Altitudinal Change in the Photosynthetic Capacity of Tropical Trees: A Case Study from Ecuador and a Pantropical Literature Analysis

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Abstract

In tropical mountains, trees are the dominant life form from sea level to above 4,000-m altitude under highly variable thermal conditions (range of mean annual temperatures: <8 to $>28^{\circ}$ C). How light-saturated net photosynthesis of tropical trees adapts to variation in temperature, atmospheric CO_2 concentration, and further environmental factors, that change along elevation gradients, is not precisely known. With gas exchange measurements in mature trees, we determined light-saturated net photosynthesis at ambient temperature (T) and $[CO_2]$ (A_{sat}) of 40 tree species from 21 families in tropical mountain forests at 1000-, 2000-, and 3000-m elevation in southern Ecuador. We tested the hypothesis that stand-level averages of A_{sat} and leaf dark respiration (R_{D}) per

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leaf area remain constant with elevation. Standlevel means of A_{sat} were 8.8, 11.3, and 7.2 µmol $CO_2 \text{ m}^{-2} \text{ s}^{-1}$; those of R_D 0.8, 0.6, and 0.7 µmol $CO_2 \text{ m}^{-2} \text{ s}^{-1}$ at 1000-, 2000-, and 3000-m elevation, respectively, with no significant altitudinal trend. We obtained coefficients of among-species variation in A_{sat} and R_{D} of 20–53% (n = 10-16 tree species per stand). Examining our data in the context of a pan-tropical A_{sat} data base for mature tropical trees (c. 170 species from 18 sites at variable elevation) revealed that area-based A_{sat} decreases in tropical mountains by, on average, 1.3 μ mol CO₂ m⁻² s⁻¹ per km altitude increase (or by $0.2 \ \mu\text{mol}\ \text{CO}_2\ \text{m}^{-2}\ \text{s}^{-1}$ per K temperature decrease). The Asat decrease occurred despite an increase in leaf mass per area with altitude. Local geological and soil fertility conditions and related foliar N and P concentrations considerably influenced the altitudinal A_{sat} patterns. We conclude that elevation is an important influencing factor of the photosynthetic activity of tropical trees. Lowered A_{sat} together with a reduced stand leaf area decrease canopy C gain with elevation in tropical mountains.

Key words: altitudinal gradient; foliar N; foliar P; leaf dark respiration; light-saturated net photo-synthesis; tropical lowland forests; mature trees; C source limitation; tropical montane forests.

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Author Contribution: Christoph Leuschner and Vivana Horna designed the study, Jürgen Homeier conducted the species identification and helped with the selection of species, Bärbel Wittich performed the field research with support of B. Wittich, V. Horna, and J. Homeier analyzed the data, and C. Leuschner, V. Horna, and B. Wittich wrote the article.

INTRODUCTION

With an estimated total of 37,000 woody plants (Odegaard 2000), tropical forests possess not only by far more tree species, but also exist under a broader spectrum of environmental conditions, than any other biome on earth. Moist forests once stretched from sea level to the alpine tree line at 3,500-4,800 m a.s.l., forming closed stands under a very broad range of mean annual temperatures (MAT; >28 to $<8^{\circ}$ C), rainfall totals (from c. 2,000 to > 8,000 mm y⁻¹) and soil fertility conditions (very low to high fertility; Whitmore 1998; Ghazoul and Sheil 2010). As a consequence, tropical forests exhibit large changes in structure, physiognomy, and species composition as one ascends from the lowlands to high elevation. Tropical mountain forests (TMFs) replace lowland forests at approximately 1,000-m elevation, where the climate becomes cooler and often moister, and radiation is frequently reduced due to cloudiness (Hamilton and others 1995; Bruijnzeel and others 2010). Other influential environmental factors that change with altitude are air pressure and the atmospheric concentrations of CO₂ and O₂, and UV-B radiation. In many altitudinal transects in tropical mountains, soil moisture tends to increase and the plant availability of nutrients, in particular of nitrogen and phosphorus, to decrease with altitude (Soethe and others 2008; Benner and others 2010; Bruijnzeel and others 2010; Moser and others 2008).

Tropical trees seem to respond to altitudinal gradients by modifications in plant form and function, notably adaptation in leaf morphology and physiology, tree stature, carbon allocation patterns, and productivity (for example, Cordell and others 1999; Moser and others 2007, 2008, 2011) but the within-species variation in morphological and functional traits in response to increasing elevation is not well studied. More information exists about community level changes in tree stature, leaf form and function, and forest productivity along elevation transects in tropical mountains which result from species turnover along the slope. One of the most obvious changes is the reduction in tree size (Lieberman and others 1996; Raich and others 1997; Aiba and Kitayama 1999; Pollmann and Hildebrand 2005; Shi and others 2008), which is accompanied by a reduction in above ground net primary production (NPP) from tropical lowland to upper montane forests (Hawaii: Raich and others 1997; Sabah, Malaysia: Kitayama and Aiba 2002; Puerto Rico: Weaver and Murphy 1990, Wang and others 2003; Peru: Girardin and others 2010; Ecuador: Moser and others 2011;

Leuschner and others, in press). One of the possible underlying causes is the temperature decrease, but N limitation of tree growth may also be involved in certain mountains where a decrease of foliar N concentration and an increase in leaf longevity with elevation was found (for example, Tanner and others 1998; Letts and Mulligan 2005; Moser and others 2010). Grubb and Tanner (1976) and Grubb (1977) identified smaller and thicker leaves with lower N concentrations as being characteristic for the trees at high elevations in tropical mountains.

Leaf level photosynthesis, the process that defines canopy carbon gain, has only rarely been investigated in TMFs (for example, Hikosaka and others 2002; Rada and others 2009; van de Weg and others 2012). In general, we know more about the photosynthetic activity of trees at the alpine tree line and their carbon relations than about altitudinal change in tree photosynthetic capacity in mountain forests, both in the tropical and the temperate regions (for example, Rada and others 1996; Hoch and others 2002; Körner 2003; Smith and Johnson 2009). However, examining how light-saturated net photosynthesis (Asat) and canopy C gain change along mountain slopes from lowland to upper montane elevation is crucial for a better understanding of the long-term adaptation of trees to changes in temperature, atmospheric CO₂ concentration, and other abiotic factors, and it may also help to answer the question about the causes of tropical alpine tree lines.

