



Original article

BGP expression in gastric biopsies may predict the development of new lesions after local treatment for early gastric cancer

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Abstract

Background. Our previous studies have demonstrated the significant role of the generative cells of intestinal metaplasia (IM) expressing brain (fetal)-type glycogen phosphorylase (BGP) (BGP-IM) as a premalignant lesion of intestinal-type adenocarcinoma. The aims of the present study were to investigate the incidence of BGP-IM in gastric biopsy specimens and to establish BGP-IM as a predictor of the coexistence of accessory carcinoma and/or metachronous cancers before and after local treatment for early gastric carcinoma.

Methods. We studied the incidence of BGP-IM in eight endoscopic biopsy specimens of methylene blue-positive mucosa of the stomach obtained from patients with multiple gastric carcinomas ($n = 14$), a single carcinoma ($n = 25$), and atrophic gastritis ($n = 20$).

Results. BGP positivity was 93.3% in the multiple carcinomas and 80.0% in the single carcinomas. The incidences of BGP-IM (mean percentage \pm SD) in the stomachs with multiple carcinomas, single carcinoma, and atrophic gastritis were $83.2\% \pm 22.8\%$, $36.5\% \pm 41.3\%$, and $7.1\% \pm 18.0\%$, respectively. The incidence was significantly higher in the stomachs with multiple carcinomas than in those with a single carcinoma or those with atrophic gastritis ($P < 0.001$).

Conclusion. It is suggested that the frequent appearance of BGP-IM reflects the high potential of carcinogenesis of intestinal-type gastric cancer, and that the involvement of BGP-IM in more than 50% of the eight biopsies may be a predictor of the coexistence of accessory and/or metachronous carcinoma before and after local treatment for early gastric carcinoma.

Key words Gastric carcinoma · Endoscopic mucosal resection · Local treatment · Intestinal metaplasia · Glycogen phosphorylase · Predictor

Introduction

The recent advent of endoscopic and laparoscopic local treatments has offered a better quality of life to patients with early gastric carcinoma involving no lymph node metastasis [1–5]. These treatments, however, incur increasing risks of missing the coexistence of accessory (microscopic) carcinomas and/or developing new cancers in the remnant stomach [6–9].

The incidence of multiple primary gastric carcinoma has been reported to be from 5% to 10% in patients who had gastrectomy for gastric cancer [10–14]. The incidence is elevated with age and male sex, and with intestinal-type tumors; frequent occurrence in the lower third, and mucosal cancers, were significantly correlated with multiple early gastric cancer. However, these accessory lesions were missed preoperatively in approximately 30%–40% of the patients with multifocal early gastric cancers. Furthermore, considerable numbers of microscopic cancers could have been overlooked. Almost all of these unrecognized lesions, however, should be co-resected by distal gastrectomy, because those concomitant lesions were detected significantly more frequently in the lower part of the stomach on the distal side of the atrophic border than in the upper part [15,16]. Also, microscopic cancers may grow into clinically significant lesions within several years when a gastric carcinoma is removed by local treatment [9]. In addition, it has been reported that the histological types of these carcinomas are almost all of the intestinal type. Therefore, we should always remember that other gastric lesions may also be present and/or grow when we are treating patients with gastric cancer by local treatment such as endoscopic mucosal resection (EMR) or laparoscopic wedge resection.

Local treatment for early gastric cancer is currently indicated mainly for intestinal-type carcinoma. If there were some indicators that predicted the frequent coexistence of multiple gastric cancers (including micro-

scopic carcinoma) and/or the metachronous growth of another gastric cancer of the intestinal type, these would be very useful to identify the high-risk group and would contribute to the follow-up studies after local treatment of gastric cancer.

We have previously demonstrated the frequent abnormal expression of brain (fetal)-type glycogen phosphorylase (BGP) in gastric carcinoma, especially in intestinal-type cancer [17–21]. Furthermore, with regard to the carcinogenesis of gastric carcinoma, we have also pointed out the significant role of the generative cells of intestinal metaplasia (IM) expressing BGP (BGP-IM) as a premalignant lesion of intestinal-type adenocarcinoma.

