS (2008) Tumor angiogenesis. *N Engl J Med* 358(19):2039–2049
66. Bertrand JY et al (2010) Haematopoietic stem cells derive directly from aortic endothelium during development. *Nature* 464(7285):108–111
67. Kissa K, Herbomel P (2010) Blood stem cells emerge from aortic endothelium by a novel type of cell transition. *Nature* 464(7285):112–115
68. Zeisberg EM et al (2007) Endothelial-to-mesenchymal transition contributes to cardiac fibrosis. *Nat Med* 13(8):952–961
69. Maddaluno L et al (2013) EndMT contributes to the onset and progression of cerebral cavernous malformations. *Nature* 498(7455):492–496
70. Zeisberg EM et al (2007) Discovery of endothelial to mesenchymal transition as a source for carcinoma-associated fibroblasts. *Cancer Res* 67(21):10123–10128
71. Fan Y (2019) Vascular detransformation for cancer therapy. *Trends Cancer* 5(8):460–463
72. Lamplugh Z, Fan Y (2021) Vascular microenvironment, tumor immunity and immunotherapy. *Front Immunol* 12:811485
73. Huang M et al (2016) c-Met-mediated endothelial plasticity drives aberrant vascularization and chemoresistance in glioblastoma. *J Clin Invest* 126(5):1801–1814
74. Amersfoort J, Eelen G, Carmeliet P (2022) Immunomodulation by endothelial cells—partnering up with the immune system? *Nat Rev Immunol* 22(9):576–588
75. Cleveland AH, Fan Y (2023) Reprogramming endothelial cells to empower cancer immunotherapy. *Trends Mol Med*. <https://doi.org/10.1016/j.molmed.2023.11.002>
76. Ma W et al (2021) Targeting PAK4 to reprogram the vascular microenvironment and improve CAR-T immunotherapy for glioblastoma. *Nat Cancer* 2(1):83–97
77. Birdsey GM et al (2015) The endothelial transcription factor ERG promotes vascular stability and growth through Wnt/ β -catenin signaling. *Dev Cell* 32(1):82–96
78. Kabir AU et al (2021) Dual role of endothelial Myc1 in tumor angiogenesis and tumor immunity. *Sci Transl Med*. <https://doi.org/10.1126/scitranslmed.abb6731>
79. O'Connor MN et al (2021) LRG1 destabilizes tumor vessels and restricts immunotherapeutic potency. *Med* 2(11):1231–1252.e10
80. Huijbers EJM et al (2022) Tumors resurrect an embryonic vascular program to escape immunity. *Sci Immunol* 7(67):eabm6388
81. Hellebrekers DM et al (2007) Identification of epigenetically silenced genes in tumor endothelial cells. *Cancer Res* 67(9):4138–4148
82. Bazou D et al (2016) Flow-induced HDAC1 phosphorylation and nuclear export in angiogenic sprouting. *Sci Rep* 6:34046
83. Hu C et al (2021) HDAC1 and 2 regulate endothelial VCAM-1 expression and atherogenesis by suppressing methylation of the GATA6 promoter. *Theranostics* 11(11):5605–5619
84. Deroanne CF et al (2002) Histone deacetylases inhibitors as anti-angiogenic agents altering vascular endothelial growth factor signaling. *Oncogene* 21(3):427–436

85. Kim MS et al (2001) Histone deacetylases induce angiogenesis by negative regulation of tumor suppressor genes. *Nat Med* 7(4):437–443
86. Hellebrekers DM et al (2006) Epigenetic regulation of tumor endothelial cell anergy: silencing of intercellular adhesion molecule-1 by histone modifications. *Cancer Res* 66(22):10770–10777
87. Maleszewska M et al (2015) Enhancer of zeste homolog-2 (EZH2) methyltransferase regulates transgelin/smooth muscle-22 α expression in endothelial cells in response to interleukin-1 β and transforming growth factor- β 2. *Cell Signal* 27(8):1589–1596
88. Glaser SF et al (2020) The histone demethylase JMJD2B regulates endothelial-to-mesenchymal transition. *Proc Natl Acad Sci USA* 117(8):4180–4187
89. Kim DJ et al (2023) Priming a vascular-selective cytokine response permits CD8(+) T-cell entry into tumors. *Nat Commun* 14(1):2122
90. De Bock K et al (2013) Role of PFKFB3-driven glycolysis in vessel sprouting. *Cell* 154(3):651–663
91. Bruning U et al (2018) Impairment of angiogenesis by fatty acid synthase inhibition involves mTOR malonylation. *Cell Metab* 28(6):866–880.e15