Altitudinal change in A_{sat} has been investigated in a few temperate and subtropical mountains (for example, Benecke and others 1981; Zhang and others 2005; Premoli and Brewer 2007; Wieser and Tausz 2007; Bresson and others 2009) showing either no change (Benecke and others 1981; Wieser and Tausz 2007; Bresson and others 2009; Wieser and others 2010), an increase (Premoli and Brewer 2007) or a decrease with increasing altitude (Slayter and Morrow 1977; Zhang and others 2005). Thus, no consistent pattern has yet been detected. Even less is known about altitudinal change in the A_{sat} of tropical trees. A notable exception is the altitudinal transect study in Metrosideros polymorpha in the tropical island forests of Hawaii between 100 and 2,500 m a.s.l. (Cordell and others 1998, 1999).

In this transect study in southern Ecuadorian TMFs, we examined patterns of altitudinal variation in light-saturated net photosynthesis of mature (or pre-mature) trees along a transect from 1,000 to 3,000 m a.s.l. covering a large number of tree species (40). We further explored the possible dependence of A_{sat} on foliar nutrient concentration and leaf

morphology along the elevation gradient. Complementary to this field study, we conducted a pantropical literature survey of published photosynthesis data from mature tropical trees at contrasting elevation (lowland to upper montane; 12 studies from 22 sites covering more than 150 tree species, excluding seedling and sapling studies). The main objectives of the study were (i) to clarify the position of tropical montane forest trees relative to tropical lowland, subtropical, and temperate trees in terms of leaf form and function and (ii) to search for a significant temperature and altitude dependence of Asat and leaf dark respiration R_D in tropical trees. We tested the hypotheses that (1) stand-level averages of mass-based A_{sat} are decreasing with elevation due to decreases in foliar N and P, whereas (2) area-based Asat remains unchanged because of the LMA increase with altitude. We also hypothesized that (3) area-based $R_{\rm D}$ does not change with elevation.

Methods

Study Sites and Selection of Trees

The measurements were conducted along a 2,000m elevation transect in TMFs on the eastern slope of the southern Ecuadorian Andes between February and May 2009. The study sites were located at about 1000, 2000, and 3000 m a.s.l. in the Podocarpus National Park and the Reserva Biológica San Francisco in the Provinces of Loja and Zamora-Chinchipe. The maximum distance between the sites was 30 km (Figure 1).

The research area has a tropical humid climate with an extremely wet season from April to July and a less humid period from September to December (Bendix and others 2006). Regularly occurring longer dry periods do not exist. Further details on the climatic conditions at the study sites are given in Table 1.

All three stands are located in protected forest sections. Natural disturbances in the past may have included landslides in the steeply sloped terrain. The three elevations along the gradient correspond to three different forest types (Homeier and others 2008): (1) at 1,000 m (4°7′S 78°58′W), in the transition zone between tropical lowland and lower montane forest, we find evergreen premontane forest whose trees attain heights of up to 40 m. Common tree families of this forest type are Fabaceae, Melastomataceae, Moraceae, Myristicaceae, Rubiaceae, and Sapotaceae. In this forest, we selected one tree individual per species, totaling to 15 species, at elevations between 950 and 1,050 m a.s.l. Ten species were identified to the species level

(see Table 3), the remaining to the genus level. (2) The evergreen lower montane forest at 2,000 m (3°58'S 79°04'W) achieves a canopy height of 18-22 m. Characteristic tree families are Euphorbiaceae, Lauraceae, Melastomataceae, and Rubiaceae. Sixteen tree species were investigated at elevations between 1,800 and 1,900 m a.s.l. Fourteen of the sixteen species could be identified to the species level. (3) The evergreen elfin-forest at 3,000 m (4°7'S 79°11'W) extends up to the tree line, and the canopy height is rarely higher than 8-10 m. Dominant tree families are Aquifoliaceae, Clusiaceae, Cunionaceae, Lauraceae, and Melastomataceae. Ten tree species were investigated at elevations between 2,850 and 3,000 m a.s.l. situated about 100-200 m below the tree line; nine were identified to the species level. Stand structural characteristics of the three stands are summarized in Table 2.

In the three stands, photosynthesis measurements were conducted on a total of 41 trees representing 40 different species (in one species, *Clethra revoluta*, we selected one individual at 2,000 and at 3,000 m). Only medium to tall trees with a minimum breast height diameter (dbh) of 10 cm were investigated (the mean dbh of the sampled trees was 16 ± 2 cm at 1,000 m, 19 ± 2 cm at 2,000 m, and 12 ± 1 cm at 3,000 m). The size of the measured trees was 10-20 m at 1.000 m. 8-15 m at 2,000 m, and 4-12 m at 3,000 m. To gain access to the sun-lit parts of the tree canopies, we selected tree individuals in the forest that grew on the steep slope beneath walking paths or below ribs on the slope, so that part of the canopy exposed to the sun could be approached from the ground.

We compiled data on the geographical distribution of the sampled families and the known altitudinal distribution of the investigated species using the online databases established by Stevens (2010) and the Missouri Botanical Garden (2010). We used APG III (2009) for family classification.

Photosynthesis Measurements

Light-saturated net photosynthesis A_{sat} was measured on about 50 days between February and May 2009 with a portable IRGA system (LI-6400, LI-COR Biosciences, Lincoln, Nebraska, USA) with a standard leaf chamber equipped with a LED red/ blue light source (type 6400-02B). All measurements were performed between 10:00 a.m. and 4:00 p.m. Three fully expanded leaves of the most distal insertion on intact twigs exposed to full sunlight were investigated per tree. Thus, every species was represented by one tree individual and three leaves. Light-saturated net photosynthesis

Elevation (m a.s.l.)	1,000	2,000	3,000
Rainfall (mm y^{-1})	c. 2,230	c. 1,950	c. 4,500
Air temperature (°C)			
Mean	19	16	9
Max	30	29	19
Min	12	8	3
Air humidity (%)			
Mean	86	91	94
Max	100	100	100
Min	16	29	29
[CO ₂] (Pa)	33	30	27
pH (H ₂ O)	4.3 ± 0.6^{a}	4.8 ± 0.5^{a}	3.7 ± 0.1^{a}
$C/N (g g^{-1})$	$19.0 \pm 2.5^{\mathrm{ab}}$	15.6 ± 0.6^{a}	$23.9\pm1.4^{\rm b}$
$P_{\rm av}$ (kg ha ⁻¹)	11.8 ± 5.3^{a}	12.5 ± 4.9^{a}	5.1 ± 0.6^{a}
Net N mineralization (kg N ha^{-1} 10 d^{-1})	4.0 ± 1.6^{a}	$1.4\pm0.7^{ m ab}$	$0.6\pm0.4^{ m b}$

Table 1. Climatic and Edaphic Characteristics of the Sampled Stand

Data from Moser and others (2007) and Wolf and others, unpublished. Mean annual air temperature and relative air humidity were measured at 1.5 m height inside the stands at 1050, 1890, and 3060 m. Rainfall data are extrapolated from measurements in a forest gap at ~1,050 m (measuring period May 2003–May 2004), and from measurements in gaps at 1,950 and 3,170 m performed by P. Emck (3-year means, unpublished). $[CO_2]$ is the CO_2 concentration of the air above the boundary layer as estimated from air pressure and by assuming a constant mixing ratio of 370 µmol CO_2 mol air⁻¹ along the slope. C/N ratio, available phosphorus (P_{av}) and net N mineralization rate (in situ buried bag method) refer to the topsoil (0–10 cm, after K. Wolf, unpublished). Available P was determined by the modified Hedley fractionation (extraction with anion exchange resins combined with NaHCO₃ percolation). For the edaphic parameters, means \pm SE of four soil profiles dug at midslope position in the stands are given.