In the current study, using gastric biopsy specimens, BGP expression in IM was evaluated as a predictive indicator showing the coexistence of accessory carcinoma of the human stomach, and as a possible predictor of metachronous gastric carcinoma.

Patients and methods

Patients and specimens

Between 1997 and 2000, 59 patients with intestinal-type early gastric cancer and endoscopic atrophic gastritis were analyzed in this study. Of these patients, 14 had synchronous multiple gastric carcinomas, 25 had a single cancer, and 20 had endoscopic atrophic gastritis without any localized lesions. During endoscopic examination, the lower two-thirds of the stomach was dyed with methylene blue [22,23] and eight endoscopic biopsies were made from the stained mucosa (Fig. 1) in the anterior, posterior, greater and lesser curvature wall of the antrum and lower body of the stomach, respectively. Informed consent was obtained from all the patients.

The biopsy specimens were put on small filters, fixed in 10% buffered formaldehyde for 1 day, embedded in paraffin to make longitudinal sections of gastric mucosa including the generative cells of IM, and cut into three serial sections for histological and immunohistochemical examinations.

Antibody

Antibody against human BGP was raised as previously reported by Ignacio et al. [24], with modification. Briefly, a synthesized 13-residue peptide (CDLQIPPPNIPRD) corresponding to cysteine coupled to the 12-carboxyterminal residues of BGP, was chosen for the immunogen, and this had no significant homology with other protein including human liver and muscle-type GP, determined using Gene Work's homol-

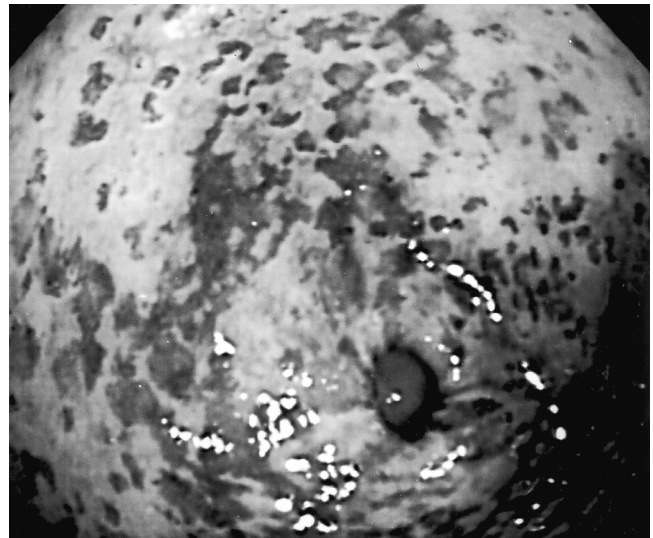


Fig. 1. Endoscopic dye staining of the stomach using methylene blue. Intestinal metaplasia was macroscopically visualized as blue islands in the gastric mucosa

ogy search. The N-terminal cysteine of the peptide was coupled with activated keyhole limpet hemocyanin (KLH; Pierce, Rockford, IL, USA). Adult rabbits were immunized with four subcutaneous injections at 1-week intervals, with 100 µg of the KLH-peptide with Freund's complete or incomplete adjuvant. One week after the last injection, the same amount of the KLH-peptide was injected subcutaneously as a booster, and 1 week later, blood samples were collected from the jugular vein. Immunoglobulin G (IgG) fractions of pooled antisera (approximately 100 ml) were precipitated by adding saturated $(\text{NH}_4)_2\text{SO}_4$ (50% saturation), dissolved in 100 ml of phosphate-buffered saline (PBS) and dialyzed extensively in the same buffer. Part of this IgG fraction (2 ml) was then applied to a column of BGP-coupled Sepharose (1 mg of the BGP peptide was coupled with 1 ml of Hi Trap NHS-activated Sepharose) (Pharmacia Biotech, Uppsala, Sweden) at a flow rate of 0.5 ml/min at room temperature. The column was washed successively with buffer A (0.5 M ethanolamine, 0.5 M NaCl, pH 8.3) and buffer B (0.1 M acetate buffer, 0.5 M NaCl, pH 4.0). Then the antibodies bound in the column were eluted with 0.1 M glycine-HCl buffer, pH 2.5, containing 1 M NaCl. Fractions containing antibodies, which were detected by measuring the absorbency at 280 nm, were collected, neutralized immediately with 1 M NaOH, and dialyzed against PBS buffer.

