



Male's age and plumage coloration predicts brood sex ratio in the Common Kingfisher (*Alcedo atthis*)

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Abstract

Plumage ornaments of birds, both pigment-based and structural, are considered age or condition-dependent, honestly signalling male quality, sexually selected traits, influencing the variation in breeding performance as well as adaptive sex allocation. In the present study, we examine the effect of age of males on the plumage structural colour in Common Kingfisher, and evaluate the breeding output (laying date, total number of young per breeding season, mean number of young per nest, total number of broods) and sex ratio in relation to two ornamental traits (forehead and rump structural colour). During the three years 2016–2018 in the Danube river system (south-western Slovakia), we collected data from 49 males, 102 broods and 645 nestlings. Our data demonstrate that structural colour is a condition-related and age-sensitive signal, while males with increased weight expressed less saturated blue forehead feathers, and old males displayed more saturated cyan rump feathers than young ones. Moreover, the brood sex ratio varies with male coloration in an age-dependent manner. The plumage colour of young males did not affect the brood sex ratio, whereas old males with increasing intensity of cyan rump had more sons than duller ones. Neither single ornamental trait nor age predicted breeding output of kingfisher males. Nevertheless, our results indicate that the blue structural plumage of kingfisher males may serve as an indicator of age and a certain kind of quality with a consequence on adaptive sex allocation in this species.

Keywords Age · Breeding success · Common Kingfisher · Feather ornamentation · Number of sons and daughters

Zusammenfassung

Alter und Gefiederfärbung des Männchens bestimmen das Geschlechterverhältnis beim Eisvogel (*Alcedo atthis*).

Der Gefiederschmuck von Vögeln, sowohl pigmentbasiert als auch strukturell, gilt als alters- oder konditionsabhängig und signalisiert ehrlich die Qualität der Männchen, sexuell selektierte Merkmale, die die Variation der Brutleistung sowie die adaptive Geschlechterverteilung beeinflussen. In der vorliegenden Studie untersuchen wir die Auswirkung des Alters der Männchen auf die Gefiederstrukturfarbe des Eisvogels und bewerten den Bruterfolg (Legedatum, Gesamtzahl der Jungen pro Brutsaison, durchschnittliche Anzahl der Jungen pro Nest, Gesamtzahl der Bruten) und das Geschlechterverhältnis in Bezug auf zwei Ziermerkmale (Stirn- und Bürzelstrukturfarbe). Während der drei Jahre 2016–2018 haben wir im Donauraum (SW Slowakei) Daten von 49 Männchen, 102 Bruten und 645 Nestlingen gesammelt. Unsere Daten zeigen, dass die Strukturfarbe

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ein konditionsabhängiges und altersempfindliches Signal ist, wobei Männchen mit höherem Gewicht weniger gesättigte blaue Stirnfedern und alte Männchen mehr gesättigte cyanfarbene Bürzelfedern aufweisen als junge. Außerdem variiert das Geschlechterverhältnis bei der Brut mit der Färbung der Männchen in einer altersabhängigen Weise. Die Gefiederfarbe der jungen Männchen hatte keinen Einfluss auf das Geschlechterverhältnis in der Brut. Hingegen hatten alte Männchen mit zunehmender Intensität des cyanfarbenen Bürzels mehr Söhne als mattere Männchen. Weder ein einzelnes Ziermerkmal noch das Alter sagten die Brutleistung von Eisvogel-Männchen voraus. Dennoch deuten unsere Ergebnisse darauf hin, dass das blaue Strukturgefieder der Eisvogelmännchen als Indikator für das Alter und eine bestimmte Art von Qualität dienen kann, was wiederum Auswirkungen auf die adaptive Geschlechtszuweisung bei dieser Art hat.

Introduction

Birds are the most colourful group of terrestrial vertebrates (Stoddard and Prum 2011). Their spectacular plumage coloration, often more elaborate in males, is produced by pigments—mostly melanin (providing structural support to feather, Burt 1981) and carotenoids (essential vitamin A precursors, Underwood 1984), physical interaction of light with feather structures (Prum and Torres 2003) or a combination of those two mechanisms (Hill 2006).

The role of pigment-based coloration as a signal of male quality is well studied in birds (Hill 2006; McGraw 2008; Guindre-Parker and Love 2014). On the other side, structure-based coloration has received much less attention (e.g. McGraw et al. 2002; Hill et al. 2005) and caused more controversy, since the self-assembly of the nanostructure reduces the potential for condition-dependent trait expression in structural coloration during feather development (Prum 2006; Prum et al. 2009). However, other features of the barb's cortex with melanosomes distribution may influence the colour variation produced by the feather (Shawkey et al. 2003). Hence, the honesty of structural signals may be maintained by the physiologically costly investment. The view that structure-based coloration can serve as a honest signal of male quality was supported by results of previous studies, founding more elaborate structure-based coloration in older males, those in better body condition or in males maintaining high-quality territories (Keyser and Hill 1999, 2000; Doucet 2002; Siefferman et al. 2005; Griggio et al. 2010a; Hyun-Young et al. 2016). Evidence also suggests that structural coloration may serve as a reliable indicator of individual quality (White 2020) in male's interactions (Alonso-Alvarez et al. 2004) as well as in mate choice (Andersson et al. 1998; Johnsen et al. 1998; Keyser and Hill 2000; Pearn et al. 2001; but see Ballentine and Hill 2003; Liu et al. 2007, 2009; Griggio et al. 2010b).

If the structural-based coloration serves as honest signal of male quality, females paired with more ornamented males should experience greater breeding performance benefiting from high-quality territory (Keyser and Hill 2000), high male investment in parental care (Siefferman and Hill 2003) and/or high-quality genotype inherited by their offspring from superior males (Norris 1993). Further, females can increase

investment into offspring produced with high-quality males and achieve high reproductive success (Harris and Uller 2009).

Additionally, these females could manipulate the sex ratio of offspring in an attempt to increase lifetime reproductive success (Ellegren et al. 1996). This may be true when one sex is more valuable than other (Fisher 1958; Hamilton 1967; Charnov 1982; Sheldon 1998). In such cases, females in better condition, breeding in high-quality environments or paired with high-quality males or males who are able or willing to invest more in parental care thus should produce high-quality sons who might be more successful in competing for access to females (Trivers and Willard 1973; Clutton-Brock 1984; Svensson and Nilsson 1996; Bowers et al. 2013). Similarly, females mated with more attractive males should preferably produce sons which would inherit attractiveness of their fathers (Burley 1981; Ellegren et al. 1996; Sheldon et al. 1999; Griffith et al. 2003, but see Parker 2013). If inherited traits have a greater effect on the reproductive fitness of male than on female, sons would benefit more than daughters by inheriting attractiveness of their father (West and Sheldon 2002, but see Ewen et al. 2004; Booksmythe et al. 2017).