Different small letters indicate significant differences between the three stands.

Table 2. Structural Characteristics of the Stands at 1000, 2000, and 3000 m

Elevation (m a.s.l.)	Canopy height (m)	DBH (cm) Mean ± SE	Stem density (n ha ⁻¹) Mean \pm SE	Basal area (m ² ha ⁻¹) Mean \pm SE	LAI (m ² m ⁻²) Mean \pm SE	Leaf lifespan (months) Mean \pm SE	$\begin{array}{l} AGB \\ (Mg \ ha^{-1}) \\ Mean \ \pm \ SE \end{array}$	BGB (Mg ha ⁻¹)
1,000 2,000 3,000	25–30 16–20 8–10	19 ± 1^{a} 20 ± 1^{a} 18 ± 1^{a}	822 ± 50^{a} 900 ± 62^{a} $1,061 \pm 84^{a}$	$\begin{array}{c} 29 \pm 4^a \\ 34 \pm 3^a \\ 30 \pm 3^a \end{array}$	$\begin{array}{c} 6.0 \pm 0.4^{a} \\ 5.7 \pm 0.5^{a} \\ 2.2 \pm 0.2^{b} \end{array}$	16 ± 3^{a} 24 ± 2^{b} 25 ± 2^{b}	177 ± 28^{a} 158 ± 22^{a} 89 ± 10^{b}	32.1 26.1 62.8

After Homeier and others, unpublished; LAI, leaf lifespan, BGB, and leaf biomass: Moser and others 2007, 2011. Presented are mean ± 1 SE for the three elevations (except for BGB). Means of DBH, stem density, basal area, and AGB were calculated for 9–18 permanent plots (400 m² each) covering the whole range of topographic positions at the respective elevations (trees > 10 cm DBH).

AGB aboveground biomass, BGB belowground biomass (coarse and fine roots), DBH diameter at breast height, LAI leaf area index. Different small letters indicate significant differences between the three stands.



Figure 1. Location of the study area in southern Ecuador with the three stands at 1000, 2000, and 3000 m a.s.l.

was determined at a photon flux density of 1,500 μ mol m⁻² s⁻¹ under ambient CO₂ concentration and temperature. Before starting the measurement cycle of a light response curve,

the leaves were exposed to high irradiance $(1,500 \ \mu mol \ m^{-2} \ s^{-1})$ until apparent photosynthesis was stable (CV $\leq 10\%$); this was achieved after 5–20 min. In the subsequently measured light

levels, the gas exchange system was programmed to 5 min for stable values before taking a reading. The measurements were conducted at mixing ratios of 370 ppm CO₂ at all sites and at cuvette temperatures of 22–24°C at 1,000 m, 19–21°C at 2,000 m, and 15-17°C at 3,000 m to simulate the typical local sun-canopy temperatures on a sunny day at noon (based on temperature measurements by Rollenbeck and Peters, unpublished data). The vapor pressure deficit was held constant at ambient conditions at the three sites. The CO₂ release in the dark was used as an estimate of leaf dark respiration (R_D) . Prior to R_D measurement, the intact leaves on the branches were allowed to acclimate to the dark in the chamber for 2-5 min. At the beginning of the measuring campaign, the CO_2 analyzer was calibrated against a gas standard of 400 ppm CO₂ in N₂. The IRGA channels were matched before each measurement. We did not check the respiration data for the possible occurrence of post-illumination burst effects; however, our $R_{\rm D}$ data compare well with leaf dark respiration rates reported from other neo- and paleotropical tree species (for example, Eschenbach and others 1998; Carswell and others 2000; Kenzo and others 2004; Meir and others 2007).

Morphological and Chemical Leaf Traits

All investigated leaves were harvested for analysis of foliar N and P concentrations. Total concentrations of foliar N were determined with a C/N elemental analyzer (Vario EL III, Elementar, Hanau, Germany). Total P concentrations were analyzed using an Inductively Coupled Plasma Analyzer (Optima 5300DV ICP-OES, Perkin Elmer, Waltham, Massachusetts, USA) after digesting the samples with concentrated HNO₃.

Data Analysis

The relationship between net photosynthesis rate and PPFD was described with a non-rectangular hyperbolic function; 90 % of the CO₂ assimilation rate at 1,500 µmol photons $m^{-2} s^{-1}$ was taken as A_{sat} . The initial slope of the light response curve was used to calculate the apparent quantum yield of CO₂ assimilation (α). The data were analyzed at the stand level by pooling 10–16 species of a study site. Analysis of variance and Scheffé's test were used to conduct multiple comparisons among the means of the three stands. If the data were not normally distributed according to a Shapiro–Wilk test, the Mann–Whitney two-sample test (Wilcoxon *U* test) was used instead of Scheffé's test. All calculations were conducted with SAS software (version 9.1; SAS Institute, Cary, North Carolina, USA). A significance level of 5% was used throughout the analyses.

Pan-tropical Literature Survey

A literature survey was conducted to compile a data base of A_{sat}, LMA and leaf nutrient values of trees from all over the tropics covering sites at variable altitudes from lowland to upper montane elevation (see Table 1 in Online supplementary material). This data base covers 13 studies (including the present one) with 169 more than 185 tree species in 25 stands located at elevations between 100 and 3,700 m. Only measurements referring to mature or pre-mature tree individuals of non-pioneer stands were considered. Studies referring to seedlings or saplings were excluded. All study sites are located in moist tropical forests with more than 1,500 mm rainfall y^{-1} . A_{sat} values obtained at irradiances below 1,200 μ mol photons m⁻² s⁻¹ were only included if that flux density was identified as being saturating. If information was available, only data referring to sun-lit, fully expanded leaves of the upper canopy were included. Wet season data were given preference over dry season data if both were available.

