



Trait-dependent resemblance of the flowering phenology and floral morphology of the allopolyploid *Cardamine flexuosa* to those of the parental diploids in natural habitats

Reiko Akiyama¹ · Stefan Milosavljevic¹ · Matthias Leutenegger¹ · Rie Shimizu-Inatsugi¹

Received: 28 December 2018 / Accepted: 8 December 2019 / Published online: 10 January 2020
© The Author(s) 2020

Abstract

Allopolyploids possess complete sets of genomes derived from different parental species and exhibit a range of variation in various traits. Reproductive traits may play a key role in the reproductive isolation between allopolyploids and their parental species, thus affecting the thriving of allopolyploids. However, empirical data, especially in natural habitats, comparing reproductive trait variation between allopolyploids and their parental species remain rare. Here, we documented the flowering phenology and floral morphology of the allopolyploid wild plant *Cardamine flexuosa* and its diploid parents *C. amara* and *C. hirsuta* in their native range in Switzerland. The flowering of *C. flexuosa* started at an intermediate time compared with those of the parents and the flowering period of *C. flexuosa* overlapped with those of the parents. *Cardamine flexuosa* resembled *C. hirsuta* in the size of flowers and petals and the length/width ratio of petals, while it resembled *C. amara* in the length/width ratio of flowers. These results provide empirical evidence of the trait-dependent variation of allopolyploid phenotypes in natural habitats at the local scale. They also suggest that the variation in some reproductive traits in *C. flexuosa* is associated with self-fertilization. Therefore, it is helpful to consider the mating system in furthering the understanding of the processes that may have shaped trait variation in polyploids in nature.

Keywords *Cardamine* · Floral morphology · Flowering phenology · Polyploid · Reproductive traits · Trait variation

Introduction

Allopolyploids (organisms possessing complete sets of genomes derived from different parental species) exhibit large variation in traits (Ramsey and Ramsey 2014). In controlled conditions, the phenotypes of synthetic neo-allopolyploids and natural allopolyploids were either similar, intermediate, or transgressive compared with those of the parental species (Abbott and Lowe 2004; Alexander-Webber et al. 2016; Davis 1943; Schranz and Osborn 2004). Such findings do not necessarily translate to natural allopolyploids

in natural habitats, because their trait variation is likely influenced by environmental conditions and potentially by adaptive evolution and genetic drift in the local habitats. The few studies that have focused on individuals in natural habitats report that the phenotypes of allopolyploids are intermediate or transgressive (Allen 2001; Vallejo-Marín 2012). However, trait variation in allopolyploids and their parental species in natural habitats remains largely unexplored (Ramsey and Ramsey 2014), partly because of the difficulty to identify the diploid progenitors of an allopolyploid (Buggs et al. 2014).

Trait variation in allopolyploids may be shaped by inheritance, polyploidization, and natural selection. The influence of inheritance and polyploidization should be prominent in allopolyploids shortly after emergence, while natural selection should affect trait variation over a longer time. Strong inheritance is observed in *Senecio cambliensis*, which is estimated to have emerged within the last 200 years (Brennan and Hiscock 2010). Allopolyploidization may confer large organ size and delayed flowering (Garbutt and Bazzaz 1983; Stebbins 1971; te Beest et al. 2012), although such effects may diminish after several generations (Gaeta et al. 2007).

Electronic supplementary material The online version of this article (<https://doi.org/10.1007/s10265-019-01164-0>) contains supplementary material, which is available to authorized users.

✉ Rie Shimizu-Inatsugi
rie.inatsugi@ieu.uzh.ch

¹ Department of Evolutionary Biology and Environmental Studies, University of Zurich, Winterthurerstrasse 190, 8057 Zurich, Switzerland

In allopolyploids with an old origin, on the other hand, trait variation is likely shaped by natural selection (Ramsey and Ramsey 2014), although it is impossible to determine the relative contributions of inheritance, allopolyploidization, and natural selection to the current trait variation in allopolyploids.

Variation in reproductive traits in polyploids may be shaped by their mating system. Many polyploids are self-fertilizing (Barringer 2007), which can contribute to the persistence of polyploids through reproductive assurance (Mable 2004; Miller and Venable 2000) and through reproductive isolation from the parental species because the success of reproduction of polyploids is independent from parents (Soltis et al. 2010 and references therein). Compared with outcrossing species, it is less important for self-fertilizing species to have large floral organs or synchronized flowering, which can be advantageous for attracting pollinators (e.g., Conner and Rush 1993; Elzinga et al. 2007; Forsyth 2003; Parachnowitsch and Kessler 2010). In fact, self-fertilizing species typically exhibit the selfing syndrome, in which the floral organs are smaller (Foxe et al. 2009; Shimizu and Tsuchimatsu 2015; Sicard and Lenhard 2011; Tedder et al. 2015a) compared with those of their outcrossing counterparts. Self-fertilizing species also exhibit an earlier onset of flowering on calendar days in the same season (Martin and Willis 2007; Mazer et al. 2004). Small floral organs and early onset of flowering is expected from self-fertilizing allopolyploids with old origin. Given the long amount of time since allopolyploidization, selfing-syndrome should have manifested stronger, whereas there may no longer be a large effect of allopolyploidization, which results in large organ size and delayed flowering. To examine whether self-fertilizing allopolyploids exhibit variation in reproductive traits in accordance with the selfing syndrome, it is essential to gather empirical data on allopolyploids and parental species with a known mating system and estimated time of allopolyploidization.

The genus *Cardamine* provides a convenient system for evolutionary ecological studies of allopolyploids. *Cardamine* is one of the largest genera in Brassicaceae and has experienced recurrent polyploid speciation, with more than half of the species estimated to be polyploid (Carlsen et al. 2009; Howard 1948; Hussein 1948; Kučera et al. 2005; Lihová and Marhold 2006; Lihová et al. 2006a, b; Mandáková et al. 2014; Zozomová-Lihová et al. 2014). The genus has been the subject of study from several aspects, such as genetics (Hay et al. 2014), genomics (Gan et al. 2016), ecological transcriptomics (Shimizu-Inatsugi et al. 2017), and morphology (e.g., Lihová et al. 2006b, 2007; Marhold 1992, 1998). While multiple species have been surveyed in *Cardamine*, there is little documentation of trait variation specifically in allopolyploids compared with their diploid parents in natural habitats.

