




Rate accelerations in nuclear 18S rDNA of mycoheterotrophic and parasitic angiosperms

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Abstract Rate variation in genes from all three genomes providing evidence that the efficiency of the translational apparatus in non-photosynthetic plants has not been affected. While the affected loss of photosynthetic ability leads to a relaxation of evolutionary constraints in genes involved in the photosynthetic apparatus, it remains to be determined how prevalent increased substitution rates are in nuclear DNA of non-photosynthetic angiosperms. In this study we infer rates of molecular evolution of 18S rDNA of all parasitic and mycoheterotrophic plant families (except Lauraceae and Polygalaceae) using relative rate tests. In several holoparasitic and mycoheterotrophic plant lineages extremely high substitution rates are observed compared to other photosynthetic angiosperms. The position and frequency of these substitutions have been identified to understand the mutation dynamics of 18S rRNA in achlorophyllous plants. Despite the presence of significantly elevated substitution rates, very few mutations occur in major functional and structural regions of the small ribosomal molecule.

Keywords 18S rDNA Mycoheterotrophy Parasitism Substitution rates Relative rate test

Introduction Flowering plants, a fully heterotrophic mode of life is an exceptional trait. Little over 1% of all described species derive all of their carbon from other organisms (Kuhlenbaker and Jørgensen 2008). Based on the partners involved in the interaction, two groups of heterotrophic plants can be distinguished: parasitic and mycoheterotrophic plants. Parasitic plants directly penetrate host plants via their haustoria to obtain water and (in)organic solutes (Nickrent et al. 1998). These plants include hemiparasites and holoparasites, a division based on the presence or absence of chlorophyll during at least one part of their life cycle, respectively. Parasitic plants are mainly restricted to eudicots, with the exception of three magnoliid genera *Hydnora* (Hydnoraceae) and *Cassytha* (Lauraceae). A parasitic lifestyle has evolved at least 11 times independently in angiosperms, occurring in 20 families and close to 4,500 extant species (Nickrent et al. 1998; Barkman et al. 2007; Heide-Jørgensen 2008). The phylogenetic positions of many parasitic lineages are still unknown at low taxonomic level (APG 2003).

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These plants include hemiparasites and holoparasites, a division based on the presence or absence of chlorophyll during at least one part of their life cycle, respectively. Parasitic plants are mainly restricted to eudicots, with the exception of three magnoliid genera *Hydnora* (Hydnoraceae) and *Cassytha* (Lauraceae). A parasitic lifestyle has evolved at least 11 times independently in angiosperms, occurring in 20 families and close to 4,500 extant species (Nickrent et al. 1998; Barkman et al. 2007; Heide-Jørgensen 2008). The phylogenetic positions of many parasitic lineages are still unknown at low taxonomic level (APG 2003).

In contrast to parasitic plants, mycoheterotrophic plants derive carbon from fungi (Leake 2005). With a few exceptions, most mycoheterotrophic plant species exploit mycorrhizal fungi that are simultaneously mycorrhizal with neighboring photosynthetic plants. Because all carbon in

this tripartite relationship is ultimately derived from auto-trophic plants, mycoheterotrophic plants are considered to be related to the loss of photosynthesis, effective population size, generation time and host specificity have been proposed to explain this phenomenon in parasitic plants. However, none of these hypotheses could unequivocally explain the rate increases in all parasitic plants (dePamphilis 2009; Selosse et al. 2010). Over 400 fully mycoheterotrophic plant species have been reported in eudicots (Ericaceae and Gentianaceae and Polygonaceae) and monocots (Burmanniaceae, Corsiaceae, Iridaceae, Orchidaceae, Petrosaviaceae, Thismiaceae and Triuridaceae) (Leakey et al. 1994; Heide-Jørgensen 2008; Merckx et al. 2009). In addition, over 20,000 flowering plant species are thought to be at least partially mycoheterotrophic, mostly initial mycoheterotrophs in the autotrophic lineages with relative rate tests (Robinson-Orchidaceae (Leakey 2005; Selosse and Roy 2009).

With the loss of photosynthetic ability, genes required for the photosynthetic apparatus will undergo random mutation under relaxed natural selection. *Cynopholis americana* (Wimpee et al. 1991; Cuscuta Funk et al. 2007; McNeal et al. 2009; Revill et al. 2005; Epifagus virginianus dePamphilis and Palmer 1990; Wolfe et al. 1992; dePamphilis et al. 1997; Lathraea clandestina Delavaut et al. 1995; Orobanchaceae hederacea Thalouarn et al. 1994). Although some chloroplast genes are retained and functional in several holoparasitic plants (Bungard 2004), many holoparasites have a reduced plastid genome due to excessive gene loss and increased substitution rates in the remaining genes (Wolfe et al. 1992). In chloroplast genome analysis of non-photosynthetic plants, different explanations have been proposed why a reduced set of genes have been retained and need to be translated within the chloroplast as opposed to replacement by a product from cytosolic orthologues: (1) import of proteins with hydrophobic membranes from the cytosol back into the organelle would be impossible; (2) rate of synthesis of specific proteins can be regulated within a individual plastids preventing deadly side-effects of oxidative stress or lethal effects of accumulating toxic products; (3) synthesis and assembly of components of the photosynthetic complexes are tightly regulated within plastids (see review by Barbrook et al. 2006). The plastid genome of fully mycoheterotrophic plants remains largely unstudied, but data suggest that the genome is prone to similar relaxation of purifying selection (Caddick et al. 2002; Barrett and Freudenstein 2008). These observations demonstrate that non-photosynthetic plants are under reduced selective constraints, which affect the structure and function of genes involved in photosynthetic reactions.

Somewhat surprisingly, extreme variation in rates of evolution has also been observed in the nuclear and mitochondrial genes of some fully mycoheterotrophic plants (Merckx et al. 2006, 2009; Petersen et al. 2006) and parasitic plants (Nickrent and Starr 1994; Nickrent et al. 1998; Davis et al. 2004; Chase 2004; Barkman et al. 2007). The causes for increased substitution rates in nuclear and mitochondrial DNA of achlorophyllous plants are still unclear. In this study, we infer the variation of substitution rates of nuclear 18S rDNA of nearly all angiosperm families by comparing branch length variation of 37 fully mycoheterotrophic species, 33 parasitic plant species, and related autotrophic lineages with relative rate tests (Robinson-Rechavi and Hucho 2000; Wilcox et al. 2004). Estimating rate variation across taxa, representing most groups of angiosperms, gives us the opportunity to address the following questions: How frequent can increased substitution rates be observed in nuclear loci of hemiparasitic, holoparasitic and mycoheterotrophic plants? Are individual achlorophyllous taxa or whole heterotrophic plant families prone to substitution rates? Is loss of chlorophyll in angiosperms associated with longer branches? Can we provide evidence for another hypothesis, which could cause the increased substitution rates in 18S rDNA of mycoheterotrophic and parasitic plants? What are the exact nucleotide positions of the substitution in 18S rRNA and are they interfering with functional sites?

In total, 18S rDNA sequences of 178 angiosperm species were used for this study, representing the 45 orders of the APGII classification (APGII 2003). All families with parasitic and fully mycoheterotrophic species are represented in our study by one or more taxa, with the exception of Polygalaceae (Epipirixanthus, fully mycoheterotrophic) and Lauraceae (Cassytha, hemiparasitic). To evaluate the substitution rates of mycoheterotrophic and parasitic taxa, 18S rDNA sequences of related autotrophic plants were obtained from Genbank and included as reference points to measure substitution rates. The outgroups used for the heterotrophic lineages, according to recent phylogenetic studies are showed in Table 1. All species sampled with voucher information and Genbank accession numbers are listed in the Appendix 1.