RESULTS

Among the 40 tree species from 21 families investigated, light-saturated net photosynthesis at ambient temperature and [CO₂] conditions (A_{sat}) varied in a broad range from 2.1 to 12.9 µmol CO₂ m⁻² s⁻¹ (Table 3, lowest value in *Ilex teratopis* (3,000 m), highest value in *Tibouchina ochipetala*, 1,000 m). A_{sat} varied from 3.4 to 16.0 µmol m⁻² s⁻¹ in the stand at 1,000 m (15 species), from 7.7 to 15.4 µmol m⁻² s⁻¹ at 2,000 m (16 species), and from 2.6 to 10.3 µmol m⁻² s⁻¹ in the uppermost stand at 3,000 m (10 species). We obtained coefficients of variation (standard deviation expressed as percent of mean) for the species collectives investigated of 36% (1,000 m), 20% (2,000 m), and 38% (3,000 m) for the three stands.

We found no evidence in support of the assumption that trees from families restricted to the tropics generally had higher A_{sat} values than members from families with tropical and extratropical distribution range. The 21 families investigated (each represented with 1–6 species in the study) are arranged in Figure 2 according to their mean photosynthetic capacity. Relatively, high A_{sat} values were determined in members from the Clethraceae, Rubiaceae, Siparunaceae, and

Table 3. Light-Saturated Net 1 Photosynthesis (α) , Leaf Dark Re and Leaf Conductance at Light 5	Photosynthesis of spiration (R _D), Fo Saturation (Mean	f Sun Leaves Mo bliar Nitrogen (<i>N</i> 1 and Maximum	easured at Ambi m) and Phosphor Values, g _{mean} ar	ent Temperus Concent ad g _{max}) of	rature and [(rations (<i>P</i> _m) 40 Tree Spec	2O ₂] (A _{sat}), (mass basec ies at Three	Apparent Qua 1), Leaf Mass po e Elevations	ntum Yield of rt Area (LMA),
	$A_{ m sat}$ (µmol m ⁻² s ⁻¹)	$\alpha \pmod{\mathrm{mol}^{-1}}$	$R_{\rm D}$ (µmol m ⁻² s ⁻¹)	$N_{\rm m}$ (mg g ⁻¹)	$P_{ m m}$ (mg g ⁻¹)	LMA (g m ⁻²)	$ extsf{mean} (extsf{mol} \ extsf{mol}^{-2} \ extsf{s}^{-1})$	g_{max} (mol m ⁻² s ⁻¹)
1,000 m a.s.l. $(n = 15)$								
Saurauia spec.1 (s)	11.0 ± 0.8	0.054 ± 0.001	0.47 ± 0.15	18	0.8	102 ± 5	0.16 ± 0.01	0.22
Guatteria pastazae (c)	11.3 ± 0.3	0.066 ± 0.001	0.77 ± 0.04	18	0.5	137 ± 3	0.13 ± 0.01	0.33
Hedyosmum sprucei (s)	12.0 ± 0.5	0.080 ± 0.007	1.33 ± 0.16	18	0.8	79 ± 2	0.08 ± 0.01	0.12
Inga spec. (c)	9.4 ± 2.5	0.060 ± 0.001	0.95 ± 0.12	26	0.7	119 ± 3	0.08 ± 0.02	0.20
Lozania klugii (s)	9.4 ± 0.3	0.070 ± 0.002	0.66 ± 0.05	19	0.5	86 ± 2	0.17 ± 0.01	0.19
Licaria cf terminalis (c)	6.1 ± 1.0	0.054 ± 0.002	0.46 ± 0.06	26	0.5	94 ± 1	0.08 ± 0.02	0.13
Mollia cf gracilis (c)	5.4 ± 0.4	0.054 ± 0.003	0.56 ± 0.01	21	0.6	58 ± 1	0.14 ± 0.01	0.18
Centronia laurifolia (s)	7.9 ± 0.7	0.070 ± 0.001	0.58 ± 0.06	10	0.2	138 ± 4	0.08 ± 0.01	0.11
Miconia spec. (s)	6.9 ± 0.5	0.058 ± 0.006	0.51 ± 0.15	14	0.4	166 ± 10	0.11 ± 0.01	0.14
Tibouchina ochipetala (s)	16.0 ± 1.0	0.063 ± 0.002	1.05 ± 0.05	16	0.5		0.20 ± 0.02	0.29
Ficus cervantesiana (c)	7.3 ± 0.6	0.060 ± 0.006	0.82 ± 0.08	13	0.6	155 ± 18	0.11 ± 0.02	0.15
Ficus spec. (c)	8.5 ± 0.6	0.059 ± 0.001	1.06 ± 0.13	11	0.5		0.19 ± 0.01	0.21
Neea cf divaricata (s)	6.0 ± 0.6	0.072 ± 0.003	0.67 ± 0.08	42	0.7	73 ± 1	0.06 ± 0.01	0.09
Cinchona spec. (c)	11.1 ± 0.5	0.062 ± 0.002	1.03 ± 0.07	14	0.4	120 ± 4	0.11 ± 0.02	0.21
Pouteria torta (c)	3.4 ± 1.7	0.054 ± 0.009	0.35 ± 0.03	14	0.4	103 ± 9	0.05 ± 0.02	0.07
Mean	$8.8\pm0.8^{ m a}$	0.062 ± 0.002^{a}	0.75 ± 0.07^{a}	19 ± 2^{ab}	$0.5\pm0.04^{\mathrm{a}}$	110 ± 6^{a}	0.12 ± 0.01	
2,000 m a.s.l. $(n = 16)$								
Saurauia spec.2 (s)	8.6 ± 1.3	0.051 ± 0.003	0.41 ± 0.13	20	1.2	121 ± 6	0.12 ± 0.01	0.15
Piptocoma discolor (c)	12.8 ± 1.0	0.063 ± 0.003	1.52 ± 0.47	26	1.9	78 ± 9	0.12 ± 0.00	0.17
Critoniopsis zamorensis (c)	8.4 ± 0.8	0.071 ± 0.003	0.73 ± 0.18	23	1.9	74 ± 9	0.06 ± 0.01	0.11
Clethra revoluta (c)	13.0 ± 1.6	0.052 ± 0.004	0.37 ± 0.08	13	0.5	196 ± 2	0.16 ± 0.00	0.37
Vismia cf tomentosa (c)	12.2 ± 0.4	0.059 ± 0.003	0.41 ± 0.05	15	0.9	168 ± 2	0.14 ± 0.01	0.19
Aniba spec. (c)	12.1 ± 1.3	0.059 ± 0.005	0.32 ± 0.04	17	1.2	135 ± 3	0.22 ± 0.00	0.30
Rhodostemonodaphne kunthiana (c)	11.4 ± 1.0	0.071 ± 0.002	0.28 ± 0.10	26	0.9	137 ± 4	0.13 ± 0.08	0.44
Heliocarpus americanus (c)	14.7 ± 1.0	0.064 ± 0.001	1.10 ± 0.20	33	2.5	59 ± 5	0.17 ± 0.02	0.27
Meriania hexamera (c)	8.5 ± 1.3	0.058 ± 0.008	0.32 ± 0.07	18	0.8	121 ± 6	0.14 ± 0.02	0.23
Tibouchina lepidota (s)	15.4 ± 1.2	0.054 ± 0.002	0.48 ± 0.06	17	0.6	144 ± 2	0.18 ± 0.03	0.25
Ficus pertusa (c)	11.3 ± 1.6	0.058 ± 0.001	0.44 ± 0.05	19	1.1	119 ± 6	0.11 ± 0.01	0.14
Ficus citrifolia (c)	10.1 ± 1.8	0.064 ± 0.007	0.78 ± 0.15	21	0.9	146 ± 2	0.05 ± 0.01	0.12
Isertia laevis (c)	13.4 ± 2.1	0.048 ± 0.004	0.62 ± 0.22	16	0.8	215 ± 5	0.13 ± 0.02	0.20
Ladenbergia acutifolia (c)	9.8 ± 2.0	0.071 ± 0.007	0.68 ± 0.33	18	0.8	98 ± 9	0.18 ± 0.07	0.45
Siparuna aspera (s)	11.4 ± 1.7	0.054 ± 0.002	0.60 ± 0.05	29	1.5	96 ± 7	0.17 ± 0.04	0.51
Cecropia andina (c)	7.7 ± 1.6	0.057 ± 0.002	0.57 ± 0.02	18	0.7	126 ± 1	0.08 ± 0.01	0.10
Mean	11.3 ± 0.6^{b}	0.060 ± 0.002^{a}	0.60 ± 0.08^{a}	21 ± 2^a	1.1 ± 0.15^{b}	127 ± 6^{a}	0.13 ± 0.01	