We conducted a local-scale field observation to quantify the variation in floral morphology and flowering phenology of a *Cardamine* allopolyploid and its parents in their native habitats in Switzerland. Populations of the three species included in the study occurred sympatrically in the same climatic region within a distance of 12 km. *Cardamine flexuosa* is an allotetraploid originating from the diploids *C. amara* and *C. hirsuta*, presumably around 10^5 – 10^6 years ago (Mandáková et al. 2014). While *C. amara* is predominantly outcrossing (Tedder et al. 2015b) and is pollinated by insects such as beetles and flies (Kentaro K. Shimizu, Akiko Yasumoto, and Reiko Akiyama, personal observation), *C. hirsuta* and *C. flexuosa* are self-fertilizing (Hay et al. 2014; Reiko Akiyama, personal observation). The three species have different preferences to drought and submergence (Shimizu-Inatsugi et al. 2017). This seemingly reflects the environmental conditions of each species' habitat, with the diploid parents at either extreme and the allopolyploid in an intermediate and wide range (Akiyama et al. 2019). In this study, we addressed the following questions. (1) How do the reproductive traits of the allopolyploid compare with those of the parents in natural habitats? (2) Is the relationship in trait variation between the allopolyploid and its parents consistent for all reproductive traits, or does it vary among traits? If the latter is true, how does it vary?

Materials and methods

Study species

Cardamine amara L. (large bittercress) is a perennial herb that is widespread across central Europe and has served recurrently as a parent of polyploid species (Grime et al. 2007; Lihová et al. 2000, 2006b; Mandáková et al. 2014; Marhold 1998; Zozomová-Lihová et al. 2014). In Switzerland, the populations of *C. amara* in lowland areas (< 1,000 m) are reported to consist of diploids ($2n=2x=16$), and autotetraploid populations of the subsp. *austriaca* have been observed at higher altitudes in the inner Alpine valleys (Marhold 1995, 1999; Marhold et al. 2002). *Cardamine amara* propagates clonally and sexually (Tedder et al. 2015b). In addition, in many populations in Switzerland, the coexistence of female-sterile individuals with short pistils and hermaphrodite individuals with normal-length pistils has been reported (Tedder et al. 2015b), which is a rare reproductive system called androdioecy. *Cardamine amara* typically grows 10–60 cm in height and does not form a compact rosette (Lauber et al. 2012). The color of anthers is red for some individuals, but occurrences of yellow anthers have been observed (Reiko Akiyama, personal observation). The petal is white in most cases, though it can be purplish at

basal parts in some cases (Reiko Akiyama, personal observation; Lauber et al. 2012).

Cardamine hirsuta L. (hairy bittercress) is a diploid ($2n = 2x = 16$) annual herb native to Europe (Grime et al. 2007; Lihová et al. 2006b; Marhold 1995; Yatsu et al. 2003). It grows 5–30 cm in height and forms a rosette (Lauber et al. 2012). The species has yellow anthers and white petals (Reiko Akiyama, personal observation).

Cardamine flexuosa With. (wavy bittercress) is a tetraploid ($2n = 4x = 32$) annual or perennial herb native to Europe (Lihová et al. 2006b; Reiko Akiyama, personal observation) derived from *C. hirsuta* and *C. amara* (Grime et al. 2007; Mandáková et al. 2014). It is distinguished from the formerly named Asian *C. flexuosa*, which now belongs to a distinct octoploid species, *C. occulta* (Lihová et al. 2006b; Marhold et al. 2016). It grows up to 30 cm in height and forms a rosette (Post et al. 2011). The species has yellow anthers and white petals (Reiko Akiyama, personal observation).

Study sites

The study was conducted in three areas: Irchel (IR, N 47° 23', E 08° 33'; altitude, 484–505 m), Wehrenbach (WBH, N 47° 21', E 08° 33'; altitude, 422–429 m), and Küsnacht-Tobel (KT, N 47° 19', E 08° 38'; altitude, 615–664 m) in and around Zurich, Switzerland (Fig. S1a). These areas are located within 4–12 km of each other (Fig. S1a). Each area had two species of the three possible combinations of co-occurring species and all combinations were observed (Fig. S1b–d). For studying flowering phenology, we selected 18 sites (IR1–9, WBH1–3, and KT1–5, 7) in the three areas (Fig. S1b–d) in 2013. WBH1–3 are geographically close, but WBH 3 is separated from WBH 1–2 by a hedge and a paved road. In 2014, flowering phenology was recorded in the same sites as in 2013, with the exception of KT6, which was newly added, and IR9, which was excluded because of the disappearance of the site due to land use. The number of sites studied was 11 for *C. hirsuta*, eight for *C. flexuosa*, and six for *C. amara* in 2013 and 10 for *C. hirsuta*, eight for *C. flexuosa*, and five for *C. amara* in 2014 (Table S1). Table S1 summarizes the composition of the species and the number of individuals of each species per site. For floral morphology, two or three sites per species were subjected to the survey: IR2 and IR8 for *C. hirsuta*; IR2, KT5, and KT6 for *C. flexuosa*; and WBH1 and KT5 for *C. amara* (Fig. S1, Table S1).

When not self-evident, *C. flexuosa* and *C. hirsuta* were identified using morphological keys (Post et al. 2011). We also screened a total of 115 plants in the study sites regarding the ploidy level using flow cytometry (CyFlow® Space, Sysmex Europe GmbH, Norderstedt, Germany). For this analysis, the median of the sample size of each species per

site was three. As we found no triploids, the incidence of backcrossing seemed to be low, although gene flow between species cannot be completely excluded (Kolář et al. 2017; Petit et al. 1999).