92. Schoors S et al (2015) Fatty acid carbon is essential for dNTP synthesis in endothelial cells. *Nature* 520(7546):192–197
93. Huang H et al (2017) Role of glutamine and interlinked asparagine metabolism in vessel formation. *Embo J* 36(16):2334–2352
94. Zhang D et al (2023) PHGDH-mediated endothelial metabolism drives glioblastoma resistance to chimeric antigen receptor T cell immunotherapy. *Cell Metab* 35(3):517–534.e8
95. Cantelmo AR et al (2016) Inhibition of the glycolytic activator PFKFB3 in endothelium induces tumor vessel normalization, impairs metastasis, and improves chemotherapy. *Cancer Cell* 30(6):968–985
96. Shan Y et al (2022) Targeting tumor endothelial hyperglycolysis enhances immunotherapy through remodeling tumor microenvironment. *Acta Pharm Sin B* 12(4):1825–1839
97. Zahalka AH et al (2017) Adrenergic nerves activate an angiometabolic switch in prostate cancer. *Science* 358(6361):321–326
98. Riabov V et al (2014) Role of tumor associated macrophages in tumor angiogenesis and lymphangiogenesis. *Front Physiol* 5:75
99. Ribatti D et al (2007) Macrophages and tumor angiogenesis. *Leukemia* 21(10):2085–2089
100. Vetsika EK, Koukos A, Kotsakis A (2019) Myeloid-derived suppressor cells: major figures that shape the immunosuppressive and angiogenic network in cancer. *Cells* 8(12):1647
101. Tazzyman S, Lewis CE, Murdoch C (2009) Neutrophils: key mediators of tumour angiogenesis. *Int J Exp Pathol* 90(3):222–231
102. Liang W, Ferrara N (2016) The complex role of neutrophils in tumor angiogenesis and metastasis. *Cancer Immunol Res* 4(2):83–91
103. Ozel I et al (2022) The good, the bad, and the ugly: neutrophils, angiogenesis, and cancer. *Cancers (Basel)* 14(3):536
104. Lewis CE, Harney AS, Pollard JW (2016) The multifaceted role of perivascular macrophages in tumors. *Cancer Cell* 30(1):18–25
105. Pollard JW (2004) Tumour-educated macrophages promote tumour progression and metastasis. *Nat Rev Cancer* 4(1):71–78
106. Yin M et al (2016) Tumor-associated macrophages drive spheroid formation during early transcoelomic metastasis of ovarian cancer. *J Clin Invest* 126(11):4157–4173
107. Jetten N et al (2014) Anti-inflammatory M2, but not pro-inflammatory M1 macrophages promote angiogenesis in vivo. *Angiogenesis* 17(1):109–118
108. Seo KH et al (2004) Essential role for platelet-activating factor-induced NF-kappaB activation in macrophage-derived angiogenesis. *Eur J Immunol* 34(8):2129–2137
109. Opzoomer JW et al (2021) Macrophages orchestrate the expansion of a proangiogenic perivascular niche during cancer progression. *Sci Adv* 7(45):eabg9518
110. Stockmann C et al (2008) Deletion of vascular endothelial growth factor in myeloid cells accelerates tumorigenesis. *Nature* 456(7223):814–818
111. Barbera-Guillem E et al (2002) Vascular endothelial growth factor secretion by tumor-infiltrating macrophages essentially supports tumor angiogenesis, and IgG immune complexes potentiate the process. *Cancer Res* 62(23):7042–7049
112. Hughes R et al (2015) Perivascular M2 macrophages stimulate tumor relapse after chemotherapy. *Cancer Res* 75(17):3479–3491
113. Leibovich SJ et al (1987) Macrophage-induced angiogenesis is mediated by tumour necrosis factor-alpha. *Nature* 329(6140):630–632
114. Voronov E, Carmi Y, Apte RN (2014) The role IL-1 in tumor-mediated angiogenesis. *Front Physiol* 5:114
115. Carmi Y et al (2009) The role of macrophage-derived IL-1 in induction and maintenance of angiogenesis. *J Immunol* 183(7):4705–4714
116. Koch AE et al (1992) Interleukin-8 as a macrophage-derived mediator of angiogenesis. *Science* 258(5089):1798–1801
117. Kimura YN et al (2007) Inflammatory stimuli from macrophages and cancer cells synergistically promote tumor growth and angiogenesis. *Cancer Sci* 98(12):2009–2018