Immunohistochemistry

ABC Elite kits (Vector Laboratories, Burlingame, CA, USA) for rabbit IgG were used. Sections of formalin-fixed and paraffin-embedded tissue (3-µm-thick) were

deparaffinized and hydrated through xylene and graded ethanol. The sections were incubated with normal horse serum for 30min following the blocking of endogenous peroxidase activity with 0.3% H₂O₂ in methanol for 30min, incubated overnight at 4°C with optimally diluted primary antibody, and subsequently incubated with biotinylated anti-rabbit and anti-mouse antibody and avidin-biotin peroxidase complex for 30min at room temperature. They were washed in 10mM PBS (pH 7.2) between each incubation step. The sites of peroxidase binding were visualized by the diaminobenzidine method. The sections were counterstained with hematoxylin for microscopic examination. The working dilutions of the primary antibodies employed in this study were 1 µg/ml of the affinity purified anti-BGP. As a negative control, nonimmunized rabbit IgG was used instead of the primary antibody.

Statistics

The incidence of BGP-IM per patient was expressed as the percentage of BGP-IM appearance in the eight biopsy specimens, excluding the specimen without the generative cells of IM. Statistical analyses were performed using Student's *t*-test and Fisher's exact probability test.

Results

Clinicopathological features of synchronous gastric carcinoma

Table 1 shows the clinicopathological data of the patients. The patients with multiple early gastric carcinomas were significantly older than those with single early

gastric carcinoma ($P = 0.041$). However, there was no significant difference between the patients with single carcinoma and those with atrophic gastritis. The incidence of multiple gastric carcinomas had a tendency to be higher in males than that of single carcinoma or that of atrophic gastritis.

BGP expression in endoscopic biopsy specimens of gastric carcinoma and IM

The anti-BGP antibody against specific peptide of BGP demonstrated good reactivity in the conventional paraffin sections at a low antibody concentration. Furthermore, no immunohistochemical staining of BGP was observed in normal gastric mucosa, even in the proliferating zone. The reactivity of gastric carcinoma to anti-BGP antibody in endoscopic biopsy specimens is shown in Fig. 2. Strongly positive reactivity was observed in the cytoplasm of cancer cells. In 93.3% (28/30) of the multiple carcinomas and 80.0% (20/25) of the single carcinomas, the biopsy specimens showed positive staining for BGP. The percentage of immunohistochemical positivity for anti-BGP antibody in the intestinal-type carcinoma corresponded well with previous reports [18,19]. Figure 3 shows BGP-IM in a biopsy specimen. The IM glands had structural deformity to a slight degree, but no cellular atypia. The generative cell zone of IM showed positive reactivity. Strong reactivity, similar to that in the cancer cells, was observed in the cytoplasm of the generative cells of IM.

Incidence of BGP-IM in stomachs with multiple carcinoma, single carcinoma, and atrophic gastritis

As shown in Fig. 4, the distribution of the plots showing BGP-IM positivity in the stomach was extremely char-

Table 1. Clinicopathological factors in patients with multiple and single early gastric carcinoma and atrophic gastritis

Factors	Multiple; <i>n</i> = 14 (30 lesions)	Single; <i>n</i> = 25	Atrophic gastritis; <i>n</i> = 20	<i>P</i> value
Age (years)	72.5 ± 9.3*	64.0 ± 13.3*	63.0 ± 18.4	0.041*
Sex				NS
Male	10 (71.4)	15 (60.0)	10 (50.0)	
Female	4 (28.6)	10 (40.0)	10 (50.0)	
Macroscopic type				NS
Elevated	6 (20.0)	4 (16.0)	NA	
Depressed	24 (80.0)	21 (84.0)	NA	
Tumor location				NS
Middle	16 (53.3)	16 (64.0)	NA	
Lower	14 (46.7)	9 (36.0)	NA	
BGP positivity	28 (93.3)	20 (80.0)		NS