In this study, we focused on structural plumage coloration in the Common Kingfisher (*Alcedo atthis*). We used data collected across three breeding seasons to assess the relationship between males' structure-based coloration, condition, age, breeding output and sex ratio. If plumage coloration is a good predictor of male quality, we expect to find superior males (those in better body condition/older) having more elaborate plumage coloration. Further, if male coloration is a sexually selected trait, we predict higher reproductive success in males with more elaborate plumage. Finally, if inherited coloration will have a greater positive effect on the reproductive fitness of sons than daughters, we expect to find the sex ratio shifted towards sons in broods of males with more elaborate plumage coloration.

Material and methods

Studied species

In the Common Kingfisher, both males and females express a very distinctive cyan feather ornament, which

is mainly structural. Cyan feathers of the head and back consist of cyan and blue barbs containing a keratinous spongy nanostructure. Nanostructures create iridescence and are likely to have a structural basis. These feathers have a high reflectance below 500 nm and a low reflectance above 600 nm (Stavenga et al. 2011). Males are known to erect blue feathers on their foreheads during aggressive male–male interactions and to show female brightly coloured rump feathers as a part of the courtship display (Cramp 1985).

The distribution of the Common Kingfisher extends across the whole Palearctic, breeding in most of Europe (Keller et al. 2020). It depends on the open water for feeding and the steep soil banks, in which they dig a breeding tunnel (Cramp 1985; Turčoková et al. 2016). Populations in Central Europe breed from March to September (Čech 2010; Rubáčová et al. 2020). They are mostly socially monogamous, with the occurrence of polygamy and production of extra-pair young (Cramp 1985; Woodall 2001; Čech 2009a; Libois 2018; Cepková et al. 2022a). Females lay up to four clutches per breeding season with two being the most common (Rubáčová et al. 2020), while their consecutive breeding attempts usually overlap (Čech 2010; Rubáčová et al. 2020). High offspring production apparently compensates for the rather low winter survival (Rubáčová et al. 2021). Both sexes dig a breeding tunnel, incubate eggs and care for young (Cramp 1985; Woodall 2001). Nestling sex ratio is equal within the population, while an obvious shift in individual broods remains unexplained (Cepková et al. 2019, 2022b).

Fieldwork

We studied Common Kingfishers from the beginning of April to late September during the breeding seasons 2016–2018 in the Danube river system (south-western Slovakia), particularly between 1868.7 (Bratislava, 48° 06' 13.5" N 17° 09' 31.3" E) and 1819.0 river km (Gabčíkovo, 47° 52' 32.1" N 17° 31' 18.0" E). We were looking for nests from the boat. Active nests were inspected weekly using a miniature camera (Probe Maxivideo MV 201).

After hatching, parents were captured using mist-nets installed near their nesting hole, and both members of the breeding pair were ringed and blood sampled. Males were subsequently aged as a second year (hereafter young) and after second year (hereafter old) following Čech (2009b), weighted and their tarsus length was measured. To obtain feathers for later spectral analyses, we cut ten feathers from the forehead and rump of each male. We stored the feathers in separate envelopes in a climate-controlled environment until further processing.

When nestlings were at least 12 days old (range 12–25, mean 16 days, $N=645$), we used bent iron wire to gently

take them out of the nest chamber. After ringing, we took a small blood sample (25 μ l) from each nestling for later sex determination and paternity analyses. Blood samples were subsequently suspended in 96% ethanol and frozen in a plastic tube.

To assess the number of broods per breeding season within a breeding pair, we continuously checked all marked breeding pairs until the end of the breeding season. The fieldwork was completed at the end of September after all the young had fledged.

Body condition

Individual condition was assessed by calculating values for body condition (defined by scaled mass index; SMI). SMI was calculated for each individual using a scaling exponent (estimated by the standardised major axis regression of body mass on linear body measurement, in our study tarsus length), arithmetic mean value of tarsus length and log-transformed body mass (Peig and Green 2009).

Spectral analyses

Ten feathers from both patches (forehead, rump) were used for spectral analyses. We arranged feathers so that they overlapped extensively and took three readings from different parts of each set of feathers. All readings were taken using an Ocean Optics JAZ spectrometer (Ocean Optics Inc., Dunedin, FL, USA) and the micron fibre-optic probe at a 90° angle to the feather surface. We averaged patch-specific spectra for each individual, which resulted in one average reflectance spectrum for each patch and bird. We used the R package “pavo” (Maia et al. 2013) to calculate summary spectral characteristics for each feather patch. In particular, we calculated the hue (spectral location) of the rump and forehead as a wavelength of maximum reflectance $\lambda_{(R_{max})}$. Chroma (spectral purity) was calculated as a relative contribution of a spectral range to the total brightness. Since patches of blue feathers are also reflected in the UV range, we calculated “blue chroma” ($R_{400-510}/R_{320-700}$) and “UV chroma” ($R_{320-400}/R_{320-700}$) (White and Cristol 2014). For both patches, we calculated brightness (spectral intensity) as mean relative reflectance over the entire spectral range (Montgomerie 2006).

Molecular procedures

In total, 645 nestlings from 49 males and 102 nests were sexed (Table 1). DNA was extracted from blood samples by E.Z.N.A.® Tissue DNA Kit. Subsequently, PCR amplification and electrophoresis were performed to identify the sex of nestlings. The volume of PCR mix was 10 μ l. Each sample contained: 2 μ l DNA, 0.2 μ l of primer sex1 and sex2, 5 μ l

Master Mix (VWR Red Taq DNA Polymerase Master Mix) and 2.6 µl deionized water. We also performed PCR reaction with primers P2 and P8 (Griffiths et al. 1998), but the sex1, sex2 method had a better outcome. Primers and procedures were done according to Wang and Zhang (2009). Electrophoresis was performed 60 min at 100 V on 2% agarose gel containing GoodView. Then, UV light was used to visualize the presence of bands: two bands identifying female and one band identifying male. Sex was determined for all nestlings we sampled. There was also a control sample of one adult male and one adult female for each electrophoresis.