DNA extraction, amplification and sequencing
Ten and six 18S rDNA sequences of mycoheterotrophic and autotrophic plants, respectively, were newly obtained

Table 1 List of mycoheterotrophic/parasitic taxa and outgroups investigated in this study

Mycoheterotroph/parasite Outgroup	BRT (constrained)	BRT (unconstrained)	References
Hemiparasites			
	Convolvulaceae		
<i>Cuscuta gronovii</i>	<u>0.332/0.390/0.471</u>	<u>0.334/0.401/0.466</u>	McNeal et al 2007
Convolvulus arvensis	0.164/0.202/0.252	0.167/0.216/0.260	
	Krameriaceae		
<i>Krameria ixine</i>	0.117/0.155/0.208	0.137/0.173/0.221	Simpson et al 2004
Guaiacum sanctum	0.096/0.137/0.176	0.118/0.152/0.201	
	Orobanchaceae		
<i>Pedicularis racemosa</i>	0.128/0.164/0.206	0.120/0.157/0.192	Olmstead et al 2010
<i>Orthocarpus erianthus</i>	0.143/0.184/0.231	0.142/0.179/0.219	
Paulownia tomentosa	0.123/0.154/0.194	0.115/0.151/0.189	
Lamium amplexicaule	0.139/0.171/0.214	0.128/0.165/0.202	
	Santalales (Loranthaceae)		
<i>Nuytsia floribunda</i>	0.114/0.152/0.195	0.106/0.139/0.181	Der and Nickrent 2009
<i>Tupeia antarctica</i>	0.102/0.139/0.184	0.094/0.130/0.174	Male and Nickrent 2008
	Santalales (Misodendraceae)		
<i>Misodendrum linearifolium</i>	0.193/0.242/0.308	0.194/0.245/0.298	
	Santalales (Olacaceae)		
<i>Erythralium scandens</i>	0.092/0.124/0.161	0.081/0.116/0.150	
<i>Olax aphylla</i>	0.156/0.207/0.261	0.130/0.172/0.216	
	Santalales (Opiliaceae)		
<i>Opilia amentacea</i>	0.179/0.227/0.293	0.161/0.205/0.267	
<i>Lepionurus sylvestris</i>	0.170/0.215/0.269	0.149/0.196/0.250	
	Santalales (Santalaceae)		
<i>Lepidoceras chilense</i>	0.170/0.208/0.270	0.166/0.210/0.262	
<i>Santalum album</i>	0.126/0.166/0.231	0.138/0.175/0.221	
	Santalales (Schoepfiaceae)		
<i>Schoepfia arenaria</i>	0.158/0.197/0.242	0.133/0.184/0.229	
	Santalales (Viscaceae)		
<i>Arceuthobium verticilliflorum</i>	<u>0.365/0.438/0.512</u>	<u>0.363/0.445/0.521</u>	
<i>Dendrophthora domingensis</i>	<u>0.233/0.288/0.351</u>	<u>0.244/0.300/0.368</u>	
Gunnera manicata	0.131/0.167/0.212	0.112/0.150/0.189	
Hamamelis virginiana	0.127/0.166/0.206	0.132/0.169/0.222	
Plumbago auriculata	0.142/0.182/0.219	0.144/0.186/0.238	
Holoparasites			
	Apodanthaceae		
<i>Pilostyles thurberi</i>	0.189/0.245/0.296	0.189/0.235/0.282	Nickrent et al 2004
Celastrus scandens	0.162/0.213/0.272	0.158/0.202/0.246	
	Balanaphoraceae		
<i>Ombrophytum subterraneum</i>	<u>0.386/0.457/0.547</u>	<u>0.383/0.456/0.555</u>	Steven 2001
Gunnera manicata	0.131/0.167/0.212	0.112/0.150/0.189	
Hamamelis virginiana	0.127/0.166/0.206	0.132/0.169/0.222	
Plumbago auriculata	0.142/0.182/0.219	0.144/0.186/0.238	
	Cynomoriaceae		
<i>Cynomorium coccineum</i>	0.189/0.236/0.285	0.182/0.226/0.285	Nickrent et al 2005
Gunnera manicata	0.131/0.167/0.212	0.112/0.150/0.189	
Hamamelis virginiana	0.127/0.166/0.206	0.132/0.169/0.222	
Plumbago auriculata	0.142/0.182/0.219	0.144/0.186/0.238	

Table 1 continued

Mycoheterotroph/parasite Outgroup	BRT (constrained)	BRT (unconstrained)	References
	Cytinaceae		
<i>Bdallophyton americanum</i>	0.256/0.303/0.346	0.248/0.308/0.374	Nickrent 2007
<i>Cytinus hypocistis</i>	0.322/0.406/0.467	0.343/0.410/0.479	
Muntingia calabura	0.136/0.172/0.217	0.147/0.187/0.233	
	Euphorbiaceae		
<i>Rafflesia keithii</i>	0.930/1.091/1.274	0.942/1.085/1.267	Davis et al 2007
<i>Rhizanthus infanticida</i>	0.982/1.136/1.310	0.977/1.132/1.317	
Euphorbia pulcherrima	0.114/0.156/0.203	0.107/0.145/0.182	
	Hydnoraceae		
<i>Hydnora africana</i>	0.293/0.359/0.410	0.364/0.432/0.497	Nickrent et al 2002
<i>Prosopanche americana</i>	0.351/0.425/0.495	0.397/0.487/0.566	
Aristolochia macrophylla	0.074/0.099/0.130	0.103/0.164/0.215	
	Lennoaceae		
<i>Pholisma arenarium</i>	0.242/0.290/0.346	0.236/0.276/0.320	Olmstead and Ferguson 2001
Hydrophyllum fendleri	0.173/0.215/0.270	0.162/0.204/0.243	
	Mitrastemonaceae		
<i>Mitrastemon yamamotoi</i>	0.351/0.420/0.508	0.363/0.439/0.542	Davis et al 2007
Vaccinium macrocarpon	0.143/0.183/0.225	0.133/0.170/0.207	
	Orobanchaceae		
<i>Boschniakia rossica</i>	0.126/0.169/0.214	0.127/0.165/0.199	Olmstead et al 2010
<i>Conopholis americana</i>	0.140/0.178/0.221	0.132/0.173/0.210	
<i>Epifagus virginiana</i>	0.135/0.173/0.214	0.130/0.168/0.204	
<i>Harveya speciosa</i>	0.155/0.195/0.241	0.158/0.195/0.238	
<i>Lathraea clandestina</i>	0.154/0.194/0.242	0.146/0.190/0.229	
<i>Orobanche fasciculata</i>	0.178/0.220/0.276	0.171/0.217/0.256	
Paulownia tomentosa	0.123/0.154/0.194	0.115/0.151/0.189	
Lamium amplexicaule	0.139/0.171/0.214	0.128/0.165/0.202	
Mycoheterotrophs	Family		
	Burmanniaceae		
<i>Apteria aphylla</i>	0.221/0.279/0.349	0.155/0.212/0.304	Merckx et al 2006
<i>Burmannia oblonga</i>	0.263/0.311/0.358	0.188/0.238/0.288	
<i>Burmannia sphagnoides</i>	0.198/0.254/0.303	0.136/0.187/0.249	
<i>Campylosiphon purpurascens</i>	0.147/0.188/0.246	0.083/0.122/0.160	
<i>Cymbocarpa refracta</i>	0.201/0.254/0.326	0.138/0.192/0.256	
<i>Dictyostega orobanchoides</i>	0.186/0.227/0.298	0.105/0.157/0.210	
<i>Gymnosiphon aphyllus</i>	0.178/0.232/0.286	0.115/0.159/0.215	
<i>Gymnosiphon divaricatus</i>	0.163/0.215/0.281	0.099/0.144/0.207	
<i>Hexapterella gentianoides</i>	0.141/0.187/0.243	0.075/0.118/0.166	
Burmannia bicolor	0.329/0.395/0.479	0.269/0.325/0.377	
Dioscorea rockii	0.139/0.189/0.239	0.083/0.123/0.169	
	Corsiaceae		
<i>Arachnitis uniflora</i> *	0.599/0.705/0.805	0.584/0.698/0.849	Fay et al 2006
Luzuriaga latifolia	0.102/0.140/0.183	0.037/0.075/0.107	
	Ericaceae		
<i>Hemitomes congestum</i> *	0.149/0.199/0.256	0.147/0.187/0.238	Cullings 2000
<i>Monotropa uniflora</i>	0.191/0.242/0.293	0.151/0.198/0.248	
<i>Pityopus californicus</i> *	0.165/0.210/0.252	0.184/0.233/0.283	
<i>Pterospora andromedea</i>	0.157/0.200/0.241	0.156/0.191/0.232	