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	$A_{ m sat}$ ($\mu m mol m^{-2}~ m s^{-1}$)	$\alpha \pmod{\mathrm{mol}^{-1}}$	$R_{\rm D}$ (µmol m ⁻² s ⁻¹)	$N_{ m m}$ (mg g ⁻¹)	$P_{ m m}$ (mg g ⁻¹)	LMA (g m ⁻²)	g_{mean} (mol m ⁻² s ⁻¹)	g_{max} (mol m ⁻² s ⁻¹)
3,000 m a.s.l. $(n = 10)$,				
llex teratopis (c)	2.6 ± 0.2	0.054 ± 0.008	0.25 ± 0.15	6	0.2	326 ± 1	0.03 ± 0.00	0.04
Hedyosmum purpurascens (c)	7.5 ± 1.0	0.064 ± 0.005	0.45 ± 0.08	16	0.5	140 ± 6	0.07 ± 0.01	0.11
Clethra revoluta (c)	10.0 ± 0.4	0.050 ± 0.002	0.54 ± 0.04	16	0.7	171 ± 6	0.41 ± 0.09	1.11
Clusia alata (c)	4.4 ± 1.2	0.059 ± 0.020	1.09 ± 0.06	10	0.5	286 ± 5	0.03 ± 0.02	0.06
Clusia elliptica (c)	5.8 ± 1.5	0.068 ± 0.002	0.77 ± 0.19	13	0.6	164 ± 7	0.05 ± 0.01	0.08
Weinmannia pubescens (c)	8.8 ± 1.1	0.058 ± 0.004	0.76 ± 0.11	13	0.5	113 ± 4	0.10 ± 0.01	0.17
Persea ferruginea (c)	8.0 ± 1.2	0.038 ± 0.005	0.67 ± 0.17	10	0.4	487 ± 9	0.08 ± 0.00	0.09
Miconia spec. (c)	4.6 ± 1.3	0.071 ± 0.001	0.56 ± 0.22	20	0.7	98 ± 2	0.06 ± 0.01	0.09
Myrica pubescens (c)	10.3 ± 0.6	0.064 ± 0.005	0.74 ± 0.29	22	0.5	156 ± 5	0.11 ± 0.01	0.33
Styrax foveolaria (s)	10.2 ± 0.9	0.044 ± 0.002	0.78 ± 0.04	10	0.4	304 ± 2	0.11 ± 0.03	0.27
Mean	7.2 ± 0.9^{a}	0.057 ± 0.003^{a}	0.66 ± 0.07^{a}	$14 \pm 1^{\rm b}$	0.5 ± 0.05^{a}	216 ± 19^{b}	0.11 ± 0.04	
c canopy species, s subcanopy species. Presented are mean + SF for three leaves	each ner snecies (snecies leur	el) and mean + SF over al	ll snecies of a stand Sianifi	cantly different st	orthis mean level-but	elevations) are ind	icated by different letters	(n < 0.05)

Table 3. continued

Anonaceae; low rates, in trees from the Clusiaceae, Nyctaginaceae, Sapotaceae, and Aquifoliceae.

In our species sample from southern Ecuador, we found no clear elevational trend in A_{sat} with stand means of 8.8 (1,000 m), 11.3 (2,000 m), and 7.2 µmol CO₂ m⁻² s⁻¹ (3,000 m). However, the 16 species of the montane stand at 2,000 m had on average significantly higher photosynthetic capacities than the trees at 1,000 or 3,000 m (Table 3). This is also visible in Figure 2 when comparing the species of a family at 2,000 m (triangles) with the members from 1,000 or 3,000 m (squares and circles).

We found apparent quantum yields (α , the initial slope of the light response curve under ambient T and [CO₂]) in the range of 0.037–0.080 mol - CO₂ mol quanta⁻¹ for the 40 species (Table 3). The tree samples from the stands at 1000, 2000, and 3000 m were not significantly different with respect to their α means.

Leaf dark respiration (R_D) at ambient temperature varied between 0.25 and 1.52 µmol CO₂ m⁻² s⁻¹ with most species means ranging between 0.4 and 0.9 µmol m⁻² s⁻¹ (Table 3). The stand means of R_D did not differ significantly at 1000, 2000, and 3000 m a.s.l.

DISCUSSION

Among-Species Variation in A_{sat} in Species-Rich Tropical Forests

This study focuses on the photosynthetic capacity of mature trees with a size of at least 4 m



Figure 2. Net photosynthesis rate of sun leaves at ambient temperature and $[CO_2]$ (A_{sat}) averaged over all species (morphotypes) of a family at a given elevation. The 21 families are arranged from left to right according to their mean A_{sat} rate. *T* indicate families with restriction to the tropics.