For parentage analyses, the forward primers were fluorescently labelled, and multiplex PCR kit (QIAGEN Multiplex PCR Plus kit) was used to amplify four microsatellite loci (AACC-106, Be2.46, Bb111, CAM17). Fragment analysis was carried out commercially in the Comenius University Science Park (Bratislava, Slovakia) and its results were visualised by software GeneMarker. Alleles of putative parents were compared to those of nestlings to determine whether they were or were not within-pair offspring. Nestlings failed to be considered as within-pair if their alleles mismatched with those of their parent at least in one locus (see Cepková et al. 2022a).

Statistical analyses

We tested data for normality (Shapiro–Wilk *W* test). Data lacking normal distribution were ln-transformed (brightness forehead, UV chroma rump, laying date). Transformed data approached a normal distribution.

To reduce spectral data recorded from two body regions to a limited number of variables, we performed principal component analyses (PCA) with varimax rotation, based on correlation matrices of standardized plumage characteristics (Table 2). The varimax rotation simplified the loadings of items by removing the middle ground and more specifically identified the factor upon which data load. Principal component analysis reduced the number of colour variables into a more manageable number (from eight variables to two components). In total, two principal components explained 74.2% of variation among colour variables. First principal component (PC1) explained 40.6% of the variation in data and received strong loadings from forehead structural blue coloration variables. An individual with a high positive PC1 score has more blue as well as more UV saturation in forehead feathers (colour appears more intense blue and brighter). Second

Table 2 Principal component analysis (PCA) loadings

Variable	PC1	PC2
Brightness_forehead	0.745	−0.314
Hue_forehead	−0.691	0.089
UV chroma_forehead	0.741	0.239
Blue chroma_forehead	0.908	0.192
Hue_rump	0.051	−0.960
Blue chroma_rump	0.088	0.956

principal component (PC2) explained 33.6% of the variation and received strong loadings from rump structural blue coloration variables. An individual with a high positive PC2 score for rump colour has feathers with greater blue saturation and with a more left-shifted hue (colour appears more intense cyan).

To determine which parameter best explains variation in phenotypic characteristics of males we used generalised linear models. We set year of the study, male age and scaled mass index (SMI) as predictors and two principal components as a continuous dependent variable.

First egg-laying date was defined as the date when the female laid her first egg. It was measured as Julian date, while in each year we determined the start of the breeding season by the earliest day of the first egg-laying date in population, and assign this day as number 1 (hereafter we will use the laying date as the parameter name). We quantified seasonal reproductive output as the total number of offspring fledged from all broods per breeding season to a specific male (hereafter nestling number), and total number of broods belonging to a specific male per breeding season (hereafter brood number). As these two characteristics were highly correlated (Pearson's $r=0.976$; $p<0.001$; $n=49$), we used only nestling number/breeding season in the analyses.

To investigate whether the year of study, age and phenotypic characteristics of males predicted variation in their seasonal reproductive output (response variable: laying date, nestling number/breeding season), we used generalized linear models with normal distribution and identity link function (McCullagh and Nelder 1989). Year of study and the age of a male were set as a categorical predictors and two principal components were set as continuous predictors. As each male was used only once in the analyses, we did not set male identity as a random factor. In addition, we performed generalized linear mixed model with normal distribution and

Table 1 Number of males, broods and nestlings of Common Kingfishers used in sex ratio analysis

Season	Males	Broods	Young	Sons	Daughters	Sex ratio
2016	23	46	290	140	150	0.48
2017	13	23	144	73	71	0.51
2018	13	33	211	103	108	0.49
Total	49	102	645	316	329	0.49

identity link function to examine the correlation between the number of young in each brood (nestling number/nest) and male age, year of study and phenotypic traits of the males. Male age and year of study were set as categorical predictors, two principal components of plumage coloration were set as continuous predictors and pull number/nest was set as a response variable. As males have typically more than one brood per breeding season, male identity was set as a random factor.

Sex ratio was defined as the proportion of males to the total number of nestlings in the nest. Brood sex ratio was analysed using generalized linear mixed model with binomial error distribution and logit link function (Krackow and Tkadlec 2001). Sex ratio was set as a response variable. Male's age and year of study (fixed effects) were set as a categorical predictors and two principal components obtained from colour measurements were set as continuous predictors. All statistical analyses were performed using the statistical program STATISTICA for WINDOWS v. 8 (StatSoft Inc., Tulsa, OK, USA) and IBM SPSS Statistics v. 23 (Armonk, NY: IBM Corp.).

Results

Kingfisher males had mean 1.91 ± 0.10 SE broods per breeding season (range 1–5 broods, $N = 102$ broods) with the mean number of nestlings per brood was 6.32 ± 0.94 SE (range 3–8 nestlings, $N = 102$ broods).

Male ornamentation in relation to age and body condition

Male body condition was negatively correlated with blue structural coloration of the forehead (PC1). Males with a higher PC1 score (greater blue and UV chroma in forehead feather) have a lower value of SMI (Fig. 1). No correlation has been found between the body condition and structural coloration of the rump (PC2).

Male age was not correlated with blue structural coloration of the forehead (PC1), but was a significant predictor of blue structural coloration of the rump (PC2). Old males displayed significantly more elaborate rump coloration consisting of more cyan-coloured (more left-shifted hue) and more saturated (with greater blue chroma) and brighter feathers, than young males (Fig. 2a). There was no difference in male coloration between the 3 years the research was conducted (Table 3).

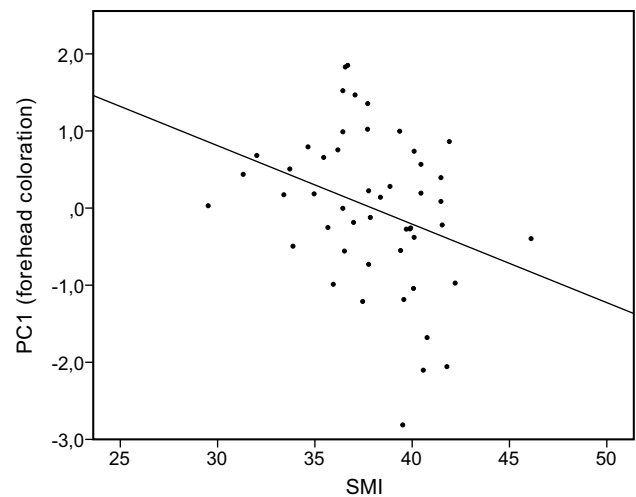


Fig. 1 Correlation between the forehead coloration (PC1) plotted against the body condition (scaled mass index, SMI) of kingfisher males. Circles intersected by regression fit indicate values of forehead coloration in each data point

Male age and ornamentation in relation to breeding success and brood sex ratio

Seasonal male reproductive output was not affected by male age, forehead coloration (PC1) or rump coloration (PC2) and did not differ between the three years of research (Table 4).