Table 1 continued

Mycoheterotroph/parasite Outgroup	BRT (constrained)	BRT (unconstrained)	References
<i>Sarcodes sanguinea</i>	0.151/0.196/0.237	0.139/0.185/0.231	
Arctostaphylos uva-ursi	0.104/0.142/0.182	0.097/0.135/0.174	
Pyrola picta	0.120/0.159/0.197	0.114/0.150/0.191	
Gentianaceae			
<i>Exacum paucisquamum</i> *	0.251/0.294/0.352	0.229/0.269/0.315	Yuan et al. (2003)
<i>Sebaea oligantha</i> *	0.195/0.241/0.303	0.177/0.217/0.255	
<i>Voyria caerulea</i> *	0.241/0.294/0.361	0.218/0.268/0.320	
<i>Voyria corymbosa</i> *	0.275/0.327/0.387	0.257/0.303/0.352	
<i>Voyria aurantiaca</i> *	0.255/0.299/0.366	0.227/0.273/0.319	
<i>Voyriella parviflora</i> *	0.170/0.221/0.284	0.152/0.198/0.238	
Sebaea grandis	0.187/0.244/0.308	0.174/0.219/0.263	
Iridaceae			
<i>Geosiris</i> sp.	0.095/0.134/0.175	0.067/0.095/0.129	Reeves et al. (2001)
Aristea glauca	0.122/0.157/0.199	0.096/0.123/0.155	
Orchidaceae			
<i>Corallorhiza maculata</i>	0.165/0.205/0.246	0.112/0.171/0.221	Freudenstein et al. (2004)
Aplectrum hyemale	0.161/0.200/0.246	0.108/0.167/0.217	Molvray et al. (2000)
<i>Erythrorchis cassythoides</i>	0.288/0.335/0.385	0.226/0.297/0.361	
Cyrtosia septentrionalis	0.277/0.329/0.378	0.231/0.293/0.367	
<i>Lecanorchis multiflora</i>	0.265/0.305/0.361	0.216/0.270/0.335	
Vanilla aphylla	0.273/0.313/0.366	0.219/0.279/0.346	
<i>Neottia nidus-avis</i>	0.201/0.245/0.293	0.167/0.217/0.272	
Eburophyton austinae	0.196/0.236/0.285	0.154/0.205/0.262	
<i>Rhizanthella gardneri</i>	0.436/0.490/0.555	0.378/0.443/0.529	
Diuris sulphurea	0.223/0.265/0.310	0.175/0.225/0.286	
<i>Wulfschlaegelia calcarata</i>	0.175/0.222/0.265	0.137/0.189/0.239	
Orchis quadripunctata	0.237/0.274/0.324	0.177/0.236/0.295	
Petrosaviaceae			
<i>Petrosavia stellaris</i>	0.083/0.131/0.204	0.052/0.112/0.162	Cameron et al. (2003)
Japonolirion osense	0.073/0.117/0.180	0.056/0.102/0.152	
Thismiaceae			
<i>Afrothismia hydra</i>	0.469/0.554/0.657	0.431/0.522/0.638	Merckx et al. (2006)
<i>Afrothismia winkleri</i>	0.472/0.557/0.658	0.428/0.518/0.632	
<i>Haplothismia exannulata</i>	0.285/0.333/0.394	0.210/0.268/0.333	
<i>Thismia rodwayi</i>	0.332/0.385/0.459	0.253/0.327/0.428	
<i>Thismia aseroe</i>	0.475/0.551/0.630	0.379/0.467/0.569	
Tacca chantieri	0.148/0.195/0.248	0.095/0.135/0.185	
Dioscorea rockii	0.139/0.189/0.239	0.083/0.123/0.169	
Triuridaceae			
<i>Kupea martinetugei</i>	0.378/0.472/0.557	0.341/0.416/0.500	Steven (2001)
<i>Sciaphila ledermannii</i> *	0.133/0.185/0.237	0.077/0.128/0.177	
<i>Sciaphila densiflora</i>	0.131/0.188/0.246	0.076/0.132/0.175	
Stemona japonica	0.103/0.156/0.206	0.075/0.119/0.164	

95% confidence intervals and median values of constrained and non-constrained Bayesian relative rates test (BRT) are indicated between $\hat{\theta}$ and $\hat{\theta}_0$. Mycoheterotrophic/parasitic taxa are indicated in bold. Lineages with significant elevated substitution rates are underlined. Choice of outgroup were based on previous phylogenetic studies. Newly obtained 18S rDNA sequences for this study are indicated with an asterisk

for this study (Table 1; Appendix). Total DNA was extracted from silica-dried material with the Puregene DNA extraction kit (Gentra Systems, Landgraaf, The Netherlands) following the manufacturer's instructions. The 18S rDNA region was amplified using primers NS1, NS2, NS3, NS4, NS5 and NS6 (White et al. 1990). Each amplification reaction was performed in 25 µl reaction mix containing 5 µl DNA, 4 µl H₂O, 2.5 µl 10× PCR buffer, 0.75 µl 25 mM MgCl₂, 10 µl of 2.2 mM forward and reverse primers, 2.5 mM dNTP and 0.2 µl TaqDNA polymerase. The polymerase chain reactions were run on a GeneAmp PCR System 9700 (Applied Biosystems, Foster City, CA, USA) for 30 cycles starting at 94°C for 1 min, followed by denaturation (94°C for 30 s), annealing (44°C for 30 s), extension (72°C for 1 min) and a final extension (72°C for 7 min). PCR products were cleaned using a Nucleospin Extraction II kit (Machery-Nagel, Germany) according to the manufacturer's instructions. Sequencing reactions were done using the ABI PRISM BigDye Terminator Cycle sequencing kit (Applied Biosystems) with the same primers as listed above. All the samples were sequenced on an ABI 310 Genetic Analyzer (Applied Biosystems, Foster City, CA, USA).

Phylogenetic analyses

The sequences were assembled and edited using Staden (1998). A preliminary sequence alignment was created with Clustal X (Thompson et al. 1997) followed by manual adjustments with MacClade 4.04 (Maddison and Maddison 2001) resulting in an unequivocal alignment with a length of 1708 nucleotide positions. The best fitting model of DNA substitution was chosen by performing hierarchical likelihood-ratio tests in MrModeltest v3.06 (Posada and Crandall 1998). The Likelihood-ratio tests and Akaike Information Criterion selected the GTR+I+G model of evolution. To reduce calculation time a starting tree for the Bayesian analyses was generated under Maximum Likelihood using Garli v0.951 (Zwickl 2006) with the GTR+I+G model of evolution. An initial unconstrained Bayesian analysis of the 18S rDNA dataset retrieved a moderate resolved topology with conflicting nodes as compared to the relationships of APG II (Huchon 2000). RRTree computes differences in molecular evolution rates between non-coding DNA sequences using Kimura's two parameter (K2P) (Kimura 1980) and Jukes and Cantor's one parameter (JC) model. The K2P substitution model was selected for this study because it is the most complex model presently implemented in RRTree. According to the analysis of patterns of nucleotide substitutions in 18S rDNA of mycoheterotrophic, parasitic and autotrophic angiosperms (Ronquist and Huelsenbeck 2003) (Fig. 1a, b). The constraints resolved some major informal groups (e.g. magnoliids, monocots, commelinids, eudicots, core eudicots, rosids) conserved and variable nucleotides in the 18S rDNA dataset were identified using the chart option in MacClade