118. Lin L et al (2015) CCL18 from tumor-associated macrophages promotes angiogenesis in breast cancer. *Oncotarget* 6(33):34758–34773
119. Lin EY et al (2006) Macrophages regulate the angiogenic switch in a mouse model of breast cancer. *Cancer Res* 66(23):11238–11246
120. Sica A, Mantovani A (2012) Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest* 122(3):787–795
121. Mantsounga CS et al (2022) Macrophage IL-1 β promotes arteriogenesis by autocrine STAT3- and NF- κ B-mediated transcription of pro-angiogenic VEGF-A. *Cell Rep* 38(5):110309
122. Nam EH, Park SR, Kim PH (2010) TGF-beta1 induces mouse dendritic cells to express VEGF and its receptor (Flt-1) under hypoxic conditions. *Exp Mol Med* 42(9):606–613
123. Fang HY et al (2009) Hypoxia-inducible factors 1 and 2 are important transcriptional effectors in primary macrophages experiencing hypoxia. *Blood* 114(4):844–859
124. Afik R et al (2016) Tumor macrophages are pivotal constructors of tumor collagenous matrix. *J Exp Med* 213(11):2315–2331
125. Egeblad M, Werb Z (2002) New functions for the matrix metalloproteinases in cancer progression. *Nat Rev Cancer* 2(3):161–174
126. Arroyo AG, Iruela-Arispe ML (2010) Extracellular matrix, inflammation, and the angiogenic response. *Cardiovasc Res* 86(2):226–235
127. Vu TH et al (1998) MMP-9/gelatinase B is a key regulator of growth plate angiogenesis and apoptosis of hypertrophic chondrocytes. *Cell* 93(3):411–422
128. Bergers G et al (2000) Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis. *Nat Cell Biol* 2(10):737–744
129. Coussens LM et al (2000) MMP-9 supplied by bone marrow-derived cells contributes to skin carcinogenesis. *Cell* 103(3):481–490
130. Du R et al (2008) HIF1alpha induces the recruitment of bone marrow-derived vascular modulatory cells to regulate tumor angiogenesis and invasion. *Cancer Cell* 13(3):206–220
131. Zhao Y et al (1998) PCR display identifies tamoxifen induction of the novel angiogenic factor adrenomedullin by a non estrogenic mechanism in the human endometrium. *Oncogene* 16(3):409–415

132. Kamiyama M et al (2006) EP2, a receptor for PGE₂, regulates tumor angiogenesis through direct effects on endothelial cell motility and survival. *Oncogene* 25(53):7019–7028
133. Basile JR et al (2007) MT1-MMP controls tumor-induced angiogenesis through the release of semaphorin 4D. *J Biol Chem* 282(9):6899–6905
134. Akiyama S et al (2004) The role of thymidine phosphorylase, an angiogenic enzyme, in tumor progression. *Cancer Sci* 95(11):851–857
135. Stepanova V et al (2016) Urokinase-type plasminogen activator (uPA) promotes angiogenesis by attenuating proline-rich homeodomain protein (PRH) transcription factor activity and de-repressing vascular endothelial growth factor (VEGF) receptor expression. *J Biol Chem* 291(29):15029–15045
136. Shao R (2013) YKL-40 acts as an angiogenic factor to promote tumor angiogenesis. *Front Physiol* 4:122
137. Baer C et al (2013) Reciprocal interactions between endothelial cells and macrophages in angiogenic vascular niches. *Exp Cell Res* 319(11):1626–1634
138. Fantin A et al (2010) Tissue macrophages act as cellular chaperones for vascular anastomosis downstream of VEGF-mediated endothelial tip cell induction. *Blood* 116(5):829–840
139. Tammela T et al (2011) VEGFR-3 controls tip to stalk conversion at vessel fusion sites by reinforcing Notch signalling. *Nat Cell Biol* 13(10):1202–1213
140. Stefater JA 3rd et al (2011) Regulation of angiogenesis by a non-canonical Wnt-Flt1 pathway in myeloid cells. *Nature* 474(7352):511–515
141. De Palma M et al (2005) Tie2 identifies a hematopoietic lineage of proangiogenic monocytes required for tumor vessel formation and a mesenchymal population of pericyte progenitors. *Cancer Cell* 8(3):211–226