Family	Geographical	1,000-m stand		2,000-m stand		3,000-m stand	
	distribution	Sampled species	Altitudinal distribution (m a.s.l.)	Sampled species	Altitudinal distribution (m a.s.l.)	Sampled species	Altitudinal distribution (m a.s.l.)
Actinidiaceae	Largely tropical, not Africa	Saurauia spec.1		Saurauia spec.2			
Annonaceae	Largely tropical, pan- tropical	Guatteria pastazae	390–2,120				
Aquifoliaceae Asteraceae	Cosmopolitan Cosmopolitan			Critoniopsis zamorensis Dinnoona discolor	100_3 890	Ilex teratopis	1,350–3,316
Chloranthaceae	Tropical and subtropi-	Hedyosmum sprucei	40-3,250	t throatin miscorol		Hedyosmum purpurascens	2,400–3,300
Clethraceae	Largely tropical mon-			Clethra revoluta	456-3,450	Clethra revoluta	456-3,450
Clusiaceae	tane or subtropical Pantropical			Vismia cf tomentosa	100-2,615	Clusia alata Clusia allintica	100-3,200
Cunoniaceae	Largely temperate and tropical, S. hemi- sphere, few African					verimannia pubescens	1,350–2,800
Fabaceae Lacistemataceae	species Cosmopolitan Neotropical	Inga spec. Lozania klugii	100–2,140				
Lauraceae	Pantropical (temperate)	Licaria terminalis	1,050–1,440	Aniba spec. Rhodostemonodaphne kunthiana	1–2,150	Persea ferruginea	1,400–3,950
Malvaceae	Pantropical, also tem-	Mollia gracilis	100-910	Heliocarpus americanus	0-2,615		
Melastomataceae	perate Largely tropical, also subtropical	<i>Centronia laurifolia</i> <i>Miconia</i> spec.	600–2,050	Meriania hexamera Tibouchina lepidota	1,100–2,900 400–3,230	<i>Miconia</i> spec.	
Moraceae	Mostly tropical to warm temperate	Tibouchina ochypetala Ficus cervantesiana Ficus spec.	140–,2900 100–2,400	Ficus citrifolia Ficus pertusa	0-3,000 0-2,100		
Myricaceae Nyctaginaceae	Cosmopolitan Pantropical to warm	Neea divaricata	45-2,500			Myrica pubescens	1,500–3,605
Rubiaceae	temperate Cosmopolitan, but especially tropical	<i>Cinchona</i> spec.		Isertia laevis Ladenbergia acutifolia	0-2,100 220-1,630		

Family	Geographical	1,000-m stand		2,000-m stand		3,000-m stand	
	distribution	Sampled species	Altitudinal distribution (m a.s.l.)	Sampled species	Altitudinal distribution (m a.s.l.)	Sampled species	Altitudinal distribution (m a.s.l.)
Sapotaceae Sinarinaceae	Pantropical Neotronical and W	Pouteria torta	0-2,440	Sinaruna asnera	30-3 300		
	African			n inden min indio			
Styracaceae	Warm N. temperate to tropical					Styrax foveolaria	2,400–3,430
Urticaceae	Cosmopolitan, but mainly tropical			Cecropia andina	1,540–2,800		
Geographical distributic	on of the families according to Stevens (201	0).					

Table 4. continued

(uppermost stand), typically 8–20 m, to examine the maximum assimilation rate of sun-canopy leaves in their natural position in the canopy. We excluded seedling and sapling studies because leaf physiology may be subject to a considerable ontogenetic shift with tree aging (for example, Thomas and Winner 2002; Mediavilla and Escudero 2003). Measurements conducted in more than 155 species in about 18 stands in the Americas, South-east Asia, and Africa met the criteria defined in this study (see Table 1 in Online supplementary material).

In terms of species numbers, our study is the most comprehensive investigation conducted so far on the leaf-level photosynthesis of tropical trees; nevertheless, 155 species examined from a pool of approximately 37,000 taxa is still a tiny number. Similarly, our in-depth study on the 40 species of the Ecuadorian TMFs represents only about 5% of the 800 or so tree species in this study area (J. Homeier, unpublished data). Thus, the two species samples investigated may well not be representative of the local or the pantropical tree flora. The among-species variation in A_{sat} in the three Ecuadorian stands was indeed considerable (coefficients of variation of 20-38%). Further bias may have been added to our data sets by the fact that we were not able to conduct a random selection of species or to choose them according to dominance in the three stands. Rather, species selection in Ecuador was confined to tree individuals growing on steep slopes because this allowed us to access part of their sun canopies with gas exchange equipment. This procedure guaranteed that the measured leaves were exposed to full sunlight at least part of the day and thus must have possessed partial or full sun leaf adaptation. The selection of tree species was based on extensive knowledge of the ecology and distribution of the taxa in the Loja region of South Ecuador (Homeier and others 2008); we chose only typical members of the closed natural forest and excluded shade-intolerant pioneer tree species. Furthermore, the phylogenetic diversity covered by our species sample was relatively large, including taxa from 21 families in the whole study, or 10–16 species from 9 to 12 families in each of the three stands (Table 4). However, we cannot rule out the possibility that a larger survey covering additional species and families would lead to different A_{sat} stand averages than the figures reported here for the Ecuadorian and pantropical analysis.

Constant or Decreasing Photosynthetic Capacity with Elevation?

From our 40-species sample in Ecuador, a significant altitudinal trend in photosynthetic capacity at

ambient [CO₂] and temperature did not manifest. However, we found a significantly higher A_{sat} stand mean for the mid-elevation stand at 2,000 m than at lower and higher elevation, corresponding to higher foliar P and N concentrations at this elevation. Correspondingly, species with relatively high A_{sat} values (>10 µmol CO₂ m⁻² s⁻¹) occurred mostly in this mid-elevation stand. To the best of our knowledge, our investigation is the first that examined altitudinal changes in A_{sat} in species-rich tropical moist forests by measuring a large number of species in the adult stage and thus accounting for the characteristic species turnover along the slope. A few studies on altitudinal change in photosynthetic performance measured daily means of actual photosynthesis but not A_{sat} (for example, Cavieres and others 2000) or derived estimates of photosynthetic activity from productivity data (for example, Kitayama and Aiba 2002). The only other transect study on altitudinal patterns in A_{sat} in mature tropical trees focused on a single, polymorphic tree species (Metrosideros polymorpha, Myrtaceae) in the species-poor Hawaiian rainforests (Cordell and others 1999). Similar to our study, these authors found no change in Asat under ambient temperature and [CO2] conditions between 100 and 2,500 m a.s.l. However, Asat varied at unusually low levels (2.3–3.8 μ mol CO₂ m⁻² s⁻ ¹) across the Hawaiian transect. It may well be that this rather unproductive species reflects the specific conditions of isolated island forests, and that the results cannot simply be extrapolated to tropical forests on the continents. Moreover, altitudinal patterns in A_{sat} derived from a single species with wide altitudinal distribution should differ from trends obtained from transect studies that cover a large number of species of different phylogenies, each adapted to the specific site conditions at the respective elevation level.