Flowering phenology

To compare the onset of flowering in the allopolyploid *C. flexuosa* with that of the parents *C. amara* and *C. hirsuta*, we recorded the flowering status (whether the plant had an open flower(s) on the day of the census) during the season in 2013 and 2014 using different methods. In 2013, prior to the onset of flowering, we marked up to 55 plants per species per site from March to June by randomly selecting plants of a representative size across each site. Because the plants were originally marked for another purpose, the census intervals were irregular (intervals of 1–14 days) and, in most of the censuses, not all marked plants were scored. The total number of plants scored at each census ranged from 13 to 77 for *C. amara*, 37–106 for *C. flexuosa*, and 72–398 for *C. hirsuta* (Fig. S2). In 2014, we marked up to 48 plants per species per site by randomly selecting plants of a representative size across each site on March 4 and recorded the flowering status of these plants weekly from March 5 to June 26 at 6–8-day intervals. When marked plants were lost because of, for example, flooding or removal of the marking by disturbance during the period of study, they were excluded from the analyses. The total number of plants analyzed was 103 for *C. amara*, 136 for *C. flexuosa*, and 195 for *C. hirsuta* (Table S1). In addition, we recorded the flowering status of all plants per site on April 10 (Julian date 100) and May 21 (Julian date 141), to assess the representativeness of the data from the weekly census of a limited number of individuals. In total, we surveyed 4,275 *C. amara* individuals, 816 *C. flexuosa* individuals, and 1,480 *C. hirsuta* individuals on April 10; and 2,920 *C. amara* individuals, 418 *C. flexuosa* individuals, and 188 *C. hirsuta* individuals on May 21.

Floral morphology

To compare the floral morphology of the study species, we sampled and quantified the size of flowers and petals during the growth season in 2016. The date of survey ranged from April 5 to May 25, according to the flowering phenology of the species. For each site, we haphazardly sampled ~20 plants of a representative size for *C. hirsuta* and *C. flexuosa*, and the male and hermaphrodite forms of *C. amara* were distinguished based on pistil length. We collected one to four flowers per plant and photographed them from above at 7 cm (Fig. S3). If possible, we avoided the first and last three flowers and any flowers that seemed abnormal, to capture representative flowers of the plant. Once photographed, the flowers were placed on 0.5% agarose gel

to prevent wilting and transported to the laboratory at the University of Zurich, where they were dissected for petal specimens. The specimens were then scanned with a ruler using Epson Perfection® V600 Photo (Epson America Inc.) at 800 dpi (Fig. S3). Using the photographs and scans, the length and width of flowers and petals were measured as the longest axis of the flower seen from above and the direction perpendicular to it, respectively, using ImageJ 1.51a (Rasband 2016). We measured one to four flowers per plant and one representative petal per flower, to calculate the average trait value of the individual. The number of individuals for flower measurement was as follows: flower data, *C. amara* hermaphrodite, $N=31$; *C. amara* male, $N=28$; *C. flexuosa*, $N=55$; *C. hirsuta*, $N=38$. The number of individuals for petal measurement was as follows: *C. amara* hermaphrodite, $N=39$; *C. amara* male, $N=32$; *C. flexuosa*, $N=55$; *C. hirsuta*, $N=40$.

Statistical analysis

All analyses were conducted using R version 3.3.3 (R Core Team 2017). To examine whether flowering phenology differed between species, we conducted two types of tests: the Kolmogorov–Smirnov test and a linear model. A two-sample Kolmogorov–Smirnov test was conducted to compare the distribution of the proportion of the flowering plants between a pair of species each in 2013 and 2014 using the package *stats*, with 0.017 as a threshold P value to adjust for multiple tests (0.05 divided by three pairwise tests). In the linear model, the Julian date on which the first flower of the plant opened was regressed upon the species using the package *stats*. Only the data from 2014 were subjected to this analysis. Because of unequal number of individuals in different groups, the data were analyzed with Type III ANOVA using the package *car* (Fox and Weisberg 2011). We conducted a post hoc test with Tukey's HSD adjustment using the package *multcomp* (Hothorn et al. 2008).

To examine the variation of floral morphology among species and sex, we ran linear models with four groups of species using sex combination (*C. amara* hermaphrodite, *C. amara* male, *C. flexuosa* hermaphrodite, and *C. hirsuta* hermaphrodite) as an explanatory variable and the mean flower length, mean flower width, length/width ratio of a flower, mean petal length, mean petal width, and mean length/width ratio of a petal as response variables. The means were calculated as the average among individuals. In the linear models, the response variables were log transformed prior to the analysis, to meet the assumption of normality. As the data consisted of unequal number of individuals in different groups, we used Type III ANOVA with the package *car* (Fox and Weisberg 2011). When significant difference was detected, post hoc tests were

conducted using Tukey's HSD adjustment with the package *multcomp* (Hothorn et al. 2008).

Results

Flowering phenology

In the two seasons, the flowering phenology of *C. flexuosa* was intermediate compared with *C. amara* and *C. hirsuta*. In 2014, the peak of flowering occurred first in *C. hirsuta*, followed by *C. flexuosa* and *C. amara* (Fig. 1). The onset of flowering varied significantly among species (Type III ANOVA, $F_{2,303}=207.75$, $P<0.0001$), with *C. hirsuta* flowering significantly earlier than *C. flexuosa* (Tukey's HSD test, $P<0.0001$), which started flowering significantly earlier than *C. amara* (Tukey's HSD test, $P<0.0001$). The distribution of the flowered plants did not differ between species (Kolmogorov–Smirnov test, *C. amara* vs *C. flexuosa*: $Z=0.33$, $P=0.27$; *C. flexuosa* vs *C. hirsuta*: $Z=0.17$, $P=0.96$; and *C. hirsuta* vs *C. amara*: $Z=0.28$, $P=0.49$). The census on all plants showed that, on Julian date 100 (April 10), the proportion of plants with open flowers was highest for *C. hirsuta*, followed by *C. flexuosa* and *C. amara*, and that this order was reversed on Julian date 141 (May 21) (Fig. 1). This trend was consistent with that of marked plants on Julian dates 100 and 142 (April 10 and May 22). Thus, the marked plants captured a representative phenology of the study species. Similar to that observed for 2014, in 2013 the peak of flowering was observed first in *C. hirsuta*, followed by *C. flexuosa* and *C. amara* (Fig. S2) and there was no between-species difference in the distribution of the flowered plants (Kolmogorov–Smirnov test, *C. amara* vs *C. flexuosa*: $Z=0.41$, $P=0.36$; *C. flexuosa* vs *C. hirsuta*: $Z=0.27$, $P=0.98$; and *C. hirsuta* vs *C. amara*: $Z=0.55$, $P=0.26$).