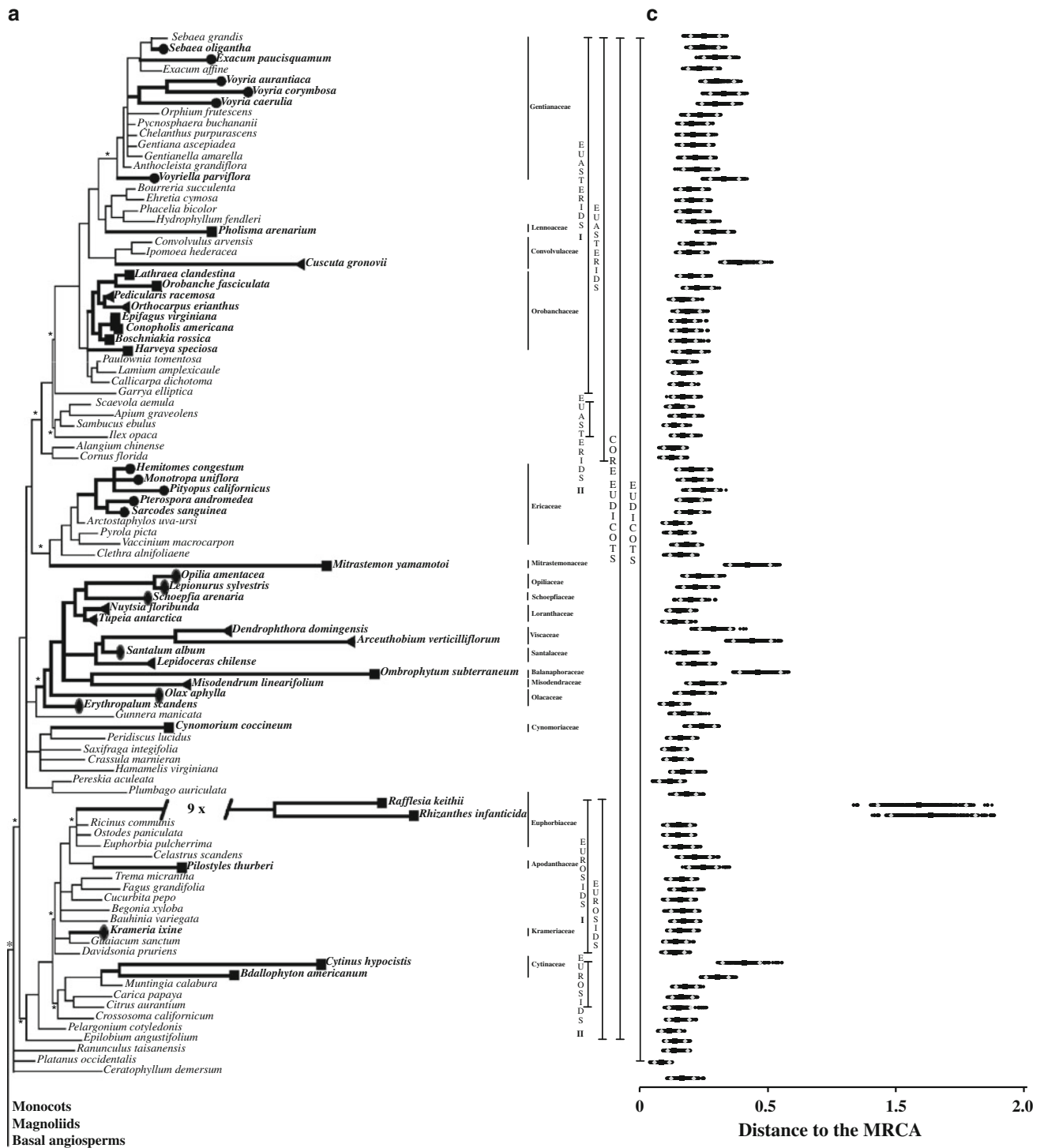


Fig. 1 a, b The majority-rule consensus phylogram of the Bayesian hemiparasitic lineages; c, d The Bayesian relative rates test shows analysis of the 18S rDNA data under topology constraints. The relative branch lengths of all ingroup taxa using a gamma distribution as implemented constraints are indicated with asterisks. Branches in bold indicate the most recent common ancestor (MRCA). The squares indicate the mean branch lengths. The triangles delimit the 95% confidence intervals

4.04 (Maddison and Maddison 2001). Each character with the Bayesian analyses. This approach provides a minimum estimate of change for each site. The values of the numbers of character substitutions or steps for every nucleotide site

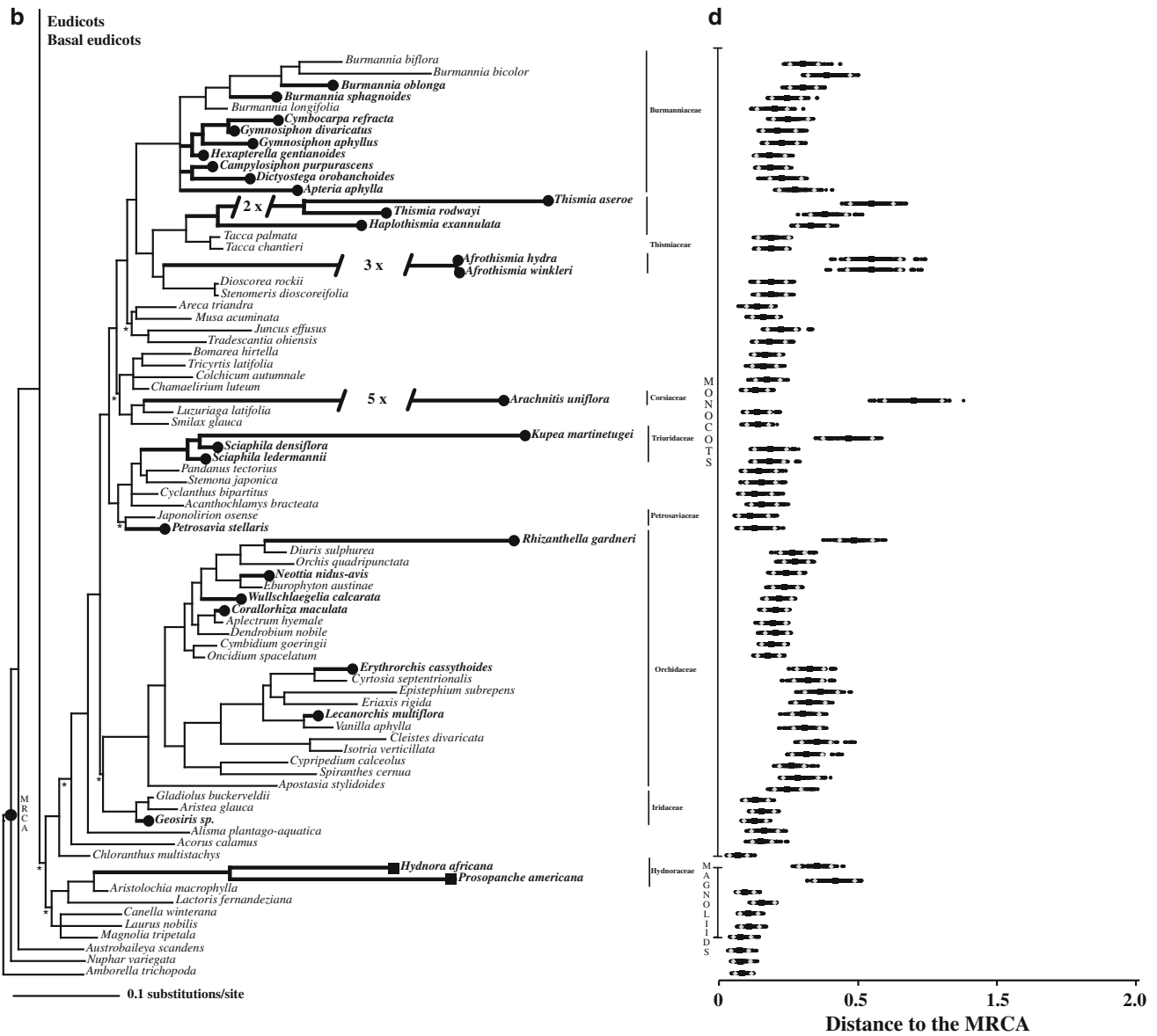


Fig. 1 continued

are shown on a histogram constructed with Excel 2004 species, respectively. Furthermore, nucleotide substitu- (for Mac version 11.4.1). This pattern of conserved andions of each nucleotide site were associated with the variable nucleotides was examined within heterotrophic secondary structures and functional regions proposed by (strict mycoheterotrophic and holoparasitic taxa) and Wuyts et al. (2000) and Caetano-Anolite (2002). Nucle- autotrophic species separately, both groups including 54 otides within the 18S rDNA alignment were divided into taxa. The autotrophic species included within this anal-915 stem and 793 loop positioie, according to secondary ysis are all sister group lineages of the heterotrophic taxa. structures obtained by the European Small Subunit Hemiparasitic and partly mycoheterotrophic taxa were Ribosomal RNA database (Van de Peer et al. 2000 removed from the analysis. In addition, the difference Wuyts et al. 2004). Stem regions are defined as those between both groups was calculated and values of steps bases that typically participate in base pairing, with the per nucleotide site were depicted on a histogram. Positive remainder occurring in non- base pairing loop structures. and negative values represent character substitutions. In this study, base pairing was restricted to Watson-Crick contributed by heterotrophic lineages and autotrophic nucleotide pairs and GDU pairs.