Males age and rump coloration (PC2) were a good predictors of the brood sex ratio (Table 5), since broods of young males (Fig. 2b) and those with more intense cyan rump feathers (PC2) were more skewed towards sons. In the group of young males, no correlation was found between forehead coloration (PC1), rump coloration (PC2) and brood sex ratio. In the group of old males, those with more intense rump coloration (PC2) had a sex ratio skewed towards sons (Fig. 3). Forehead coloration (PC1) did not significantly affect the brood sex ratio in this group of males (Table 5).

Discussion

In our population of Common Kingfisher, the expression of male structural plumage ornamentation was shown to be sensitive to body condition. Males in lower body condition were also those with a more pronounced blue forehead. This runs contrary to previous studies, which found a positive correlation between male structural ornamentation and body condition (Doucet 2002; Siefferman et al. 2005; Griggio et al.

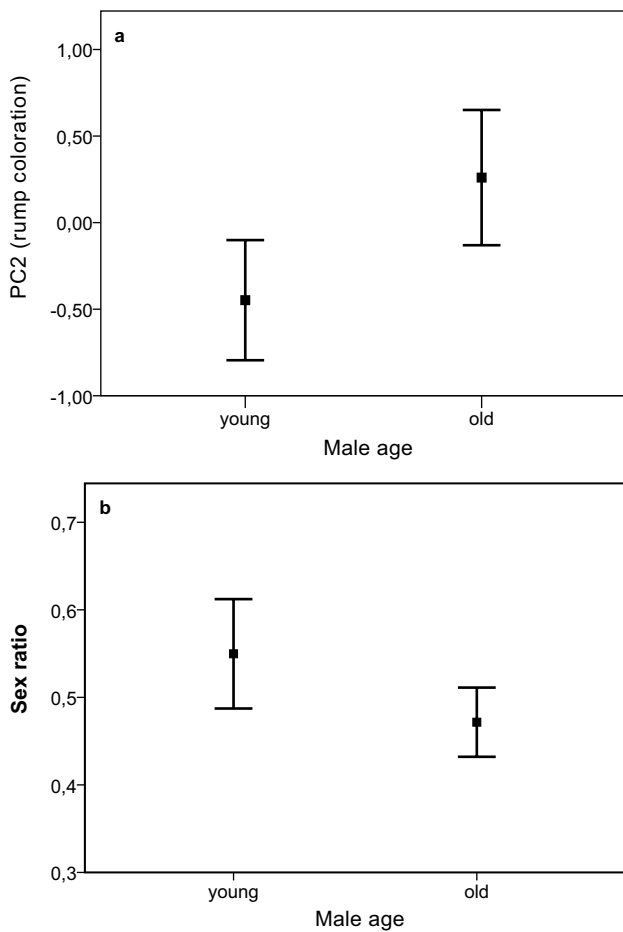


Fig. 2 Differences between age groups of kingfisher males in **a** rump coloration (PC2), **b** nestling sex ratio. Means and confidence intervals are presented

Table 3 Relationship between PC factors of plumage ornament (PC1=forehead structural color, PC2=rump structural color, set as continuous response variables) and year (2016, 2017, 2018), age (2K, +2K set as categorical predictor) and scaled mass index (SMI, set as continuous predictor)

Model	Predictor	Estimate	SE	<i>t</i>	<i>P</i>
PC1	Year	-0.274	0.360	0.579	0.447
	Age	-0.137	0.283	0.233	0.630
	SMI	-0.104	0.042	6.147	0.013
PC2	Year	0.440	0.355	1.533	0.216
	Age	-0.630	0.279	5.074	0.024
	SMI	-0.029	0.041	0.479	0.489

Parameter estimate, standard error and significance for predictor variables used in generalized only GLM

Table 4 Relationship between laying date, nestling number/season, nestling number/nest (set as continuous response variables) and year, age (set as categorical predictor), PC factors of plumage ornament (PC1=forehead structural color, PC2=rump structural color, set as continuous predictors)

Model	Predictor	Estimate	SE	<i>t</i>	<i>P</i>
Laying date					
	Year	10.262	9.058	1.133	0.264
	Age	9.098	7.857	1.158	0.253
	PC1	0.001	3.575	0.000	1.000
	PC2	-0.284	3.824	-0.336	0.739
Nestling/season					
	Year	-2.900	2.574	-1.127	0.266
	Age	-0.696	2.232	-0.312	0.757
	PC1	0.028	1.016	0.027	0.978
	PC2	0.525	1.086	0.484	0.631
Nestling/nest					
	Year	-0.092	0.218	-0.420	0.675
	Age	0.087	0.187	0.468	0.641
	PC1	-0.120	0.082	-1.461	0.147
	PC2	-0.150	0.091	-1.658	0.101

Parameter estimate, standard error and significance for predictor variables used in generalized maybe only GLM

Table 5 Relationship between sex ratio (set as binomial response variable), year, age (categorical predictors) and PC factors of plumage ornament (PC1=forehead structural color, PC2=rump structural color, set as continuous predictors). Model (1) analysed both age categories of males (2K, +2K), model (2) only young males (2K), model (3) only old males (+2K)

Model	Predictor	Estimate	SE	<i>t</i>	<i>P</i>
Sex ratio (all males)					
	Year	-0.139	0.129	-1.075	0.289
	Age	0.327	0.119	2.754	0.009
	PC1	-0.036	0.050	-0.718	0.477
	PC2	0.190	0.059	3.196	0.003
Sex ratio (young males)					
	Year	0.605	0.430	1.407	0.183
	PC1	0.140	0.131	1.069	0.305
	PC2	-0.097	0.274	-0.356	0.728
Sex ratio (old males)					
	Year	-0.230	0.134	-1.718	0.098
	PC1	-0.039	0.059	-0.656	0.518
	PC2	0.189	0.061	3.113	0.004

Parameter estimate, standard error and significance for predictor variables used in GLMM. Significant results are in bold