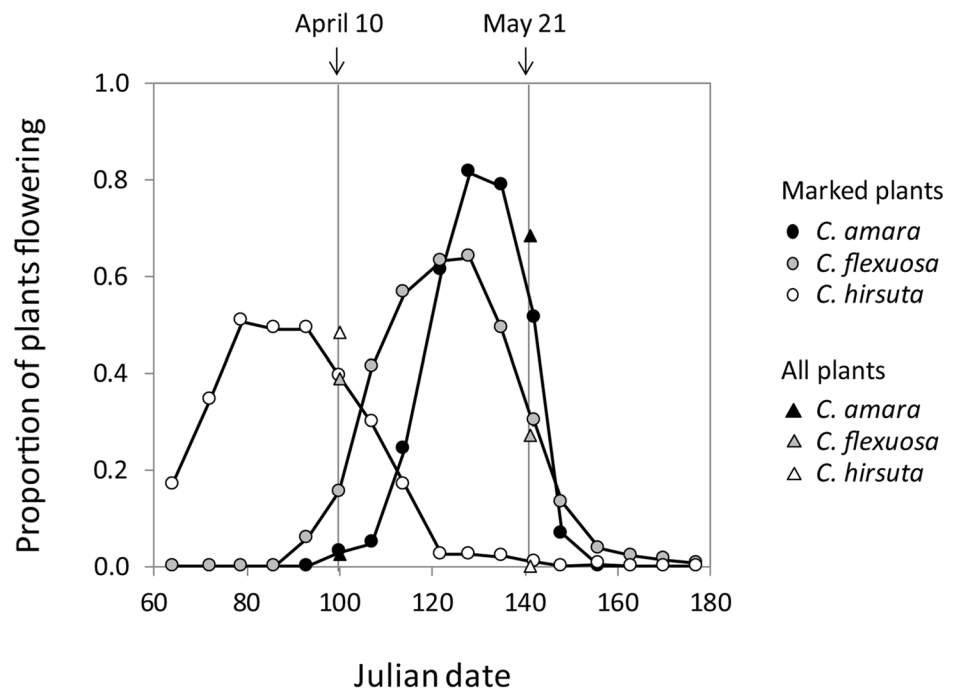
In sites that were cohabited by two species, *C. hirsuta* flowered earlier than *C. flexuosa* and *C. flexuosa* flowered earlier than *C. amara* (Fig. S4), indicating that the difference detected using species-level comparisons (Fig. 1) reflected differences between species, rather than between sites, as far as the study sites are concerned.

In *C. amara*, the period of flowering was similar among sites, with a different composition of males and hermaphrodites (Fig. S4): almost all individuals (>95%) in WBH1 were male, whereas males and hermaphrodites were mixed at a higher ratio in the other populations (Bachmann et al., unpublished).

Floral morphology

The size of flowers and petals varied among species and between sexes (Type III ANOVA, length: $F_{3,159}=706.13$, $P<0.0001$; width: $F_{3,159}=777.68$, $P<0.0001$). The length

Fig. 1 Proportion of flowering individuals of *Cardamine amara*, *C. flexuosa*, and *C. hirsuta* in all study sites in Switzerland in 2014. The circles represent weekly census on marked individuals (*C. amara*, $N=113$; *C. flexuosa*, $N=136$; *C. hirsuta*, $N=195$), while triangles represent census on all individuals at the study sites on April 10 (*C. amara*, $N=4,275$; *C. flexuosa*, $N=812$; *C. hirsuta*, $N=8,140$) and May 21 (*C. amara*, $N=2,920$; *C. flexuosa*, $N=418$; *C. hirsuta*, $N=188$) (Table S1)



and width of flowers were largest in the hermaphrodite *C. amara*, followed by the male *C. amara*, *C. flexuosa*, and *C. hirsuta* (Tukey's HSD tests, Fig. 2a, c). The hermaphrodite and male flowers of *C. amara* were approximately twice as large as the flowers of *C. flexuosa* and *C. hirsuta* (Fig. 2a, c). The shape of flowers also varied among species and sexes (Type III ANOVA, $F_{3,159} = 116.12$, $P < 0.0001$). The flowers of *C. amara* and *C. flexuosa* exhibited a shape that was close to a square, while the flowers of *C. hirsuta* were more rectangular (Tukey's HSD tests, Fig. 2e, g).

The petal size varied among species (Type III ANOVA, length: $F_{3,162} = 930.48$, $P < 0.0001$; width: $F_{3,162} = 1204.1$, $P < 0.0001$). The length and width of petals were largest in *C. amara*, followed by *C. flexuosa* and *C. hirsuta* (Tukey's HSD tests, Fig. 2b, d). The petal shape of *C. amara* was different from the petal shapes of *C. flexuosa* and *C. hirsuta* (Type III ANOVA, $F_{3,162} = 160.03$, $P < 0.0001$), as *C. amara* petals were proportionally wider compared with the other two species (Tukey's HSD test, Fig. 2f, h). The hermaphrodites and males of *C. amara* did not differ in petal size (Fig. 2b, d), indicating that the difference in flower size observed between sexes (Fig. 2a, c) can be attributed to the extent of petal opening.

The examination of the sites individually, especially the ones in which *C. flexuosa* cohabited with either of the parents, revealed that the difference, or lack thereof, between species regarding flower and petal morphologies persisted (Figs. S5, S6). These results indicate that the differences detected among species (Fig. 2) reflected differences between species, rather than between sites, at least for the sites included in this study.

Discussion

The results of the present study provide empirical documentation of trait variation in an allopolyploid and its parents in natural habitats at the local scale. Overall, *C. flexuosa* resembled either of the parents, but there was between-trait difference regarding which parent *C. flexuosa* resembled more. The peak flowering time was closer to that of *C. amara*, while the duration of flowering and the size of flowers and petals were closer to those of *C. hirsuta* (Figs. 1, 2). This trait-dependent divergence of the allopolyploid from the parental species is in accordance with the findings obtained by a study of *Nicotiana* in the laboratory (McCarthy et al. 2016), in which multiple species collected worldwide were grown in a uniform condition. Therefore, the reproductive traits of an allopolyploid and its diploid parents can diverge in both controlled and natural conditions. No environment was controlled in the present study; however, because it was conducted at a local scale in the same climatic region, the evolutionary significance of the divergence observed between the allopolyploid and its diploid parents is likely attributed to the difference in the mating system, genetics, natural selection, or microhabitat rather than to environmental differences between sites. In the following sections, we discuss the evolutionary and ecological implications of variations in the traits studied.