Results

Evolutionary rates

The constrained phylogram (Fig. a, b) reveals heterogeneity in branch lengths among the taxa investigated. Several mycoheterotrophic and parasitic plant species show extremely elevated substitution rates resulting in long terminal branches (e.g. *Rafflesia keithii*, *Rhizanthus infanticida* and *Arachnitis uniiformis*). The results of the Bayesian relative rates test are shown in Fig. (panels c, d) and listed in Table 1. The values of the confidence intervals for every mycoheterotrophic and parasitic lineage with the associated outgroups are given in Table 1 in order to analyze the effect of the implementation of constraints on the substitution rates of mycoheterotrophic and parasitic lineages, a second Bayesian relative rates test was conducted on the same dataset using trees from the unconstrained analysis. Overall, the confidence intervals of the substitution rates in the constrained and non-constrained phylogenetic analyses showed no pronounced differences, which mean that the implementation of topology constraints did not affect branch lengths (Table 1).

According to the Bayesian relative rate test the substitution rates of several hemiparasitic (3 out of 16), holoparasitic (8 out of 17) and mycoheterotrophic (8 out of 37) species are significantly higher compared to their autotrophic relatives (Table 1). In general, most holoparasitic plant families share substantial elevated substitution rates which are on average 1.8-fold (*Allophytum americanum*, *Rhizanthus infanticida*, *Euphorbia* biaceae) higher compared to their autotrophic relatives. Only the holoparasitic families, Apodanthaceae, Cynomoriaceae, Lennoaceae and Orobanchaceae, do not have significantly increased substitution rates. Hemiparasitic plants are characterized by less pronounced substitution rates where only three species are placed on significantly longer branches: average of 1.6-fold (*Andropogon domingensis*, *Viscaceae*) to 2.4-fold (*Arceuthobium verticilliformis*, *Viscaceae*) higher compared to their autotrophic relatives. The (facultative) hemiparasites of the families Krameriaceae and Orobanchaceae and the remaining Santalalean lineages are not prone to increased substitution rates in 18S rDNA. In mycoheterotrophic plants only eight out of 37 fully mycoheterotrophic plants have significantly elevated substitution rates with an average increase of between 1.7-fold (*Haplothismia exannulata*, *Thismiaceae*) and 5.0-fold (*Arachnitis uniiformis*, *Corsiaceae*). The most pronounced increase of substitution rates among mycoheterotrophic species included all five taxa of *Thismiaceae* and the single taxon of *Corsiaceae*. Although the branch lengths of the two mycoheterotrophic species

gardneri (Orchidaceae) and *Kupea martinetugae* (Triuridaceae) are considerably shorter they are still significantly longer compared to their autotrophic sister groups. The Bayesian relative rates test reveals no significant variation in rates of molecular evolution in species of the mycoheterotrophic families Burmanniaceae, Ericaceae, Gentianeaceae, Iridaceae and Petrosaviaceae. However, depending on whether *Burmannia bicolor* or *Dioscorea rockii* was used as autotrophic relative, the mycoheterotrophic *Burmannia oblonga* is either placed on a significant or non-significant long branch than its green relatives. The RRTree test revealed similar results compared to the Bayesian relative rate test (Table 1). However, significant increased substitution rates were observed in *Peucedanum paucisquamum* and the genus *Voyria*) and holoparasitic groups *Cynomorium coccineum*, *Pholisma arenarium* and *Pilostyles thurberi*. Furthermore, mycoheterotrophic/parasitic clades (i.e. the Triuridaceae and the Santalales clade), which contain both lineages with and without significantly increased substitution rates according to the Bayesian relative rates test, are indicated by significant P-values for the whole clade.

Patterns of nucleotide substitution in 18S rDNA Alignment of 18S rDNA angiosperm sequences showed an alternation of conserved and variable regions. Using the chart option in MacClade, patterns of nucleotide substitutions were generated, describing a mosaic of conserved and variable nucleotides across the angiosperms (Fig. 2). Due to missing data the first and last nucleotide positions of the alignment could not be compared among all sequences. This resulted in a substitution pattern starting and ending in helix structure number 8 and 49, respectively. Variable nucleotides tend to occur both in loop and stem structures in both autotrophs and holoparasites/mycoheterotrophs, but highly variable regions (≥ 10 steps) are more abundant in loop structures (e.g. 11, 17, E23_2, E23_7, E23_12, E23_13 and 49), than in stem structures (e.g. E10_1, 11, 43 and 49) (Fig. a, b). Conserved regions with no or few mutations appear both in stem and loop structures (8, 9, 13, 14, 15, 16, 4, 19, 20, 21, 3, 22, 23, E23_8, E23_9, E23_10, E23_11, 26, 23, 27, 28, 30, 31, 2, 32, 33, 34, 35, 36, 38, 39, 40, 41, 42 and 34) (Fig. a, b). The loop and stem structures comprised 1943 and 1745 total steps, respectively. Patterns of substitutions were separately analyzed in autotrophic (Fig. 2a) and holoparasitic/mycoheterotrophic lineages (Fig. 2b), showing a similar distribution pattern: the positions of conserved and variable nucleotides are almost identical in the autotrophic and achlorophyllous

Table 2 Values of the relative rate test for comparing molecular evolutionary rates between parasitic/mycoheterotrophic lineages (Lineage 1) and autotrophic relatives (Lineage 2)

Family	Lineage 1	Lineage 2	K1	K2	K1/K2	K1/K2P value ^a	
Apodanthaceae	Pilostyle ^b	Celastrus	0.055	0.043	0.012	1.3	2.2×10^{-2}
Burmanniaceae	Apteria	B. bispora B. bicolor	0.006	0.068	-0.062	0.1	3.0×10^{-1}
	Burmannia oblonga	B. bispora B. bicolor	0.059	0.068	-0.009	0.9	4.5×10^{-2}
	Burmannia sphagnoides	B. bispora B. bicolor	0.033	0.053	-0.020	0.6	8.0×10^{-5}
	Campylosiphon, Dictyostega	B. bispora B. bicolor	0.063	0.093	-0.029	0.7	7.7×10^{-5}
	Cymbocarpa Gymnosiphon clade, Hexapterella	B. bispora B. bicolor	0.044	0.068	-0.024	0.6	1.0×10^{-7}
Convolvulaceae	Cuscuta	Convolvulus pomoea	0.077	0.044	0.033	1.8	1.0×10^{-7}
Corsiaceae	Arachnitis	Luzuriaga Smilax	0.105	0.020	0.084	5.2	1.0×10^{-7}
Cynomoriaceae	Cynomorium	Hamamelis, Peridiscus	0.053	0.039	0.014	1.4	3.9×10^{-3}
Cytinaceae	Baldophyton Cytinus	Muntingia	0.072	0.041	0.031	1.8	1.0×10^{-7}
Ericaceae	Ericaceae clade	Arctostaphylos Pyrola, Vaccinium	0.054	0.053	0.001	1.0	7.4×10^{-1}
Euphorbiaceae	Rafflesia Rhizanthus	Euphorbia Ostodes, Ricinus	0.150	0.035	0.115	4.3	1.0×10^{-7}
Gentianaceae	Exacum	Sebaea grandis	0.064	0.050	0.014	1.3	1.5×10^{-3}
Gentianaceae	Sebaea oligantha	Sebaea grandis	0.051	0.050	0.001	1.0	5.2×10^{-1}
Gentianaceae	Voyria clade ^b	Sebaea grandis Orphium	0.063	0.052	0.011	1.2	1.4×10^{-3}
Gentianaceae	Voyriella	Sebaea grandis Orphium	0.049	0.050	-0.001	1.0	8.3×10^{-1}
Hydnoraceae	Hydnora Prosopanche	Aristolochia Lactoris	0.086	0.039	0.047	2.2	1.0×10^{-7}
Iridaceae	Geosiris	Aristea Gladiolus	0.034	0.035	-0.001	1.0	7.6×10^{-1}
Krameriaceae	Krameria	Guaiacum	0.039	0.039	0.000	1.0	9.9×10^{-1}
Lennoaceae	Pholisma	Hydrophyllum Phacelia Ehretia	0.060	0.044	0.016	1.4	7.7×10^{-4}
Mitrostemonaceae	Mitrostemon	Arctostaphylos Pyrola, Vaccinium	0.086	0.043	0.044	2.0	1.0×10^{-7}
Orchidaceae	Corallorhiza	Aplectrum Dendrobium	0.048	0.045	0.003	1.1	1.2×10^{-1}
Orchidaceae	Erythrorchis	Cyrtosia Eriaxis, Epistephium	0.060	0.062	-0.002	1.0	6.0×10^{-1}
Orchidaceae	Lecanorchis	Vanilla Eriaxis, Epistephium	0.050	0.054	-0.004	0.9	2.7×10^{-1}
Orchidaceae	Neottia	Europhyton	0.060	0.055	0.006	1.1	1.2×10^{-1}
Orchidaceae	Rhizanthella	Diuris Orchis	0.088	0.050	0.038	1.8	2.0×10^{-7}
Orchidaceae	Wulfschlaegelia	Europhyton Diuris	0.044	0.054	-0.010	0.1	7.4×10^{-3}
Orobanchaceae	Orobanchaceae clade	Paulownia Lamium Callicarpa	0.044	0.425	0.002	0.1	5.6×10^{-1}
Petrosaviaceae	Petrosavia	Japonolirion	0.035	0.031	0.004	1.1	2.5×10^{-1}
Santalales	Santalales clade	Gunnera Peridiscus Plumbago	0.051	0.039	0.012	1.3	6.1×10^{-6}
Thismiaceae	Afrothismiaceae	Dioscorea Stenomeris Tacca	0.113	0.041	0.072	2.8	1.0×10^{-7}
Thismiaceae	Haplothismia Thismiaceae	Dioscorea Stenomeris Tacca	0.082	0.042	0.040	1.9	1.0×10^{-7}
Triuridaceae	Kupea Sciaphila clade	Pandanus Stemona	0.056	0.037	0.019	1.5	5.7×10^{-6}