142. De Palma M et al (2003) Targeting exogenous genes to tumor angiogenesis by transplantation of genetically modified hematopoietic stem cells. *Nat Med* 9(6):789–795
143. Pucci F et al (2009) A distinguishing gene signature shared by tumor-infiltrating Tie2-expressing monocytes, blood “resident” monocytes, and embryonic macrophages suggests common functions and developmental relationships. *Blood* 114(4):901–914
144. Coffelt SB et al (2010) Angiopoietin-2 regulates gene expression in TIE2-expressing monocytes and augments their inherent proangiogenic functions. *Cancer Res* 70(13):5270–5280
145. Ji J et al (2013) The frequency of tumor-infiltrating Tie-2-expressing monocytes in renal cell carcinoma: its relationship to angiogenesis and progression. *Urology* 82(4):974.e9–13
146. Du S et al (2022) Tumor cell-derived exosomes deliver TIE2 protein to macrophages to promote angiogenesis in cervical cancer. *Cancer Lett* 529:168–179
147. Matsubara T et al (2013) TIE2-expressing monocytes as a diagnostic marker for hepatocellular carcinoma correlates with angiogenesis. *Hepatology* 57(4):1416–1425
148. Chen L et al (2016) Tie2 expression on macrophages is required for blood vessel reconstruction and tumor relapse after chemotherapy. *Cancer Res* 76(23):6828–6838
149. Kioi M et al (2010) Inhibition of vasculogenesis, but not angiogenesis, prevents the recurrence of glioblastoma after irradiation in mice. *J Clin Invest* 120(3):694–705
150. Kozin SV et al (2010) Recruitment of myeloid but not endothelial precursor cells facilitates tumor regrowth after local irradiation. *Cancer Res* 70(14):5679–5685
151. Welford AF et al (2011) TIE2-expressing macrophages limit the therapeutic efficacy of the vascular-disrupting agent combretastatin A4 phosphate in mice. *J Clin Invest* 121(5):1969–1973
152. Venneri MA et al (2007) Identification of proangiogenic TIE2-expressing monocytes (TEMs) in human peripheral blood and cancer. *Blood* 109(12):5276–5285
153. Coffelt SB et al (2011) Angiopoietin 2 stimulates TIE2-expressing monocytes to suppress T cell activation and to promote regulatory T cell expansion. *J Immunol* 186(7):4183–4190
154. Hongu T et al (2022) Perivascular tenascin C triggers sequential activation of macrophages and endothelial cells to generate a prometastatic vascular niche in the lungs. *Nat Cancer* 3(4):486–504
155. Yang F et al (2021) Synergistic immunotherapy of glioblastoma by dual targeting of IL-6 and CD40. *Nat Commun* 12(1):3424
156. Ren X et al (2023) Macrophage-endothelial cell crosstalk orchestrates neutrophil recruitment in inflamed mucosa. *J Clin Invest*. <https://doi.org/10.1172/JCI170733>
157. Li Q, Barres BA (2018) Microglia and macrophages in brain homeostasis and disease. *Nat Rev Immunol* 18(4):225–242
158. Kisler K, Nikolakopoulou AM, Zlokovic BV (2021) Microglia have a grip on brain microvasculature. *Nat Commun* 12(1):5290
159. Yotsumoto F et al (2015) NG2 proteoglycan-dependent recruitment of tumor macrophages promotes pericyte-endothelial cell interactions required for brain tumor vascularization. *Oncimmunology* 4(4):e1001204
160. Thurgur H, Pinteaux E (2019) Microglia in the neurovascular unit: blood-brain barrier-microglia interactions after central nervous system disorders. *Neuroscience* 405:55–67
161. Zhu C et al (2017) CECR1-mediated cross talk between macrophages and vascular mural cells promotes neovascularization in malignant glioma. *Oncogene* 36(38):5356–5368
162. Senger DR et al (1983) Tumor cells secrete a vascular permeability factor that promotes accumulation of ascites fluid. *Science* 219(4587):983–985
163. Harney AS et al (2015) Real-time imaging reveals local, transient vascular permeability, and tumor cell intravasation stimulated by TIE2hi macrophage-derived VEGFA. *Cancer Discov* 5(9):932–943
164. Zhang S et al (2021) Macrophage-mediated vascular permeability via VLA4/VCAM1 pathway dictates ascites development in ovarian cancer. *J Clin Invest*. <https://doi.org/10.1172/JCI140315>