We recorded the flowering phenology of an allopolyploid in two consecutive years in natural habitats. *Cardamine flexuosa* consistently flowered at an intermediate time, showing a similar trend to the flowering of allopolyploids observed under controlled conditions (Anssour et al. 2009;

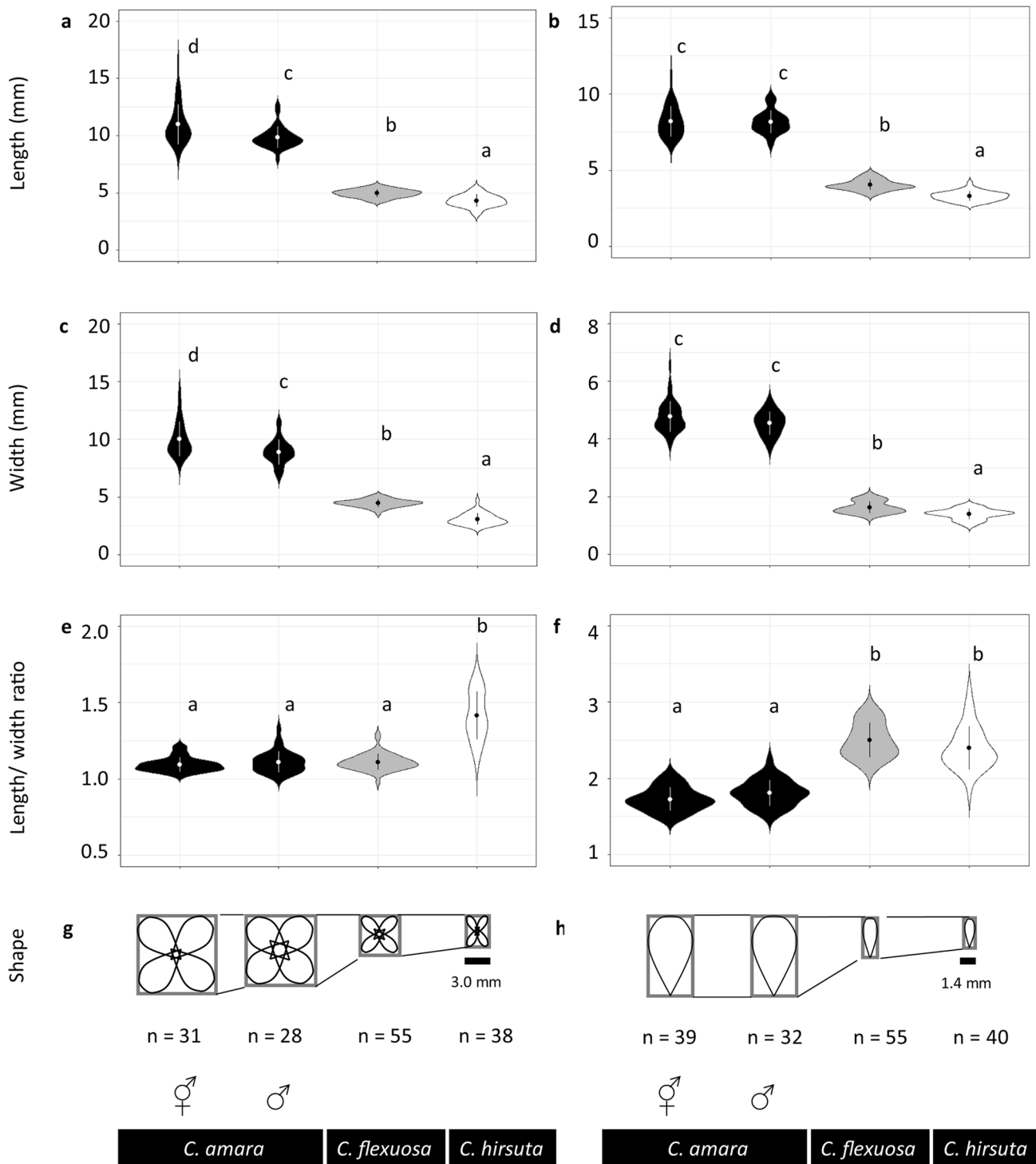


Fig. 2 Dimension, length/width ratio of flowers and petals, and schematic drawings of flower and petal morphology of *Cardamine amara* (hermaphrodite and male), *C. flexuosa*, and *C. hirsuta*. **a** Flower length, **b** petal length, **c** flower width, **d** petal width, **e** length/width ratio of the flowers, **f** length/width ratio of the petals, **g** schematic drawing of flower shape based on the mean length and width of the

flowers, and **h** schematic drawing of petal shape based on the mean length and width of the petals. For **a–f**, the dots and vertical lines within violin plots indicate the mean and SD, and the lowercase letters in each figure indicate statistical differences based on Tukey's HSD test

Davis 1943; Schranz and Osborn 2004). In both years, *C. hirsuta* was the first to flower, followed by *C. flexuosa* and *C. amara*. This is similar to previous findings from *Mimulus* and *Clarkia*, in which self-fertilizing species flowered earlier compared with the outcrossing counterparts (Martin and Willis 2007; Mazer et al. 2004). Although this result was not statistically significant, the peak of flowering was narrower in the outcrossing species *C. amara* than it was in the other two selfing species in the present study. This trend persisted when the species were resolved into sites (Fig. S4) and corresponded to the known difference between outcrossing and selfing species; that is, while pollinators impose natural selection on synchronized flowering in outcrossing species (Forsyth 2003), selfing species are expected to be free from such natural selection. The large overlap between the flowering period of *C. flexuosa* and those of the parents (Fig. 1) should not hinder the reproductive isolation of *C. flexuosa*, given that it is self-fertilizing. Consistent with this interpretation, no triploids were observed in the cohabited study sites (see “Materials and methods”).