In the absence of other transect studies on gas exchange in tropical mountains, additional information on altitudinal change in A_{sat} is provided by our pan-tropical literature survey (Table 1 in Online supplementary material). The data incorporated in this analysis were selected according to strict criteria with respect to tree size (mature or pre-mature trees only), successional status (no pioneer species), canopy position (sun-exposed branches only), measuring conditions (saturating light, typically >1,200 μ mol m⁻² s⁻¹ PPFD; ambient temperature and [CO₂]), and climate (tropical moist); this protocol excluded a number of studies on maximum photosynthesis of tropical trees that either worked with saplings, understorey or pioneer trees or apparently used non-saturating irradiances. According to this data compilation, sun leaf-A_{sat} significantly decreases from tropical lowland to upper montane elevation which contradicts our first hypothesis. The linear regression equation indicates a decrease in A_{sat} by about 1.3 µmol $CO_2 \text{ m}^{-2} \text{ s}^{-1}$ per km altitude (Figure 3: left column) or by about 0.20 μ mol CO₂ m⁻² s⁻¹ per K air temperature reduction (Figure 3: center column). The trees from lowland forests reached average photosynthetic capacities of about 10-11 µmol $CO_2 \text{ m}^{-2} \text{ s}^{-1}$, those of the upper montane forests of about 7 μ mol CO₂ m⁻² s⁻¹. The decrease in areabased A_{sat} coincides not only with the temperature decrease but also with a significant decrease in foliar P whereas no significant $N_{\rm m}$ decrease with elevation was detected in the pantropical survey. Not only area-based, but mass-based Asat decreased also from lowland to upper montane elevation (P = 0.004; data of Table 1 in Online supplementary material); this indicates that the altitudinal increase in LMA, which occurred between lowland and montane elevation, was on the average not large enough to compensate the mass-based A_{sat} reduction and thus to prevent a decrease in areabased A_{sat} .

Local edaphic and climatic conditions seem to modify this more general picture of altitudinal change in area- and mass-based A_{sat} considerably. Although the Ecuadorian data generally fit quite well into the patterns extracted from the pantropical analysis, A_{sat} deviates with relatively high rates at mid-slope position (2,000 m) from the overall trend which is explained by the relatively high soil P availability in this stand. With a mean foliar P concentration of 1.1 mg g^{-1} , the 2,000-m stand fits well into the "moderately fertile" category of Vitousek and Sanford (1986) for lowland forests, whereas the 1,000- and 3,000-m stands with 0.5 mg P g^{-1} must be classified as "infertile." The very high annual rainfall amount in the 3,000-m stand, in combination with high cloudiness, poor soil aeration, and low decomposition rates, is another site-specific factor in the Ecuador transect that may have influenced A_{sat}. Upper montane forests studied in Peru at a similar elevation received only half the rainfall amount (van de Weg and others 2009) and thus grew under less extreme conditions.

The lowered A_{sat} of TMF trees was evident from our analysis, even though the among-species variation was large and local edaphic and climatic factors seem to modify the effect of elevation and temperature considerably. The relatively low A_{sat} differentiates TMF trees functionally from tropical lowland trees. This difference matches with the



Figure 3. Dependence of light-saturated net photosynthesis at ambient temperature and $[CO_2]$ (A_{sat}), foliar nitrogen (N_m), and phosphorus (P_m) concentrations (mass based), and leaf mass per area (LMA) on elevation, temperature, and precipitation in a pan-tropical data set of c. 170 tree species and 18 forest stands. Only data referring to mature or premature tree individuals (no seedlings or saplings) of non-pioneer stands were included. For further explanations and references see Table 1 in online supplementary material. *Filled gray symbols* data from lowland forests; *black filling* data from this study in southern Ecuador (*squares* 1,000-m stand, *triangles* 2,000-m stand, and *circles* 3,000-m stand).

MAT and MAP characteristics of the investigated TMF stands which place this tree group closer to the trees of temperate evergreen rainforests or temperate deciduous forests than to tropical lowland forests (Figure 4); certain TMF stands with extremely high precipitation are even falling outside any biome range as defined by Wright and others (2004). A more detailed analysis of leaf trait



Figure 4. Grouping of the tropical montane forests considered in this study in the major biomes of the world (after Wright and others 2004) using annual precipitation and mean annual temperature as criteria. Accordingly, these tropical montane forests have affiliations to temperate rain forests (*A*), tropical rainforests (*B*), temperate forests (*C*), or tropical seasonal forests (*D*). Only sites located between 1,000 and 3,600 m a.s.l. are included. Locations: Ecuador (EC1000, EC2000, and EC3000, this study), Peru (PE1000, PE1500, PE1855, and PE2990, van de Weg and others 2012), Colombia (CO1445, CO2145, Letts and Mulligan, 2005), Venezuela (VE2400, Rada and others), and Indonesia (ID1100, Hölscher and others 2006).

relationships in the TMF tree sample shows that this group is partly deviating from the pattern of A_{sat} -foliar N-LMA inter-relationships as they appear from a global perspective in the leaf economics spectrum of Wright and others (2004) (Figure 5). One striking difference is the large scatter of the data in the plot of mass-based A_{sat} against SLA and N_{m} in TMF trees. The surprisingly low mass-based

A_{sat} rates in several leaves of the TMF sample with high SLA might point at plant-internal N allocation patterns that are directed to an optimization of light capture at the expense of maximum carboxylation rate (Evans 1989), in a similar manner as has been found in tropical understorey plants (Santiago and Wright 2007). In contrast to tropical lowland trees, many montane forest trees are exposed to continuously high cloudiness and low-light conditions which could explain this apparent functional difference from lowland trees. High variation in SLA at a relatively narrow range of mass-based Asat rates may offer a greater potential for optimizing CO₂ assimilation in this cool, often light-limited environment (Givnish 1988; Sims and others 1994; Santiago and Wright 2007).