165. Yang Y et al (2021) M2 macrophage-derived exosomes promote angiogenesis and growth of pancreatic ductal adenocarcinoma by targeting E2F2. *Mol Ther* 29(3):1226–1238
166. Lu Y et al (2023) M2 macrophage-secreted exosomes promote metastasis and increase vascular permeability in hepatocellular carcinoma. *Cell Commun Signal* 21(1):299
167. Veglia F, Sanseviero E, Gabrilovich DI (2021) Myeloid-derived suppressor cells in the era of increasing myeloid cell diversity. *Nat Rev Immunol* 21(8):485–498
168. Hegde S, Leader AM, Merad M (2021) MDSC: markers, development, states, and unaddressed complexity. *Immunity* 54(5):875–884
169. Veglia F, Perego M, Gabrilovich D (2018) Myeloid-derived suppressor cells coming of age. *Nat Immunol* 19(2):108–119
170. Binsfeld M et al (2016) Granulocytic myeloid-derived suppressor cells promote angiogenesis in the context of multiple myeloma. *Oncotarget* 7(25):37931–37943
171. Kujawski M et al (2008) Stat3 mediates myeloid cell-dependent tumor angiogenesis in mice. *J Clin Invest* 118(10):3367–3377
172. Yang L et al (2004) Expansion of myeloid immune suppressor Gr⁺CD11b⁺ cells in tumor-bearing host directly promotes tumor angiogenesis. *Cancer Cell* 6(4):409–421
173. Yang L et al (2008) Abrogation of TGF beta signaling in mammary carcinomas recruits Gr-1+CD11b+ myeloid cells that promote metastasis. *Cancer Cell* 13(1):23–35
174. Guedez L et al (2012) TIMP-2 targets tumor-associated myeloid suppressor cells with effects in cancer immune dysfunction and angiogenesis. *J Immunother* 35(6):502–512

175. Qu X et al (2012) Induction of Bv8 expression by granulocyte colony-stimulating factor in CD11b+Gr1+ cells: key role of Stat3 signaling. *J Biol Chem* 287(23):19574–19584
176. Shojaei F et al (2007) Bv8 regulates myeloid-cell-dependent tumour angiogenesis. *Nature* 450(7171):825–831
177. Hsu YL et al (2019) CXCL17-derived CD11b(+)Gr-1(+) myeloid-derived suppressor cells contribute to lung metastasis of breast cancer through platelet-derived growth factor-BB. *Breast Cancer Res* 21(1):23
178. Shi H et al (2017) Recruited monocytic myeloid-derived suppressor cells promote the arrest of tumor cells in the premetastatic niche through an IL-1 β -mediated increase in E-selectin expression. *Int J Cancer* 140(6):1370–1383
179. Mucha J et al (2014) MDSCs mediate angiogenesis and predispose canine mammary tumor cells for metastasis via IL-28/IL-28RA (IFN- λ) signaling. *PLoS ONE* 9(7):e103249
180. Pingwara R et al (2017) Interferon lambda 2 promotes mammary tumor metastasis via angiogenesis extension and stimulation of cancer cell migration. *J Physiol Pharmacol* 68(4):573–583
181. Toh B et al (2011) Mesenchymal transition and dissemination of cancer cells is driven by myeloid-derived suppressor cells infiltrating the primary tumor. *PLoS Biol* 9(9):e1001162
182. Gabrilovich DI, Ostrand-Rosenberg S, Bronte V (2012) Coordinated regulation of myeloid cells by tumours. *Nat Rev Immunol* 12(4):253–268
183. Li Y et al (2021) Targeting myeloid-derived suppressor cells to attenuate vasculogenic mimicry and synergistically enhance the anti-tumor effect of PD-1 inhibitor. *iScience* 24(12):103392
184. Amulic B et al (2012) Neutrophil function: from mechanisms to disease. *Annu Rev Immunol* 30:459–489
185. Jaillon S et al (2020) Neutrophil diversity and plasticity in tumour progression and therapy. *Nat Rev Cancer* 20(9):485–503
186. Mueller MD et al (2000) Neutrophils infiltrating the endometrium express vascular endothelial growth factor: potential role in endometrial angiogenesis. *Fertil Steril* 74(1):107–112
187. Heryanto B, Girling JE, Rogers PA (2004) Intravascular neutrophils partially mediate the endometrial endothelial cell proliferative response to oestrogen in ovariectomised mice. *Reproduction* 127(5):613–620