In addition to the mating system and genetic differences between species, the fine-scale environment may have affected the flowering phenology of the studied individuals. Early flowering is often associated with dry, bright, and nutrient-rich environments (Martin and Willis 2007; Mazer et al. 2004; Nord and Lynch 2009; Stanton et al. 2000; but see Callahan and Pigliucci 2002 for early onset of reproduction in shade). In fact, the habitat in the study area of the early-flowering parent *C. hirsuta* was characterized by the presence of less water, more nitrogen, and more light, while the opposite was observed for the late-flowering parent *C. amara*; the environmental condition of the *C. flexuosa* habitat was intermediate (Akiyama et al. 2019). Thus, the allopolyploid *C. flexuosa* was intermediate in both flowering phenology and habitat at the fine geographical scale.

The size of the floral organs of *C. flexuosa* was closer to that of the self-fertilizing parent *C. hirsuta* than to that of the outcrossing parent *C. amara* (Fig. 2). This result corresponds to the evolution of the selfing syndrome; selfing species have smaller flowers than the outcrossing counterparts, as a consequence of natural selection on floral morphology (Foxe et al. 2009; Shimizu and Tsuchimatsu 2015; Sicard and Lenhard 2011; Tedder et al. 2015a). Conversely, the findings of previous studies of recently emerged allopolyploids in natural habitats suggest that inheritance and/or allopolyploidization influence floral organ size. The floral organs of *Mimulus peregrinus* were larger than those of the parental diploids (Vallejo-Marín 2012). The discrepancy between this result and findings of the present study may be attributed to the time since allopolyploidization. Having emerged 10^5 – 10^6 years ago, *C. flexuosa* may exhibit a number of generations that is sufficient for the manifestation of the selfing syndrome. Whether the floral organ size of

self-fertilizing allopolyploids is negatively correlated with time since allopolyploidization can be examined by a comparative field study of multiple taxa featuring different mating systems, different time since allopolyploidization, and different locations (laboratory and natural habitat). Such a study should provide a perspective on the interpretation of previous studies performed in the field or laboratory that reported that the floral organs of allopolyploids are larger than, not different from, or smaller than those of the parents (Abbott and Lowe 2004; Alexander-Webber et al. 2016; Allen 2001; Anssour et al. 2009; Benedict et al. 2012; McCarthy et al. 2016; Perný et al. 2005; Vallejo-Marín 2012; Table 2 in Vamasi et al. 2007).

In summary, the present study provides empirical quantitative data on the reproductive traits of a wild allopolyploid and its parental species in natural habitats. At the local scale, we examined the variation in floral morphology and flowering phenology in *Cardamine*. The allopolyploid exhibited a trait-dependent resemblance to the parental species. We also discussed a scenario in which the small floral organs of the self-fertilizing allopolyploid *C. flexuosa* are a product of the selfing syndrome. Such an evolutionary scenario, as well as the underlying molecular mechanisms, can be examined effectively in *Cardamine*. The genus has the advantages of consisting of different ploidy levels, of having a known time since polyploidization and mating systems for multiple species, and of the availability of rich genomic and genetic resources. Our understanding of trait variation in allopolyploids may be advanced by considering not only the inheritance and allopolyploidization, but also the mating system of these species.

Acknowledgements We thank Peter Enz, Bernhard Hirzel, Daniel Schlagenhauf, and Nadine Kofmehl at the Botanical Garden of the University of Zurich, Jiro Sugisaka at Kyoto University, Diana Shneebeli, Samuel Moor, Sofia van Moorsel at the University of Zurich, and Masaomi Hatakeyama, Matthias Helling, Lucas Mohn, Aki Morishima, Hiroyuki Kakui, Kentaro K. Shimizu, Reinhold Stockenhuber, Alejandro Morales Suarez, Andrew Tedder, Misako Yamazaki, Nicole Zweifel, and the other members of the Shimizu Laboratory at the University of Zurich for their assistance at various phases of the study. We also thank the Botanical Garden of the University of Zurich; private landowners at the Irchel Campus; the City of Zurich, and Gemeinde Küsnacht-Tobel for permits for field studies; and Swisstopo for geographical data. We highly appreciate comments from two anonymous reviewers and linguistic advice from Gwyneth Halstead-Nussloch.