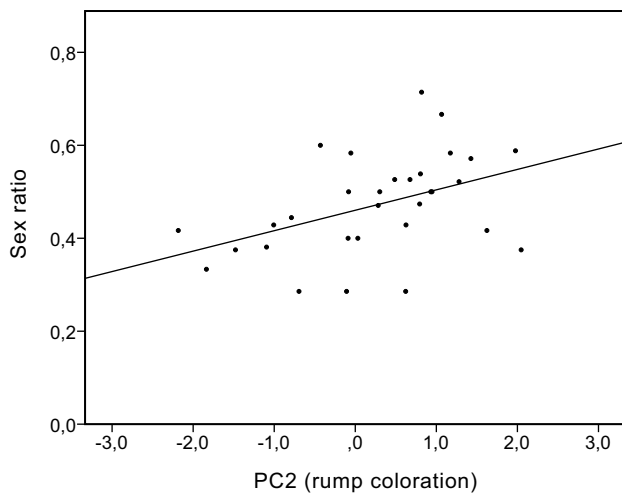


Fig. 3 Correlation between nestling sex ratio and rump coloration (PC2) in old males of Common Kingfisher. Circles intersected by regression fit indicate values of sex ratio in each data point

2010a). As body condition interprets variations in energy reserves for a given body size (Peig and Green 2009), a possible explanation is that increased weight, which is traditionally considered better condition in birds (Wendeln and Becker 1999; Robinson et al. 2005; Peig and Green 2009) may indeed be a sign of lower condition for some species. For example, kingfisher males are known to aggressively attack intruders in aerial combat and also perform aerial display to attract females (Cramp 1985; Woodall 2001). It is probably more advantageous for such behaviour to weigh less and be more agile (Székely et al. 2006).

In addition, we found evidence that structural plumage coloration is related to age, with old males displaying more intense cyan rump than the young ones. Those results are in agreement with other studies, showing that young birds display less elaborate plumage compared to old ones (Siefferman et al. 2005; Delhey and Kempenaers 2006; Bitton and Dawson 2008). Such an age-related differences in expression of ornamentation may be due to a within individual increase in ornamentation, correlation between intensity of ornamentation and condition, or correlation between signalling and survival (Jennions et al. 2001; Proulx et al. 2002; Lindström et al. 2009; Grunst et al. 2014; Moore et al. 2015). More specifically, age-related differences in intensity of ornamentation can be caused by differential survival of more ornamented individuals, meaning that often the more ornamented, presumably high-quality individuals show higher apparent survival than less ornamented and low-quality ones (Jennions et al. 2001). More ornamented individuals are thus over-represented in older age classes (Forslund and Pärt 1995). Since body condition does not seem to correlate with rump coloration, and we do not have enough data on coloration and survival analyses, nor data to test individual

changes in ornamentation, we can only speculate how this works in kingfishers. Either way, a female kingfisher seems to be able to assess the age of her potential mate using structural colour of his rump as an indicator.

Surprisingly, the breeding output of kingfisher males was not affected by their age with older and younger males did not differ in laying date, number of young per breeding season or number of young per nest. The effect of the male's age on breeding output can be obscured by the lower survival of this species, leading to a rather short lifespan (Rubáčová et al. 2021). Although it is reported in the literature that kingfishers can live up to 15 years (Woodall 2001), the long-term ringing schemes in Slovakia and Czechia show that only very few males older than four years are represented in the population (Rubáčová unpubl. data, Čech pers. com.). With such a short lifespan, the differences in amount of parental experience between males can be expected to be quite small.

Furthermore, the breeding output of kingfisher males was not affected by their structure-based ornamentation. A possible explanation is that in kingfisher males parental performance is not related to their structure-based ornamentation, similarly as was found in the Bluethroat (*Luscinia svecica*, Smiseth et al. 2001). However, our results do not mean that females do not have any apparent advantage from mating with more ornamented males. Furthermore, it is necessary to also mention the importance of female and her parental abilities. Although there is no evidence of condition-dependent mate preferences in kingfishers as it can be in other birds (Cotton et al. 2006; Holveck et al. 2011), we do know that investment into parental care and subsequently breeding output in kingfishers is affected by the combination of age and actual condition of both partners (Cepková et al. 2022b), which could obscure the effect of male parental performance on breeding output.

Interestingly, males with more elaborate rump coloration had more sons than daughters, as was also found in other studies (Sheldon et al. 1999; Griffith et al. 2003; Delhey et al. 2007). This is consistent with the sex-allocation theory according to which more attractive males produce more valuable sex, inheriting the attractiveness of their father. The attractiveness is more beneficial for sons as they usually have a higher variance in breeding performance than females (Ellegren et al. 1996; West and Sheldon 2002, but see Ewen et al. 2004; Booksmythe et al. 2017). This may be due to extra-pair mating, polygamy or higher survival rates (Johnsen et al. 1998; Örnborg et al. 2001; Griffith et al. 2003; Barenger et al. 2009), which also applies to the Common Kingfisher (Rubáčová et al. 2021; Cepková et al. 2022a).

Surprisingly, in our population, the relationship between brood sex ratio and male coloration was found only in old males. Young males produced more sons than old males, regardless of the rump coloration. Age-related adjustment

of the brood sex ratio was documented also in other studies, but explanation remains unclear. Specifically in Blue Tit (*Cyanistes caeruleus*), a positive effect of female but not male age on brood sex ratio was found only in some years, while not in others (Griffith et al. 2003). In another study, it was the age of the male that affected the sex ratio of the offspring. Young males with more elaborate plumage had more sons, while old brightly ornamented males had more daughters (Delhey et al. 2007).

In conclusion, our results demonstrate that plumage structural coloration of kingfisher males is condition related and age dependent. We also show that the brood sex ratio varies with the age and structural coloration of males. Our work also opens new directions for future studies, specifically in the field of relationship between plumage structural colour of kingfisher males and survival, mate choice, paternal care or habitat quality.

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Study methods comply with the current laws of the country in which they were performed. Permissions to carry out the study were granted by the relevant national authorities of the Ministry of the Environment of the Slovak Republic.

Data availability The datasets analysed during the current study are available from the corresponding author on reasonable request.

Declarations

Conflict of interest The authors declare no conflict of interests.