Our literature survey further suggests that the range of A_{sat} species means occurring in lowland forests (3.7–20.3 µmol CO₂ m⁻² s⁻¹) is larger than that in montane and upper montane stands (2.1–13.2 µmol m⁻² s⁻¹). The apparent decrease in A_{sat} variation with increasing elevation is partly caused by the uneven distribution of data over the elevation range with fewer trees being studied at altitudes above 1,500 m than at lower elevation. However, the upslope decrease in the number of investigated species mirrors the general decrease in tree species richness from tropical lowland to montane elevation (Gentry 1988, 1995; Aiba and Kitayama 1999; Slik and others 2009) and thus seems to be justified.

A functional explanation of the upslope reduction in A_{sat} variation is provided by stand differences in canopy structure. We expect that taller multilayered lowland forests should generate a spatially more heterogeneous light climate than the structurally simpler high-elevation forests. This



Figure 5. Relationships between photosynthetic rate per mass (A_{sat}) and (*left*) specific leaf area (SLA) or (*right*) leaf nitrogen per mass (N_{m}) for the tree species in the survey. *Filled symbols* are for species from montane locations with *black filling* indicating the Ecuadorian species of this study (*squares* 1,000-m stand, *triangles* 2,000-m stand, *circles* 3,000-m stand) and *gray filling* species from other montane sites, *open symbols* stand for all other species in the survey. *Dashed lines* represent model II regressions with 95% confidence intervals for the global data set of Santiago and Wright (2007) (GLOPNET).

would allow at lower altitude the coexistence of more tree species with different strategies of light use and carbon assimilation. We assume that the large A_{sat} variation in the lowland data of our literature survey (maxima up to 20 μ mol CO₂ m⁻² s^{-1}) is mirroring the co-existence of species from the full spectrum of tree functional types that are present in old-growth lowland forests, notably shade-tolerant late-successional and lightdemanding gap species (Turner 2001). High-elevation forests of low stature, in contrast, are typically dense with only a single main canopy layer in which the among-species variation in photosynthetic capacity (A_{sat} typically <8 µmol CO₂ m⁻² s⁻¹) is indeed reduced. In the neotropical high-elevation tree genus Polylepis, which forms the highest forests of the world, the largest observed A_{sat} values do not exceed 9 μ mol CO₂ m⁻² s⁻¹ in Argentina (2,100 m), 7 μ mol CO₂ m⁻² s⁻¹ in Venezuela (4,200 m), and 3 μ mol CO₂ m⁻² s⁻¹ in Bolivia (4,300 m, Rada and others 1996; Azocar and others 2007). Accordingly, it is not very likely that a future more extensive set of photosynthesis data from tropical high-elevation tree species would result in higher mean A_{sat} values than those reported in this study.

What are the underlying causes of the apparent A_{sat} decrease with elevation? The direct or indirect air pressure effects on A_{sat} are probably small, in particular at low temperatures, as indicated by modeling studies of Terashima and others (1995) and Smith and Hughes (2009). A more likely explanation would be the effect of lowered temperature on the rate of RuBP carboxylation, that is, reduced light-saturated photosynthesis rates of montane and upper montane tree species under the lowered ambient temperatures of their growing sites. Low temperatures can reduce enzyme activity in the photosynthetic apparatus or may cause feedback inhibition of photosynthesis because of impaired sucrose metabolism and phloem loading. However, alpine and arctic plants have been found to compensate effectively for low temperatures by increasing Rubisco concentration and carboxylation capacity which is reflected by the generally relatively high foliar N concentrations (Körner and Larcher 1988; Körner 1989). A prerequisite of such a compensatory response is a sufficiently high N and P supply which existed in the Ecuador TMF at 2,000 m, but N and P were in short supply at 3,000 m (Moser and others 2011). Suppression of photosynthetic capacity by shortage of N or P (or other elements) in montane and upper montane forests is a possible scenario because the rate of nutrient cycling generally decreases at higher

elevations with a reduction in temperature (Benner and others 2010). We found markedly smaller average foliar N concentrations in the 3000-m stand than in the other two forests (difference significant to the 2000-m stand). Growing deterioration of N and/or P supply with increasing elevation could limit the photosynthetic capacity of high-altitude trees by two mechanisms, by restricting the amount of N and P available for allocation to the photosynthetic machinery or by increasing mean leaf longevity and thus mean leaf age; longer living leaves are typically more sclerophyllous and physiologically less active (Reich and others 1991; Wright and others 2004). In fact, Moser and others (2007) found tree leaf longevity to increase from 16 to 25 months on average between 1,000- and 3,000-m elevation. Further studies are needed to identify both the biotic and the abiotic factors causing A_{sat} to decrease with elevation in tropical mountains.

As predicted, the stand means of leaf dark respiration measured at ambient temperature remained constant along the elevation transect indicating effective adaptation of mitochondrial respiration of the montane and upper montane trees to the 10-K temperature gradient in Ecuador. Similar homeostatic responses of leaf dark respiration were reported from gradient studies with *Eucalyptus pauciflora* and herbaceous plants growing at different elevations (Larigauderie and Körner 1995; Atkin and others 2000a, b, 2006).

CONCLUSIONS

Altitudinal gradient studies may provide valuable insights into the temperature dependence of tree growth and the forest carbon balance. Canopy C gain is a key element of the C balance, but how this flux is altered with altitude in response to decreasing temperature, atmospheric [CO₂], and other factors, that change with elevation, is not well understood. The detection of more general patterns of altitudinal change in Asat in tropical forests has long been hindered by the often high species diversity and difficult canopy access. Our combined approach of gas exchange screening in a large sample of pre-montane to upper montane neotropical tree species and a pan-tropical literature survey of carefully selected A_{sat} data from low to high elevation revealed (1) a linear decrease from sea level to the alpine tree in the stand-level average of (mass- and area-related) A_{sat} measured at ambient conditions, and (2) a decreasing range of among-species A_{sat} variation with altitude. However, the data also suggest that (3) mountain transects with different geology and thus soil N and P availabilities might differ in their altitudinal A_{sat} patterns. Our findings have implications for the search for the causes of alpine tree lines in tropical mountains. It appears that stands close to the tropical alpine tree line are built by tree species with reduced A_{sat} rates, but, in addition, a reduced stand leaf area also contributes to a decrease in canopy C gain with increasing altitude in tropical mountains (Moser and others 2007). We interpret this result as a strong hint to carbon source limitation in tropical high-elevation forests, not supporting the hypothesis of C sink limitation as the principal cause of alpine tree lines (Körner 2003). Our results are in accordance with several recent studies that questioned the idea of carbon saturation in trees at high elevations (for example, Millard and others 2007; Susiluoto and others 2007; Li and others 2008).

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