Funding This study was funded by grants from the University Research Priority Program (URPP) Evolution in Action from the University of Zurich to Reiko Akiyama and Rie Shimizu-Inatsugi and by The Japan Science and Technology Agency, Core Research for Evolutionary Science and Technology Grant Number JPMJCR16O3.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Abbott RJ, Lowe AJ (2004) Origins, establishment and evolution of new polyploidy species: *Senecio cambrensis* and *S. eboracensis* in the British Isles. *Biol J Linnean Soc* 82:467–474
- Akiyama R, Sun J, Hatakeyama M, Lischer HEL, Briskine RV, Hay A, Gan X, Tsiantis M, Kudoh H, Kanaoka MM, Sese J, Shimizu KK, Shimizu-Inatsugi R (2019) Fine-scale ecological and transcriptomic data reveal niche differentiation of an allopolyploid from diploid parents in *Cardamine*. *bioRxiv*. <https://doi.org/10.1101/600783>
- Alexander-Webber D, Abbott RJ, Chapman MA (2016) Morphological convergence between an allopolyploid and one of its parental species correlates with biased gene expression and DNA loss. *J Hered* 107:445–454
- Allen GA (2001) Hybrid speciation in *Erythronium* (Liliaceae): a new allotetraploid species from Washington State. *Syst Bot* 26:263–272
- Anssour S, Krügel T, Sharbel TF, Saluz HP, Bonaventure G, Baldwin IT (2009) Phenotypic, genetic and genomic consequences of natural and synthetic polyploidization of *Nicotiana attenuata* and *Nicotiana obtusifolia*. *Ann Bot* 103:1207–1217
- Barringer BC (2007) Polyploidy and self-fertilization in flowering plants. *Am J Bot* 94:1527–1533
- Benedict BG, Modliszewski JL, Seiart AL, Martin NH, Ganders FR, Willis JH (2012) *Mimulus sookensis* (Phrymaceae), a new allotetraploid species derived from *Mimulus guttatus* and *Mimulus nasutus*. *Madroño* 59:29–43
- Brennan AC, Hiscock SJ (2010) Expression and inheritance of sporophytic self-incompatibility in synthetic allohexaploid *Senecio cambrensis* (Asteraceae). *New Phytol* 186:251–261
- Buggs JA, Wendel JF, Doyle JJ, Soltis DE, Soltis PS, Coate JE (2014) The legacy of diploid progenitors in allopolyploid gene expression patterns. *Philos Trans R Soc B* 369:20130354
- Callahan HS, Pigliucci M (2002) Shade-induced plasticity and its ecological significance in wild populations of *Arabidopsis thaliana*. *Ecology* 83:1965–1980
- Carlsen T, Bleeker W, Hurka H, Elven R, Brochmann C (2009) Biogeography and phylogeny of *Cardamine* (Brassicaceae). *Ann Missouri Bot Garden* 96:215–236
- Conner JK, Rush S (1993) Effects of flower size and number on pollinator visitation to wild radish, *Raphanus raphanistrum*. *Oecologia* 105:509–516
- Davis BM (1943) An amphidiploid in the F1 generation from the cross *Oenothera franciscana* x *Oenothera biennis*, and its progeny. *Genetics* 28:275–285
- Elzinga JA, Atlan A, Biere A, Gigord L, Weis AE, Bernasconi G (2007) Time after time: flowering phenology and biotic interactions. *Trends Ecol Evolut* 22:432–439
- Forsyth SA (2003) Density-dependent seed set in the Haleakala silversword: evidence for an Allee effect. *Oecologia* 136:551–557
- Fox J, Weisberg S (2011) An R companion to applied regression. Sage, Thousand Oaks. <https://socserv.socsci.mcmaster.ca/jfox/Books/Companion>. Accessed 12 Dec 2018
- Foxe JP, Slotte T, Stahl EA, Neuffer B, Hurka H, Wright SI (2009) Recent speciation associated with the evolution of selfing in *Capsella*. *Proc Natl Acad Sci USA* 106:5241–5245
- Gaeta RT, Pires JC, Iniguez-Luy F, Leon E, Osborn TC (2007) Genomic changes in resynthesized *Brassica napus* and their effect on gene expression and phenotype. *Plant Cell* 19:3403–3417
- Gan X, Hay A, Kwantes M, Haberer G, Hallab A, Ioio RD, Hofhuis H, Pieper B, Cartolano M, Neumann U, Nikolov LA, Song B, Hajheidari M, Briskine R, Kougioumoutzi E, Vlad D, Broholm S, Hein J, Meksem K, Lightfoot D, Shimizu KK, Shimizu-Inatsugi R, Imprialou M, Kudrna D, Wing R, Sato S, Huijser P, Filatov D, Mayer KFX, Mott R, Tsiantis M (2016) The *Cardamine hirsuta* genome offers insight into the evolution of morphological diversity. *Nat Plants* 2:16167
- Garbutt K, Bazzaz FA (1983) Leaf demography, flower production and biomass of diploid and tetraploid populations of *Phlox drummondii* Hook. on a soil moisture gradient. *New Phytol* 93:129–141
- Grime JP, Hodgson JG, Hunt R (2007) Comparative plant ecology: a functional approach to common British species. Castlepoint Press, Colvend
- Hay AS, Pieper B, Cooke E, Mandáková T, Cartolano M, Tattersall AD, Ioio RD, McGowan SJ, Barkoulas M, Galinha C, Rast MI, Hofhuis H, Then C, Plieske J, Ganal M, Mott R, Martinez-Garcia JF, Carine MA, Scotland RW, Gan X, Filatov DA, Lysak MA, Tsiantis M (2014) *Cardamine hirsuta*: a versatile genetic system for comparative studies. *Plant J* 78:1–15
- Hothorn T, Bretz F, Westfall P (2008) Simultaneous inference in general parametric models. *Biom J* 50:346–363
- Howard HW (1948) Chromosome number of *Cardamine pratensis*. *Nature* 161:277
- Hussein F (1948) Chromosome number of *Cardamine pratensis*. *Nature* 161:1015
- Kolář F, Čertner M, Suda J, Šchönswetter P, Husband BC (2017) Mixed-ploidy species: progress and opportunities in polyploid research. *Trends Plant Sci* 22:1041–1055
- Kučera J, Valco I, Marhold K (2005) On-line database of the chromosome numbers of the genus *Cardamine* (Brassicaceae). *Biologia* 60:473–476
- Lauber K, Wagner G, Gygax A (2012) Flora helvetica. Haupt Verlag AG, Berne
- Lihová J, Marhold K (2006) Phylogenetic and diversity patterns in *Cardamine* (Brassicaceae)—a genus with conspicuous polyploid and reticulate evolution. In: Sharma AK, Sharma A (eds) Plant genome biodiversity and evolution. Part C Phanerogams, vol 1. Science Publishers Inc., Enfield, pp 149–186
- Lihová J, Marhold K, Neuffer B (2000) Taxonomy of *Cardamine amara* (Cruciferae) in the Iberian Peninsula. *Taxon* 49:747–763
- Lihová J, Shimizu KK, Marhold K (2006a) Allopolyploid origin of *Cardamine asarifolia* (Brassicaceae): incongruence between plastid and nuclear ribosomal DNA sequences solved by a single-copy nuclear gene. *Mol Phylogenet Evol* 39:759–786
- Lihová J, Marhold K, Kudoh H, Koch MA (2006b) Worldwide phylogeny and biogeography of *Cardamine flexuosa* (Brassicaceae) and its relatives. *Am J Bot* 93:1206–1221
- Lihová J, Kučera J, Perný M, Marhold K (2007) Hybridization between two polyploid *Cardamine* (Brassicaceae) species in North-western Spain: discordance between morphological and genetic variation patterns. *Ann Bot* 99:1083–1096
- Mable BK (2004) Polyploidy and self-compatibility: is there an association? *New Phytol* 162:803–811