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References

- Alonso-Alvarez C, Doutrelant C, Sorci G (2004) Ultraviolet reflectance affects male–male interactions in the blue tit (*Parus caeruleus ultramarinus*). *Behav Ecol* 15:805–809. <https://doi.org/10.1093/beheco/arih083>
- Andersson S, Ornborg J, Andersson M (1998) Ultraviolet sexual dimorphism and assortative mating in blue tits. *Proc R Soc Lond B* 265:445–450. <https://doi.org/10.1098/rspb.1998.0315>
- Ballentine B, Hill GE (2003) Female mate choice in relation to structural plumage coloration in Blue Grosbeaks. *Condor* 105(3):593–598. <https://doi.org/10.1093/condor/105.3.593>
- Barenger SL, Johnson LS, Masters BS (2009) Sexual selection in a socially monogamous bird: male color predicts paternity success in the Mountain Bluebird, *Sialia currucoides*. *Behav Ecol Sociobiol* 63:403–411. <https://doi.org/10.1007/s00265-008-0674-5>
- Bitton PP, Dawson RD (2008) Age-related differences in plumage characteristics of male tree swallows *Tachycineta bicolor*: hue and brightness signal different aspects of individual quality. *J Avian Biol* 39:446–452. <https://doi.org/10.1111/j.0908-8857.2008.04283.x>
- Booksmythe I, Mautz B, Davis J, Nakagawa S, Jennions MD (2017) Facultative adjustment of the offspring sex ratio and male attractiveness: a systematic review and meta-analysis. *Biol Rev* 92:108–134. <https://doi.org/10.1111/brv.12220>
- Bowers EK, Munclinger P, Bures S, Kucerova L, Nadvornik P, Krist M (2013) Cross-fostering eggs reveals that female collared flycatchers adjust clutch sex ratios according to parental ability to invest in offspring. *Mol Ecol* 22:215–228. <https://doi.org/10.1111/mec.12106>
- Burley N (1981) Sex-ratio manipulation and selection for attractiveness. *Science* 211:721–722. <https://doi.org/10.1126/science.211.4483.721>
- Burt EH Jr (1981) The adaptiveness of animal colors. *Bioscience* 31:723–729. <https://doi.org/10.2307/1308778>
- Čech P (2010) Length of the breeding season of the Eurasian Kingfisher (*Alcedo atthis*) in the Czech Republic. *Sylvia* 46:53–61
- Čech P (2009a) Paper on knowledge about nesting biology of Kingfisher (*Alcedo atthis*). In: Čech P (ed) Ledňáček říční (*Alcedo atthis*), jeho ochrana a výzkum. Sborník referátů z II. Mezinárodního semináře, Vlašim, pp 118–125
- Čech P (2009b) Paper on age and sex determination of Kingfisher (*Alcedo atthis*). In: Čech P (ed) Ledňáček říční (*Alcedo atthis*), jeho ochrana a výzkum. Sborník referátů z II. Mezinárodního semináře, Vlašim, pp 118–125 (in Czech, English abstract)
- Cepková M, Balážová M, Melišková M, Rubáčová-Turčoková L (2019) No seasonal variation of the sex ratio in the Common Kingfisher *Alcedo atthis* broods. *Acta Ornithol* 54:149–155. <https://doi.org/10.3161/00016454AO2019.54.2.002>
- Cepková M, Melišková M, Rubáčová L (2022a) Low extra-pair paternity and polygamy in the common Kingfisher *Alcedo atthis*. *Ardeola* 70:41–58. <https://doi.org/10.13157/arla.70.1.2023.ra2>
- Cepková M, Balážová M, Melišková M, Rubáčová L (2022b) Influence of age and body condition on breeding performance in Common Kingfisher *Alcedo atthis*. *J Ornithol* 163:251–261
- Charnov EL (1982) The theory of sex allocation. Princeton University Press, Princeton
- Clutton-Brock TH, Albon SD, Guinness FE (1984) Maternal dominance, breeding success and birth sex ratios in red deer. *Nature* 308:358–360. <https://doi.org/10.1038/308358a0>
- Cotton S, Small J, Pomiankowski A (2006) Sexual selection and condition-dependent mate preferences. *Curr Biol* 16:R755–R765. <https://doi.org/10.1016/j.cub.2006.08.022>
- Cramp S (1985) The Birds of the Western Palearctic, vol IV: Terns to Woodpeckers. Oxford University Press, Oxford