188. Jablonska J et al (2010) Neutrophils responsive to endogenous IFN- β regulate tumor angiogenesis and growth in a mouse tumor model. *J Clin Invest* 120(4):1151–1164
189. Scapini P et al (2004) CXCL1/macrophage inflammatory protein-2-induced angiogenesis in vivo is mediated by neutrophil-derived vascular endothelial growth factor-A. *J Immunol* 172(8):5034–5040
190. Gordon-Weeks AN et al (2017) Neutrophils promote hepatic metastasis growth through fibroblast growth factor 2-dependent angiogenesis in mice. *Hepatology* 65(6):1920–1935
191. Shojaei F et al (2008) Role of Bv8 in neutrophil-dependent angiogenesis in a transgenic model of cancer progression. *Proc Natl Acad Sci USA* 105(7):2640–2645
192. Bordbari S et al (2022) SIRT1-mediated deacetylation of FOXO3a transcription factor supports pro-angiogenic activity of interferon-deficient tumor-associated neutrophils. *Int J Cancer* 150(7):1198–1211
193. Numasaki M et al (2005) IL-17 enhances the net angiogenic activity and in vivo growth of human non-small cell lung cancer in SCID mice through promoting CXCR-2-dependent angiogenesis. *J Immunol* 175(9):6177–6189
194. Nozawa H, Chiu C, Hanahan D (2006) Infiltrating neutrophils mediate the initial angiogenic switch in a mouse model of multistage carcinogenesis. *Proc Natl Acad Sci USA* 103(33):12493–12498
195. Queen MM et al (2005) Breast cancer cells stimulate neutrophils to produce oncostatin M: potential implications for tumor progression. *Cancer Res* 65(19):8896–8904
196. Ardi VC et al (2007) Human neutrophils uniquely release TIMP-free MMP-9 to provide a potent catalytic stimulator of angiogenesis. *Proc Natl Acad Sci USA* 104(51):20262–20267
197. Gaudry M et al (1997) Intracellular pool of vascular endothelial growth factor in human neutrophils. *Blood* 90(10):4153–4161
198. Sorin M et al (2023) Single-cell spatial landscapes of the lung tumour immune microenvironment. *Nature* 614(7948):548–554
199. Nadkarni S et al (2016) Neutrophils induce proangiogenic T cells with a regulatory phenotype in pregnancy. *Proc Natl Acad Sci USA* 113(52):E8415–e8424
200. Teuwen LA et al (2021) Tumor vessel co-option probed by single-cell analysis. *Cell Rep* 35(11):109253
201. Tsai YM et al (2021) Cooperation between cancer and fibroblasts in vascular mimicry and N2-type neutrophil recruitment via Notch2-Jagged1 interaction in lung cancer. *Front Oncol* 11:696931
202. Papayannopoulos V (2018) Neutrophil extracellular traps in immunity and disease. *Nat Rev Immunol* 18(2):134–147
203. Jung HS et al (2019) Cancer cell-induced neutrophil extracellular traps promote both hypercoagulability and cancer progression. *PLoS ONE* 14(4):e0216055
204. Yang S et al (2023) Neutrophil extracellular traps promote angiogenesis in gastric cancer. *Cell Commun Signal* 21(1):176
205. Aldabbous L et al (2016) Neutrophil extracellular traps promote angiogenesis: evidence from vascular pathology in pulmonary hypertension. *Arterioscler Thromb Vasc Biol* 36(10):2078–2087
206. Lavoie SS et al (2018) Synthesis of human neutrophil extracellular traps contributes to angiopoietin-mediated in vitro proinflammatory and proangiogenic activities. *J Immunol* 200(11):3801–3813
207. Browning JL et al (2005) Lymphotoxin- β receptor signaling is required for the homeostatic control of HEV differentiation and function. *Immunity* 23(5):539–550
208. Veerman K et al (2019) Single-cell analysis reveals heterogeneity of high endothelial venules and different regulation of genes controlling lymphocyte entry to lymph nodes. *Cell Rep* 26(11):3116–3131.e5
209. Allen E et al (2017) Combined antiangiogenic and anti-PD-L1 therapy stimulates tumor immunity through HEV formation. *Sci Transl Med*. <https://doi.org/10.1126/scitranslmed.aak9679>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.