P values of lineages with significant increased substitution rates are indicated in bold

K1 mean divergence between lineage 1 and the most recent common ancestor of lineages 1 and 2, and K2 mean divergence between lineage 2 and the most recent common ancestor of lineages 1 and 2

^a Significance of the P values < 0.05

^b Lineages with significant increased substitution rates which are not observed in the Bayesian relative rates test

histograms. However, more and highly variable nucleotides are presented on the latter histogram. Figure 2 shows this result by calculating the difference of steps between mycoheterotrophic/holoparasitic taxa and autotrophic species. The distribution pattern showed that most of the substitutions occurred within achlorophyllous taxa, revealing that mainly heterotrophic taxa contribute to the observed mutations. Both Bayesian relative rate test and the distance based relative rate test showed similar results (Tables 2), providing evidence for their accuracy and robustness.

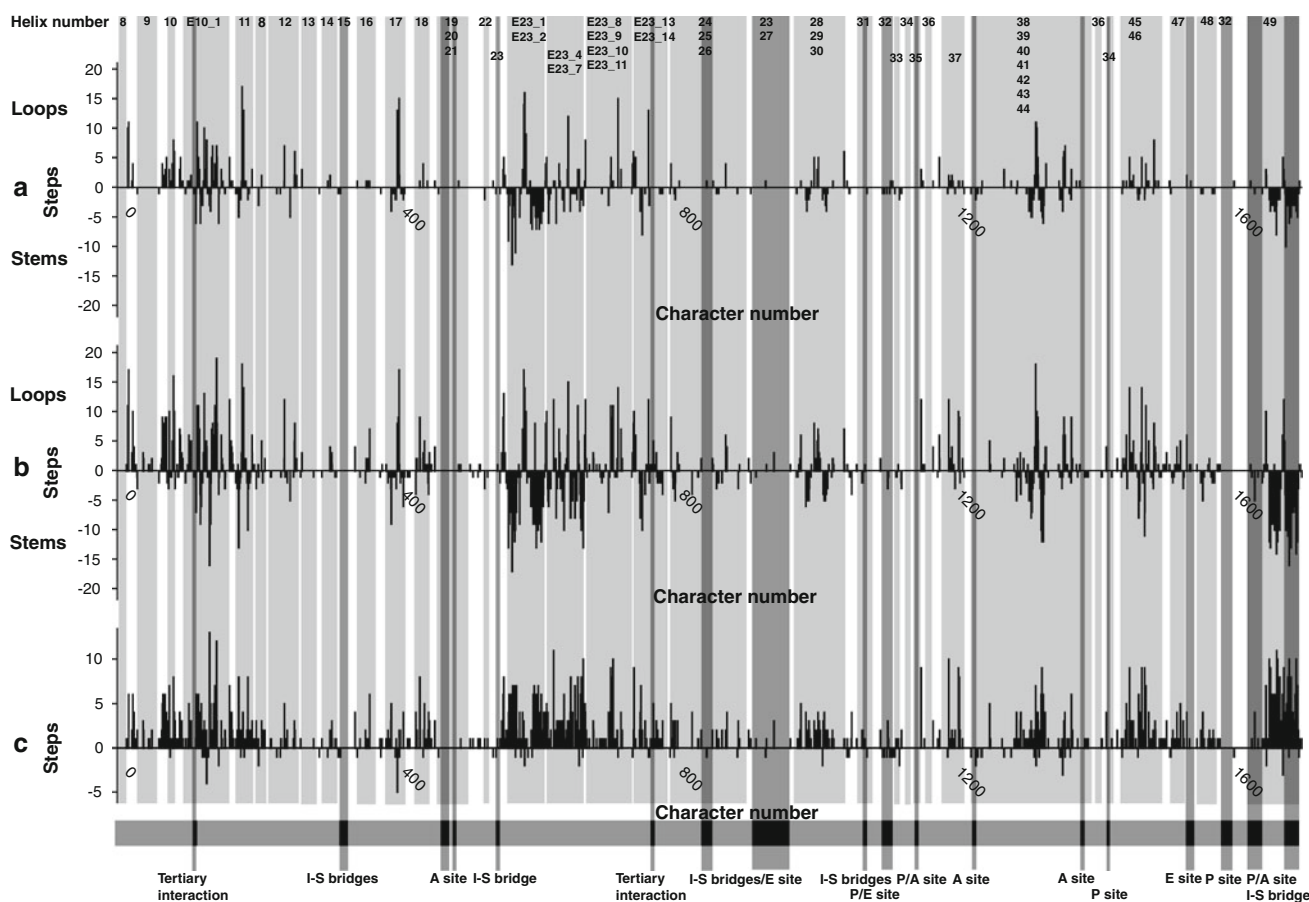


Fig. 2 Histograms showing the pattern of nucleotide substitutions in nucleotide substitution of 54 autotrophic taxa. The histogram describes for each nucleotide site the amount of parsimony steps or character substitutions (y-axis; a, b loops above, stems below; c heterotrophic minus autotrophic). Helix numbering according to Wuyts et al. (2000) is shown at the top. The major functional and structural sites according to Caetano-Anolles (2002) are indicated at the bottom. Pattern of nucleotide substitution of 37 mycoheterotrophic and 17 holoparasitic taxa. Difference between mycoheterotrophic/parasitic and autotrophic values of character substitutions per nucleotide site is shown in light gray and dark gray shading displays the different helix structures and functional regions, respectively