- Delhey K, Kempenaers B (2006) Age differences in blue tit plumage colour: within-individual changes or colour-biased survival? *J Avian Biol* 37:339–348. <https://doi.org/10.1111/j.2006.0908-8857.03655.x>
- Delhey K, Peters A, Johnsen A, Kempenaers B (2007) Brood sex ratio and male UV ornamentation in blue tits (*Cyanistes caeruleus*): correlational evidence and an experimental test. *Behav Ecol Sociobiol* 61:853–862. <https://doi.org/10.1007/s00265-006-0314-x>
- Doucet S (2002) Structural plumage coloration, male body size, and condition in the blue-black grassquit. *Condor* 104:30–38. <https://doi.org/10.1093/condor/104.1.30>
- Ellegren H, Gustafsson L, Sheldon BC (1996) Sex ratio adjustment in relation to paternal attractiveness in a wild bird population. *PNAS* 93:11723–11728. <https://doi.org/10.1073/pnas.93.21.11723>
- Ewen JG, Cassey P, Møller AP (2004) Facultative primary sex ratio variation: a lack of evidence in birds? *Proc R Soc Lond B* 271:1277–1282. <https://doi.org/10.1098/rspb.2004.2735>
- Fisher RA (1958) *The Genetical Theory of Natural Selection*. Clarendon, Oxford
- Forslund P, Pärt T (1995) Age and reproduction in birds—hypotheses and tests. *Trends Ecol Evol* 10:374378. [https://doi.org/10.1016/S0169-5347\(00\)89141-7](https://doi.org/10.1016/S0169-5347(00)89141-7)
- Griffith SC, Örnborg J, Russell AF, Andersson S, Sheldon BC (2003) Correlations between ultraviolet coloration, overwinter survival and offspring sex ratio in the blue tit. *J Evol Biol* 16:1045–1054. <https://doi.org/10.1046/j.1420-9101.2003.00550.x>
- Griffiths R, Double MC, Orr K, Dawson RJG (1998) A DNA test to sex most birds. *Mol Ecol* 7:1071–1075. <https://doi.org/10.1046/j.1365-294x.1998.00389.x>
- Griggio M, Zanollo V, Hoi H (2010a) UV plumage color is an honest signal of quality in male budgerigars. *Ecol Res* 25:77–82. <https://doi.org/10.1007/s11284-009-0632-3>
- Griggio M, Hoi H, Pilastro A (2010b) Plumage maintenance affects ultraviolet colour and female preference in the budgerigar. *Behav Process* 84:739–744. <https://doi.org/10.1016/j.beproc.2010.05.003>
- Grunst AS, Rotenberry JT, Grunst ML (2014) Age-dependent relationships between multiple sexual pigments and condition in males and females. *Behav Ecol* 25:276–287. <https://doi.org/10.1093/beheco/art124>
- Guindre-Parker S, Love OP (2014) Revisiting the condition dependence of melanin-based plumage. *J Avian Biol* 45:29–33. <https://doi.org/10.1111/j.1600-048X.2013.00190.x>
- Hamilton WD (1967) Extraordinary sex ratios. *Science* 156:477–488. <https://doi.org/10.1126/science.156.3774.477>
- Harris WE, Uller T (2009) Reproductive investment when mate quality varies: differential allocation versus reproductive compensation. *Philos Trans R Soc B Biol Sci* 364:1039–1048. <https://doi.org/10.1098/rstb.2008.0299>
- Hill GE (2006) Environmental regulation of ornamental coloration. In: Hill GE, McGraw KJ (eds) *Bird coloration vol. I: mechanisms and measurements*. Harvard University Press, Cambridge, pp 507–560
- Hill GE, Doucet SM, Buchholz R (2005) The effect of coccidial infection on iridescent plumage coloration in wild turkeys. *Anim Behav* 69:387–394. <https://doi.org/10.1016/j.anbehav.2004.03.013>
- Holveck MJ, Riebel K (2010) Low-quality females prefer low-quality males when choosing a mate. *Proc R Soc B Biol Sci* 277:153–160. <https://doi.org/10.1098/rspb.2009.1222>
- Hyun-Young N, Lee S, Lee J, Choi Ch, Choe JC (2016) Multiple structural colors of the plumage reflect age, sex, and territory ownership in the Eurasian Magpie *Pica pica*. *Acta Ornithol* 51:83–92. <https://doi.org/10.3161/00016454AO2016.51.1.007>
- Jennions MD, Møller AP, Petrie M (2001) Sexually selected traits and adult survival: a meta-analysis. *Q Rev Biol* 76:3–36. <https://doi.org/10.1086/393743>
- Johnsen A, Andersson T, Örnborg F, Lifjeld J (1998) Ultraviolet plumage coloration affects social mate choice and sperm competition in bluethroats (Aves: Luscinia s. svecica): a field experiment. *Proc R Soc Lond B* 265:1313–1318. <https://doi.org/10.1098/rspb.1998.0435>
- Keller V, Herrando S, Voríšek P, Franch M, Kipson M, Milanese P, Martí D, Anton M, Klvaňová A, Kalyakin MV, Bauer HG (2020) *European breeding bird atlas 2: distribution, abundance and change*. Lynx Edition, Barcelona
- Keyser A, Hill G (1999) Condition-dependent variation in the blue ultraviolet coloration of a structurally-based plumage ornament. *Proc R Soc Lond B* 265:771–777. <https://doi.org/10.1098/rspb.1999.0704>
- Keyser A, Hill G (2000) Structurally-based plumage color is an honest indicator of quality in male blue grosbeaks. *Behav Ecol* 11:202–209. <https://doi.org/10.1093/beheco/11.2.202>
- Krackow S, Tkadlec E (2001) Analysis of brood sex ratios: implications of offspring clustering. *Behav Ecol Sociobiol* 50:293–301. <https://doi.org/10.1007/s002650100366>
- Libois R (2018) *Plumes d’azur. Histoire naturelle du martin-pêcheur d’Europe*. Presses Universitaires de Liège, Gembloux
- Lindström J, Pike T, Blount J, Metcalfe N (2009) Optimization of resource allocation can explain the temporal dynamics and honesty of sexual signals. *Am Nat* 174:515–525. <https://doi.org/10.1086/606008>
- Liu M, Siefferman LM, Hill GE (2007) An experimental test of female choice relative to male structural coloration in eastern bluebirds. *Behav Ecol Sociobiol* 61:623–630. <https://doi.org/10.1007/s00265-006-0292-z>
- Liu M, Siefferman L, JrH M, Steffen JE, Hill GE (2009) A field test of female mate preference for male plumage coloration in eastern bluebirds. *Anim Behav* 78:879–885. <https://doi.org/10.1016/j.anbehav.2009.07.012>
- Maia R, Eliason CM, Bitton PP, Doucet SM, Shawkey MD (2013) pavo: an R package for the analysis, visualization and organization of spectral data. *Methods Ecol Evol* 4:906–913. <https://doi.org/10.1111/2041-210X.12069>
- McCullagh P, Nelder J (1989) *Generalized linear models*, 2nd edn. Chapman and Hall, Boca Raton
- McGraw KJ (2008) An update on the honesty of melanin-based color signals in birds. *Pigment Cell Melanoma Res* 21:133–138. <https://doi.org/10.1111/j.1755-148X.2008.00454.x>
- McGraw KJ, Mackillop EA, Dale J, Hauber ME (2002) Different colors reveal different information: how nutritional stress affects the expression of melanin and structurally based ornamental plumage. *J Exp Biol* 205:3747–3755. <https://doi.org/10.1242/jeb.205.23.3747>
- Montgomery R (2006) Analyzing colors. In: Hill GE, McGraw KJ (eds) *Bird coloration: mechanisms and measurements, vol 2*. Harvard University Press, Massachusetts, pp 90–147
- Moore FR, Ćirule D, Kivleniece I, Vrublevska J, Rantala MJ, Sild E, Sepp T, Hõrak P, Krama T, Krams I (2015) Investment in a sexual signal results in reduced survival under extreme conditions in the male great tit (*Parus major*). *Behav Ecol Sociobiol* 69:151–158. <https://doi.org/10.1007/s00265-014-1828-2>
- Norris K (1993) Heritable variation in a plumage indicator of viability in male great tits *Parus major*. *Nature* 362(6420):537–539. <https://doi.org/10.1038/362537a0>
- Örnborg J, Andersson S, Johnsen A, Lifjeld J, Amundsen T (2001) Male characteristics and fertilisation success in bluethroats. *Behaviour* 138:1371–1390. <https://doi.org/10.1163/156853901317367645>