Values in parentheses are percentages

NA, Not applicable; NS, not significant; BGP, brain (fetal)-type glycogen phosphorylase

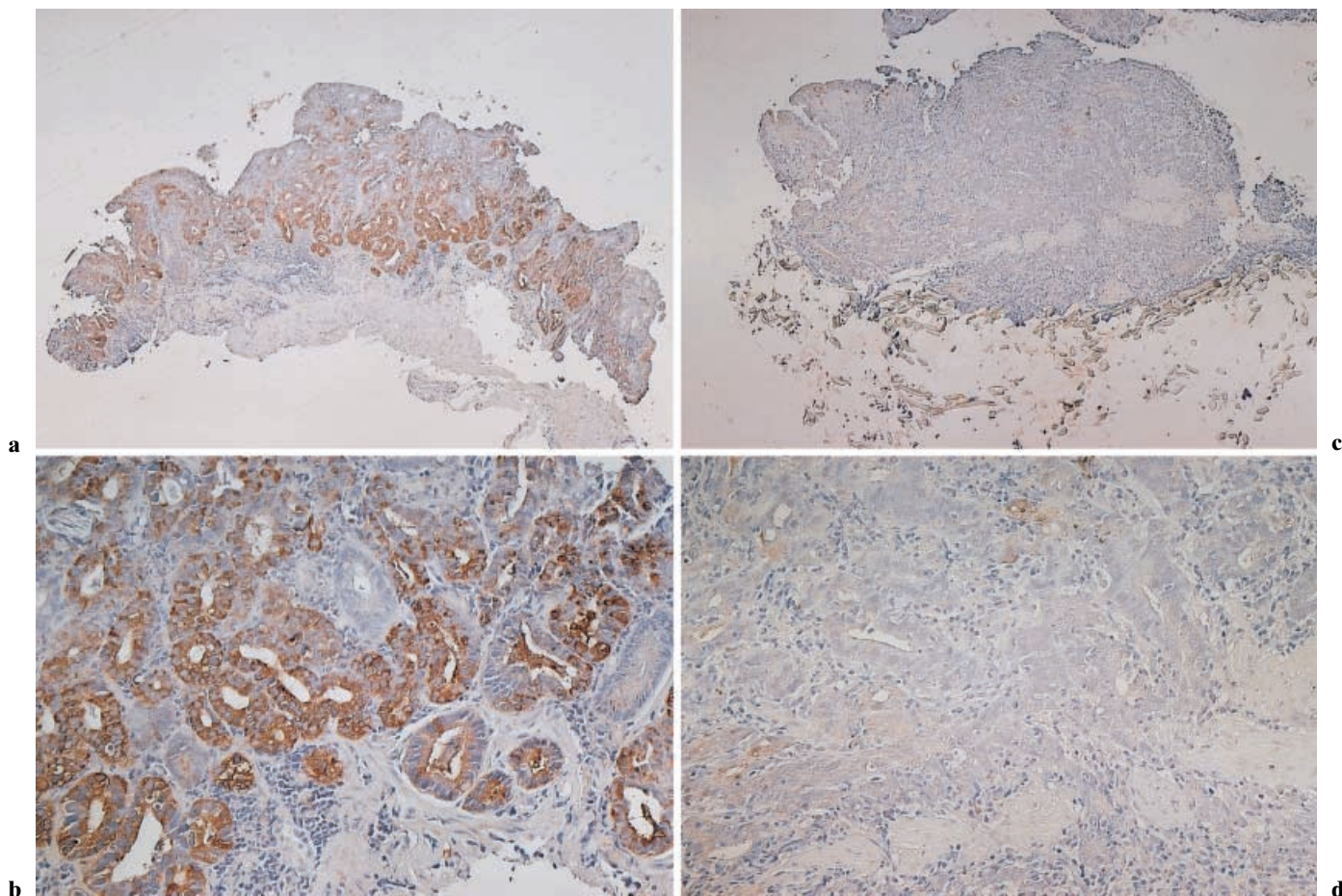


Fig. 2a–d. Immunohistochemical staining of biopsy specimens from gastric cancer foci with affinity-purified anti-brain type glycogen phosphorylase (BGP) antibody. **a** BGP-positive carcinoma and **b** high-power view; **c** BGP-negative carcinoma, and **d** high-power view. Strongly positive reactivity was observed in the cytoplasm of cancer cells, but none of the staining was detected in the nonneoplastic gastric gland (**b**). **a** $\times 60$; **b** $\times 260$; **c** $\times 60$; **d** $\times 260$