The statistical tests showed remarkably long terminal branches for the heterotrophic taxa *Raflesia keithii*, *Rhizanthus infanticida*, *Arachnitis uniflora*, *Afrothismia hydra*, *A. winkleri* and *Thismia aseroe* compared to other angiosperms. Long terminal branches were not observed in chlorophyllous relatives of these taxa. These significantly increased substitution rates suggest that a non-photosynthetic mode of life could be a necessary condition for the presence or absence of chlorophyll and elevated substitution rates in 18S rDNA. However, our results show that the loss of photosynthesis in 29 mycoheterotrophic and nine holoparasitic representatives does not necessarily imply significantly increased substitution rates in 18S rDNA. Several non-photosynthetic lineages in Apodanthaceae, Burmanniaceae, Cynomoriaceae, Ericaceae, Gentianaceae, Iridaceae, Leucorhizaceae, noaceae, Orobanchaceae, Orchidaceae, Petrosaviaceae and Triuridaceae lack significantly accelerated substitution rates, according to the Bayesian relative rates test. On the other hand all sampled mycoheterotrophic and holoparasitic Balanophoraceae, Corsiaceae, Cytinaceae, Euphorbiaceae, Hydnoraceae, Mitrastemonaceae and Thismiaceae are characterized by elevated substitution rates (Fig. 2). Previous studies investigating the substitution rates in mycoheterotrophic and parasitic plants confirm the observation that there is no evolutionary trend between the presence or absence of chlorophyll and elevated substitution rates (mycoheterotrophs: Cameron and Chase 2000; Molvray et al. 2000; Merckx et al. 2006, 2009 and parasitic plants: Nickrent and Starmer 1994; Nickrent et al. 1998). Currently it is still unclear why some heterotrophic non-photosynthetic plants accumulate much more substitutions in their ribosomal DNA than autotrophic plants, but numerous potential hypotheses explaining causes of accelerated substitution rates have been proposed. The long-term effects of a small effective population size resulting in a genetic bottleneck effect (Wu and Li 1985), the influence of a short generation

time and the correlated higher number of mutation-generational reproductive events (Wu and Li 1985), an increased tolerance of mutations due to a relaxation of selective constraints, variations in mutation rate (Sniegowski et al. 2000), DNA repair efficiency (Modrich and Lahure 1996) and speciation rates (Barraclough and Savolainen 2001) are possible factors that trigger high substitution rates in mycoheterotrophic and parasitic plants. None of these hypotheses could unequivocally explain higher substitution rates in parasitic plants (Nickrent and Starr 1994; dePamphilis et al. 1997; Young and dePamphilis 2005). However, in mycoheterotrophic plants, the effective population size hypothesis may have played an important role in many cases of elevated substitution rates. The genera *Triuridaceae* (Triuridaceae), *Afrothismia* (Thismiaceae) and *Arachnitis* (Corsiaceae) have very limited distribution ranges. *Arachnitis martinetugei* has only been reported from two localities in Cameroon (Cheek et al. 2003), *Afrothismia* is restricted to extremely scattered populations in the tropical rain forests in Africa (Maas-van de Kamer 1998; Franke et al. 2004) and *Arachnitis* is restricted in its distribution to disjunct areas in the southern part of South America (Ibisch et al. 1996). A very limited geographic distribution or reduced effective population size may indeed explain the significantly increased substitution rates of 18S rDNA in these mycoheterotrophic taxa. In parasitic lineages, however, Nickrent and Starr (1994) postulated that the effective population size hypothesis could not be the only cause of higher substitution rates. A very restricted distribution pattern has been observed in the holoparasites *Rafflesia* (Euphorbiaceae), *Rhizanthus* (Euphorbiaceae) and *Prosopanche* (Hydnoraceae), which have accumulated significantly more substitutions compared to their autotrophic relatives. In contrast, *Arceuthobium* (Viscaceae) species have large and extremely widespread populations and show also significantly increased evolutionary rates. However, it remains to be studied whether population ranges are still mating (see Taylor et al. 2004).

An alternative hypothesis to explain the absence of accelerated substitution rates in mycoheterotrophic plants is the absolute time when photosynthesis capacity was lost. Non-photosynthetic plants without significant increased substitution rates might have lost their chlorophyll only recently. However dating analysis on *Burmanniaceae*, which show no significant increase in substitution rates, suggests a Late Cretaceous/Eocene origin of mycoheterotrophy, rejecting this hypothesis in that case (Merckx et al. 2008). Probably no single mentioned hypothesis will be able to explain unequivocally the substitution patterns observed in mycoheterotrophic and parasitic plants, but most likely a combination of several mechanisms is affecting divergent rDNA sequences.

Nucleotide variability and functionality of 18S rDNA The number of substitutions per site in 18S rDNA differs greatly among sites. Alignment of 18S rDNA sequences reveals the presence of conserved areas with few variable substitutions interspersed (Nickrent and Soligo 1995; Van Peer et al. 1993, 1996). We observed similar results in our dataset, showing a mosaic pattern with long stretches of conserved nucleotides and short variable regions in 18S rDNA (Fig. 2). Both stem and loop structures contain variable and conservative nucleotides. A higher proportion of highly variable nucleotides (≥ 10 steps) and a higher number of steps occurred in loops compared to stem structures, suggesting differences in selective constraints between both structures. This result confirms the pattern found in eukaryotes, where loops evolve considerably faster than stems (Smit et al. 2007).

More important is the similar distribution pattern between heterotrophic and related autotrophic plants (Fig. 2a, b). Despite a high number of variable nucleotides in mycoheterotrophic and holoparasitic plants (Fig. 2), mutations seem to be strictly positioned to specific nucleotide sites, suggesting fixed selective constraints in the 18S rRNA molecule. These variable nucleotide sites occur in regions that are known to be highly variable within eukaryotes (Wuyts et al. 2000, 2001). The variable regions include mainly peripherally located eukaryotic-specific structures attached to a conserved core structure. Consequently, most conserved nucleotide sites are near the ribosome centre with increased nucleotide variability towards the ribosome surface (Wuyts et al. 2001). Nucleotides at the periphery of the ribosome, which are prone to substitutions, were located in structure number 6, 10, E10_1, 11, 17, 18, E23, 29, 43, 45 and 46. These variable nucleotides are under less functional constraints compared to centrally located sites with a catalytic or binding activity, which have less freedom to mutate. In Fig. 2, functional structures involved in peptidyl transferase activity, the translational cycle and interaction between rRNA subunits delimited by Caetano-Anolles (2002) are presented. As expected, substitutions are rare in those functional sites even within the faster evolving heterotrophic taxa. Based on these observations we can conclude that no major interference with the functions of the ribosome occurs and that the small ribosomal subunit retains its functionality, despite extremely high substitution rates in chlorophyllous taxa detected.

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Appendix

List of sampled species, Genbank accession numbers and voucher information arranged into the major Angiosperm groups. Newly obtained 18S rDNA sequences for this study are indicated with an asterisk. Sequences obtained from Genbank do not list voucher information.

Taxon-Genbank accession; Voucher.

OUTGROUP

Amborella trichopoda Baill.-U42497

BASAL ANGIOSPERMS

Austrobaileya scandens C.T. White-AF206858, *Nuphar variegata* Engelm. ex Clinton-AF206972

MAGNOLIIDS

Aristolochia macrophylla Lam.-AF206855, *Canella winterana* Gaertn.-AF206879, *Chloranthus multistachys* P.Oei-AF206885, *Hydnora africana* Thunb.-L25681, *Lactoris fernandeziana* Phil.-LFU42783, *Laurus nobilis* Cav.-AF197580, *Magnolia tripetala* L.-AF206956, *Prospanche americana* (R.Br.) Kuntze-L24047

MONOCOTS

Acanthochlamys bracteata P.C.Kao-AY952411, *Acorus calamus* L.-L24078, *Afrothismia hydra* Sainge & T.Franke-DQ786083, *Afrothismia winkleri* Sainge & T.Franke-EU420992, *Alisma plantago-aquatica* L.-AF197585, *Aplectrum hyemale* Torr.-U59937, *Apostasia stylidioides* Rchb.f.-AF135207, *Arachnitis uniflora** Phil.-HQ448758; Cocucci 2122, *Aristea glauca* Klatt-AF206854, *Areca triandra* Roxb.-AY952409, *Apteris aphylla* (Nutt.) Barnh. ex Small-DQ786035, *Bomarea hirtella* Herb.-AF206871, *Burmanna bicolor* Mart.-DQ786072, *Burmanna biflora* L.-DQ786070, *Burmanna longifolia* Becc.-AF309398, *Burmanna oblonga* Ridl.-DQ786064, *Burmanna sphagnoides* Becc.-AF309400, *Campylosiphon purpurascens* Benth.-EU420996, *Chamaerilium luteum* Miq.-AF206884, *Cleistes divaricata*