- Parker TH (2013) What do we really know about the signaling role of plumage colour in blue tits? A case study of impediments to progress in evolutionary biology. *Biol Rev* 88:511–536. <https://doi.org/10.1111/brv.12013>
- Pearn SM, Bennett AT, Cuthill IC (2001) Ultraviolet vision, fluorescence and mate choice in a parrot, the budgerigar *Melopsittacus undulatus*. *Proc Biol Sci* 268:2273–2279. <https://doi.org/10.1098/rspb.2001.1813>
- Peig J, Green AJ (2009) New perspectives for estimating body condition from mass/length data: the scaled mass index as an alternative method. *Oikos* 118:1883–1891. <https://doi.org/10.1111/j.1600-0706.2009.17643.x>
- Proulx SR, Day T, Rowe L (2002) Older males signal more reliably. *Proc R Soc Lond B* 269:2291–2299. <https://doi.org/10.1098/rspb.2002.2129>
- Prum RO (2006) Anatomy, physics and evolution of avian structural colours. In: Hill GE, McGraw KJ (eds) *Bird coloration: mechanisms and measurements*, vol 2. Harvard University Press, Cambridge
- Prum RO, Torres RH (2003) A Fourier tool for the analysis of coherent light scattering by bio-optical nanostructures. *Integr Comp Biol* 43:591–602. <https://doi.org/10.1093/icb/43.4.591>
- Prum RO, Dufresne ER, Quinn T, Waters K (2009) Development of colour-producing keratin nanostructures in avian feather barbs. *J R Soc Interface* 6:S253–S265. <https://doi.org/10.1098/rsif.2008.0466.focus>
- Robinson S, Chiaradia A, Hindell MA (2005) The effect of body condition on the timing and success of breeding in Little Penguins *Eudyptula minor*. *Ibis* 147:483–489. <https://doi.org/10.1111/j.1474-919x.2005.00431.x>
- Rubáčová L, Čech P, Melišková M, Balážová M (2020) The length of breeding season in two populations of the Common Kingfisher (*Alcedo atthis*). *Sylvia* 56:39–48
- Rubáčová L, Čech P, Melišková M, Čech M, Procházka P (2021) The effect of age, sex and winter severity on return rates and apparent survival in the Common Kingfisher *Alcedo atthis*. *Ardea* 109:15–25. <https://doi.org/10.5253/arde.v109i1.a2>
- Shawkey MD, Estes AM, Siefferman LM, Hill GE (2003) Nanostructure predicts intraspecific variation in structural plumage colour. *Proc R Soc Lond B* 270:1455–1460. <https://doi.org/10.1098/rspb.2003.2390>
- Sheldon BC (1998) Recent studies of avian sex ratios. *Heredity* 80:397–402. <https://doi.org/10.1046/j.1365-2540.1998.00374.x>
- Sheldon BC, Andersson S, Griffith SC, Örnborg J, Sendecka J (1999) Ultraviolet colour variation influences blue tit sex ratios. *Nature* 402:874–877. <https://doi.org/10.1038/47239>
- Siefferman LM, Hill GE (2003) Structural and melanin plumage coloration indicate parental effort and reproductive success in male eastern bluebirds. *Behav Ecol* 14:855–861. <https://doi.org/10.1093/beheco/arg063>
- Siefferman LM, Hill GE (2005) Blue structural coloration of male eastern bluebirds *Sialia sialis* predicts incubation provisioning to females. *J Avian Biol* 36:488–493. <https://doi.org/10.1111/j.0908-8857.2005.03659.x>
- Siefferman L, Hill GE, Dobson S (2005) Ornamental plumage coloration and condition are dependent on age in eastern bluebirds *Sialia sialis*. *J Avian Biol* 36:428–435. <https://doi.org/10.1111/j.0908-8857.2005.03401.x>
- Smiseth P, Örnborg J, Andersson S, Amundsen T (2001) Is male plumage reflectance correlated with paternal care in bluethroats? *Behav Ecol* 12:164–170. <https://doi.org/10.1093/beheco/12.2.164>
- Stavenga DG, Tinbergen J, Leertouwer HL, Wilts BD (2011) Kingfisher feathers coloration by pigments, spongy nanostructures and thin films. *J Exp Biol* 214:3960–3967. <https://doi.org/10.1242/jeb.062620>
- Stoddard MC, Prum RO (2011) How colorful are birds? Evolution of the avian plumage color gamut. *Behav Ecol* 22:1042–1052. <https://doi.org/10.1093/beheco/arr088>
- Svensson E, Nilsson JA (1996) Mate quality affects offspring sex ratio in blue tits. *Proc R Soc Lond B* 263:357–361. <https://www.jstor.org/stable/50621>
- Székely T, Thomas GH, Cuthill IC (2006) Sexual conflict, ecology, and breeding systems in shorebirds. *Bioscience* 56:801–808. [https://doi.org/10.1641/0006-3568\(2006\)56\[801:SCEABS\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2006)56[801:SCEABS]2.0.CO;2)
- Trivers RL, Willard DE (1973) Natural selection of parental ability to vary the sex ratio of offspring. *Science* 179:90–92. <https://doi.org/10.1126/science.179.4068.90>
- Turčoková L, Melišková M, Balážová M (2016) Nest site location and breeding success of Common kingfisher (*Alcedo atthis*) in the Danube river system. *Folia Oecol* 43:74–82
- Underwood BA (1984) Vitamin A in animal and human nutrition. In: Sporn MB, Roberts AB, Goodman DS (eds) *The retinoids*. Academic Press, New York, pp 281–392
- Wang N, Zhang ZW (2009) The novel primers for sex identification in the brown eared-pheasant and their application to other species. *Mol Ecol Resour* 9:186–188. <https://doi.org/10.1111/j.1755-0998.2008.02177.x>
- Wendeln H, Becker PH (1999) Effects of parental quality and effort on the reproduction of common terns. *J Anim Ecol* 68:205–214. <https://doi.org/10.1046/j.1365-2656.1999.00276.x>
- West SA, Sheldon BC (2002) Constraints in the evolution of sex ratio adjustment. *Science* 295:1685–1688. <https://doi.org/10.1126/science.1069043>
- White TE (2020) Structural colours reflect individual quality: a meta-analysis. *Biol Lett* 16:20200001. <https://doi.org/10.1098/rsbl.2020.0001>
- White AE, Cristol DA (2014) Plumage coloration in belted kingfishers (*Megaceryle alcyon*) at a mercury-contaminated river. *Waterbirds* 37(2):144–152. <https://doi.org/10.1675/063.037.0203>
- Woodall PF (2001) Family Alcedinidae (Kingfishers). In: del Hoyo J, Elliott A, Sargatal J (eds) *Handbook of the birds of the world*. Vol. 6: mousebirds to hornbills. Lynx Edicions, Barcelona

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