- Mandáková T, Marhold K, Lysak MA (2014) The widespread crucifer species *Cardamine flexuosa* is an allotetraploid with a conserved subgenomic structure. *New Phytol* 201:982–992
- Marhold K (1992) A multivariate morphometric study of the *Cardamine amara* group (Cruciferae) in the Carpathian and Sudeten mountains. *Biol J Linn Soc* 110:121–135
- Marhold K (1995) Taxonomy of the genus *Cardamine* L. (Cruciferae) in the Carpathians and in Pannonia. III. *Folia Geobot Phytotaxon* 30:397–434
- Marhold K (1998) Morphometric comparison of diploid populations of *Cardamine amara* (Brassicaceae) from Central Europe and the Balkan Peninsula. *Thaiszia J Bot* 8:19–32
- Marhold K (1999) Taxonomic evaluation of the tetraploid populations of *Cardamine amara* (Brassicaceae) from the Eastern Alps and adjacent areas. *Bot Helv* 109:67–84
- Marhold K, Huthmann M, Hurka H (2002) Evolutionary history of the polyploid complex of *Cardamine amara* (Brassicaceae): isozyme evidence. *Plant Syst Evol* 233:15–28
- Marhold K, Šlenker M, Kudoh H, Zozomova-Lihová J (2016) *Cardamine occulta*, the correct species name for invasive Asian plants previously classified as *C. flexuosa*, and its occurrence in Europe. *PhytoKeys* 62:57–72
- Martin NH, Willis JH (2007) Ecological divergence associated with mating system causes nearly complete reproductive isolation between sympatric *Mimulus* species. *Evolution* 61:68–82
- Mazer J, Paz H, Bell MD (2004) Life history, floral development, and mating system in *Clarkia xantiana* (Onagraceae): do floral and whole-plant rates of development evolve independently? *Am J Bot* 91:2041–2050
- McCarthy EW, Chase MW, Knapp S, Litt A, Leitch AR, Le Comber SC (2016) Transgressive phenotypes and generalist pollination in the floral evolution of *Nicotiana* polyploids. *Nat Plants* 2:16119
- Miller JS, Venable DL (2000) Polyploidy and the evolution of gender dimorphism in plants. *Science* 289:2335–2338
- Nord EA, Lynch JP (2009) Plant phenology: a critical controller of soil resource acquisition. *J Exp Bot* 60:1927–1937
- Parachnowitsch AL, Kessler A (2010) Pollinators exert natural selection on flower size and floral display in *Penstemon digitalis*. *New Phytol* 188:393–402
- Perný M, Tribsch A, Stuessy TF, Marhold K (2005) Allopolyploid origin of *Cardamine silana* (Brassicaceae) from Calabria (southern Italy): karyological, morphological and molecular evidence. *Bot J Linn Soc* 148:101–116
- Petit C, Bretagnolle F, Felber F (1999) Evolutionary consequences of diploid–polyploid hybrid zones in wild species. *Trends Ecol Evol* 14:306–311
- Post AR, Krings A, Xiang J, Sosinski BR, Neal JC (2011) On the identity of the weedy bittercresses (*Cardamine*: Brassicaceae) in United States nurseries: evidence from molecules and morphology. *Weed Sci* 59:123–135
- Ramsey J, Ramsey TS (2014) Ecological studies of polyploidy in the 100 years following its discovery. *Philos Trans R Soc B* 369:20130352
- Rasband WS (2016) ImageJ. U.S. National Institutes of Health, Bethesda. <https://imagej.nih.gov/ij/>. Accessed 1 Apr 2016
- R Core Team (2017) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna. <https://www.r-project.org>. Accessed 3 May 2018
- Schranz ME, Osborn TC (2004) *De novo* variation in life-history traits and responses to growth conditions of resynthesized polyploidy *Brassica napus* (Brassicaceae). *Am J Bot* 91:174–183
- Shimizu KK, Tsuchimatsu T (2015) Evolution of selfing: recurrent patterns in molecular adaptation. *Annu Rev Ecol Evol Syst* 46:593–622
- Shimizu-Inatsugi R, Terada A, Hirose K, Kudoh H, Sese J, Shimizu KK (2017) Plant adaptive radiation mediated by polyploid plasticity in transcriptomes. *Mol Ecol* 26:193–207
- Sicard A, Lenhard M (2011) The selfing syndrome: a model for studying the genetic and evolutionary basis of morphological adaptation in plants. *Ann Bot* 107:1433–1443
- Soltis DE, Buggs RJA, Doyle JJ, Soltis PS (2010) What we still don't know about polyploidy. *Taxon* 59:1387–1403
- Stanton ML, Roy BA, Thiede DA (2000) Evolution in stressful environments I Phenotypic variability, phenotypic selection, and response to selection in five distinct environmental stresses. *Evolution* 54:93–111
- Stebbins GL (1971) Chromosomal evolution in higher plants. Edward Arnold, London
- te Beest M, Le Roux JJ, Richardson DM, Brysting AK, Suda J, Kubešová PP (2012) The more the better? The role of polyploidy in facilitating plant invasions. *Ann Bot* 109:19–45
- Tedder A, Carleial S, Gołębiewska M, Kappel C, Shimizu KK, Stift M (2015a) Evolution of the selfing syndrome in *Arabis alpina* (Brassicaceae). *PLoS ONE* 10:e0126618
- Tedder A, Helling M, Pannell JR, Shimizu-Inatsugi R, Kawagoe T, van Campen J, Sese J, Shimizu KK (2015b) Female sterility associated with increased clonal propagation suggests a unique combination of androdioecy and asexual reproduction in populations of *Cardamine amara* (Brassicaceae). *Ann Bot* 115:763–776
- Vallejo-Marín M (2012) *Mimulus peregrinus* (Phrymaceae): a new British allopolyploid species. *PhytoKeys* 14:1–14
- Vamosi JC, Goring SJ, Kennedy BF, Mayberry RJ, Moray CM, Neame LA, Tunbridge ND, Elle E (2007) Pollination, floral display, and the ecological correlates of polyploidy. *Funct Ecosyst Commun* 1:1–9
- Yatsu Y, Kachi N, Kudoh H (2003) Ecological distribution and phenology of an invasive species, *Cardamine hirsuta* L., and its native counterpart, *Cardamine flexuosa* With., in central Japan. *Plant Species Biol* 18:35–42
- Zozomová-Lihová J, Krak K, Mandáková T, Shimizu KK, Španiel S, Vít P, Lysak MA (2014) Multiple hybridization events in *Cardamine* (Brassicaceae) during the last 150 years: revisiting a textbook example of neoallopolyploidy. *Ann Bot* 113:817–830

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.