Almes-AF135205, *Colchicum autumnale* L.-U42072, *Corallorhiza maculata* Greene-U59940, *Cyclanthus bipartitus* Poit.-AF168837, *Cymbidium goeringii* (Rchb.f.) Rchb.f.-AJ271248, *Cymbocarpa refracta* Miers-DQ786038, *Cypripedium calceolus* L.-AF069208, *Cyrtosia septentrionalis* (Rchb.f.) Garay-AF135198, *Dendrobium nobile* Lindl.-AB027309, *Dictyostega orobanchoides* (Hook.) Miers-DQ786056, *Dioscorea rockii* Prain & Burkill-DQ786090, *Diuris sulphurea* R.Br.-AF135196, *Eburophyton austinae* A. Heller-U59949, *Epistephium subrepens* Hoehne-AF135200, *Eriaxis rigida* Rchb.f.-AF135201, *Elythrorchis cassythoides* (A. Cunn. ex Lindl.) Garay-AF135199, *Geosiris sp.* Bail-EU816707, *Gladiolus buckeri* Goldblatt L54062, *Gymnosiphon aphyllus* Blume-AF309402, *Gymnosiphon divaricatum* (Benth.) Benth. & Hook.-DQ786043, *Haplothismia exannulata* Airy Shaw-DQ786082, *Hexapterella gentianoides* Urb.-DQ786057, *Isotria verticillata* Raf.-AF135204, *Japonolirion osense* Nakai-AF206942, *Juncus effusus* L.-AF206944, *Kupea martinetegei* Cheek & Williams-EU816706, *Lecanorchis multiflora* J.J.Sm.-AF135203, *Luzuriaga latifolia* Poir.-AF233091, *Musa acuminata* Colla-AF069226, *Neottia nidus-avis* (L.) Rich.-U59948, *Oncidium sphacelatum* Lindl.-U59939, *Orchis quadripunctata* Cirillo ex Ten.-AF135206, *Pandanus tectorius* Parkinson ex J.P.du Roi-AY952391, *Petrosavia stellaris* Becc.-AF206987, *Rhizanthella gardneri* R.S.Rogers-AF135198, *Sciaphila densiflora* Schltr.-EU816704, *Sciaphila ledermannii** Engl.-HQ448766; Merckx 128, *Smilax glauca* Walter-AF207022, *Spiranthes cernua* Rich.-AF135195, *Stemona japonica* Franch. & Sav.-AF207028, *Stenomeres dioscoreifolia* Planch.-DQ786087, *Tacca chantrieri* André-DQ786086, *Tacca palmata* Blume-EU421000, *Thismia aseroe* Becc.-AF309404, *Thismia rodwayi* F.Muell.-AF309403, *Tradescantia ohiensis* Raf.-AF069213, *Tricyrtis latifolia* Maxim.-AF207046, *Vanilla aphylla* Wight-AF135202, *Wulschlaegelia calcarata* Benh.-EU816708

BASAL EUDICOTS

Ceratophyllum demersum L.-U42517, *Gunnera manicata* Linden-U43787, *Platanus occidentalis* L.-U42794, *Ranunculus taisanensis* Hayata-D29780

CORE EUDICOTS

Arceuthobium verticilliflorum Engelm.-L24042, *Crassula marnieriana* Huber & Jacobsen-U42525, *Cynomorium coccineum* L.-AF039069, *Dendrophthora domingensis* Eichler-X16601, *Erythralium scandens* Blume-DQ790111, *Hamamelis virginiana* L.-AF094551, *Lepidoceras chilense* (Molina) Kuijt-EF464459, *Lepionurus sylvestris* Blume-DQ790101, *Misodendrum linearifolium*

DC.-L24397, *Nuytsia floribunda* R.Br.-DQ790103, *Olap aphylla* R.Br.-L24405, *Ombrophytum subterraneum* (Asplund) B.Hansen-L24406, *Opilia amentacea* Wall.-L24407, *Pereskia aculeata* Mill.-AF206986, *Peridiscus lucidus* Benth.-AY372815, *Plumbago auriculata* Lam.-U42795, *Santalum album* L.-DL24416, *Saxifraga integrifolia* Hook.-U42810, *Schoepfia arenaria* Britton & X16606, *Tupeia antarctica* Cham. & Schltldl.-L24425

ROSIDS

Bauhinia variegata L.-AF525295, *Begonia oxyloba* Welw. ex. Hook.f.-AY968392, *Bdallophyton americanum* (R. Br.) Harms-AY739089, *Carica papaya* L.-U42514, *Celastrus scandens* L.-AY674581, *Citrus aurantium* L.-U38312, *Crossosoma californicum* Nutt.-U42529, *Cucurbita pepo* L.-AF206895, *Cytinus hypocistis* L.-AY739092, *Davidsonia pruriens* F.Muell.-AF206897, *Epilobium angustifolium* L.-AF206907, *Euphorbia pulcherrima* Willd. ex Klotzsch-L37582, *Fagus grandifolia* Ehrh.-AF206910, *Guaiacum sanctum* L.-U42824, *Krameria ixine* L.-AF206948, *Muntingia calabura* L.-U42539, *Ostodes paniculata* Blume-AB268104, *Pelargonium cotyledonis* L.-AF206982, *Pilostyles thurberi* A.Gray-AY739081, *Rafflesia keithii* Meijer-L24041, *Rhizanthus infanticida* H.Benziger & B. Hansen-L24048, *Ricinus communis* L.-AB233559, *Trema micrantha* Blume-AF207044

ASTERIDS

Alangium chinense (Lour.) Harms-AF206843, *Anthocleista grandiflora* Gilg-AJ236026, *Apium graveolens* L.-AF206852, *Arctostaphylos uva-ursi* Spreng.-L49272, *Boschniakia rossica* (Cham. & Schltldl.) Standl.-U59951, *Bourreria succulenta* Jacq.-U38319, *Callicarpa dichotoma* Raeusch.-AJ236048, *Chelonanthus purpurascens** (Aubl.) Struwe, S.Nilsson & V.A.Albert-HQ448759; BINCO-FG43, *Clethra alnifolia* L.-AF419793, *Convolvulus arvensis* L.-AJ236013, *Conopholis americana* (L.) Wallr.-U59954, *Cornus florida* L.-X17370, *Exacum paucisquamum** (C.B.Clarke) Klack-HQ448760; Yuan CN2k1-3C, *Cuscuta gronovii* Wild. ex Roem. & Schult. DL2474, *Ehretia cymosa* Thonn.-U59938, *Epifagus virginiana* (L.) W.P.C.Barton-U59955, *Exacum affine* Balf.f.-AJ236023, *Garrya elliptica* Dougl. ex Lindl.-U42540, *Gentiana asclepiadea** L.-HQ448773; 19801931, *Gentianella amarella** (L.) H. Sm.-HQ448761; Fay 14626, *Harveya speciosa* Bernh.-U59950, *Hemitomes congestum** A.Gray-HQ448762; Bidartondo s.n., *Hydrophyllum fendleri* A. Heller-AJ236019, *Ilex opaca* Soland.-AF206938, *Lipomoea hederacea* Jacq.-U38310, *Lamium amplexicaule* L.-L49287, *Lathraea clandestina* L.-U59941, *Mitrastemon yamamotoi* Makino -AY739090, *Monotropa uniflora* L.-L25680, *Orobanche*

fasciculata Nutt.-U59961, *Orthocarpus erianthus* Benth.-U38316, *Orphium frutescens** E.Mey.-HQ448763; J. Kissling and L. Zeltner 44, *Paulownia tomentosa* Steud.-AJ236039, *Pedicularis racemosa* Dougl. ex Hook.-U59959, *Phacelia bicolor* Torr. Ex S.Watson-L49292, *Pholisma arenarium* Nutt.-U59935, *Pityopus californicus** (Eastw.) Copel.-HQ448764; Bidartondo s.n., *Pterospora andromedea* Nutt.-U59943, *Pycnosphaera buchananii** N.E.Br.-HQ448765; Bingham MG937, *Pyrola picta* Sm.-U59936, *Sambucus ebulus* L.-AJ236005, *Sarcodes sanguinea* Torr.-U59945, *Scaevola aemula* R.Br.-AJ236008, *Sebaea grandis** Steud.-HQ448767; Dessein et al. 75, *Sebaea oligantha** Schinz-HQ448768; Merckx 103, *Vaccinium macrocarpon* Ait.-AF419808, *Voyria aurantiaca** Splitq.-HQ448769; Maas et al. 9610, *Voyria caerulea** Aubl.-HQ448770; Maas et al. 9636, *Voyria corymbosa** Splitq.-HQ448771; Maas et al. 9611, *Voyriella parviflora** Miq.-HQ448772; Maas et al. 9678.

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