acteristic in each group. The distribution was almost symmetrical in the multiple carcinoma and the atrophic gastritis groups. Although almost all stomachs with atrophic gastritis had no BGP-IM in any biopsy specimen, all the stomachs with multiple carcinoma had BGP-IM in each of the biopsy specimens. Furthermore, all the carcinomas in the multiple carcinoma group had high percentages of BGP-IM appearance, except for two in which BGP was negative in the cancer foci. On the other hand, a bipolarized distribution of the plots was observed in the single-carcinoma group; that is, about a quarter of the group had BGP-IM at high percentages, but about half of the group did not have it at all. The incidences of BGP-IM (mean percentage \pm SD) in the stomachs with multiple carcinomas, single carcinoma, and atrophic gastritis were $83.2\% \pm 22.8\%$, $36.5\% \pm 41.3\%$, and $7.1\% \pm 18.0\%$, respectively (Fig. 4). The incidence of BGP-IM in the stomachs with multiple carcinomas was significantly higher than that in

those with a single carcinoma ($P < 0.001$) or those with atrophic gastritis ($P < 0.001$). The incidence in stomachs with a single carcinoma was significantly higher than that in those with atrophic gastritis ($P = 0.011$).

Discussion

One of the major problems with the local treatment of gastric cancer is that of the metachronous carcinomas in other parts of the stomach being different from the initial site of the carcinoma. A recent molecular biological study has suggested that high microsatellite instability in gastric tumors had a relationship with synchronous and/or metachronous gastric cancer compared with single carcinoma, whereas there was no difference in proliferative ability, carcinogenetic pathway through p53 or K-ras, and various mismatch repair genes, although the mechanism was unclear [25]. How-

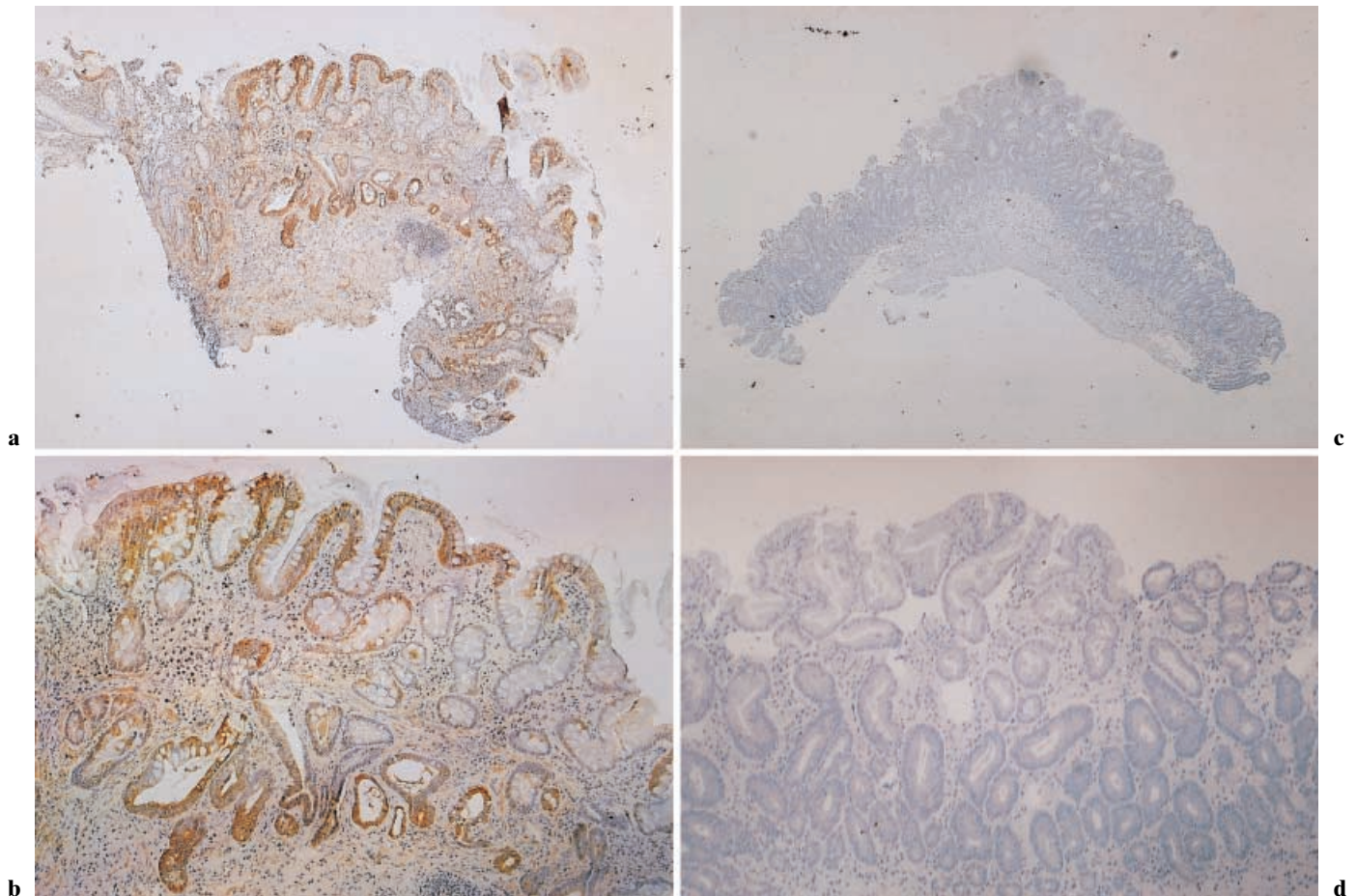


Fig. 3a–d. Immunohistochemical staining of biopsy specimens from gastric mucosa stained by methylene blue dye with affinity-purified anti-brain type glycogen phosphorylase (BGP) antibody. **a** BGP-positive intestinal metaplasia and **b** high-power view; **c** BGP-negative intestinal metaplasia and **d**

high-power view. Strongly positive reactivity was observed in the generative cells of intestinal metaplasia and its epithelium, but none of the staining was detected in the neighboring normal gastric mucosa (pyloric gland). **a** $\times 60$; **b** $\times 230$; **c** $\times 60$; **d** $\times 230$

ever, the application of molecular genetics in the screening and surveillance of patients for gastric carcinoma is still in its infancy. Arima et al. [8] reported that metachronous recurrence was found in 6 of 76 endoscopically treated patients, and it was detected significantly more frequently in patients whose synchronous multiple lesions were found during the initial treatment; they stressed the importance of the detection of gastric mucosal recurrence by frequent periodic endoscopic examinations during the follow-up period after the endoscopic treatment. Early detection of the metachronous cancer is beneficial for the subsequent treatment of the new lesion, for which minimally invasive therapy, including EMR, can be used. The necessity for frequent endoscopic follow-up, however, affects the quality of life for the patient and increases the overall medical cost. Therefore, a reliable predictive indicator of patients with a high risk of metachronous recurrence

is very important for determining the schedule of endoscopic follow-up after the initial endoscopic treatment. Because metachronous recurrence was detected significantly more frequently in patients with synchronous multiple lesions [2,3,8], a predictive indicator for metachronous recurrence would correspond with the indicator for synchronous multiple gastric carcinoma.

Wittekind et al. [14] analyzed 61 patients with synchronous gastric carcinoma from among 1664 patients, and suggested that multiple primary tumors arose from precancerous conditions leading to similar genetic alterations. It is generally accepted that IM in the stomach increases the risk of gastric cancer [26–29]. However, it has been suggested that only 0.1%–0.2% of IM is related to the carcinogenesis of intestinal-type gastric cancer worldwide [30]. Therefore, the IM significantly correlated with carcinogenesis of intestinal-type cancer should be selected for use as an appropriate marker.

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